

# Strain Identification and Drug Resistance Analysis of Matrix-Assisted Laser Desorption/Ionization Time-of-Flight Mass Spectrometry in Nontuberculous Mycobacterial Lung Disease

Jichan Shi<sup>1,\*</sup>, Gexin Gao<sup>2,\*</sup>, Jing Pan<sup>3</sup>, Lian-Peng Wu<sup>4</sup>, Hongye Ning<sup>1</sup>, Zhengxin Wu<sup>1</sup>, Xinchun Ye<sup>1</sup>, Xiangao Jiang<sup>1</sup>

<sup>1</sup>Department of Infectious Disease, The Theorem Clinical College of Wenzhou Medical University, Wenzhou Central Hospital, Wenzhou, People's Republic of China; <sup>2</sup>Department of Nursing, Wenzhou Medical University, Wenzhou, People's Republic of China; <sup>3</sup>Department of Geriatric Medicine, The Theorem Clinical College of Wenzhou Medical University, Wenzhou Central Hospital, Wenzhou, People's Republic of China; <sup>4</sup>Department of Laboratory, The Dingli Clinical Institute of Wenzhou Medical University, Wenzhou Central Hospital, Wenzhou, People's Republic of China

\*These authors contributed equally to this work

Correspondence: Xiangao Jiang, Department of Infectious Disease, The Theorem Clinical College of Wenzhou Medical University, Wenzhou Central Hospital, 252 BaiLiDong Road, Wenzhou, Zhejiang, 325000, People's Republic of China, Tel +86 13676788085, Email xiangaojiang09@163.com

**Objective:** To evaluate the clinical value of matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) in detecting *Nontuberculous mycobacteria* (NTM).

**Methods:** The clinical data of 172 patients with suspected NTM lung disease were collected from our hospital from January 1, 2018, to December 30, 2021. The results were compared with those of BACTEC MGIT 960 in liquid culture and gene chip. This study also utilised MALDI-TOF MS to detect macrolide (MA) and amikacin (Am) mutations.

**Results:** One hundred thirty-seven cases of NTM pulmonary disease were confirmed by identifying the NTM gene chip in bronchoalveolar lavage fluid and/or MALDI-TOF MS detection. The positive predictive value and negative predictive value were 100% (131/131) and 85.37% (35/41), respectively, and the consistency of the two methods was high ( $\kappa=0.899$ ). For the drug resistance detection of MAs, the consistency rate between MALDI-TOF MS detection and drug sensitivity detection was 97.71% (128/131), the sensitivity was 81.25% (13/16) and the specificity was 100% (115/115). The positive and negative predictive values were 100% (13/13) and 93.75% (115/118), respectively. There was no coincidental consistency between the two methods, and the consistency was high ( $P<0.001$ ,  $\kappa=0.884$ ). For the drug resistance test of Am, the consistency rate between the MALDI-TOF MS test and the drug sensitivity test was 93.13% (122/131), the sensitivity was 93.52% (101/108), the specificity was 90.91% (21/23) and the positive predictive value and negative predictive value were 98.06% (101/103) and 75.00% (21/28), respectively. The two methods had high consistency, and the consistency was not coincidental ( $P<0.001$ ,  $\kappa=0.781$ ).

**Conclusion:** Utilising MALDI-TOF MS has a good consistency with the drug resistance gene chip method and can be a rapid and effective method to identify strains and drug resistance of NTM. Therefore, it has certain clinical application value in patients with suspected NTM lung disease.

**Keywords:** *Nontuberculosis mycobacteria*, matrix-assisted laser desorption/ionization time-of-flight mass spectrometry, strain identification, resistance

## Introduction

The name *Nontuberculous mycobacteria* (NTM) refers to mycobacteria other than *Mycobacterium tuberculosis* complex (*tuberculosis mycobacterium*, MTBC) and *Mycobacterium leprae*, which are widely present in nature and belong to opportunistic pathogens.<sup>1</sup> They can cause disease in various systems, including the respiratory tract, digestive tract, skin and mucous membranes. An infection of the human body with NTM is called NTM disease, which causes lesions of related

tissues and organs.<sup>2</sup> In recent years, global *NTM* disease has rapidly increased and has become a major public health problem.<sup>3</sup> It is also one of China's rising public health concerns, with more *NTM* pulmonary disease observed in south and coastal China.<sup>4</sup> My country has not yet formed a unified scheme for identifying *NTM*, which mainly relies on molecular detection technology; there is a great deal of controversy over the primers used. For some rare *NTMs*, whole-genome sequencing is required for identification.<sup>5,6</sup> In addition, *NTM* lung disease and tuberculosis are very similar in clinical and imaging manifestations,<sup>7</sup> increasing the misdiagnosis rate. Therefore, finding new etiological detection methods, improving the identification of *NTM*, and the ability to diagnose drug resistance would result in earlier diagnosis and treatment, reducing the amount of damage *NTM* disease does to the human body.

Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) was first used to identify mycobacteria in 2004. The use of MALDI-TOF MS in the analysis of oligonucleotides is a new approach and has developed rapidly in recent years. Compared with traditional identification methods, MALDI-TOF MS simplifies the identification process, shortens the identification time and has the advantages of accuracy, rapidity, sensitivity and high throughput.<sup>8</sup> This study evaluated the clinical value of MALDI-TOF MS in detecting *NTM* compared with the BACTEC MGIT 960 liquid culture system and gene chip.

## Materials and Methods

### General Information

The clinical data of 172 patients with suspected *NTM* lung disease who visited Wenzhou Central Hospital, a tertiary Class A general hospital, from January 1, 2018, to December 30, 2021, were collected. All subjects met the diagnostic criteria for suspected *NTM* lung disease formulated by the Tuberculosis Branch of the Chinese Medical Association.<sup>9</sup> Inclusion criteria (one of the following conditions was sufficient): (1) The sputum test for acid-fast bacilli was positive, and the clinical manifestations were inconsistent with pulmonary tuberculosis; (2) Those with positive acid-fast bacilli in sputum or other specimens and negative in molecular biology of *MTBC*; (3) Patients for whom regular anti-tuberculosis treatment was ineffective and repeatedly excreted bacteria; and whose lung lesions were mainly bronchiectasis, multiple small nodules or thin-walled cavities.

At the same time, two bronchoalveolar lavage fluid (BALF) specimens from each patient were collected. All patients signed informed consent forms, and the research protocol was approved by the hospital's medical ethics committee (ethics approval ID: L2021-03-083).

### Method

Flow charts provided quick access to relevant information (Figure 1).

#### Gene Chip Testing

The BALF specimens were identified using the Boao gene chip method.<sup>10</sup> The length of the 16S rRNA primer was about 1500 bp, the upstream primer was 5'-AGTTTGATCCTGGCTCAG-3' and the downstream primer was 5'-GGTACCTTGTTACGACTT-3'. The length of the *hsp65* primer was 441 bp, the upstream primer was 5'-ACCAACGATGGTGT-GTCCAT-3' and the downstream primer was 5'-CTTGTCGAACCG-CATACCCT-3'.

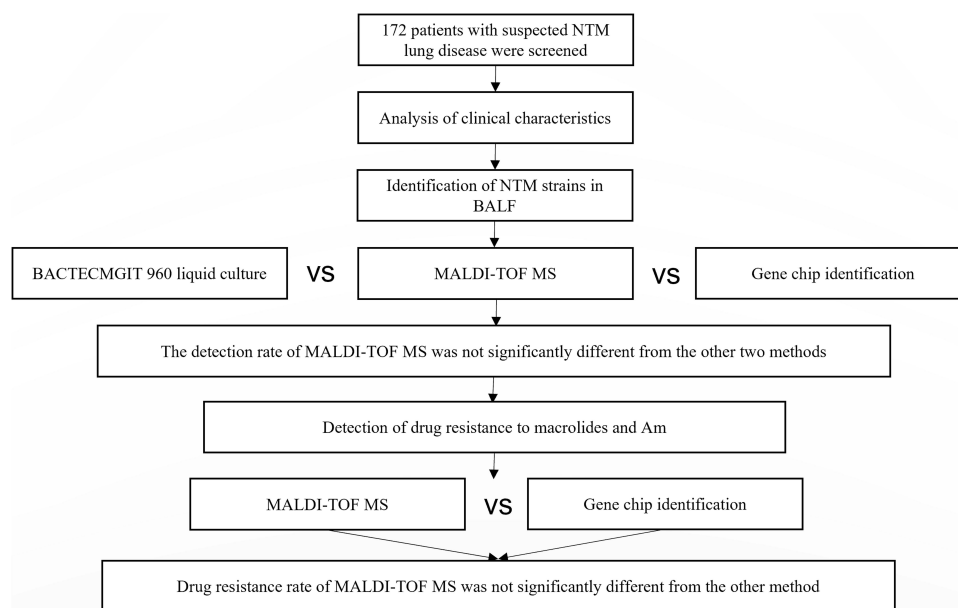
#### Mycobacterium Culture

BACTEC MGIT 960 was used for mycobacterial culture. After the BALF was thoroughly mixed, 5 mL was put into a centrifuge tube, and the digestion solution was added. It was vortexed until the digestion was complete. The supernatant was discarded after separation, and a small amount of polybutylene succinate was added to the pellet to resuspend it. The suspension was added to the MGIT tube (CAT #70937T) for culture. Culture-positive tubes were reconfirmed by acid-fast staining and subsequent identification of bacterial species.

#### Polymerase Chain Reaction and MALDI-TOF MS

##### Specimen DNA Extraction

The 50-mL collection tube was marked with the serial number and name and centrifuged at 10,000 rpm for 15 min. Then, the supernatant was discarded to keep the pellet, and 1.5–2 times the pellet volume of 4% sodium hydroxide (NaOH) was



**Figure 1** The flow chart of research.

added to the corresponding 50-mL tube. This was shaken and mixed for 30s, incubated in a 45°C water bath to make it completely liquefied (the liquefaction time was controlled within 1 h). If the liquefaction had not been completed in 1 h, the clear and transparent part was taken for the experiment. The fully liquefied sample was transferred to a 1.5-mL Eppendorf (EP) tube. The sample was centrifuged at 15,000 rpm for 5 min. The supernatant was carefully aspirated after centrifugation. One mL of normal saline was added to each tube; each tube was shaken, mixed, and centrifuged at 15,000 rpm for 5 min. The supernatant was carefully aspirated after centrifugation. Next, 200–600 uL of lysis solution F5 was added to all samples according to the precipitation volume, then transferred to 1.5-mL cryopreservation tubes (if there were two tubes in one sample, they were combined into one tube), shaken and mixed well, and incubated in a metal bath at 99°C for 10 min. After the metal bath was complete, they were centrifuged briefly. All the liquid in the 1.5-mL EP tube was transferred to the first well of the reagent strip of Lab-Aid 824 *Mycobacterium tuberculosis* nucleic acid extraction, Maxi reagent. All samples treated with 4% NaOH had 10 uL of enhancement fluid added. The corresponding serial number of the sample was written on the edge of the tenth hole of the reagent strip. After ensuring that all reagent strips and trays were fixed on the sample rack, the magnetic rod sleeve was inserted into the groove of the magnetic rod. After the magnetic rod sleeve was locked in place, the extraction was performed, and after the extraction was completed, the liquid was aspirated from the last reaction well for subsequent polymerase chain reactions (PCRs).

### Nucleic Acid Amplification

The PCR solution (prepared in advance) was added to a 96- or 384-well plate. Then 2 uL of the sample to be tested was added. The DNA extracted from the negative and positive samples was added to the reaction plate for product quality control during use. Then, the PCR product was purified. The reaction system was SAP Buffer 0.17 uL, SAP Enzyme (1.7 U/uL) 0.3 uL, and ultrapure water was added to make up 2 uL of solution. Next, 2 uL of SAP reaction solution was added to each reaction well. The reaction conditions were 37°C for 40 min and 85 °C for 5 min. After the SAP reaction, the extension reaction was carried out. The reaction system was: iPLEX Buffer 0.2 uL, iPLEX Termination mix 0.2 uL, Extend Primer Mix 0.94 uL and iPLEX Enzyme 0.041 uL. Finally, ultrapure water was added so the total volume equaled 2 uL, and 2 uL of the extension reaction solution was added to each reaction well.

3) Genotype interpretation according to the peak map of nucleic acid mass spectrometry detection by MALDI-TOF MS was performed and compared with the wild-type genotype to determine whether drug resistance occurred at the detection site as reported.<sup>11</sup>

Interpretation of results: See Table 1 for examples of the test samples.

**Table I** Examples of MALDI-TOF MS Detection of Drug Resistance Genes in *Nontuberculous mycobacteria*

Non-tuberculous mycobacteria (NTM)						
Positive						
<i>(Mycobacterium abscessus)</i>						
NTM drugs						
Macrolides (clarithromycin, azithromycin)	M1	M2	M3	M4	M5	M6
Aminoglycosides (Amikacin, kanamycin, gentamicin)	●	○		○	○	○

**Notes:** Note 1: ○ May be sensitive. Note 2: ● May be resistant. Note 3: - Not sure. Note 4: This assay contains the following mutation sites: The drug resistance gene loci of NTM drug detection are: M1: erm 28; M2: rrl 2058; M3: rrl 2059; M4: rrs 1406; M5: rrs 1408; M6: rrs 1409; among them, erm 28 only has guiding significance for the infection of *Mycobacterium abscessus* complex flora. Note 5: The mutation of erm 284 was detected in this sample, indicating that *Mycobacterium abscessus* in this sample is resistant to macrolides.

## Statistical Analysis

SPSS software version 22.0 was used for statistical analysis of the experimental data, and the enumeration data were expressed as percentages (%). The consistency between groups was compared by the kappa consistency test, and  $P < 0.05$  indicated that the consistency between different testing methods was not coincidental. The BACTEC MGIT 960 cultured gene chip drug resistance detection was used as a reference, and the efficacy of MALDI-TOF MS detection to identify *NTM* strains was evaluated. The agreement between the two methods was judged by the kappa test. A kappa value of 0.81–1 is almost perfect; 0.61–0.80 is highly consistent (substantial); 0.4–0.60 is moderate; 0.21–0.40 is fair; and 0.0–0.20 indicates very low consistency (slight).<sup>12</sup>

## Results

### General Information

Among the 172 patients with suspected *NTM* lung disease, ages ranged from 36 to 78 years, with an average age of 57. This included 47 males and 125 females, a body mass index of  $22 \pm 5$  kg/m<sup>2</sup>. Out of the 172 total patients, 108 (62.79%) had underlying lung diseases, including 54 cases (31.39%) *bronchiectasis*, 28 cases (16.28%) had chronic obstructive pulmonary disease; 21 cases (12.21%) had a history of pulmonary tuberculosis; 9 cases (5.23%) had silicosis; and 4 cases (2.33%) had pulmonary aspergillosis. In addition, 60 cases had comorbidities (34.88%), including 57 cases of diabetes (33.14%), 18 cases of hypertension (10.47%); 3 cases of long-term immunosuppressive therapy after kidney transplantation (1.74%); and 2 cases of rheumatoid arthritis (2.91%). Of the 172 patients, 137 cases of *NTM* lung disease tested positive by BALF *NTM* gene chip identification and/or MALDI-TOF MS.

### Identification Results of *NTM* Strains

Among 172 patients with suspected *NTM* lung disease, 152 were positive for *Mycobacterium* in BACTEC MGIT 960 liquid culture, and 137 (79.65%, 137/172) were identified as *NTM* by gene chip. Thirteen isolates were identified. *Mycobacterium avium* complex (MAC) was the most commonly isolated strain, including *Mycobacterium intracellulare*, *Mycobacterium avium*, *Mycobacterium marseillense* and *Mycobacterium colombiense*, accounting for 70.07% (96/137), followed by *Mycobacterium abscessus*, accounting for 21.17% (29/137). Other *NTMs* only accounted for 8.76% (12/137) (Table 2).

### Efficacy Comparison of MALDI-TOF MS and BALF Culture

Among the 172 patients, 148 were positive for *Mycobacterium* by MALDI-TOF MS, and 131 (76.61%, 131/172) were identified as *NTM*. There was no coincidental consistency in the positive detection rate of *Mycobacterium* between MALDI-TOF MS and the BACTEC MGIT 960 liquid culture method ( $\chi^2=131.57$ ,  $P < 0.001$ ). The sensitivity and specificity were 97.37% (148/152) and 100% (20/20), respectively. The positive predictive value and negative predictive value were 100% (148/148) and 83.33% (20/24), respectively, and the two detection methods were highly consistent (kappa=0.896). See Table 3.

**Table 2** The Bacterial Species Identification Results of 137 Patients with Confirmed NTM Lung Disease

Cultivation Result Classification	Number of Plants (n)	Composition Ratio (%)
Mycobacterium avium complex	96	70.07
Mycobacterium abscessus	29	21.17
Other	12	8.76
Total	137	100

**Table 3** Efficacy of MALDI-TOF MS Detection of Mycobacteria in BALF Specimens with the Results of Liquid Culture as the Reference Standard

		BACTEC960 Liquid Culture		Sensitivity (%)	Specificity (%)	Coincidence Rate (%)	Positive Predictive Value (%)	Negative Predictive Value (%)	Kappa Value
		Positive	Negative						
MALDI-TOF MS	Positive Negative	148 4	0 20	97.37	100	94.19	100	83.33	0.896

**Notes:** The left side of the dotted line is the crosstab form. To the right of the dotted line is a three-line table.

## Comparison of Detection Efficiency of MALDI-TOF MS and the Gene Chip Method

There was no coincidental consistency in the positive detection rate of *NTM* strains detected by MALDI-TOF MS and the gene chip method ( $\chi^2=140.40$ ,  $P<0.001$ ). The sensitivity and specificity were 95.67% (131/137) and 100% (35/35), respectively. The positive predictive value and negative predictive value were 100% (131/131) and 85.37% (35/41), respectively, and the two detection methods were highly consistent ( $\kappa=0.899$ ). See [Table 4](#).

## Efficacy Analysis of MALDI-TOF MS for Detection of Drug Resistant Mutations

The efficacy analysis of 131 samples identified as *NTM* by the gene chip method and MALDI-TOF MS was performed. For the detection of drug resistance to macrolides (MAs), the consistency rate between MALDI-TOF MS detection and drug susceptibility detection was 97.71% (128/131). The sensitivity was 81.25% (13/16), the specificity was 100% (115/115) and the positive and negative predictive values were 100% (13/13) and 93.75% (115/118), respectively. There was no coincidental consistency in the detection of MA resistance rates between the two methods, and the consistency was high ( $\chi^2=94.84$ ,  $P<0.001$ ,  $\kappa=0.884$ ). For the drug resistance detection of amikacin (Am), the consistency rate between MALDI-TOF MS detection and drug susceptibility detection was 93.13% (122/131). The sensitivity was 93.52% (101/108), the specificity was 90.91% (21/23) and the positive and negative predictive values were 98.06% (101/103) and 75.00% (21/28), respectively. There was no coincidental consistency in the detection rate of Am resistance between the two methods, and the consistency was high ( $\chi^2=93.92$ ,  $P<0.001$ ,  $\kappa=0.781$ ). See [Table 5](#).

**Table 4** The Efficacy of MALDI-TOF MS Detection of NTM in BALF Specimens with the Results of Gene Chip Method as the Reference Standard

		Gene Chip Method		Sensitivity (%)	Specificity (%)	Coincidence Rate (%)	Positive Predictive Value (%)	Negative Predictive Value (%)	Kappa Value
		Positive	Negative						
MALDI-TOF MS	Positive Negative	131 6	0 35	95.67	100	96.51	100	85.37	0.899

**Notes:** The left side of the dotted line is the crosstab form. To the right of the dotted line is a three-line table.

**Table 5** Analysis of the Efficacy of MALDI-TOF MS for the Detection of Drug Resistance Mutations

			Gene Chip		Sensitivity (%)	Specificity (%)	Compliance Rate (%)	Positive (%)	Negative (%)	X <sup>2</sup>	P	Kappa Value
			Positive	Negative								
Macrolides	MALDI-TOF MS	Positive	13	0	81.25	100	97.71	100	97.46	94.84	<0.001	0.884
		Negative	3	115								
Amikacin		Positive	101	2	93.52	90.91	93.13	98.06	75.00	93.92	<0.001	0.781
		Negative	7	21								

**Notes:** The left side of the dotted line is the crosstab form. To the right of the dotted line is a three-line table.



## Discussion

The clinical data, imaging manifestations and laboratory examinations from 172 patients treated in our hospital with suspected *NTM* lung disease were statistically analysed. The results showed that most patients were middle-aged to elderly, and most suffered from chronic underlying lung diseases. The common clinical manifestations were cough and expectoration, night sweats, chest tightness, shortness of breath, sputum and blood haemoptysis and fatigue, with no obvious specificity; imaging manifestations showed various forms. Morphological lesions were mixed; sputum or other specimens were positive for acid-fast bacilli but negative by Xpert. Finally, 137 cases of *NTM* lung disease were diagnosed by BALF *NTM* gene chip identification and/or MALDI-TOF MS testing. When the results is positive, the *NTM* isolation rate is on the rise globally, and the distribution of strains varies greatly in different countries and regions.<sup>13,14</sup> A recent meta-analysis of domestic *NTM* epidemiological studies showed that the *NTM* isolation rate was higher in the south than in the north and higher in the coastal areas than inland. The bacterial species differed in different provinces and cities.<sup>15</sup> In this study, 137 isolates were identified as *NTM* by gene chip, and MAC (70.07%) was the most common isolate, including *M. intracellulare*, *M. avium*, *M. marseillense* and *M. colombiense*, followed by *M. abscessus* (21.17%); these findings are basically consistent with the prevalent strains in Hangzhou.<sup>16</sup> For patients with these clinical manifestations, who are likely to have *NTM* lung disease, molecular diagnostic techniques should be used to confirm the diagnosis as soon as possible to reduce misdiagnosis and mistreatment.

This study showed that there was no coincidental consistency in the positive detection rate of *Mycobacterium* between MALDI-TOF MS detection and the BACTEC MGIT 960 liquid culture method ( $P < 0.001$ ). The sensitivity and specificity were 97.37% (148/152) and 100% (20/20), respectively, the positive predictive value and the negative predictive value were 100% (148/148) and 83.33% (20/24), respectively, and the kappa value was 0.896. It shows that the method has a good consistency with the detection results of the culture method and has a high detection efficiency, which is suitable for the primary screening of clinical specimens for *Mycobacterium* infection. There was no coincidental consistency in the positive detection rate of *NTM* strains detected by MALDI-TOF MS and the gene chip method ( $P < 0.001$ ). The sensitivity and specificity were 95.67% (131/137) and 100% (35/35), respectively, and the positive and negative predictive values were 100% (131/131) and 85.37% (35/41), respectively. The value is 0.899. It shows that MALDI-TOF MS detection and the gene chip method have high consistency in identifying *NTM* strains and have good application value in clinical practice, consistent with previous studies.<sup>17</sup> In addition, four specimens in this study were positive for mycobacterial culture but negative for MALDI-TOF MS. The sequencing results showed that three were *MTBC* and one was *M. surga*. The reason for a negative MALDI-TOF MS test is that the specimen may contain no bacteria, or the bacterial content is below or close to the minimum detection limit of the reaction system, resulting in a negative result.

Global medical societies have jointly issued guidelines for the treatment of *NTM*, recommending that MAs and Am be used in patients with MAC lung disease and *M. abscessus* lung disease. Morimoto et al<sup>18</sup> confirmed that MA-sensitive MAC lung disease is a predictor of treatment success. Drug susceptibility testing should be performed at the time of diagnosis to determine whether MA resistance can benefit patients in treatment. This study shows that the MALDI-TOF MS detection of MA and Am drug resistance is 97.71% and 93.13%, consistent with the drug susceptibility test results. The sensitivity and specificity are 81.25%, 100% and 93.52%, 90.91%, respectively. The positive predictive value and negative predictive value were 100%, 93.75% and 98.06%, 75.00%, respectively. Comparing the two detection methods, the kappa value was above 0.61, indicating that the two methods had high consistency in detecting drug resistance of MAs and Am. Due to the obvious correlation between MAs and Am, in vitro activity, and in vivo treatment results, the drug sensitivity test can be used as an important reference for the selection of treatment options.<sup>19,20</sup>

**Limitations:** We detected a minimal amount of types and numbers of *NTM* strains, while more than 170 different *NTMs* exist. The *NTMs* clinically isolated in this study are affected by regional differences, and the statistical data may be biased. In addition, this study only included clinical *NTM* isolates from the tuberculosis laboratory of our hospital, and the types of strains cannot cover all clinical *NTM* clinical isolates. Large-scale and multi-centre collaborative research is needed to enrich the evaluation of MALDI-TOF MS's ability to identify *NTM* strains not covered in this paper.

## Conclusion

In conclusion, MALDI-TOF MS detection is fast, sensitive, accurate and economical. It has high resolution, low requirements on the original sample and can provide accurate identification of *NTM*. It is highly consistent with the identification results of BACTEC MGIT 960 liquid culture and the gene chip method and can become a fast and effective way to screen for *NTM*.<sup>21,22</sup> Moreover, it can provide drug resistance information to clinicians in a short time and provide an accurate, timely and effective laboratory test basis for clinical practice. It enables patients to receive timely and effective treatment, saves medical costs, improves patient prognosis and has good clinical application value for diagnosing and treating *NTM* lung disease.

## Data Sharing Statement

All data generated or analyzed during this study are included in this published article.

## Ethics Approval and Consent to Participate

This study was conducted in accordance with the declaration of Helsinki. This study was conducted with approval from the Ethics Committee of Wenzhou Central Hospital (No.: L2021-03-083). Written informed consent was obtained from all participants.

## Consent for Publication

The manuscript is not submitted for publication or consideration elsewhere.

## Acknowledgments

No funding or sponsorship was received for this study or publication of this article.

## Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

## Funding

No funding or sponsorship was received for this study or publication of this article.

## Disclosure

The authors declare that they have no competing interests in this work.

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