Integrated Approaches Revealed the Therapeutic Mechanisms of Zuojin Pill Against Gastric Mucosa Injury in a Rat Model with Chronic Atrophic Gastritis

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Background: The Zuojin Pill (ZJP) is widely used for treating chronic atrophic gastritis (CAG) in clinical practice, effectively ameliorating symptoms such as vomiting, pain, and abdominal distension in patients. However, the underlying mechanisms of ZJP in treating CAG has not been fully elucidated.

Purpose: This study aimed to clarify the characteristic function of ZJP in the treatment of CAG and its potential mechanism.

Methods: The CAG model was established by alternant administrations of ammonia solution and sodium deoxycholate, as well as an irregular diet. Therapeutic effects of ZJP on body weight, serum biochemical indexes and general condition were analyzed. HE staining and AB-PAS staining were analyzed to characterize the mucosal injury and the thickness of gastric mucosa. Furthermore, network pharmacology and molecular docking were used to predict the regulatory mechanism and main active components of ZJP in CAG treatment. RT-PCR, immunohistochemistry, immunofluorescence and Western blotting were used to measure the expression levels of apoptosis-related proteins, gastric mucosal barrier-associated proteins and PI3K/Akt signaling pathway proteins.

Results: The results demonstrated that ZJP significantly improved the general state of CAG rats, alleviated weight loss and gastric histological damage and reduced the serum biochemical indicators. Network pharmacology and molecular docking found that ZJP in treating CAG by inhibiting inflammation, suppressing apoptosis, and protecting the gastric mucosal barrier via the PI3K/Akt signaling pathway. Further experiments confirmed that ZJP obviously modulated the expression of key proteins involved in gastric mucosal cell apoptosis, such as Bax, Bad, Apaf-1, cleaved-caspase-3, cleaved-caspase-9, Cytochrome C, Bcl-2, and Bcl-xl. Moreover, ZJP significantly reversed the protein expression of Occludin, ZO-1, Claudin-4 and E-cadherin.

Conclusion: Our study revealed that ZJP treats CAG by inhibiting the PI3K/Akt signaling pathway. This research provided a scientific basis for the rational use of ZJP in clinical practice.

Keywords: Zuojin Pill, chronic atrophic gastritis, network pharmacology, molecular docking, PI3K/Akt signaling pathway

Introduction

Chronic atrophic gastritis (CAG) is a gastric disease with pathological characteristics of reduced gastric acid secretion, atrophic gastric mucosa and metaplasia of intestine epithelium.1,2 CAG is considered a key precancerous lesion in gastric cancer, which is the third leading cause of cancer death reported in the global cancer statistics for 2018.3 Previous studies have shown that CAG is caused by a combination of factors, including helicobacter pylori infection, inadequate immune
regulation and impaired energy metabolism, and that its pathogenesis is related to oxidative stress, inflammation, endothelial and mucosal dysfunction. The current clinical treatment of CAG relies mainly on the agents used for gastric mucosal protection and symptom relief, vitamin C supplementation and proton pump inhibitors, which limit their clinical application due to their long course, aggressiveness and side effects. Therefore, there is an urgent need for complementary and alternative treatments for CAG.

Traditional Chinese medicine (TCM) has unique advantages in the treatment of CAG due to its dual therapeutic and coordinating functions, wide range of applications and few side effects. The classic Chinese prescription Zuojin Pill (ZJP) is recorded in “Danxi prescription therapy” in the Yuan Dynasty. It is a famous TCM formula in clinic used for the treatment of CAG, which consists of two herbs including Coptidis Rhizoma and Euodiae Fructus. Modern pharmacological researches have shown that ZJP has various effects such as anti-inflammatory, antibacterial, anti-apoptosis and mucosal protection, which exhibits remarkable therapeutic effects on digestive disorders. A recent study found that ZJP acts as an inflammatory inhibitor to regulate comprehensive metabolic disorders in clinic, which is an important mechanism of ZJP in the treatment of chronic gastritis. However, the mechanism of ZJP for the treatment of chronic atrophic gastritis is not fully understood. Therefore, an exploration into the underlying mechanism of ZJP in the treatment of CAG is necessary.

With the rapid development of bioinformatics, an emerging network pharmacology has been proposed which uses mathematical and computable representations to represent the various links between herbal formulations and diseases. Network pharmacology provides a new idea for drug research, especially for TCM research based on complex system. Accumulating evidence shows that network pharmacology can elucidate the potential mechanisms of action of multi-component, multi-target drugs through the analysis of various complex, multi-level interaction networks. Molecular docking technology is a common tool used to explore the interaction sites between small molecules and macromolecules. The study aims to elucidate the characteristic effects of ZJP on the histopathological improvement of gastric tissue in CAG by integrating experimental methods, providing experimental evidence for the clinical rational application of ZJP in treating CAG; Through high-throughput analysis methods such as network pharmacology and molecular docking, it aims to reveal the molecular biological mechanisms of ZJP in treating CAG and potential active substances, providing data support for future development of new drugs based on ZJP for treating CAG (Figure 1).

Materials and Methods

Reagents
Coptidis Rhizoma (Lot: 21030801) and Euodiae Fructus (Lot: 21101401) were purchased from Beijing LVYE Pharmaceutical Co., Ltd. (Beijing, China). Sodium deoxycholate (Lot: CD33141310) and ammonia (Lot: 2023AS0328) were provided by Beijing Zhongke Ruijin Technology Co., Ltd. and Biotechnology Co., Ltd., respectively. Vitacoenzyme tablets (Lot No. 221002) were obtained from Guangxi Dahai Sunshine Pharmaceutical Co., Ltd. All antibodies and the related reagents were obtained from commercial sources.

Preparation of ZJP
Coptidis Rhizoma and Euodiae Fructus (6:1, w/w) were soaked in pure water for 30 min. Then, it was extracted twice by heating (1h at a time). After that, all filtrates were rotary evaporated, concentrated, dried into dry powder and stored at 4 °C. The weight ratio of ZJP was 18.75%.

Animals and Experimental Design
Specific pathogen free (SPF) male Sprague-Dawley rats (180–200g) were obtained from Sibeifu Biotechnology Co., Ltd. (Beijing, China, license No.: SCXK (Beijing) 2019–0010). All the rats were kept in ventilated polypropylene cages at room temperature (temperature: 25 ± 0.5 °C, humidity: 55 ± 5%, 12 h: 12 h light dark cycle), and access to water and standard laboratory diet ad libitum. All animal experiments were conducted in accordance with the Laboratory Care and Use Guidelines. The research was approved by Ethics Committee of the Chinese PLA General Hospital (Approval ID: IACUC-2021-0022).
After one week of quarantine and adaptation, 34 SD rats have randomly divided into 2 groups: 8 in control group and 26 in model group. CAG was established by a method as previously described. Briefly, the rats in model group alternatively received 0.1% ammonia and 20 mmol/L sodium deoxycholate solution, accompanied with irregular fasting cycle. Ten weeks later, 2 rats in the control group and model group were randomly selected to detect the success of CAG modeling. All the rats with CAG were divided into four groups: model group, positive drug group (Vitacoenzyme, VIT, Figure 1 Flow chart of this study. ZJPL: Zuojin Pill low dose (1.26g/kg); ZJPH: Zuojin Pill high dose (2.52g/kg); VIT: Vitacoenzyme (200 mg/kg). *P < 0.05 and **P < 0.01 vs control group. *P < 0.05 and **P < 0.01 vs model group.

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The corresponding drugs were given by intragastric administration for 4 weeks (Figure 2).

Sample Collection
At the end of the experiment, all rats were fasted for 24 h with access to water. Firstly, rats were sacrificed under 20% ethyl carbamate solution. Secondly, blood samples were collected from the abdominal aorta and gastric tissues were removed. After centrifugation, the blood serum was stored at −80 °C until analysis. The gastric tissues were divided into two parts. One part was fixed in 10% paraformaldehyde for subsequent pathological examination and the other part was stored at −80 °C for subsequent molecular measurement. Finally, the gastric juice was collected and centrifuged at 4 °C and 13,000 rpm for 10 min. The supernatant was taken and then subjected to pH determination.

Pathological Observation
The gastric tissues were fixed in 10% neutral formalin solution and then dehydrated in ethanol and xylene respectively. After dehydration, the samples were embedded in paraffin wax. 4 μm thick serial slices were obtained and stained with hematoxylin and eosin (H&E) and Alcian blue-Periodic acid-Schiff (AB-PAS) staining. The histopathology of gastric mucosa was observed by Nikon microscope (Nikon instruments, Japan).

Biochemical Detection
The serum biochemical levels of interleukin-6 (IL-6), interleukin-1β (IL-1β), and tumor necrosis factor-α (TNF-α) were measured following the manufacturer’s instructions. Additionally, ELISA kits were used to quantify the levels of Pepsinogen I (PG I), Pepsinogen II (PG II), and Gastrin17 (GAS-17).

TdT-Mediated dUTP Nick-End Labeling (TUNEL) Staining
The gastric tissues were fixed in paraformaldehyde prior to paraffin embedding, and then cut into 4 μm sections. Then, the TUNEL staining was used to evaluate the apoptosis rate as described previously. The apoptosis of cells was observed under light microscope, and the results were expressed by the average number of TUNEL positive staining cells per 200× magnification.
Measurement of Gastric PH

The accurate PH indicator paper was used to detect the PH value of gastric juice in each group after centrifugation. And the same person used a colorimetric card to immediately read out the PH value for qualitative analysis.

Data Preparation of Network Pharmacology

Selection of Target Compounds for ZJP

The compounds of ZJP were collected using the Traditional Chinese Medicine Systems Pharmacology Database (TCMSP, http://lsp.nwu.edu.cn/tcmsp.php). Candidate compounds were screened according to the criteria of oral bioavailability (OB) $\geq$ 30% and drug-likeness (DL) $\geq$ 0.18. Then, the targets of the candidate components of ZJP were screened in SwissTargetPrediction database (http://www.swisstargetprediction.ch/) (Probability > 0). Finally, Cytoscape 3.8.0 software (Cytoscape Consortium, National Institute of General Medical Sciences, USA) was used to make the network diagram of “ZJP-compounds-targets”.

Identification of CAG Targets

The CAG targets were obtained from the GeneCards (http://www.genecards.org/) and Online Mendelian Inheritance in Man (OMIM, http://www.omim.org/) databases with “chronic atrophic gastritis” or “CAG” as the keyword. By integrating these two databases, duplicate targets were removed and all targets were merged and standardized to construct a CAG disease target dataset. Finally, the network diagram of “CAG-target” was visualized by Cytoscape 3.8.0 software.

Construction of Protein-Protein Interaction (PPI) Network

Venn diagram of CAG and ZJP targets was drawn by Venn online tool (https://bioinformatics.psb.ugent.be/webtools/Venn/) and intersection was taken. The PPI network was constructed through the String (https://www.string-db.org/) database and visualized using Cytoscape3.8.0 software.

GO and KEGG Pathway Enrichment Analysis

Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis and Gene Ontology (GO) analysis were performed by linking targets to the Database for Annotation, Visualization and Integrated Discovery (DAVID, https://david.ncifcrf.gov/). GO terms and KEGG pathways with $P$ value < 0.05 were considered statistically significant. Finally, the top 20 KEGG pathways and the top 10 pathways of cell composition, biological process and molecular function in GO enrichment analysis were selected for visual analysis.

Total RNA Extraction and Real-Time PCR

Total RNAs were extracted from gastric tissues with RNA extraction kit (Huaxingbio, Beijing, China), and then the mRNA was reverse transcribed into cDNA template for PCR amplification. RT-qPCR analysis was performed using SYBR Green PCR Master Mix (Huaxingbio, Beijing, China) and executed on 7500 Fast Real-Time PCR System (Applied Biosystems, Foster City, CA, USA). The data was calculated through $2^{-\Delta\Delta CT}$ method with GAPDH as an endogenous reference. The primer sequences were listed in Supplementary Table 1.

Western Blotting Assay

The total protein of gastric tissues were extracted with ice-cold radioimmunoprecipitation assay (RIPA) buffer containing phenylmethylsulfonyl fluoride (PMSF) and the phosphatase inhibitor. The protein concentrations were determined by a BCA Protein Assay Kit. Protein samples were separated on sodium dodecyl sulphatepolyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to polyvinylidene difluoride (PVDF) membranes. Next, the membranes were blocked for 2 h at room temperature. Then, all the membranes were incubated overnight at 4 °C with corresponding primary antibodies and the details were shown in Supplementary Table 2. The next day, all membranes were washed 5 times with TBS-0.1% Tween 20 (TBST) and incubated with secondary antibody for 2 h at room temperature. Finally, the membranes were again washed 5 times with TBST and then visualized using ECL Plus detection kit. GAPDH as internal reference. The ImageJ software (National Institutes of Health, Bethesda, United States) was used for quantitative analysis of the acquired images.
Immunohistochemistry
The gastric tissue samples were first fixed in a 10% formaldehyde solution, then embedded in paraffin and sliced. The tissue sections were dewaxed and dehydrated with graded ethanol. The antigen was extracted by citric acid antigen repair buffer (pH6.0) in microwave for 16 min. After natural cooling, the tissue was placed in phosphate buffered saline (PBS) with pH7.4 for 3 times for 5min. The tissues were incubated in darkness with 3% hydrogen peroxide solution at room temperature for 25 min, then 3% bovine serum albumin was dropped into the histochemical circle, uniformly covered the tissues, and sealed at room temperature for 30 min. Then, the samples were incubated with a primary rabbit polyclonal anti-ZO-1 antibody (Huaxingbio, HX19906, 1:100), rabbit polyclonal anti-Claudin-4 antibody (Huaxingbio, HX13369, 1:100), rabbit polyclonal anti-E-cadherin antibody (Huaxingbio, HX14050, 1:300) and abbit polyclonal anti-Occludin antibody (Huaxingbio, HX20200, 1:300) at 4°C overnight. The sections were then incubated with a horseradish peroxidase (HRP)-conjugated goat anti-rabbit secondary antibody (Huaxingbio, HX2031, 1:5000) and counterstained with hematoxylin. Images of three randomly chosen fields on each slide were captured under a microscope. The quantification of protein expression was presented with integrated optical density (IOD), using ImageJ software (National Institutes of Health, Bethesda, United States).

Immunofluorescence
Paraffin sections of gastric tissues were dewaxed and dehydrated in graded ethanol, and placed in ethylenediaminetetraacetic acid (EDTA) for antigen retrieval. The sections were then washed three times with phosphate-buffered saline (PBS) and blocked for 30 min at room temperature with 1% BSA. Subsequently, the sections were incubated with anti-NF-κB p65 antibody overnight at 4°C. The next day, after incubating with the corresponding secondary antibody for 1 hour at room temperature and staining the nuclei with 4,6-diamino-2-phenylindole (DAPI), images were taken using a confocal microscope (NIKON Eclipse C1, Nikon Instruments Inc., Japan) and analyzed by NIKON DS-U3 (Nikon Instruments Inc., Japan). For quantitative analysis of the NF-κB p65 immunofluorescence staining, integral optical density (IOD) was measured by Image software.

Molecular Docking
Molecular docking was performed after screening key targets and the key ingredients from the PPI network and the “key ingredients-targets” network of ZJP in treatment of CAG. The structure of the key targets was obtained from RCSB database (https://www.rcsb.org/) and the mol2 format files of key ingredients were downloaded from the TCMSP database. AutoDock Tools 1.5.6 was used for molecular docking, and PyMol software was used to visualize the molecular docking results.

Statistical Analysis
Statistical analyses were performed with SPSS 26.0 (International Business Machines Corporation, New York, United States). All data were expressed as means ± standard deviation (SD). Statistical differences between the groups were analyzed with a two-tailed Student’s t-test or one-way analysis of variance (ANOVA). GraphPad Prism software (version 8.0) was used to visualize the results. P < 0.05 was considered statistically significant.

Results
Evaluation of CAG Model Establishment
In order to verify whether the model is successful, the rats in the model group were tested after 10 weeks of continuous modeling. The results showed that the rats in the model group had slower weight gain, listlessness, less activity, messy fur and lack of luster. The gastric mucosa of the rats in the model group became thinner, the mucosal folds were reduced and flattened, the tissue lacked elasticity (Figure 3A). HE staining showed that there was loss of inherent glands, partial mucosal detachment, as well as neutrophil infiltration in the mucosa in the model rats (Figure 3B). Compared with the control group, the body weight of rats decreased significantly after modeling (Figure 3C).
Effect of ZJP on General Condition and Body Weight in CAG Rats

The most typical feature of CAG is weight loss. With the increase of modeling time, the body weight of the model group was significantly lower than that of the control group. And the ZJP group and the vitacoenzyme group exerted well improvement on the body weight, indicating that they had similar therapeutic effect on CAG rats (Figure 4A). The pH values of gastric juice in model group were significantly higher than control group. ZJP had an excellent moderation effect on the pH of gastric juice (Figure 4B). In addition, the rats in the control group were in good condition with shiny fur and free movement, while the rats in model group presented pale yellow color hair, lack of energy, sleepiness, and

Figure 3 Evaluation of CAG model establishment. (A) General state of rats and macroscopic manifestation of gastric tissue in the control and model group. (B) HE staining of gastric tissue in control group and model group. (C) Changes of body weight of rats in control group and model group. All data were expressed as means ± standard deviation (SD). Two group comparisons were analyzed by unpaired two tailed Student’s t-test. **P < 0.01 vs control group.

Figure 4 Effect of ZJP on general condition and body weight. (A) Changes of body weight in each group. (B) The result of gastric juice pH test. (C) General state of rats in the groups. All data were expressed as means ± standard deviation (SD). Multiple comparisons were analyzed by ANOVA. ***P < 0.01 vs control group. **P < 0.01 vs model group.

Abbreviations: ZJPL, Zuojin Pill low dose (1.26g/kg); ZJPH, Zuojin Pill high dose (2.52g/kg); VIT, Vitacoenzyme (200 mg/kg).
decreased appetite. After treatment with ZJP and vitacoenzyme, the above symptoms in each group were alleviated to some extent. The improvement effect of ZJPH group was better than that of ZJPL group (Figure 4C).

### ZJP Attenuated the Pathological Changes of CAG Rats

Macroscopic and microscopic were applied to observe the integral morphology and pathological changes of gastric tissues, respectively. Macroscopic observation showed that the gastric tissue was thinner and the gastric fold was shallower in model group. The morphology of gastric mucosa of rats in the ZJPL, ZJPH and vitacoenzyme groups had deep wrinkles and rich mucosal layer (Figure 5A).

The pathological changes of gastric tissue were observed by HE staining. In the control group, the gastric mucosal cells and proper glands were neatly arranged and the structure was complete. In contrast, the gastric mucosa of the model group rats was atrophic and thin, the glands were significantly reduced, the arrangement was irregular, and the inflammatory cells infiltrated. Moreover, rats in the ZJPL, ZJPH group and the vitacoenzyme group showed orderly arrangement of gastric mucosal cells, reduced degree of glandular atrophy, relatively intact glandular structure, and less infiltration of inflammatory cells (Figure 5B). The thickness of functional gastric mucosa was characterized by AB-PAS staining. Compared with the control group, the thickness of gastric mucosa in the model group was significantly reduced, while the administration of ZJP and vitacoenzyme supplements improved the thickness of the mucosa to varying degrees (Figure 5C).

### ZJP Inhibited Inflammatory Response in CAG Rats

To characterize the inflammatory injury of CAG, inflammation-related factors including TNF-α, IL-1β, IL-6, and NF-κB p65 were detected. As shown in Figure 6A–C, the serum levels of TNF-α, IL-1β and IL-6 in the model group were
notably higher than that in the control group. Compared with the model group, the expression of TNF-α, IL-1β and IL-6 of ZJP group were down-regulated in the ZJP group. In particular, the inhibition effect of ZJPH group was more obvious than that of ZJPL group. In addition, we also analyzed the expression of NF-κB p65 by immunofluorescence, and the results indicated that in the model group, there was a significant increase the expression of NF-κB p65. However, ZJP treatment significantly decreased the expression of NF-κB p65 in a dose-dependent manner, which indicated that ZJP improved the gastric injury by reducing the inflammatory reaction (Figure 6D and E).

**ZJP Improved Biomarkers in Serum of CAG Rats**

To clarify the activities of several specific markers of CAG, the serum levels of PG I, PG II and GAS-17 were measured. As revealed in Figure 7A–C, the levels of PG I and PG II decreased markedly, and the level of GAS-17 increased significantly in model group compared to control group. Conversely, the above indexes reversed significantly after ZJP and vitacoenzyme administration, and the effect of ZJPH group was more obvious than that of ZJPL group.

**Figure 6** Effect of ZJP on inflammatory response. (A) The expression level of TNF-α in serum. (B) The expression level of IL-1β in serum. (C) The expression level of IL-6 in serum (n=6). (D) The NF-κB p65 relative expression. (E) Representative immunofluorescence staining images in gastric mucosa showing NF-κB p65 expressing (400× magnification). All data were expressed as means ± standard deviation (SD). Multiple comparisons were analyzed by ANOVA. ##P < 0.01 vs control group. *P < 0.05 and **P < 0.01 vs model group.

**Abbreviations:** ZJPL, Zuojin Pill low dose (1.26g/kg); ZJPH, Zuojin Pill high dose (2.52g/kg); VIT, Vitacoenzyme (200 mg/kg).
Network Pharmacology and Molecular Docking Predicted Potential Mechanism for ZJP Treatment of CAG
Potential Targets and Cross-Gene Screening of ZJP and CAG

We used network pharmacology technology to predict ZJP components and CAG targets, and obtained enriched signaling pathways, which provided directions for further molecular mechanism studies. A total of 33 compounds and 826 targets of ZJP and 766 targets of CAG related targets were identified after merging and removing duplicate genes (Figure 8A and B). As revealed in Figure 8C and D, 108 overlapped genes related to compounds and CAG were kept for PPI network construction using STRING database. Additionally, visualizing the PPI network through Cytoscape software, larger nodes represent higher degree values and are considered key targets. These key targets were associated with inflammation, apoptosis, and mucosal protection.

GO and KEGG Enrichment Analysis and Construction of PPI Network

To clarify the biological functions and crucial pathways among the key targets in CAG treatment, GO and KEGG pathway enrichment analysis was performed. As shown in Figure 9A–D and Supplementary Tables 3 and 4, the top important 30 signaling pathways of the KEGG and the top 10 GO terms were significantly enriched. KEGG enrichment analysis showed that key target genes were strongly associated with PI3K/Akt signaling pathway, TNF signaling pathway and so on. Among them, the PI3K/Akt signaling pathway is the most significant pathway. The GO results suggest that the top 10 GO biological processes associated with ZJP treatment of CAG included negative regulation of apoptotic process, positive regulation of transcription from RNA polymerase II promoter, positive regulation of gene expression, signal transduction, and protein phosphorylation, etc; the associated cellular components included cytoplasm, nucleus, cytosol, plasma membrane, and nucleoplasm, etc; the related molecular functions included protein binding, identical protein binding, ATP binding, protein kinase activity, and enzyme binding, etc. Next, we structured the interaction network between the top important 30 signaling pathways and GO terms information collected in enrichment analysis and the relevant genes in each signaling pathway, which were concerned with the mechanism of ZJP in treating CAG.

ZJP Regulated Key Genes in the Treatment of CAG

Network pharmacology have predicted that ZJP exerted the therapeutic effect on CAG mainly through inflammation, apoptosis, and mucosal protection. Therefore, we conducted experiments to verify these related targets. Compared with the control group, the mRNA levels of AKT1, TNF-α, CASP 3 and EGFR increased significantly, and the mRNA level of CTNNB1 decreased significantly in the model group. Conversely, different doses of ZJP administration significantly decreased the levels of AKT1, TNF-α, CASP 3 and EGFR, and significantly increased the level of CTNNB1 in a dose-dependent manner (Figure 10A–E).
To verify the role of the PI3K-Akt signaling pathway in ZJP treatment of CAG, the protein expression levels of PI3K, Akt, and p-Akt in the gastric tissue were measured by Western blot analysis. As shown in Figure 11A–C, the expression levels of PI3K and p-Akt were increased in model group compared to the control group. In contrast to the model group, the protein expression of PI3K and p-Akt in each treatment group decreased significantly. There were no significant difference in the level of total Akt expression in the control group, model group and the ZJP groups. In addition, compared with the model group, p-Akt/Akt significantly reversed in a dose-dependent manner after ZJP treatment.

ZJP Inhibited Apoptosis by Reducing Levels of Apoptosis-Related Proteins in CAG Rats

To further elucidate the anti-apoptotic effect of ZJP in the treatment of CAG, apoptotic-related proteins were detected. As shown in Figure 12A–K, the protein expressions of Bcl-2, Bcl-xl, and ratios of Bcl-2/Bax, Bcl-xl/Bad were dramatically reduced, and the Bax, Bad, Cytochrome C, cleaved-caspase-9, cleaved-caspase-3, and Apaf-1 protein expressions were significantly elevated compared with the control group. Conversely, after intragastric administration of ZJP, the abnormal expressions of Bcl-2, Bcl-xl, Bax, Bad, cleaved-caspase-9, cleaved-caspase-3, Apaf-1, Cytochrome C, ratios of Bcl-2/Bax, Bcl-xl/Bad were evidently reversed compared with the model group, which the effect was more pronounced in the ZJPH group. In addition, TUNEL staining was used to detect the effect of ZJP on the apoptosis level of gastric mucosa in rats. Compared to the model group, we detected a reduction of TUNEL-positive cells in gastric tissues after treatment with ZJP (Figure 12L).
ZJP Protected the Integrity on Gastric Mucosa Epithelium

To further evaluated the protective effect of ZJP on the gastric mucosa of CAG rats, the protein expression of Occludin, ZO-1, Claudin-4 and E-cadherin were measured to evaluate the integrity on gastric mucosa epithelium. The results showed that the protein expression of Occludin, ZO-1, Claudin-4 and E-cadherin in the model group was significantly downregulated compared with the control group. In contrast, ZJP significantly reversed the proteins mentioned above. Especially in the ZJPH group. The results of mRNA expression were consistent with those of protein expression (Figure 13A–I).

Molecular Docking Analysis

To further explore the mechanism of ZJP in the treatment of CAG, the key targets and the key ingredients were selected for molecular docking. Based on the network pharmacology results, we screened the top 5 key targets, including TNF-α, EGFR,
CASP 3, AKT1 and CTNNB1, and performed molecular docking with the top 3 key ingredients, including quercetin (QUE), obacunone (OBA) and berberine (BBR). The results showed that the PDB ID numbers of TNF, EGFR, CASP 3, AKT1 and CTNNB1 were 5UUI, 5Y9T, 5IBP, 1UNQ and 1JDH, respectively. The binding energy of most of these ingredients to the targets was less than −5 kcal/mol, indicating a good binding activity and strong stability between the key ingredients and targets (Table 1). In addition, Figure 14A–C exhibited an image of the optimal docking of ingredients and targets after visualization.

Figure 10 Effects of ZJP on key targets of CAG. (A) The TNF-α mRNA relative expression. (B) The CASP 3 mRNA relative expression. (C) The EGFR mRNA relative expression. (D) The AKT1 mRNA relative expression. (E) The CTNNB1 mRNA relative expression (n=6). All data were expressed as means ± standard deviation (SD). Multiple comparisons were analyzed by ANOVA. **P < 0.01 vs control group. *P < 0.05 and ***P < 0.01 vs model group.
Abbreviations: ZJPL, Zuojin Pill low dose (1.26g/kg); ZJPH, Zuojin Pill high dose (2.52g/kg); VIT, Vitacoenzyme (200 mg/kg).

Figure 11 Effects of ZJP on PI3K/Akt signaling pathway. (A) Western blotting images of PI3K, p-Akt and Akt. (B) The relative protein expression of PI3K. (C) Relative protein expression of p-Akt. All data were expressed as means ± standard deviation (SD). Multiple comparisons were analyzed by ANOVA. **P < 0.01 vs control group. *P < 0.05 and ***P < 0.01 vs model group.
Abbreviations: ZJPL, Zuojin Pill low dose (1.26g/kg); ZJPH, Zuojin Pill high dose (2.52g/kg); VIT, Vitacoenzyme (200 mg/kg).
Discussion

CAG is considered a characteristic precancerous lesion of gastric cancer with a high risk of developing into gastric cancer. In recent years, the incidence of CAG has increased and shown a trend towards younger onset, seriously affecting the health and quality of life of an increasing number of patients. ZJP is a commonly used traditional Chinese prescription with a wide range of pharmacological actions in the digestive system. An increasing number of studies have shown that ZJP has anti-inflammatory, antibacterial and mucosal protection effects. With the continuous development of new technology, the mechanism of traditional Chinese medicine in treating CAG is becoming increasingly profound. In this study, the potential mechanism of ZJP in the treatment of CAG was explored by using the network figure.
Figure 13 Effects of ZJP on mucosal integrity. (A) The images of Occludin, ZO-1, Claudin-4 and E-cadherin in gastric tissues of rats was measured using immunohistochemical staining (100 ×). (B) Claudin-4 protein expression level. (C) E-cadherin protein expression level. (D) Occludin protein expression level. (E) ZO-1 protein expression level. (F) The Claudin-4 mRNA relative expression. (G) The E-cadherin mRNA relative expression. (H) The Occludin mRNA relative expression. (I) The ZO-1 mRNA relative expression. All data were expressed as means ± standard deviation (SD). Multiple comparisons were analyzed by ANOVA. *P < 0.05 and **P < 0.01 vs control group. *P < 0.05 and **P < 0.01 vs model group.

Abbreviations: ZJPL, Zuojin Pill low dose (1.26g/kg); ZJPH, Zuojin Pill high dose (2.52g/kg); VIT, Vitacoenzyme (200 mg/kg).
pharmacology combined with experimental verification. The data demonstrated that ZJP exerts a good therapeutic effect on CAG by inhibiting apoptosis and inflammatory response, improving gastric mucosal damage through the PI3K/Akt signaling pathway, which provided a certain theoretical basis for the study of ZJP in the treatment of CAG (Figure 15).

It was found that weight loss is typical clinical feature of CAG. In this study, the body weight of rats in model group decreased significantly, which could be significantly improved after administration of ZJP. In addition, pepsinogen (PG) is a proenzyme secreted by gastric mucosal cells, which exists in the human body in two forms: PG I and PG II. The changes in serum levels of PG I and PG II are effective indicators for reflecting the morphology and function of gastric mucosa. It has been well-documented PGI and PGII are low levels in the CAG model group. Last but not least, GAS-17 is a peptide hormone that participates in the division, proliferation, and apoptosis of gastrointestinal mucosal cells and the secretion of gastric acid. It can indicate the status of gastric mucosal function in the body, and its level is closely related to the occurrence and development of gastric diseases. Studies have found that inactivation of NF-κB is a major redox-sensitive transcription factor that can induce the expression of various pro-inflammatory genes and regulate the expression of inflammatory cytokines. Studies have found that inactivation of

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NF-κB lead to decreased expression of inflammatory cytokines such as TNF-α, IL-6, and IL-1β. Our results suggested that ZJP could significantly inhibit the expression of TNF-α, IL-6, IL-1β, and NF-κB p65, with better effects observed in the ZJPH group. The above results indicate that ZJP exerts its therapeutic effect on CAG by inhibiting the inflammatory response.

Network pharmacology explains the biological network in which drugs play their role from the perspective of macro or global regulation, and provides new research ideas and technical means for studying the mechanism of action of TCM and TCM prescriptions. In the present study, a network pharmacology approach was applied to identify active compounds, corresponding targets, and pharmacological mechanisms of ZJP in the treatment of CAG. The PI3K/Akt signaling pathway was considered an important pathway involved in the treatment of CAG, which exerted therapeutic effects through anti-inflammatory action, inhibition of apoptosis, and protection of gastric mucosa. The PI3K/Akt signaling pathway not only regulates apoptosis, migration, differentiation and metabolism of cells, but also plays

**Figure 14** Docking complexes 3D diagram of 5 key targets along with 3 key ingredients. (A) 3D binding posture schematic diagram of berberine and 5 key targets. (B) 3D binding posture schematic diagram of obacunone and 5 key targets. (C) 3D binding posture schematic diagram of quercetin and 5 key targets.
a role in the reconstruction of gastric mucosa epithelium and promotes the occurrence and development of gastric tumors. Studies have found that epidermal growth factor receptor (EGFR) can activate the expression of the PI3K signaling pathway. Akt is a serine/threonine protein kinase located downstream of PI3K in the intracellular signal transduction system. Phosphorylation of Akt activates its kinase activity, leading to downstream molecular phosphorylation and regulation of cell apoptosis and inflammatory response. In our study, ZJP improved CAG through PI3K/Akt signaling pathway, mainly by significantly reducing the ratio of p-Akt/Akt and the level of PI3K.

It has been reported that mitochondria is considered to be regulatory centers of biological energy, metabolism, and apoptosis, which is closely regulated by a variety of internal and external signals, including chemicals, Bcl-2, and caspase families etc. In general, the mitochondrial apoptosis process is tightly regulated by the proteins of the Bcl-2 family, including anti-apoptotic proteins (Bcl-2 and Bcl-xl) and pro-apoptotic proteins (Bax and Bad). The relative balance between anti-apoptotic proteins and pro-apoptotic proteins directly affects the physiological function of mitochondria. It is well-known that activated Akt can phosphorylate the Ser136 residue of Bad, reducing the binding with Bcl-xl and Bcl-2, thus playing a role in inhibiting apoptosis. In addition, the pro-apoptotic protein induces apoptosis by disrupting the integrity of the mitochondrial membrane, causing a decrease in mitochondrial membrane potential and biochemical changes between the inner and outer mitochondrial membranes, ultimately resulting in the release of cytochrome c from the mitochondria into the cytoplasm. Subsequently, Apaf-1 is activated to form an apoptosome, which then activates caspase-3, triggering cell apoptosis. Caspases are another family of enzymes, also known as aspartate-specific cysteine proteases, which play an important role in regulating the apoptotic signaling

![Figure 15 Potential molecular mechanism of ZJP in the treatment of CAG.](https://doi.org/10.2147/DDDT.S454758)
pathway. Caspase9 is the promoter of Caspase in mitochondrial pathway, which is recruited and activated by apoptotic complex and activates downstream efferent protein Caspase 3 in turn, ultimately leading to cell apoptosis. In the present study, we found that ZJP dramatically reduced the concentration of Cytochrome c in the cytosol, Bax, Bad, cleaved-caspase-9, cleaved-caspase-3 and Apaf-1 protein expressions and significantly elevated the protein expressions of Bcl-2, Bcl-xl, concentration of Cytochrome c in the mitochondria, and ratios of Bcl-2/Bax, Bcl-xl/Bad. These findings indicated that the therapeutic effect of ZJP on CAG was closely connected with inhibiting apoptosis.

The gastric mucosal barrier is an important defense mechanism that protects the body against pathogen invasion and maintains gastric homeostasis. Once it is compromised, it can lead to the occurrence and development of gastric diseases. Cell junction is one of the main structures of gastric mucosal barrier, and tight junction is the most important membrane protein complex of cell junction, which is composed of occludin, claudin, ZOs and junction adhesion molecule (JAM). It is generally known that ZO-1 is a scaffold protein of the tight junction and is often used as an index to evaluate barrier function and permeability function. In addition, occludin is an integral transmembrane protein at tight junctions, and increasing its concentration would enhance cell-to-cell adhesion. Interestingly, in view of the evidence that claudin-4 can enrich occludin and interact with ZO-1. In addition, the expression of claudin-4 is closely related to gastric cancer. Furthermore, E-cadherin has been previously reported to mediate cell adhesion and maintain the integrity of the organizational structure. In our study, the expression of ZO-1, E-cadherin and occludin were significantly decreased, while the expression of claudin-4 was significantly increased in the model group, suggesting that CAG breached the mucosal layer and increased the permeability of gastric mucosa. However, ZJP reversed the expression of ZO-1, E-cadherin, claudin-4 and occludin, indicating that ZJP could protect the gastric mucosal barrier.

Although this study clarified the therapeutic efficacy of ZJP on CAG through integrated biological approach and explained the molecular biological mechanism of ZJP’s improvement of CAG through PI3K/Akt signaling pathway, there are still some limitations in this study. For example, the effect of ingredients of ZJP on CAG has not been verified in this study; Although this study revealed that ZJP inhibits apoptosis and improves inflammation and gastric mucosal barrier through PI3K/Akt signaling pathway, there has been no reverse validation conducted for PI3K/Akt signaling inhibitors or activators in this experiment. Therefore, we will further enrich and refine this study in the future to comprehensively and systematically elucidate the molecular mechanisms and substance basis of ZJP in treating CAG, providing a research foundation for the rational clinical application of ZJP.

Conclusion
In conclusion, our results demonstrated that ZJP exerted a therapeutic effect on CAG by regulating gastric mucosal barrier, suppressing inflammation, inhibiting apoptosis through PI3K/Akt signaling pathway. This study provided a methodological and theoretical basis for further revealing the pharmacological mechanism of ZJP in the treatment of CAG.

Abbreviations
ZJP, Zuojin Pill; IL-6, Interleukin-6; CAG, Chronic atrophic gastritis; IL-1β, Interleukin-1β; TCM, Traditional Chinese medicine; TNF-α, Tumor necrosis factor-α; PG I, Pepsinogen I; OB, Oral bioavailability; PG II, Pepsinogen II; DL, Drug-likeness; GAS-17, Gastrin17; RIPA, Radioimmunoprecipitation assay; KEGG, Kyoto Encyclopedia of Genes and Genomes; PMSF, Phenylmethylsulfonyl fluoride; GO, Gene Ontology.

Ethics Statement
All animal experiments were conducted in accordance with the Laboratory Care and Use Guidelines. The research was approved by Ethics Committee of the Chinese PLA General Hospital (Approval ID: IACUC-2021-0022).

Acknowledgments
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Author Contributions
All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Disclosure
The authors declare that they have no competing interests in this work.

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