The Potential of Natural Compounds Regulating Autophagy in the Treatment of Osteoporosis

Yiwei Zhao*, Zechao Qu*, Songchuan Zhao, Yong Zhang, Yining Gong, Bo Zhang, Xiangcheng Gao, Dong Wang, Liang Yan

Department of Spine Surgery, Honghui Hospital, Xi’an Jiao Tong University, Xi’an, People’s Republic of China

*These authors contributed equally to this work

Correspondence: Liang Yan, Department of Spine Surgery, Honghui Hospital, Xi’an Jiao Tong University, Xi’an, People’s Republic of China, Email yanliangdr5583@163.com

Abstract: The maintenance of bone homeostasis is dynamically regulated by osteoblast-mediated bone formation and osteoclast-mediated bone resorption. Abnormal differentiation of osteoclast and insufficient osteoblast production can cause bone diseases such as osteoporosis. As one of the highly conserved catabolic pathways in eukaryotic cells, autophagy plays an important role in maintaining cell homeostasis, stress injury repair, proliferation and differentiation. Numerous studies have found that autophagy activity is essential for the survival, differentiation and function of bone cells, and that regulation of autophagy can affect the metabolism of osteoblasts and osteoclasts, thus affecting bone homeostasis. Therefore, using autophagy as a theme, this review outlines the basic process of autophagy, the relationship between autophagy and osteoblasts and osteoclasts, and summarizes the latest research progress of common autophagic signaling pathways in osteoblasts and osteoclasts. The regulatory effects of protein molecules and natural compounds on the autophagy pathway of osteoblasts and osteoclasts discovered in current research are summarized and discussed. This will help to further clarify the mechanism of osteoporosis, understand the relationship between autophagy and osteoporosis, and propose new therapeutic strategies and new ideas for anti-osteoporosis.

Keywords: osteoporosis, osteoblast, osteoclast, autophagy signaling pathway, natural compounds

Introduction

Osteoporosis (OP) is a multifactorial systemic bone disease that is characterized by decreased bone mass, destruction of bone microarchitecture, and increased bone fragility. According to a myriad of scientific investigations, it has been established that OP can be attributed to a wide array of factors. At the cellular level, two principal categories of cells have been identified, as elucidated by numerous scholarly studies: Osteoblasts (OBs), originating from bone marrow mesenchymal stem cells, and osteoclasts (OCs), formed through the fusion of mononuclear macrophages, constitute two distinct cell types that hold significant relevance in the context of osteoporosis. Notably, researchers have shown a keen interest in understanding the mechanisms behind OP resulting from inadequate bone formation, orchestrated by OBs, as well as excessive bone resorption, orchestrated by OCs. At the pathogenic level, a multitude of factors, including inflammation, endocrine dysfunction, aging, oxidative stress, among others, have been identified as disruptors of the delicate equilibrium between OBs and OCs in bone metabolism. Hence, the intricate interplay and harmonious equilibrium between OBs and OCs assume paramount importance and must not be disregarded.

Autophagy, a highly conserved self-degradative process in cells, plays a vital role in preserving cellular homeostasis during nutrient scarcity or stressful conditions. It encompasses a distinctive series of processes, including the orchestrated recruitment of autophagy-related proteins, the initiation of autophagic flux, the formation of autophagosomes, and the fusion of autophagosomes with lysosomes. Therefore, any intervention that promotes or disrupts any aspect of this intricate process can modulate the level of autophagy and impact cellular homeostasis. For instance, the autophagy inhibitor bafilomycin A1 has demonstrated the ability to effectively impede the fusion of autophagosomes with lysosomes, thereby attenuating autophagy. Recent accumulating...
evidence substantiates that the modulation of autophagy can mitigate the detrimental effects triggered by hormonal fluctuations, nutrient insufficiency, chronic inflammation, aging, oxidative stress, and effectively safeguard cells against apoptosis. Moreover, dysregulated autophagic activity exerts a profound influence on the differentiation and functionality of OBs and OCs, rendering it inseparable from OP pathogenesis instigated by a myriad of factors, including aging, estrogen deficiency, glucocorticoid exposure, diabetes, and other pertinent etiological elements. Thus, modulating autophagy presents a promising therapeutic strategy for the management of bone-related disorders.

Natural compounds are the result of nature’s screening and evolution over a long period of time, and their rich stereostructures and functional groups are of great significance to the discovery of new drugs and the exploration of drug mechanisms, which have always been an important source of new drug discovery. Many clinically applied drugs are directly or indirectly derived from natural compounds, such as, penicillin, rapamycin, etc. The discovery of these natural compound drugs has made an indelible contribution to human health. In recent years, many natural compounds have been found to have great potential in regulating autophagy and treating osteoporosis.

Given the aforementioned discoveries, this article reviews the autophagy signaling pathways of OBs and OCs, furthermore, it explores the impact of protein molecules and natural compounds that regulate autophagy signaling pathways on the functionality of OBs and OCs, and this article offers a significant reference point for comprehending the pathogenesis of OP. Further provides novel insights for the treatment of OP.

**Overview of Autophagy**

Autophagy can be defined as the process by which cytoplasmic components are delivered to lysosomes for degradation and recycling. Currently, it is well-established that autophagy participates in various cellular metabolism, these include the clearance of misfolded proteins, removal of damaged organelles, elimination of intracellular pathogens, as well as the regulation of cell differentiation and specific functions. The essence lies in conferring survival advantages to cells with active autophagy compared to autophagy-deficient cells when confronted with environmental changes, nutrient scarcity, or energy deficiencies.

**Classification of Autophagy**

In mammalian cells, three types of autophagy have been identified: macroautophagy, microautophagy and chaperone-mediated autophagy. In macroautophagy, the autophagosome, characterized by its double-membrane structure, engulfs damaged organelles, proteins, and invading pathogens, facilitating the degradation of these components and providing recycled materials for the cell’s continued utilization. Macroautophagy is widely recognized as the primary form of autophagy and is often closely associated with various diseases. Therefore, for the purpose of this paper, we will use the term “autophagy” to specifically refer to macroautophagy. Autophagy can be subdivided into two distinct forms: selective autophagy and non-selective autophagy. Extensive research has demonstrated that non-selective degradation of cellular cargo in response to autophagy signals, such as starvation, is crucial for maintaining cellular stability. But now there is also much evidence that highly specific and selective degradation of cargo (such as mitochondria, endoplasmic reticulum, ribosomes, etc.) is also active under nutrient-rich conditions. It can specifically remove dysfunctional organelles, proteins or intracellular pathogens when activated. The former is called non-selective autophagy and the latter is selective autophagy. However, when the damaged mitochondria are specifically cleared by autophagosomes, it is called mitophagy.

**The Process of Autophagy**

The autophagosome, a closed lipid bilayer membrane structure, originates from endoplasmic reticulum and Golgi apparatus. However, the membrane formation entails a multi-stage process, characterized by the sequential involvement of various autophagy-related gene proteins (ATG) and their selective recruitment at distinct time points. Moreover, nearly all ATG, with rare exceptions, play a role in the formation of fully formed autophagosomes. This process encompasses several key steps, including initiation, nucleation, extension, closure, and fusion (Figure 1).

The ULK1 kinase complex (ULK1-C) consists of ULK1, RB1-inducible coiled-coil protein 1, ATG13, and ATG101. When the activation of mTORC1 is restricted, ULK1-C activation initiates autophagy. Type III phosphatidylinositol
3-kinase complexes I (PI3K-C1) is composed of beclin-1, The catalytic subunit vacuolar protein sorting 34 (VPS34), vacuolar protein sorting 15 (VPS15), ATG14, Autophagy and Beclin 1 regulator 1 (AMBRA1), and plays a central role in the autophagy nucleation stage. During phagophore elongation, PI3P effector protein WD repeat domain phosphoinositide interacting protein 2 (WIPI2) binds to the membrane, and WIPI2 binds directly to the ATG16L1 complex (ATG16L1-C: ATG12-ATG5-ATG16L). ATG16L1-C promotes the formation of LC3-II. LC3-II then binds to the phagosomal membrane, promotes the elongation of the autophagosome membrane, and helps autophagosome membrane closure and fusion with lysosomes. Finally, the small GTPase Rab7 assists the fusion of the autophagosome and the lysosome.

The Involvement of Autophagy in OBs and OCs

OBs originate from mesenchymal stem cells in the bone marrow, under certain conditions, preosteoblasts differentiate into mature OBs. Mature OBs exhibit the expression of essential genes, including alkaline phosphatase and osteocalcin. Additionally, they actively secrete crucial components such as osteocalcin and osteonectin. These secreted molecules interact with inorganic salt components, facilitating mineralization and contributing to the process of osteogenesis. Autophagy contributes to the osteogenic differentiation of mesenchymal stem cells and facilitates the secretion of crucial components of OBs. In vitro studies have demonstrated that canonical Wnt signaling inhibits the differentiation of mesenchymal stem cells into adipocytes and promotes the differentiation of MSCs into OBs.

Irisin, identified as an anti-inflammatory cytokine, has been shown to play a crucial role in promoting the differentiation of mesenchymal stem cells into OBs. This effect is achieved by activating autophagy through the Wnt/β-catenin signaling pathway. According to the literature, Runx2 is an OBs-specific transcription factor that plays a crucial role in regulating both the differentiation and function of OBs. During the process of OBs differentiation, Runx2 exhibits high expression levels in preosteoblasts. Elevating the expression of Runx2 has been shown to enhance OBs autophagy and increase the levels of p-p38MAPK. This upregulation influences the expression of osteogenesis-related genes such as alkaline phosphatase and osteocalcin. Runx2 possesses the capacity to enhance autophagy in OBs, a process that also can be achieved by regulating the expression of lysosomal-associated transmembrane protein 5, which is a regulatory mechanism that influences OBs mineralization and its subsequent influences the overall bone formation process. In addition, the targeted knockout of ATG7, a specific autophagy-related gene, results in autophagy impairments, which in turn causes endoplasmic reticulum stress in OBs and subsequently hinders osteoblast mineralization in a DNA damage-induced transcript 3-mitogen-activated protein kinase 8-SMAD1/5/8-dependent manner, demonstrating the positive role of autophagy in maintaining OBs stability and mineralization.
However, the energy generator of mitochondria plays an important role in osteoblast differentiation. In osteoblast cell line MC3T3-E1 and primary calvarial osteoblasts, the formation of mitochondria and supercomplex occurs continuously during osteoblast differentiation, and SIRT3 enhances SOD2 activity and regulates mitochondrial stress. PTEN-induced kinase 1 (PINK1) can trigger mitophagy and regulate the quality of mitochondria. The differentiation of osteoblasts is inhibited by the down-regulation of PINK1, accompanied by the damage of mitochondrial homeostasis and the increase of mitochondrial reactive oxygen species. The activation of PINK1 is a necessary condition for osteoblast differentiation.

Due to the biological activity of exosomes, exosomes from bone marrow mesenchymal stem cells (BMSC-Exos) circular RNA (circRNA) promote the differentiation of MC3T3-E1 into osteoblasts, among them, circHIPK3 was verified to be the key to promoting osteoblast differentiation in BMSC-Exos, which plays a role in mitophagy by targeting miR-29a-5p and PINK1. In addition, the expression of Kruppel-like factor 2 (KLF2) and autophagy-related genes are significantly increased during osteoblast differentiation. When mitophagy is promoted, the levels of KLF2 and osteogenic differentiation-related molecules are increased. Chromatin immunoprecipitation analysis confirmed that KLF2 and active epigenetic markers (H3K27Ac and H3K4me3) are up-regulated in the promoter region of ATG7 and play a crucial role in regulating mitophagy and osteoblast differentiation.

Furthermore, a plethora of studies have demonstrated that autophagy plays a huge role in the anti-aging and anti-apoptosis of OBs. A recent study has identified discoidin domain receptor 1 (DDR1) as a collagen receptor that actively contributes to bone development. Intriguingly, the knockout of the DDR1 gene has been shown to markedly diminish autophagy levels while concurrently triggering OBs apoptosis. Heat shock protein 60 (HSP60) has been recognized as a key regulator in preserving protein functionality within the cellular microenvironment. Notably, in OBs subjected to glucocorticoid treatment, HSP60 maintains the expression of specific autophagy-related genes, such as Atg4 and Atg12, and promotes autophagic flux. This process ultimately mitigates apoptosis in OBs, highlighting the cytoprotective effects of HSP60 in this cellular context. Similarly, in the galactose-induced OBs senescence model, glucosamine can improve the apoptosis of osteoblasts by promoting autophagy. However, in the context of hypoxia, OBs exhibit an elevation in the expression of cyclin-dependent kinase 8 (CDK8), and the overexpression of CDK8 has been found to induce both autophagy and apoptosis in this cellular setting.

The Involvement of Autophagy in OCs
OCs, derived primarily from hematopoietic progenitor cells of the monocyte-macrophage lineage, serve as the exclusive cells responsible for bone resorption within the human body. Mature OCs secrete acidic substances and proteases, which facilitate the dissolution of minerals and the digestion of collagen fibers within the bone matrix, thereby promoting the process of bone resorption. During this intricate process, a large actin ring sealing region is formed in the cytoplasm of mature multinucleated OCs to degrade the bone matrix, and the actin ring surrounds a folded boundary. The edge of the fold undergoes fusion between lysosomes and the plasma membrane, resulting in the formation of an acidic microenvironment. Within this microenvironment, OCs secrete various proteolytic enzymes, including cathepsin K (CTSK), to efficiently degrade the bone matrix. Previous studies have reported that autophagy-related proteins, including Atg5, Atg7, Atg4B, and LC3, play a crucial role in governing the formation of fold boundaries in OCs. These proteins are instrumental in guiding the fusion process between lysosomes and the plasma membrane, which is essential for facilitating OCs secretion and enabling efficient bone resorption. Specifically, Atg5 has been identified as a key regulator in facilitating the localization of LC3-mediated lysosomes at the edges of OCs folds, thereby exerting a significant influence on the bone resorption function of OCs. In addition, the migration of OCs on the bone surface is crucial to the function of bone resorption, and the migration of OCs requires the continuous and rapid assembly and disassembly of podosomes. Nevertheless, when LC3-mediated autophagy is inhibited, there is an accumulation of kindlin3, a critical adapter protein. This accumulation amplifies the interaction between kindlin3 and integrins, leading to the hyperactivation of integrins. This, in turn, impedes the degradation of podosomes and disrupts OCs migration.

During OCs differentiation, monocyte-macrophage colony stimulating factor (M-CSF) and receptor activator for nuclear factor-κB ligand (RANKL) are the key stimulating molecules to induce mature osteoclasts. RANK on the surface of OCs binds to RANKL released by OBs and promotes TNF receptor associated factor (TRAF)-mediated downstream signaling. As reported in the literature, silencing of the autophagy-related gene Beclin-1 has been shown to impede the degradation of
TRAF3, thus affecting the regulation of osteoclastogenesis through the RANKL/Beclin-1/Autophagy/TRAF3 axis.\textsuperscript{54} In addition, in a study conducted by Ke et al it was discovered that RANKL stimulation leads to the phosphorylation of BCL2 at the Ser70 (S70) site. This phosphorylation event triggers autophagy in OCs precursors (OCPs), thereby promoting OCs differentiation. Interestingly, when the S70 site of BCL2 was mutated via site-directed mutagenesis, it led to the inhibition of autophagy activity in OCPs and instead promoted apoptosis in these precursor cells.\textsuperscript{55}

Additionally, mitochondria are highly active during OCs formation, and mitochondrial dynamics (fusion, fission, and mitophagy) are at a high level. Mitofusin2 (MFN2) is a protein that can regulate adjacent mitochondrial fusion, promote mitochondrial calcium absorption, and regulate mitophagy. OCs differentiation is regulated by MFN2 through Ca\textsuperscript{2+}-NFATc1 axis.\textsuperscript{56} Mitochondrial deacetylase sirtuin-3 (Sirt3) is associated with aging-induced bone loss. Although osteoclast progenitor cells in Sirt3-deficient senescent mice can differentiate into OCs, differentiated cells show decreased levels of mitophagy and impaired bone resorption.\textsuperscript{57} In vitro, PTEN-induced kinase 1 (PINK1) -dependent mitophagy is enhanced by inhibiting signal transducer and activator of transcription 3 (STAT3) and PTEN-induced kinase 1 (PINK1) -dependent mitophagy, which affects nlrp3 activation and regulates osteoclastogenesis.\textsuperscript{58} In addition, the fate of mature OCs is related to mitophagy. By inhibiting mitophagy, osteoclast division can be inhibited and osteoclast apoptosis can be induced. By enhancing mitophagy, osteoclast-osteomorph recycle can be promoted, and the bone resorption ability of fused OCs is proved.\textsuperscript{59}

### Autophagy of Osteoblasts and Osteoclasts in Osteoporosis

Bone is a living, dynamic organ that is constantly remodeling, and in this dynamic process, it is regulated by two important cellular activities: OBs and OCs.\textsuperscript{60} During the intricate process of bone remodeling, OCs activation initiates bone resorption, while OBs activation promotes mineralization. These two processes are intricately interconnected, constantly influencing each other’s activity. Furthermore, a series of cellular molecules come into play, orchestrating the delicate balance between bone catabolism and anabolism.\textsuperscript{61} However, the essence of autophagy involved in the pathogenesis of osteoporosis is to affect the differentiation and osteogenesis of OBs and the differentiation and bone resorption function of OCs. Baseline levels of autophagy exist in all cells, maintaining homeostasis and promoting cell survival. Abnormal autophagy of OBs and OCs caused by pathological environment may be an important factor in breaking the delicate balance of bone remodeling and leading to bone loosening (Figure 2).

Primary osteoporosis involves a variety of molecular mechanisms, but oxidative stress and estrogen reduction seem to be the two most critical factors. Two types of primary osteoporosis: postmenopausal OP and age-related OP are associated with advanced oxidation protein product (AOPP: an oxidative stress product) levels.\textsuperscript{62,63} In vitro, AOPP interacts with RANK and RAGE to activate nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, induce ROS production and induce osteoclast differentiation and maturation,\textsuperscript{64} and ROS induced mitochondrial dysfunction, endoplasmic reticulum stress and Ca\textsuperscript{2+} overload in osteoblasts by inducing phosphorylation of mitogen-activated protein kinases (MAPKs), leading to bone loss.\textsuperscript{65} More importantly, ROS is involved in the regulation of autophagy of osteoblasts and osteoclasts by activating downstream molecules FOXO3, AMPK, Akt, etc., as a therapeutic target for osteoporosis.\textsuperscript{10} In postmenopausal OP, the decline in estrogen levels results in reduced cellular autophagy and increased susceptibility of OBs to apoptosis, estrogen receptor antagonist (ICI18270) combined with estrogen receptor (ER) to reduce autophagy leads to osteoblast death, and estradiol activation of ER-ERK-mTOR pathway may be a pathway to save osteoblast apoptosis.\textsuperscript{66} 17β-Estradiol can activate the downstream ERK signal to regulate osteoblast mitophagy through the G protein-coupled receptor 30 (GPR30) as one of the estrogen receptors.\textsuperscript{67} Estrogen regulatory protein RAB3GAP1 is a catalytic component of the RAB3GAP complex and plays a role in autophagosome biosynthesis. Estrogen deficiency and reduced interaction with RAB3GAP1 lead to reduced autophagy in human osteoblasts, with corresponding reductions in both osteoblast lifespan and osteoblast mineralization.\textsuperscript{68} Moreover, the hormone estrogen (specifically 17β-estradiol) directly amplifies pro-osteoclast autophagy while concurrently impeding OCs differentiation.\textsuperscript{69}

Glucocorticoids, as the main force of anti-inflammatory, have been widely used for a long time. In secondary osteoporosis, the disadvantages of glucocorticoid-induced bone loss are gradually exposed.\textsuperscript{70} It has been reported that the expression of autophagy-related genes, such as ATG7 and Beclin 1, is reduced by 1–5 times in dexamethasone-induced OP in vivo and in vitro.\textsuperscript{62} The level and activity of autophagy in osteoblasts treated with different concentrations of dexamethasone are also different. Low level of autophagy induced by ground rice contributes to the survival of osteoblasts, while high level of dexamethasone can lead to autophagy inhibition and OBs function inhibition and apoptosis.\textsuperscript{63} In addition, Rankl-induced OCs...
formation was significantly higher in the glucocorticoid (1μM) treatment group than in the hormone-free treatment group, and dexamethasone-induced ROS played a key role in regulating autophagy. Interestingly, inhibition of autophagy by 3-MA did not cause ROS accumulation.

Diabetes is also an important cause of secondary osteoporosis. It is known that hyperglycemia and insulin resistance may induce osteoblast death, and hyperglycemia and glucose fluctuation are two common states in diabetic patients, both of which inhibit osteoblast proliferation and activate autophagy, generating ROS leading to apoptosis induced by a decrease in the mitochondrial membrane potential of OBs. Non-imprinted in Prader-Willi/Angelman syndrome region protein 2 (NIPA2), a highly selective magnesium transporter, has been reported to be associated with type 2 diabetic osteoporosis. Among them, high glucose (35 mM glucose) decreased the expression of NIPA2 osteoblasts and induced mitophagy, while NIPA2 can regulate mitophagy and mitophagy is negatively correlated with osteogenic ability in high glucose environment.

**Signaling Pathways Involved in Autophagy in OBs and OCs**

**Autophagy in OBs and OCs: AMPK Signaling Pathway**

AMPK is an AMP-dependent protein kinase, controlled by the AMP: ATP ratio and its upstream signal, and is a sensor of energy metabolism. The downstream of AMPK molecules, there are mainly two molecules, mTOR and ULK1, that regulate autophagy and participate in the metabolism of bone homeostasis. In the absence of energy, phosphorylated AMPK causes tuberous sclerosis complex 2 (TSC2) phosphorylation, resulting in the formation of TSC1/TSC2 complex, resulting in the inactivation of Rheb and the loss of the ability to activate mTOR. In addition, AMPK directly phosphorylates Raptor, preventing Raptor from binding to mTOR, which in turn leads to mTOR inactivation. The inactivation of mTOR eliminates its inhibition of ULK1 phosphorylation and induces the binding of ULK1 to AMPK, one of the main mechanisms by which AMPK activates autophagy. In OBs, AMPK activation induces autophagy, as evidenced by upregulation of LC3-II, downregulation of p62 and increased autophagosome density, while enhancing OBs differentiation and mineralization.
Various factors can affect the AMPK signaling pathway in OBs. Under simulated negative pressure treatment conditions, activation of AMPK-ULK1 axis promotes MSCs differentiation into OBs. Metformin can target AMPK signaling in glucocorticoid-induced OBs apoptosis. As a small ribosomal 40S subunit S6 protein kinase, P70S6K enables the small ribosomal 40S subunit to easily bind to translation complexes and promote autophagy-related protein synthesis, metformin regulates AMPK signaling and affects downstream mTOR and P70S6K molecules to alter autophagy levels and reverse OBs apoptosis.

AMPK signaling exhibits reliable regulation of OCs autophagy levels in a high level glucose environment, inhibition of AMPK/mTOR/ULK1 can reduce the high autophagy level of OCs mediated by high concentration of glucose, and inhibit the formation of OCs, elucidating the mechanism of glucose-mediated OCs formation. In addition, osteoprotegerin (OPG), a paracrine inhibitor derived from OBs, has been shown to be an important molecule involved in osteoclastogenesis. It can bind to RANK to block the binding of RANK to RANKL and inhibit OCs formation. According to reports in the literature, OPG inhibits OCs differentiation and bone resorption by enhancing autophagy in vitro, and the mechanism is that OPG activates AMPK protein, which regulates OCs autophagy through the AMPK/mTOR/p70S6K pathway. In the field of autophagy, the effect of OPG on OCs has been elucidated, and AMPK signaling regulates autophagy in OCs.

**Autophagy in OBs and OCs: PI3K/AKT/mTOR Signaling Pathway**

Phosphatidylinositol 3-kinase (PI3K) is a lipid kinase with three types of I, II and III in mammals. It is mainly produced on the lipid membrane and recruits downstream lipid signals such as serine / threonine kinase (AKT) to participate in cell metabolism. Light has many biological effects, including promoting tissue repair, improving neuroinflammation, anti-apoptosis, etc. Photobiomodulation (PMB) treatment can reduce the phosphorylation levels of PI3K and AKT, enhance OBs autophagy by inhibiting PI3K/AKT/mTOR signaling pathway, alleviate OBs apoptosis and enhance OBs differentiation. The Jintiange (JTG) protein in the artificial tiger bone JTG can regulate the expression of key proteins in the PI3K/AKT and endoplasmic reticulum stress (ER) pathways in osteogenesis, and enhance the formation of autophagosomes, through PI3K/AKT/mTOR and ER stress signaling pathways enhance autophagy to promote OBs formation and inhibit OBs apoptosis.

High levels of cholesterol can affect the expression of PI3K signaling and autophagy proteins in OCs, and inhibit autophagy and OCs differentiation through PI3K/AKT/mTOR. The expression of PI3K, p-Akt and p-mTOR was inhibited when excessive glucocorticoid induced OCs autophagy. The PI3K/Akt/mTOR signaling pathway can increase OCs autophagy and promote OCs formation. In addition, OPG regulates OCs differentiation by altering OCs autophagy not only through AMPK signaling, but also by regulating the AKT signaling pathway. According to Zhao et al reported that phosphorylated AKT and mTOR levels were decreased in OPG-treated OCs, and OPG regulates autophagy by activating PI3K/AKT/mTOR signaling pathway leading to OCs viability, differentiation ability and bone resorption both activities are weakened.

**Autophagy in OBs: FOXO1/3 Signaling Pathway**

FOXO is a transcription factor that responds to various external stimuli, including oxidative stress, fluctuations in energy levels, and other factors. In mouse OBs, the deletion of FOXO1, FOXO3, and FOXO4 has been found to induce oxidative stress and elevate OBs apoptosis. Conversely, upregulation of FOXO3 expression in mature OBs has shown to confer resistance against oxidative stress and reduce cell death. Furthermore, FOXO3 can mitigate OBs senescence by regulating the methylation of FOXO3. These findings highlight the crucial role of FOXO-dependent oxidative defense in counteracting the detrimental effects of free radicals generated during the aerobic metabolism of OBs. According to literature reports, inhibiting the FOXO1 signaling pathway has been shown to decrease the autophagy level in OBs exposed to a high glucose environment, consequently impacting the overall function of OBs. On the other hand, it has been reported that FOXO3 induces autophagy in order to mitigate the generation of ROS during mitochondrial metabolism in OBs. This protective mechanism helps safeguard OBs from potential damage. In addition, Furthermore, it has been observed that treatment of MC3T3-E1 cells with low-dose dexamethasone leads to the upregulation of glucocorticoid-induced kinase-1 (SGK1), which subsequently facilitates the dephosphorylation of FOXO3. This dephosphorylation event triggers the activation of autophagy, thereby enhancing cell viability and improving the overall function of OBs.
Autophagy in OBs: SIRT1/3 Signaling Pathway
Silent Information Regulator (SIRT) proteins belong to the NAD⁺-dependent deacetylase family. The subtypes SIRT1-7 have been implicated in human aging-related diseases and are known to regulate various biological responses in cells, including immune responses and autophagy.⁹⁸,⁹⁹ SIRT1 and SIRT3 have been found to have a close association with autophagy in osteoblasts. Notably, SIRT1 functions as an NAD⁺-dependent histone that offers cellular protection through the process of deacetylating target proteins.¹⁰⁰ In the hFOB1.19 cell model, 17β-estradiol has been shown to induce an upregulation of SIRT1 gene expression. This increase in SIRT1 expression subsequently promotes OBs autophagy while inhibiting apoptosis.¹⁰¹ SIRT3, a prominent mitochondrial deacetylase, plays a significant role in oxidative metabolism and the response to oxidative stress. Reducing the signal of SIRT3 leads to weakening the differentiation of adipocytes into osteoblasts.¹⁰² According to the literature, increasing the expression of SIRT3 can reduce superoxide dismutase 2 (SOD2) acetylation and promote SOD2 activity, maintain mitochondrial stability, and regulate autophagy through SIRT3/SOD2 signaling pathway to resist titanium ion-induced osteoblast damage.¹⁰³

Autophagy in OBs: PINK1/Parkin Signaling Pathway
PTEN-induced kinase 1 (PINK1), a serine/threonine kinase with a mitochondrial targeting sequence, plays a critical role in regulating mitochondrial quality control. During OBs differentiation, the knockout of the PINK1 gene disrupts mitochondrial homeostasis and leads to elevated production of ROS, thereby inhibiting OBs differentiation.³⁸ In age-related OP, a large number of advanced oxidation protein products (AOPPs) promote the production of mitochondrial ROS (mROS), which aggravates oxidative stress and promotes OBs apoptosis, while PINK1/Parkin-mediated mitophagy can eliminate excessive mROS and damaged mitochondria to reverse AOPP-induced OBs apoptosis.¹⁰⁴ Excess ROS can activate mitophagy, which is an important mechanism for controlling mitochondrial quality and cell homeostasis. Interestingly, when dibutyl phthalate (DBP) treated OBs, a large amount of ROS was generated and accompanied by the activation of mitophagy. However, silencing Parkin expression inhibited mitophagy and alleviated DBP-induced MC3T3-E1 dysfunction and apoptosis.¹⁰⁵ In addition, it has been reported that dexamethasone attenuates OBs mitophagy by downregulating the expression of PINK1, Parkin genes, resulting in the inhibition of OBs differentiation and mineralization in vitro.¹⁰⁶

Autophagy in OCs: Beclin-1/Bcl-2 Signaling Pathway
Beclin-1 is an important autophagy-initiating protein with multiple roles in OCs differentiation. Interleukin-17A (IL-17A) can promote the differentiation of OCs precursors into OCs, inhibit the phosphorylation of extracellular regulated kinase (ERK) to increase the expression of Beclin-1, enhance the autophagy activity of OCs precursors through the ERK/mTOR/Beclin1 pathway, and promote OCs differentiation.¹⁰⁷ Chung et al found that knockout of Beclin-1 inhibited RANKL-mediated activation of JNK and p38, thereby inhibiting the expression of OCs-specific gene NFATc1.¹⁰⁸ In vitro, autophagy is activated during RANKL-induced OCs differentiation and requires TRAF6-mediated Beclin-1 ubiquitination, and Beclin-1 knockout mice exhibit impaired OCs bone resorption and thickened bone cortex.¹⁰⁹ Beclin-1 inhibits OPG-induced osteoclastogenesis by increasing autophagy. In the process of OPG inhibiting OCs differentiation stimulated by RANKL and M-CSF, the expression of matrix metalloproteinase 9 (MMP-9) decreased and the expression of Beclin-1 increased. Beclin-1 knockdown can reduce the level of autophagy and resist the number of OCs inhibited by OPG.¹¹⁰ In addition, it has been reported that metformin can be used as an autophagy regulator and is widely involved in the protection of various tissues. Metformin-mediated down-regulation of E2F1 in ovariectomized mice reduces the expression levels of Beclin-1 and BNIP3, which subsequently interferes with the binding of Beclin-1 to Bcl-2, inhibits the formation of autophagosomes, and leads to the inhibition of osteoclast formation, thereby reducing OVX-induced bone loss.¹¹¹

Autophagy in OCs: ROS Signaling Pathway
ROS are critical to cellular signaling and physiological functions. However, oxidative stress leading to excess ROS also induces autophagy and affects the metabolic level of osteoclasts.¹⁰,¹¹² Cholesterol in human body is positively correlated with reactive oxygen species. Cholesterol metabolite 7-ketocholesterol (7-KC) enhances autophagy and increases the number and activity of osteoclast through ROS-TFEB signaling pathway.¹¹³ Doxorubicin (DOX) is a widely used chemotherapeutic drug
that can induce excessive autophagy of osteoclasts, leading to bone loss and increasing the risk of human fractures. The mechanism is due to DOX-induced increase in mitochondrial ROS levels, causing mROS to oxidize the transient receptor potential mucin 1 (TRPML1) in lysosomes, leading to nuclear localization of TFEB, increasing autophagy levels and OCs differentiation.\textsuperscript{114}

\section*{Regulation of Protein Molecules on Autophagy of OBs and OCs}

The tumor suppressor M receptor (OMSR) is expressed on a variety of cells, including OBs and OCs. It has been reported that Osmr-/- OBs are defective in differentiation and lead to reduced bone formation.\textsuperscript{115} Zhou et al reported the direct role and mechanism of OMSR in controlling osteogenic differentiation. OSMR inhibits extracellular regulated kinase (ERK) signaling and autophagy, while silencing OSMR gene can promote OB differentiation, reduce fat accumulation and inhibit OBs differentiation.\textsuperscript{116} Taurine up-regulated gene 1 (TUG1) is one of the earliest discovered lncRNAs related to human diseases. Hypoxia can up-regulate the expression of TUG1, and knockout of TUG1 can promote SIRT1-induced mitophagy.\textsuperscript{117,118} In addition, TUG1 plays an important role in regulating OBs and improving OP. The expression of TUG1 increased during the differentiation of BMSCs into OBs, while in TUG1-silenced BMSCs, the expression of AMPK and P53 decreased and the number of autophagosomes decreased, demonstrating the potential of TUG1 to promote the differentiation of BMSCs into OBs by regulating the AMPK/mTOR/autophagy axis.\textsuperscript{119} Non-imprinted in Prader-Willi/Angelman syndrome region protein 2 (NIPA2) is a highly selective magnesium transporter. The overexpression and silencing of NIPA2 gene indicated that NIPA2 regulates PINK1/Parkin-mediated mitophagy through the PGC-1/FoxO3a pathway, affecting the function of OBs.\textsuperscript{73}

\section*{Natural Compounds Regulate OBs and OCs Autophagy Signaling Pathways to Produce Anti-OP}

\subsection*{Natural Compounds Regulate OBs Autophagy Signaling Pathway}

Paeoniflorin is a compound extracted from traditional Chinese medicine with anti-tumor, anti-inflammatory and other pharmacological effects. It can promote autophagy through the AKT/mTOR signaling pathway to reduce dexamethasone-induced OBs
Table 1 Regulatory Molecules of Autophagy in Osteoblast and Osteoclast

<table>
<thead>
<tr>
<th>Regulators</th>
<th>Target Cell</th>
<th>Pathway</th>
<th>Main Finds</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>OSMR</td>
<td>BMSCs</td>
<td>ERK/autophagy signaling pathway</td>
<td>Silencing OMSR gene can alleviate ERK signal inactivation and enhance autophagy to promote osteoblast differentiation.</td>
<td>[116]</td>
</tr>
<tr>
<td>TUG1</td>
<td>BMSCs</td>
<td>AMPK/mTOR signaling pathway</td>
<td>Silencing TUG1 gene inhibits autophagy and osteoblast differentiation via AMPK/mTOR signaling pathway.</td>
<td>[119]</td>
</tr>
<tr>
<td>NIPA2</td>
<td>hFOB1.19</td>
<td>PINK1/Parkin signaling pathway</td>
<td>Silencing NIPA2 gene enhanced PINK1/Parkin-mediated mitophagy and down-regulated the function of osteoblasts.</td>
<td>[73]</td>
</tr>
<tr>
<td>c-Fos</td>
<td>BMNs</td>
<td>Beclin-1, PI3K/Akt, TAK1/S6 signaling pathway</td>
<td>Overexpression of the c-Fos gene reverses OPG-mediated inhibition of osteoclastogenesis by activating Beclin-1-induced autophagy.</td>
<td>[121]</td>
</tr>
<tr>
<td>KIF2</td>
<td>RAW264.7</td>
<td>Beclin-1 signaling pathway</td>
<td>KIF2 knockdown significantly increases Beclin-1 expression, thereby regulating autophagy and osteoclast differentiation.</td>
<td>[123]</td>
</tr>
<tr>
<td>JNK1</td>
<td>RAW264.7</td>
<td>Beclin-1/Bcl2 signaling pathway</td>
<td>Silencing of the JNK1 gene reduces autophagy and osteoclastogenesis, and Beclin-1 silencing blocks JNK1-promoted autophagy responses in OCPs.</td>
<td>[125]</td>
</tr>
<tr>
<td>TET2</td>
<td>RAW264.7</td>
<td>Beclin-1/Bcl2 signaling pathway</td>
<td>Knockdown of TET2 gene increases Bcl2 expression and decreases Beclin-1-dependent autophagy to reduce osteoclast differentiation.</td>
<td>[126]</td>
</tr>
<tr>
<td>P2X7R</td>
<td>BMNs</td>
<td>Ca(^{2+})/Calcineurin/NFATc1 signaling pathway</td>
<td>Knockdown of P2X7R gene inhibits Ca(^{2+})/Calcineurin/NFATc1 inhibits autophagy and osteoblast bone resorption.</td>
<td>[128]</td>
</tr>
<tr>
<td>TRPV4(^{1})</td>
<td>RAW264.7</td>
<td>Ca(^{2+})/Calcineurin/NFATc1 signaling pathway</td>
<td>Knockdown of TRPV4 gene inhibits autophagy via Ca(^{2+})/calcineurin/NFATc1 pathway, thereby inhibiting osteoclast differentiation and osteoporosis.</td>
<td>[129]</td>
</tr>
</tbody>
</table>

apoptosis and promote bone formation.\(^{130}\) Leonurine is a natural compound extracted from Motherwort that has been shown to have anti-inflammatory and antioxidant effects and is effective in the treatment of many diseases including cardiovascular disease and ischaemic stroke.\(^{131}\) It can promote the differentiation of BMSCs into OBs by increasing autophagy through the PI3K/Akt/mTOR signaling pathway. Leonurine may become a new method for the treatment of OP.\(^{132}\) Resveratrol is a polyphenolic hexene compound found in plants such as grapes, mulberry, peanuts, and rhubarb. It can reduce oxidative stress and inflammation and promote autophagy.\(^{133,134}\) When resveratrol inhibits the PI3K/Akt/mTOR pathway to enhance autophagy, PI3K inhibitors or SIRT1 inhibitors can eliminate the effects of resveratrol-induced mitophagy in OBs. The expression of SIRT1 also plays an important role in resveratrol-induced autophagy to protect OBs in osteoporotic rats.\(^{135}\) Geniposide is a natural compound. It can not only inhibit some inflammatory pathways such as NF-κB, MAPK, etc., but also increase some antioxidant enzymes such as superoxide dismutase and lipid peroxidase.\(^{136}\) In OBs, gardenia glycosides can alleviate glucocorticoid-induced endoplasmic reticulum stress and OBs apoptosis by inhibiting the PI3K/Akt/mTOR signaling pathway to activate autophagy and reduce cell damage and apoptosis.\(^{137}\) β-ecdysterone is a polyhydroxysteroid hormone, known as phytoestrogen. Studies have shown that β-ecdysterone reversed the effects of dexamethasone-treated BMSCs apoptosis and autophagy in a dose-dependent manner in vitro.\(^{138}\) In addition, Tang et al found that β-ecdysterone can activate autophagy by inhibiting PI3K/Akt/mTOR signaling pathway to promote OBs differentiation.\(^{139}\) Alpinetin, a flavonoid compound, is an important active ingredient in Alpinia katsumadai. It has been found to have anti-tumour, antioxidant and anti-inflammatory pharmacological effects. In Alpinetin-treated OBs, the expression of p-PKA was most affected, while the expression levels of p-JNK, p-AKT, p-p38, p-β-catenin, p-ERK, BMP2 or p-Smad were weakly affected, regulating autophagy through the PKA/mTOR/ULK1 signalling pathway. Therefore, Alpinetin may become a potential drug for the treatment of OP by targeting the PKA-autophagy pathway.\(^{140}\)

Timosaponin BII is a steroidal saponin extracted from Anemarrhena asphodeloides Bge, which has the pharmacological activity in regulating glucose and lipid metabolism.\(^{141}\) And oxidative stress in osteoblasts under high glucose conditions can cause autophagy defects, which may be the pathogenesis of diabetic osteoporosis, Wang et al reported that Timosaponin BII can inhibit the phosphorylation of mTOR and downstream factor NF-κB, activate autophagy through mTOR/NF-κB signaling pathway, reduce oxidative stress and OBs apoptosis induced by high glucose, and alleviate the deterioration of tibial microstructure in diabetic rats.\(^{142}\) Cladrin are the main bioactive components found in the stem
bark of purple rivet, which can promote the proliferation and differentiation of OBs. Peng et al showed that the apoptosis rate of OBs treated with Cladrin in dexamethasone environment was significantly lower than that of OBs without Cladrin treatment, and the level of p-AMPK/T-AMPK was significantly up-regulated, the AMPK/mTOR signaling pathway was significantly involved in Cladrin-induced autophagy in OBs under dexamethasone. Aucubin, an iridoid glycoside mainly found in Chinese herbal medicine, can enhance OBs autophagy through the AMPK signaling pathway to counteract steroid-induced apoptosis. Trehalose, a naturally occurring non-reducing disaccharide found in several plants and algae, has been reported to enhance cellular antioxidant capacity by increasing the expression of Nrf2 in cells. Trehalose can also protect mitochondrial function by activating transcription factor EB (TFEB)-mediated autophagy, moreover, Cao et al found that trehalose alleviated OBs apoptosis induced by palmitic acid by enhancing OBs autophagy through AMPK/mTOR/ULK1 signaling pathway and up-regulating SIRT3 expression. Tetramethylpyrazine (TMP) is extracted from Ligusticum chuanxiong Hort, which has anti-apoptotic effect. During the study of TMP protecting BMSCs from Dex-induced apoptosis, it was found that TMP can activate AMPK/mTOR signaling pathway to promote autophagy of BMSCs exposed to dexamethasone. Cistanche Deserticola is usually used in the treatment of kidney deficiency in traditional Chinese medicine. Based on the theory of “kidney dominate bone”, Cistanche Deserticola can be used as an alternative drug for the intervention of osteoporosis. Cistanoside A (Cis A) is a phenylethanoid glycoside extracted from Cistanche Deserticola, which has the antioxidant effect of inhibiting ROS activity to inhibit apoptosis. The expression of Wnt, β-catenin protein and autophagy were enhanced in OBs treated with Cis A. When Dickkopf-1 (DDK-1), an inhibitor of Wnt/β-catenin pathway, was used, it showed a significant decrease in LC3 fluorescence and a decrease in autophagosomes. Cis A induces autophagy by activating the Wnt/β-catenin signaling pathway and promotes primary OBs mineralization and osteogenesis (Figure 3).

Natural Compounds Regulate OCs Autophagy Signaling Pathway
Kaempferol, a flavonoid compound renowned for its anti-inflammatory and antioxidant properties, has garnered considerable attention. Notably, literature reports have elucidated its potent capacity to inhibit the expression of p62/SQSTM1 protein in OCs, thereby suppressing autophagy and attenuating OCs differentiation and bone resorption. Curcumin, derived from the turmeric root, has shown promising potential in ameliorating glucocorticoid-induced OP. It has a dual regulatory effect on

Figure 3 Mechanism diagram of natural compounds regulating autophagy signaling pathway in osteoblasts. Different natural compounds regulate osteoblast autophagy through a variety of different molecular mechanisms, thereby affecting osteoblast differentiation or cell death.
pre-OCs. It can not only stimulate the autophagy of pre-OCs alone, but also inhibit the expression of Beclin-1 in pre-OCs induced by RANKL, and reduce the level of autophagy, resulting in a decrease in OCs formation. This action positions curcumin as a prospective candidate for targeted drug development in the realm of OP treatment.\textsuperscript{156} Orcinol glucoside (OG), a naturally occurring phenolic glycoside derived from Curculigo orchioides, exhibits a multitude of beneficial properties, including antioxidative, anxiolytic, neuroprotective, and anti-osteoporotic activities.\textsuperscript{157,158} Senile OP is associated with oxidative stress, Gong et al have reported that OG holds the potential to mitigate oxidative stress and curb OCs autophagy by activating the Nrf2/Keap1 and mTOR signaling pathways. Additionally, OG exerts a down-regulatory effect on OCs differentiation and function, thereby presenting a preventive approach against senile OP.\textsuperscript{158} Lycorine is an alkaloid isolated from Lycoraceae plants, which has anti-inflammatory and antioxidant effects.\textsuperscript{159} Notably, lycorine exhibits the ability to diminish the production of mROS, modulate the triggering signal for autophagy via the mROS/TRPML1/TFEB axis, and mitigate lipopolysaccharide-induced OCs autophagy, as well as suppress OCs activity and abundance.\textsuperscript{160} Epigallocatechin-3-gallate (EGCG) is a polyphenolic compound extracted from the leaves of Camellia sinensis. EGCG has attracted much attention because of its antioxidant, antiviral, anti-inflammatory, anti-aging and effects, and effectiveness for many diseases.\textsuperscript{161} In the mechanism of inhibiting OCs differentiation, EGCG not only blocks the binding of Rank and Rankl, but also reduces ROS and mitochondrial membrane potential, inhibits the expression of mitophagy-related molecules, and regulates mitophagy through AKT and p38MAPK pathways.\textsuperscript{162} Here we summarize the details of the possible link between natural compounds regulating autophagy in OBs and OCs (Table 2).

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Structure</th>
<th>Dose</th>
<th>Target Cell</th>
<th>Mechanism</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Paeoniflorin</td>
<td><img src="image" alt="Paeoniflorin" /></td>
<td>10, 20µM</td>
<td>MC3T3-E1</td>
<td>Paeoniflorin can attenuate osteoblast apoptosis by inhibiting Akt/mTOR pathway to enhance autophagy</td>
<td>[130]</td>
</tr>
<tr>
<td>Leonurine</td>
<td><img src="image" alt="Leonurine" /></td>
<td>2, 5, 10µM</td>
<td>BMSCs</td>
<td>Leonurine can promote osteoblast differentiation by inhibiting PI3K/Akt/mTOR pathway and activating autophagy.</td>
<td>[132]</td>
</tr>
<tr>
<td>Resveratrol</td>
<td><img src="image" alt="Resveratrol" /></td>
<td>10^6, 10^-7, 10^-6 M</td>
<td>MC3T3-E1</td>
<td>Resveratrol can enhance autophagy and SIRT1 expression through the PI3K/Akt/mTOR signaling pathway.</td>
<td>[135]</td>
</tr>
<tr>
<td>Geniposide</td>
<td><img src="image" alt="Geniposide" /></td>
<td>10, 20µM</td>
<td>MC3T3-E1</td>
<td>Geniposide can ameliorate osteoblast apoptosis by activating PI3K/ AKT/mTOR to enhance autophagy.</td>
<td>[137]</td>
</tr>
<tr>
<td>β-ecdysterone</td>
<td><img src="image" alt="β-ecdysterone" /></td>
<td>20, 40, 60µM</td>
<td>Rat primary osteoblasts</td>
<td>β-ecdysterone can inhibit PI3K/Akt/mTOR pathway to activate autophagy and reduce osteoblast apoptosis</td>
<td>[139]</td>
</tr>
</tbody>
</table>

(Continued)
Table 2 (Continued).

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Structure</th>
<th>Dose</th>
<th>Target Cell</th>
<th>Mechanism</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alpinetin</td>
<td><img src="image" alt="Structure" /></td>
<td>10, 20, 40 μM</td>
<td>BMSCs</td>
<td>Alpinetin can augment PKA/mTOR/ULK1 signaling pathway to enhance autophagy and promote osteoblast differentiation.</td>
<td>[140]</td>
</tr>
<tr>
<td>Timosaponin BII</td>
<td><img src="image" alt="Structure" /></td>
<td>0.1, 1, 10μM</td>
<td>Rat primary osteoblasts</td>
<td>Timosaponin BII can inhibit mTOR/NF-κB signaling pathway and activate autophagy to attenuate oxidative stress and apoptosis.</td>
<td>[142]</td>
</tr>
<tr>
<td>Cladrin</td>
<td><img src="image" alt="Structure" /></td>
<td>1, 10, 100 nM</td>
<td>Mice calvarial osteoblasts</td>
<td>Cladrin can induce autophagy through AMPK/mTOR signaling pathway to protect osteoblasts.</td>
<td>[144]</td>
</tr>
<tr>
<td>Aucubin</td>
<td><img src="image" alt="Structure" /></td>
<td>50, 100, 150 μM</td>
<td>MC3T3-E1</td>
<td>Aucubin can enhance autophagy by activating AMPK signaling pathway to avoid steroid-induced osteoblast apoptosis.</td>
<td>[145]</td>
</tr>
<tr>
<td>Trehalose</td>
<td><img src="image" alt="Structure" /></td>
<td>50, 100, 150, 200μM</td>
<td>hFOB 1.19</td>
<td>Trehalose can activate AMPK/mTOR/ULK1 pathway to induce autophagy and improve palmitic acid-induced apoptosis</td>
<td>[149]</td>
</tr>
<tr>
<td>Tetramethylpyrazine</td>
<td><img src="image" alt="Structure" /></td>
<td>50, 100, 200 μM</td>
<td>BMSCs</td>
<td>Tetramethylpyrazine can activate AMPK/mTOR to promote autophagy and protect BMSCs from apoptosis.</td>
<td>[150]</td>
</tr>
<tr>
<td>Cistanoside A</td>
<td><img src="image" alt="Structure" /></td>
<td>5, 10, 20 μM</td>
<td>Rat cranial osteoblasts</td>
<td>Cistanoside A can enhance osteoblast differentiation by inducing autophagy through the Wnt/β-catenin pathway.</td>
<td>[153]</td>
</tr>
<tr>
<td>Kaempferol</td>
<td><img src="image" alt="Structure" /></td>
<td>5, 10, 25, 50, 75, 100 μM</td>
<td>RAW264.7</td>
<td>Kaempferol can inhibit autophagy and activate osteoclast apoptosis through p62/SQSTM1 pathway.</td>
<td>[154]</td>
</tr>
<tr>
<td>Curcumin</td>
<td><img src="image" alt="Structure" /></td>
<td>5, 10, 15μM</td>
<td>BMMs</td>
<td>Curcumin can inhibit autophagy by inhibiting Beclin1 signaling, thereby causing an anti-osteoclast generation effect.</td>
<td>[156]</td>
</tr>
</tbody>
</table>

(Continued)
Conclusion
OP is a disease of bone metabolism imbalance, which is caused by the imbalance of OBs and OCs metabolism in bone homeostasis. Autophagy can regulate cell proliferation, differentiation, maturation and metabolic processes, and can resist cell damage caused by various reasons such as aging and stress, and provide energy and material basis for cell homeostasis and survival. Autophagy plays an important role in maintaining cell function and dynamic balance of OBs and OCs. In addition, there are a variety of autophagy signaling molecules that activate OBs and OCs. Researchers have found that a large number of protein and natural compounds molecules can play an anti-OP role by regulating the autophagy signaling pathway to affect the differentiation of OBs and OCs, showing the potential of autophagy in bone remodeling. Therefore, regulating autophagy-related signaling pathways may be a target for understanding the pathogenesis of OP and future treatment of OP, providing new methods and ideas for the treatment of OP.

Funding
National Natural Science Foundation of China (82260181), Key project of Natural Science Basic Research Plan of Shaanxi Province (2022JZ-43) and the National Key Research and Development Program of China (No. 2022YFC2407503).

Disclosure
The authors report no conflicts of interest in this work.

References

Table 2 (Continued).

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Structure</th>
<th>Dose</th>
<th>Target Cell</th>
<th>Mechanism</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Orcinol glucoside</td>
<td><img src="image1" alt="Structure" /></td>
<td>1.5 µM, 10µM</td>
<td>RAW264.7</td>
<td>Orcinol Glucoside can attenuate oxidative stress and osteoclast autophagy by activating Nrf2/Keap1 and mTOR signaling pathways.</td>
<td>[158]</td>
</tr>
<tr>
<td>Lycorine</td>
<td><img src="image2" alt="Structure" /></td>
<td>1.6 µM, 3.2µM</td>
<td>BMMs</td>
<td>Lycorine can inhibit autophagy through the mROS/TRPML1/TFEB axis, reducing the number and activity of osteoclasts.</td>
<td>[160]</td>
</tr>
<tr>
<td>Epigallocatechin-3-gallate</td>
<td><img src="image3" alt="Structure" /></td>
<td>10, 25, 50 µM</td>
<td>RAW264.7</td>
<td>Epigallocatechin-3-gallate regulates mitophagy through AKT and p38MAPK pathways, thereby inhibiting OC differentiation.</td>
<td>[162]</td>
</tr>
</tbody>
</table>
Zhao et al.


57. Ling W, Krager K, Richardson KK, et al. Mitochondrial Sirt3 contributes to the bone loss caused by aging or estrogen deficiency. JCI Insight. 2021;6(10). doi:10.1172/jci.insight.146728


Dovepress

105. Cui Y, Li B, Du J, et al. Dibutyl phthalate causes MC3T3-E1 cell damage by increasing ROS to promote the PINK1/Parkin-mediated autophagic cell death of osteoblasts via the SIRT3/SOD2 pathway.


Dovepress

Publish your work in this journal

The Journal of Inflammation Research is an international, peer-reviewed open-access journal that welcomes laboratory and clinical findings on the molecular basis, cell biology and pharmacology of inflammation including original research, reviews, symposium reports, hypothesis formation and commentaries on: acute/chronic inflammation; mediators of inflammation; cellular processes; molecular mechanisms; pharmacology and novel anti-inflammatory drugs; clinical conditions involving inflammation. The manuscript management system is completely online and includes a very quick and fair peer-review system. Visit http://www.dovepress.com/testimonials.php to read real quotes from published authors.

Submit your manuscript here: https://www.dovepress.com/journal-of-inflammation-research-journal

Journal of Inflammation Research 2023:16

Zhao et al


Submit your manuscript here: https://www.dovepress.com/journal-of-inflammation-research-journal