

The Emerging Roles of Ferroptosis in Neonatal Diseases

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Abstract: Ferroptosis is a novel type of programmed cell death involved in many diseases' pathological processes. Ferroptosis is characterized by lipid peroxidation, reactive oxygen species accumulation, and iron metabolism disorder. Newborns are susceptible to ferroptosis due to their special physiological state, which is prone to abnormal iron metabolism and the accumulation of reactive oxygen species. Recent studies have linked ferroptosis to a variety of diseases in the neonatal period (including hypoxic-ischemic encephalopathy, bronchopulmonary dysplasia, and necrotizing enterocolitis). Ferroptosis may become an effective target for the treatment of neonatal-related diseases. In this review, the ferroptosis molecular mechanism, metabolism characteristics of iron and reactive oxygen species in infants, the relationship between ferroptosis and common infant disorders, and the treatment of infant diseases targeted for ferroptosis are systematically summarized.

Keywords: ferroptosis, neonatal disease, reactive oxygen species, iron metabolism, lipid peroxidation

Introduction

The concept of ferroptosis was first proposed by Dr. Brent R Stockwell in 2012, which is a programmed cell death mode caused by iron-dependent lipid peroxidation and reactive oxygen species accumulation.¹ Unlike other forms of cell death discovered so far, it has distinct characteristics regarding cytogenetics, biochemistry, metabolism, and morphology. A wave-like spreading pattern can be seen in its spread through cellular populations.² During ferroptosis, lipid peroxides (eg, malondialdehyde (MDA) and 4-hydroxynonoleic acid (4-HNE)) increase, ferroptosis-related gene changes (eg, SLC7A11, ACSL4, TFR1 up-regulated, RGS4 down-regulated) and specific changes in cell morphology (decrease in mitochondrial volume, increase in mitochondrial membrane density, decrease or disappearance of mitochondrial cristae, rupture of mitochondrial outer membrane, despite normal nucleus size, without nuclear condensation or chromatin marginalization), which is reversed by ferroptosis inhibitors like ferrostatin-1.³ In various neonatal diseases, ferroptosis plays a significant role in the pathological process and may serve as a potential therapeutic target.⁴⁻⁷ In this article, we will review ferroptosis' molecular mechanisms and its potential role in neonatal diseases. The relationship between ferroptosis and neonatal hypoxic-ischemic encephalopathy, neonatal bronchopulmonary dysplasia, and neonatal necrotizing enterocolitis was emphatically introduced.

Molecular Mechanism of Ferroptosis

Ferroptosis is a form of programmed cell death caused by iron-dependent lipid peroxidation. Lipid peroxidation, reactive oxygen species (ROS) accumulation, and iron metabolism imbalance are the basic and central link in the molecular mechanism of ferroptosis³. (Figure 1, which modified from Zhang, X.2023)⁸.

Iron Metabolism and Ferroptosis

Iron is an essential trace element in the human body, playing a crucial role in hematopoiesis, ATP production, and DNA synthesis.⁹ Maintaining iron homeostasis is vital, as disruptions in this balance are closely associated with ferroptosis.¹⁰

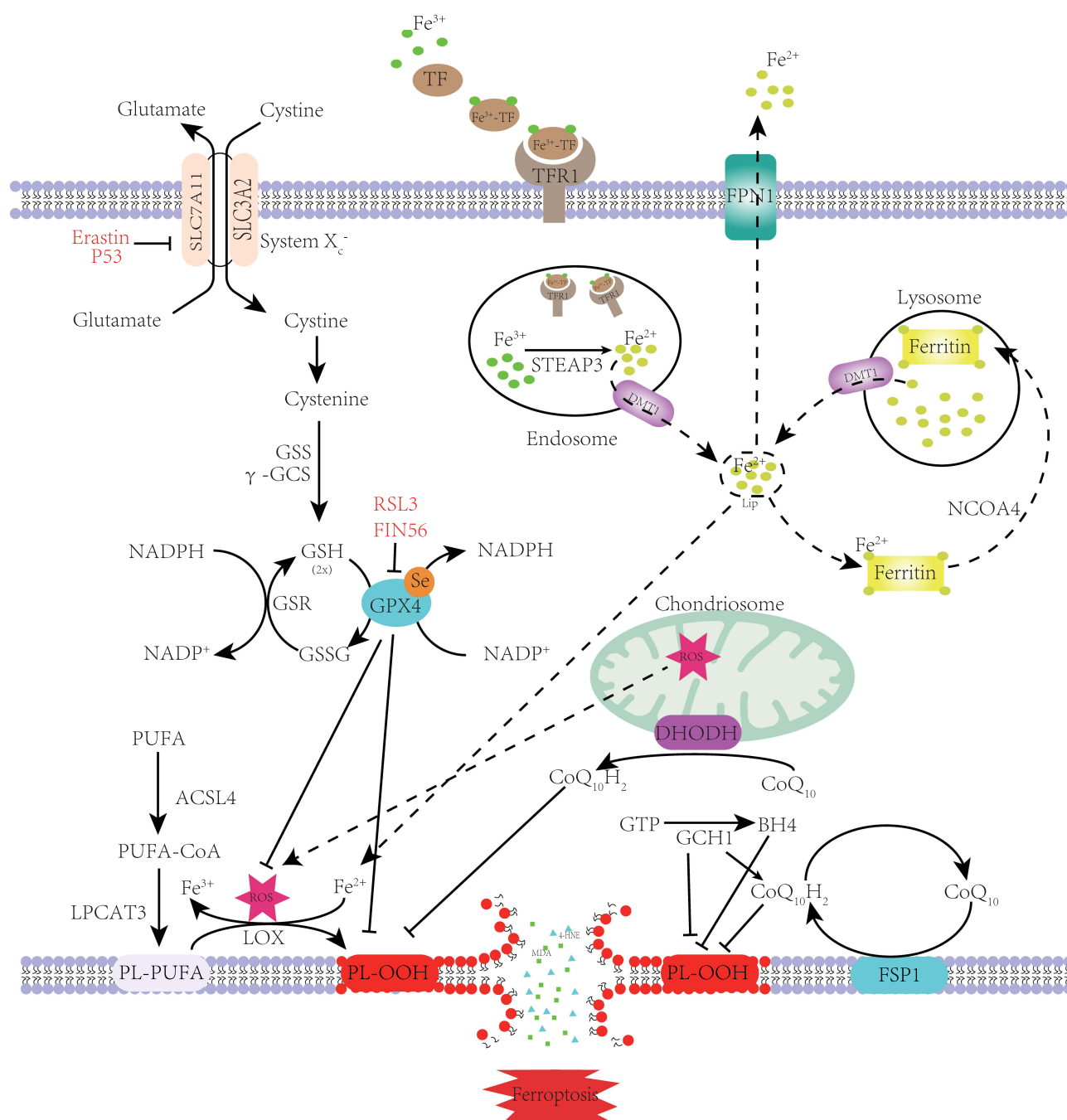


Figure 1 Mechanism of ferroptosis. PUFA became a part of the cell membrane under the action of ACSL4 and LPCAT3. Through the Fenton reaction and enzymatic reaction, PL-PUFA on the cell membrane undergoes a lipid peroxidation reaction to form PL-OOH. Due to the consumption of PL-PUFA and the toxic effect of peroxidation products, the cell membrane ruptures, eventually causing ferroptosis. Cystine enters the cytoplasm through System X_c⁻ and rapidly transforms into cysteine to participate in GSH synthesis. GPX4 inhibits lipid peroxidation while reducing GSH to GSSG. Lipid peroxidation is also inhibited by FSP1 and DHODH on the cell membrane, which reduce CoQ10 to CoQ10H₂. GCH1 can inhibit lipid peroxidation directly or indirectly. When the TF binds to the TFR on the cell membrane, it is endocytosed into the cell. Under the action of STEAP3 and DMT1, Fe²⁺ enters the cytoplasm and forms labile iron pools. Most Fe²⁺ is kept in ferritin. When intracellular iron is deficient, ferritin is transported to the lysosomes by NCOA4 to release Fe²⁺ through iron autophagy. Excess intracellular iron is excreted out of the cell through FPN1. When intracellular iron metabolism is imbalanced, the Fenton reaction and increased iron-containing enzyme activity cause ferroptosis.

Note: Adapted from Zhang X, Ma Y, Lv G, Wang H. Ferroptosis as a therapeutic target for inflammation-related intestinal diseases. *Front Pharmacol*. 2023;14:1095366.⁸

Excessive iron leads to lipid peroxidation and ferroptosis through the Fenton reaction or increased activity of iron-containing enzymes.

In newborns, breast milk and formula are the sole sources of iron.¹¹ Lactoferrin (LF) in breast milk binds to the lactoferrin receptor (LFR) and is absorbed by intestinal epithelial cells through clathrin-mediated endocytosis.^{12–14} Iron is

stored by ferritin (FTN) in the cytoplasm or exported by ferroportin-1 (FPN1).¹⁵ Transferrin (TF) transports circulating iron in complex form to various tissues. Moreover, in addition to iron absorption from the digestive tract, macrophages can phagocytize aging red blood cells and degrade Fe²⁺ in hemoglobin through heme oxygenase-1 (HO-1).¹⁶ Overexpression of HO-1, leading to increased iron release, can promote ferroptosis.¹⁷

Hepcidin regulates the balance of circulating iron. It is produced by the liver in response to high iron levels in the body. In enterocytes and macrophages, hepcidin binds to FPN1, leading to its degradation.¹⁸ This negative regulation of iron levels reduces the uptake of dietary iron and the release of iron from macrophages. Enterocytes, with a short lifespan of 3–4 days, shed the iron that is not transported into circulation.¹⁹ The synthesis of hepcidin is influenced by factors such as erythropoietin, inflammation, and iron levels in the body. High iron levels or inflammation increase the release of hepatic hepcidin while decreasing FPN1 expression. Conversely, anemia, hypoxia, and low iron levels result in decreased hepcidin expression, increasing FPN1 activity and iron mobilization.²⁰ Increased hepcidin expression can alleviate ferroptosis in hypoxia-reperfused rat cardiomyocytes.²¹

Extracellular transferrin binds to the transferrin receptor (TFR) on the cell membrane, undergoes endocytosis, and forms endosomes.²² Within the endosomes, a protein called six-transmembrane epithelial antigen of the prostate 3 (STEAP3) reduces Fe³⁺ to Fe²⁺, which is then transported to the cytoplasm by the divalent metal ion transporter 1 (DMT1).²³ The TFRs are recycled back to the cell membrane. Intracellular Fe²⁺ is utilized in enzymatic reactions and heme synthesis. Cellular iron homeostasis is maintained by either pumping excess iron out of the cell membrane through FPN1 or storing it in ferritin. When intracellular Fe²⁺ levels decrease, ferritin is transported to lysosomes via nuclear receptor coactivator 4 (NCOA4). In the lysosomes, ferritin undergoes degradation through iron autophagy, releasing iron ions that are exported to the cytoplasm via DMT1 on the lysosome.²⁴ This process helps maintain intracellular iron balance. TF and TFR promote ferroptosis by importing iron into cells.²⁵ Conversely, iron export mechanisms can render cells resistant to ferroptosis.²⁶ Iron autophagy also regulates ferroptosis.^{27,28} For example, specific knockout of NCOA4 can inhibit ferroptosis induced by Erastin, controlled by autophagy-related genes ATG5 and ATG7.²⁹

Iron regulatory proteins 1 and 2 (IRP1 and 2) are crucial for regulating intracellular iron levels.⁹ They interact with iron-responsive elements (IREs) in genes related to iron metabolism, controlling processes such as absorption, storage, utilization, and output of iron.^{30,31} When intracellular iron is deficient, IRP1 is detached from the iron-sulfur cluster and binds to IRE located at the 3′ “end of the gene, increasing gene transcription of DMT1 and TFR and promoting iron uptake. In addition, IRP1 can bind to the ferritin gene’s IRE at its 5′ end and negatively regulate FTN, thereby reducing iron storage. Upon intracellular iron excess, IRP1 rebinds to iron-sulfur clusters, resulting in increased FTN and decreased TFR and DMT1, thereby reducing iron levels. Similar to IRP1 in structure, IRP2 is degraded by ubiquitin upon iron accumulation and modulates gene expression to regulate iron metabolism.¹⁶ During ferroptosis, IRP plays a key role. The expression of IRP1 and IRP2 is increased during ferroptosis of melanoma cells, and inhibition of IRP1 can significantly inhibit ferroptosis.³² Overexpression of IRP2 in hepatic stellate cells reduces liver fibrosis through ferroptosis.³³

Lipid Peroxidation and Ferroptosis

Lipid peroxidation is a key step in ferroptosis.³⁴ It occurs when lipids have one or more carbon-carbon double bonds. Active hydrogen atoms are located between the methylene bridges (-CH₂-) on both sides of the double bond. Free radicals acquire hydrogen atoms from cell membranes through these bridges, leading to cell damage through free radical chain reactions.³⁵ Lipid oxidation results in the accumulation of lipid peroxidation products in cells, altering cell membrane fluidity and permeability, ultimately causing structural and functional damage.³⁶ Phospholipids containing polyunsaturated fatty acids (PUFAs) are particularly susceptible to lipid peroxidation and ferroptosis due to their multiple double bonds. Deuterium isotope of hydrogen, found at the bi-allylic position of PUFAs, inhibits PUFA peroxidation and blocks ferroptosis.³⁷ Increasing the synthesis of monounsaturated fatty acids (MUFAs) or adding palmitoleic acid or oleic acid can also inhibit ferroptosis.³⁸ MUFAs may displace PUFAs in cell membranes, inhibiting lipid peroxidation.³⁹ However, not all PUFAs cause lipid peroxidation.³ ROS attack phosphatidylethanolamines with specific fatty acid groups, making them major targets for lipid peroxidation.⁸

The lipid peroxidation process that causes ferroptosis can be divided into two steps: phospholipid synthesis with PUFAs as substrates and peroxidation reaction. First, Lysophosphatidylcholine acyltransferase 3 (LPCAT3) integrates PUFA-CoA into cellular membranes after synthesis by acyl-CoA synthetase 4 (ACSL4).⁴⁰ Phospholipids-polyunsaturated fatty acids (PL-PUFA) will participate in subsequent peroxidation reactions as peroxidation substrates. In the peroxidation step, either a Fenton reaction or enzymatic reaction induces lipid peroxidation.⁴¹ The Fenton reaction converts Fe²⁺ to Fe³⁺, generating ROS that attack PL-PUFA and cause lipid peroxidation.⁴² In the lipoxygenase-promoted lipid peroxidation pathway,⁴³ PL-PUFA is oxidized by 15-lipoxygenase (15-LOX), which binds to phosphatidylethanolamine-binding protein 1 (PEP1). PUFA-PE-OOH is then produced, resulting in ferroptosis in the cells.^{44,45} In the endoplasmic reticulum, NADH-cytochrome B5 reductase (CB5R) converts electrons from nicotinamide adenine dinucleotide phosphate (NADPH) into superoxide (O₂⁻) which is then used to initiate lipid peroxidation directly or indirectly by acquiring methylene hydrogen from PL-PUFA or by reducing ferric iron.⁴⁶ Since the cells are in an oxidative imbalance, Lipid peroxidation will continue until PL-PUFA is exhausted. As PL-PUFA is an integral part of cell structure, its excessive consumption will eventually result in cell ruptures and perforations. Furthermore, major lipid peroxidation products (PL-OOH, MDA, 4-HNE, etc.) produced during lipid peroxidation do further damage to the cell membrane, resulting in membrane pores and ferroptosis.⁴⁷

ROS and Ferroptosis

Lipid Peroxidation Induced by ROS

ROS, including superoxides (O₂⁻), peroxides (H₂O₂), and free radicals (HO⁻ and RO⁻), are products of O₂ gaining electrons.⁴⁸ These reactive oxygen species play a central role in promoting lipid peroxidation. Increased accumulation of ROS in cells makes them more vulnerable to ferroptosis.

Regulatory Mechanism of ROS

The body clears lipid peroxides caused by ROS through the GSH/GPX4 system and GPX4-independent regulatory mechanisms. Once these regulatory mechanisms are abnormal, intracellular ROS accumulation and lipid peroxidation will occur, leading to ferroptosis.

GSH/GPX4 Inhibits ROS

GPX4, a selenium protein, is the primary enzyme in mammalian cells responsible for reducing PL-OOH, a lipid peroxide.⁴⁹ It catalyzes the reduced glutathione (GSH) to the conversion of oxidized glutathione (GSSG) and reduces cytotoxic PL-OOH to non-toxic phospholipid alcohol (PL-OH),⁵⁰ thus inhibiting ferroptosis. However, compounds like RSL3 and FIN56 can directly inhibit or degrade GPX4, promoting ferroptosis.³

Glutathione (GSH), a tripeptide, acts as an intracellular antioxidant and inhibits ferroptosis. It is synthesized from glutamic acid, glycine, and cysteine, with cystine being crucial for GSH biosynthesis.⁵¹ Cystine enters cells through the cystine/glutamate antiporter (system Xc⁻), facilitated by the proteins SLC7A11 and SLC3A2L. Inside the cell, cystine is rapidly reduced and then converted into GSH through enzymatic reactions.⁵² GSH, under the influence of GPX4, helps to neutralize ROS, reducing lipid peroxidation and preventing ferroptosis.⁵³ Inhibitors like erastin and P53 target SLC7A11, hindering cystine uptake and suppressing GSH synthesis, ultimately leading to lipid peroxidation and ferroptosis.^{1,54} N-acetylcysteine can counteract ferroptosis by promoting GSH synthesis.⁵⁵

Regulation of Lipid ROS Independent of GPX4

In addition to GSH/GPX4, three GSH/GPX4-independent signaling pathways have been identified to regulate ferroptosis: the FSP1-CoQ10-NADPH pathway, the DHODH-CoQ10 pathway, and the GCH1-BH4 pathway.³

By using NADPH, ferroptosis suppressor protein 1 (FSP1), which is translocated to the cell membrane, converts ubiquinone (also known as CoQ10) into ubiquinol (CoQ10H₂). Antioxidant ubiquinol traps lipid-ROS to form ubiquinone, inhibiting lipid peroxidation in this process and attenuating ferroptosis.^{56,57} Researchers have found that inhibiting FSP1 can induce ferroptosis in tumor cells.³⁴

A flavin-dependent enzyme, dihydroorotate dehydrogenase (DHODH) is present in the mitochondrial membrane. DHODH can oxidize dihydro whey to whey acid and reduce CoQ10 to CoQ10H2 in the mitochondrial membrane, thus inhibiting lipid peroxidation and ferroptosis.⁵⁸ Through mTOR signaling, DHODH promotes ferroptosis in cervical cancer cells.⁵⁹

Tetrahydrobiopterin (BH4), synthesized from GTP catalyzed by GTP cyclohydrolase 1 (GCH1), is a lipophilic antioxidant with a function like CoQ10H2 to prevent lipid peroxidation.⁶⁰ In addition to catalyzing the synthesis of BH4, GCH1 can also lead to the remodeling of the lipid membrane environment, increase CoQ10H2, and consume PL-PUFA.³ Thus, by overexpressing GCH1 or supplementing with BH4, ferroptosis can be reduced.⁶¹

Neonates are Susceptible to Ferroptosis

Newborns' susceptibility to ferroptosis is due to imbalanced iron metabolism and increased ROS levels. Their delicate physiology and unique metabolism make them prone to this process. As a result, various neonatal diseases are closely linked to ferroptosis. (Figure 2, which modified from Du, X. 2022).⁶²

Newborns are Prone to Increase Free Iron

Newborns are prone to the accumulation of non-transferrin-bound iron (NTBI) due to factors such as perinatal acidosis, recurrent hypoxic ischemia, and immature antioxidant system.⁶³ Red blood cell transfusions in newborns also contribute to NTBI accumulation.⁶⁴ In addition, newborns are at risk for NTBI through red blood cell transfusions.⁶⁵ The increased extracellular NTBI can elevate intracellular iron levels,^{66,67} leading to oxidative stress and lipid peroxidation, ultimately resulting in ferroptosis.

Newborns are Prone to ROS Accumulation

The accumulation of ROS in newborns is due to an imbalance between oxidative stress and antioxidants. High oxidative stress during pregnancy, delivery, and the postnatal period exposes neonates to increased ROS production. Maternal conditions like preeclampsia, chronic hypertension, obesity, and infections contribute to elevated oxidative stress.⁶⁸ Maternal elevated oxidative stress also impact newborn ROS levels.⁶⁹ Neonates experience a transition from intrauterine hypoxia to postnatal hyperoxia, leading to ROS generation.⁷⁰ The body degrades ROS through enzymatic (SOD, CAT, GPX) and non-enzymatic antioxidants (glutathione, tocopherol, etc.).⁶⁸ Newborns, especially preterm infants, have immature antioxidant systems and imbalanced oxidation.⁷¹ Neonatal diseases like BPD, ROP, NEC, and HIE are

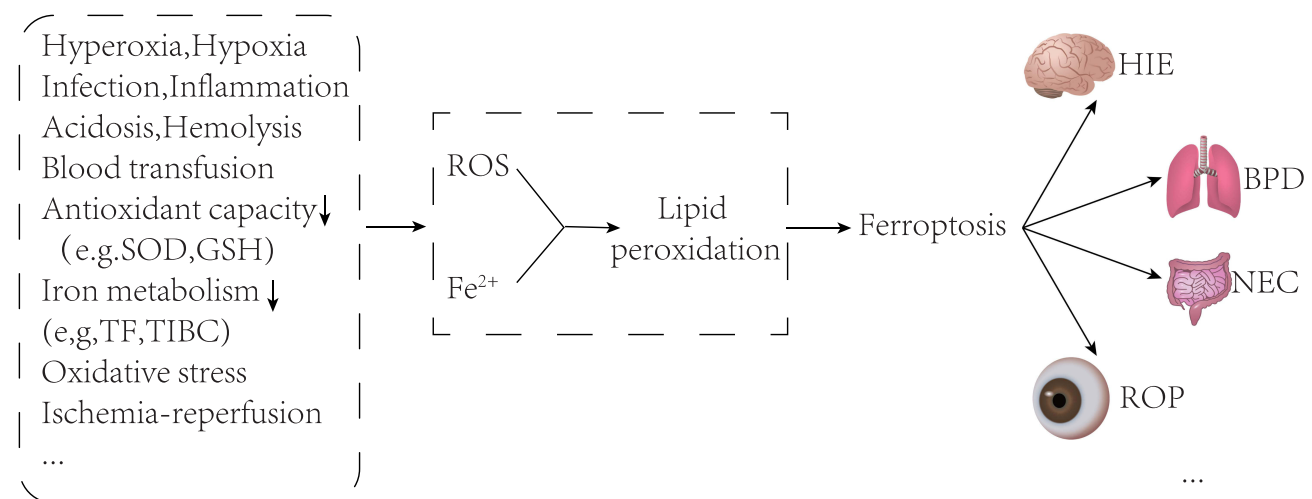


Figure 2 Neonatal disease and ferroptosis. Newborns are easily exposed to oxidative stress environments such as hypoxia, hyperoxia, infection, inflammation, acidosis, and blood transfusion after birth, and their antioxidant capacity and iron metabolism capacity are poor, which will lead to increased ROS and free iron, and then cause ferroptosis through lipid peroxidation. A variety of neonatal diseases may be caused by ferroptosis, such as hypoxic-ischemic encephalopathy, bronchopulmonary dysplasia, necrotizing enterocolitis, and retinopathy of prematurity.

Note: Adapted from Du X, Dong R, Wu Y, Ni B. Physiological effects of ferroptosis on organ fibrosis. *Oxid Med Cell Longev*. 2022;2022:5295434.⁶²

associated with increased ROS levels.⁷² Excess ROS triggers ferroptosis via lipid peroxidation.⁷³ Inhibiting the ferroptosis signaling pathway can attenuate tissue damage caused by these diseases.^{7,74–76}

The Role of Ferroptosis in Common Neonatal Diseases

The Role of Ferroptosis in Neonatal HIE

Neonatal hypoxic-ischemic encephalopathy (HIE) is a severe neurological injury caused by hypoxia-ischemia of brain tissue.⁷⁷ Despite mild hypothermia, half of infants with moderate to severe HIE still die or suffer neurological sequelae.⁷⁸

Oxidative stress is a key factor in neonatal hypoxic-ischemic encephalopathy (HIE) due to decreased activity of antioxidant enzymes in the immature brain.⁷⁹ Hypoxia-ischemia and hyperoxia in newborns lead to the production of excessive ROS in the brain, causing lipid peroxidation. Infants with perinatal asphyxia show significantly elevated levels of malondialdehyde (MDA) in cord blood and blood, indicating increased oxidative stress.⁸⁰ Elevated serum lipid peroxides in neonates can serve as a sensitive diagnostic marker for HIE.⁸¹ High concentrations of MDA in the cerebrospinal fluid of neonates with HIE suggest lipid peroxidation in nerve cells.⁸² Mild hypothermia is the main therapeutic approach for HIE and works by inhibiting ROS production in the nervous system.⁸³ Hypothermia treatment has been shown to reduce MDA levels compared to normothermic controls.⁸⁴

Due to the Fenton reaction, non-transferrin-bound iron (NTBI) generates free radicals, leading to lipid peroxidation and triggering ferroptosis. NTBI has been detected in significant amounts in the cerebrospinal fluid and serum of neonates after hypoxic-ischemic encephalopathy (HIE).^{82,85} The concentration of NTBI is directly related to the severity of brain injury and can predict long-term neurodevelopmental outcomes in infants.⁸⁵ During cerebral ischemia-reperfusion, increased levels of ROS cause the release of iron from binding proteins and red blood cells, exacerbating brain damage.^{20,86} Iron-induced free radicals disrupt the blood-brain barrier and contribute to endothelial necrosis.⁸⁷ Intracellular iron accumulation in microglia following hypoxia contributes to the death of oligodendrocytes and axonal swelling.⁸⁸ Iron released from heme disruption in hypoxic-ischemia-induced intracerebral hemorrhage leads to lipid peroxidation and ferroptosis.⁸⁹ Treatment options such as deferoxamine, an iron chelator, and erythropoietin, which promotes iron sequestration, have shown beneficial effects in reducing brain injury after HIE.^{90,91} Heat shock protein B (HSPB1) plays a role in reducing cerebral infarction and brain ferritin levels in HIE by regulating iron metabolism, promoting antioxidant enzyme expression, and alleviating ferroptosis.⁹²

The expressions of SLC7A11, GSH, and GPX4 were decreased in HIE neonatal rat brain tissues, indicating the activation of ferroptosis-related signaling pathways.⁹³ Inhibition of ferroptosis through intraventricular injection of a ferroptosis inhibitor increased GPx4 expression and reduced MDA and iron levels in the hippocampi of HIE rats. The SIRT1/NRF2/GPx4 signaling pathway plays a crucial role in this process.⁹⁴ TLR4 also contributes to ferroptosis in HIE, and the use of a TLR4 inhibitor called TAK-242 elevated SLC7A11 and GPX4 levels in hippocampal tissue, both in vivo and in vitro. It also reduced the expression of genes associated with ferroptosis.⁹⁵ Lipocalin 2 (LCN2) was identified as a critical gene associated with HIE, promoting ferroptosis through the NF- κ B/STAT3 pathway. LCN2 can serve as a biomarker for the diagnosis of HIE.⁹⁶

The Role of Ferroptosis in Neonatal BPD

Bronchopulmonary dysplasia (BPD) is a common lung disorder in premature infants. Oxidative stress plays a significant role in its development.^{68,97} Premature infants have immature type II alveolar cells, leading to the accumulation of intracellular ROS.⁹⁸ Excessive ROS damages the lung epithelium and vascular endothelium, impairing lung development and contributing to BPD.⁹⁹ The pathogenesis of BPD involves lipid peroxidation, supporting the association with ferroptosis. Lipid hydrogen peroxide (LOOH) is generated during lipid peroxidation, causing damage to biological macromolecules. Infants with BPD exhibit higher levels of LOOH in their alveolar lavage fluid, likely due to ROS overload.¹⁰⁰

Hyperoxia is a risk factor for bronchopulmonary dysplasia (BPD) and is commonly used to create an animal model of BPD.¹⁰¹ Ferroptosis has been observed in neonatal rats experiencing hyperoxia-induced lung injury, indicating its involvement in lung development impairment.¹⁰² The increased transcription and activity of P53, along with enhanced

acetylated P53 expression, are observed as a result of hyperoxia exposure.¹⁰³ P53 inhibits SLC7A11, leading to decreased glutathione (GSH) synthesis and GPX4 activity, resulting in ROS accumulation and triggering ferroptosis.¹⁰⁴

Excessive iron supplementation in infants with very low birth weight increases the risk of developing BPD.¹⁰⁵ Experiments on a mouse model of lung injury demonstrated the accumulation of iron ions in the lung, which can induce ferroptosis.¹⁰⁶ Inflammation also plays a role in BPD, and the activation of the KEAP1-NRF2/HO-1 pathway by panaxydol inhibits ferroptosis, thereby reducing LPS-induced pneumonitis.^{107,108}

In summary, hyperoxia, oxidative stress, inflammation, and excessive iron are key factors contributing to the development of BPD, and their association with ferroptosis suggests a significant role of ferroptosis in BPD pathology. However, more research is required to elucidate the specific regulatory mechanisms underlying ferroptosis in BPD.

The Role of Ferroptosis in Neonatal NEC

Neonatal necrotizing enterocolitis (NEC) is a severe gastrointestinal disorder. Factors such as oxidative stress, ischemia-reperfusion, and inflammation play a role in its development. Ferroptosis is closely associated with the pathogenesis of NEC and may contribute to its progression.

First, NEC is characterized by intestinal ischemia.¹⁰⁹ Ischemia-reperfusion generates ROS and free iron, worsening intestinal injury.¹¹⁰ Elevated free iron levels enhance the Fenton response, leading to the production of highly toxic free radicals.¹¹¹ Free iron and ROS cause ferroptosis through lipid peroxidation, contributing to tissue damage in NEC.

Second, NEC development is linked to lipid peroxidation damage. NEC-affected infants had higher total oxidative state and oxidative stress index, which correlated with NEC severity.¹¹² NEC infants' cord blood had higher levels of NTBI and ROS compared to non-NEC infants.¹¹³ High levels of NTBI and ROS lead to tissue damage through lipid peroxidation. Lipid peroxidation products in cord blood strongly correlate with NEC.¹¹³ Preterm infants with NEC had increased intestinal MDA.^{114,115} Animals with NEC exhibited decreased levels of GPX, SOD, and GSH in intestinal tissues, along with significantly elevated MDA levels.¹¹⁶ Recent research discovered that *Lactobacillus* can alleviate NEC by reducing lipid peroxidation in enterocytes.¹¹⁷

Third, Ferroptosis is linked to TLR4, which is important in NEC. The TLR4 signaling pathway plays a role in embryonic crypt development and intestinal epithelial cell differentiation.¹¹⁸ Persistent TLR4 pathway hyperactivation in intestinal epithelial cells after birth is associated with gut damage in NEC. Inhibiting TLR4 signaling effectively reduces the occurrence and severity of NEC.¹¹⁹ TLR4 is also closely related to ferroptosis. Inhibition of TLR4 attenuates ferroptosis in hippocampal neurons during hypoxic-ischemic injury.⁹⁵ TLR4 promotes ferroptosis in heart failure cardiomyocytes.¹²⁰ During renal ischemia-reperfusion injury, TLR4/NOX4 induce oxidative stress and ferroptosis.¹²¹

Fourth, excessive inflammation is a major pathological change in NEC. In NEC intestinal tissue, immune cells release inflammatory factors like IL-6 and TNF- α . IL-6 promotes lipid peroxidation and ferroptosis in epithelial cells by increasing ferrous ions.¹²² TNF- α promotes ferroptosis by increasing the production of acyl-CoA synthetase long-chain family member 1 (ACSL1), which enhances the production of polyunsaturated lipids and subsequent lipid peroxidation in human endothelial cells.¹²³ IFN- γ downregulates SLC3A2 and SLC7A11, causing lipid peroxidation and ferroptosis.¹²⁴ Ferroptosis of intestinal epithelial cells further triggers an inflammatory response by releasing cellular contents. For instance, the lipid peroxidation product 4-HNE activates the NF- κ B pathway, promoting inflammation.¹²⁵ Release of HMGB1, a protein found in ferroptosis cells, also contributes to NEC-related inflammation by activating the TLR4-mediated NF- κ B signaling pathway.^{126,127} The evidence suggests a close association between inflammation and ferroptosis in NEC.⁸

In addition, In an NEC model created by anemia-blood transfusion, elevated levels of free iron and MDA were observed in the intestines, along with mitochondrial dysfunction. Ferroptosis inhibitors were found to alleviate intestinal injury in NEC.⁶ The analysis of genetic data in premature infants with NEC revealed that specific genes related to NEC were significantly enriched in the ferroptosis signaling pathway.¹²⁸ While there is evidence linking NEC to ferroptosis, the precise mechanisms by which ferroptosis contributes to NEC and how it can be regulated to impact NEC are not fully understood.

The Role of Ferroptosis in Other Common Neonatal Diseases

Premature infants are susceptible to retinal diseases such as retinopathy of prematurity (ROP), which can result in blindness. The mouse model of oxygen-induced retinopathy showed lipid peroxidation and ferroptosis markers in the mouse retina were found to increase significantly, suggesting that ferroptosis is critical in the pathological process of ROP, and ferroptosis inhibitors can alleviate the process of ROP middle retinal vascular occlusion and pathological neovascularization.^{5,129} Intracranial hemorrhage is a common complication in neonates, especially premature infants. In the intracranial hemorrhage model of neonatal rats, ferroptosis-related genes were detected higher 72 hours after hemorrhage, implicating ferroptosis in hemorrhage-induced brain injury pathology.¹³⁰ Lipocalin 2 (LCN2) inhibition attenuated inflammation and ferroptosis in neonate acute respiratory distress syndrome (ARDS) through MAPK/ERK signaling.¹³¹ In addition, ferroptosis may play a critical role in the neuronal damage caused by hemolytic hyperbilirubinemia in the brain.¹³²

New Therapeutic Targets for Neonatal Diseases Based on Ferroptosis

In neonatal diseases, ferroptosis plays an important role, so targeting ferroptosis may be a promising treatment strategy (Table 1). For example, Vitamin D is an important antioxidant, which can increase the production of NRF2 and HO-1, lead to the increase of GPX4, SOD and GSH, reduce MDA and ROS, and reduce ferroptosis in brain injury.¹³³

Table 1 Targeted Drugs for Ferroptosis and Their Mechanisms in Common Diseases of Newborns

Study	Disease	Mechanism	Dilemmas	Potential Solutions for Therapy
Cai, Y. 2022 ¹³³	HIE	Increase NRF2 and HO-1 production, decrease ferroptosis	Rat model; Uncertainty regarding efficacy in the human body	Vitamin D
Zhu, K. 2022 ¹³⁴	HIE	Inhibit Ferroptosis and Neuroinflammation via the HMGB1/GPX4 Pathway	Rat model; Uncertainty regarding efficacy in the human body	Glycyrrhizin
Gou, Z. 2020 ¹³⁵	HIE	Inhibit ferroptosis by activating the Akt/Nrf2/Gpx4 pathway	Rat model; Uncertainty regarding efficacy in the human body	Melatonin
Li, C. 2022 ⁹⁴	HIE	Inhibit ferroptosis via activating the SIRT1/Nrf2/GPx4 pathway	Rat model; Uncertainty regarding efficacy in the human body	Intraventricular injection Ferrostatin-I or Resveratrol
Chou, H. 2022 ¹³⁶	BPD	Attenuates Hyperoxia-Induced Lung Injury by Inhibiting Ferroptosis	The specific mechanism of action is unclear	Cathelicidins
Lelli, J. 1993 ¹³⁷	NEC	Chelating free iron reduces ischemia-reperfusion injury in NEC	Rat model; Uncertainty regarding efficacy in the human body	Deferoxamine
Feng, YD. 2022 ¹³⁸	NEC	Inhibition of HO-1 and MAO-B activity suppresses ferroptosis	Rat model; Uncertainty regarding efficacy in the human body	Apigenin-7-O- β -D-(6"-p-coumaroyl)-glucopyranoside
Ozdemir, R. 2013 ¹¹⁴	NEC	Anti-inflammatory and antioxidant	The specific mechanism of action is unclear	All-trans retinoic acid
Yazici, S. 2014 ¹³⁹	NEC	Inhibit peroxidation	The specific mechanism of action is unclear	Boric acid and 2-aminoethoxydiphenyl borate
Liu, CQ. 2023 ⁵	ROP	Inhibition of ferroptosis and pathological angiogenesis.	The specific mechanism of action is unclear; mouse model	Intravitreal injection Ferrostatin-I
Wang J 2022 ¹²⁹	ROP	Promoting angiogenesis by inhibiting ferroptosis through the xCT/GPX4 axis	Rat model; Uncertainty regarding efficacy in the human body	Elabela

Intraventricular injection of ferroptosis inhibitor (Ferrostatin-1 or Resveratrol) can reduce neonatal cerebral ischemia-reperfusion injury.⁹⁴ Glycyrrhizin attenuated RSL3-induced ferroptosis and inflammatory response in neonatal rat brain tissue by inhibiting HMGB1.¹³⁴ Melatonin can inhibit the process of ferroptosis in hypoxic-ischemic encephalopathy by activating GPX4 activity and playing a protective role in the brain.¹³⁵ Hyperoxia-induced lung damage can be prevented by cathelicidins, which have both antioxidant and ferroptosis inhibition properties.¹³⁶ Deferoxamine would be able to attenuate intestinal damage in neonatal necrotizing enterocolitis by binding excess free iron in neonates.¹³⁷ Flavonoid glycosides are potent antioxidants that attenuate endothelial ROS production and free iron accumulation by specifically binding to HO-1 and MAO-B, thereby inhibiting ferroptosis in the ischemia-reperfused intestine in a dose-dependent manner.¹³⁸ All-trans retinoic acid treatment decreased intestinal MDA, increased SOD and GPX activity, and attenuated NEC by inhibiting lipid peroxidation.¹¹⁴ Boric acid and 2-aminoethoxydiphenyl borate can reduce NEC by increasing GSH expression and inhibiting lipid peroxidation.¹³⁹ Further clinical studies are required to confirm the safety and effectiveness of novel medications targeting ferroptosis for the treatment of newborn illnesses.

Conclusion

Ferroptosis is a newly discovered form of programmed cell death triggered by free iron and ROS, leading to tissue damage through lipid peroxidation. Newborns, due to their unique physiological state and metabolic characteristics, are prone to accumulate free iron and ROS, resulting in organ damage via ferroptosis. Studies have shown that ferroptosis is involved in the development of neonatal diseases such as hypoxic-ischemic encephalopathy, bronchopulmonary dysplasia, and necrotizing enterocolitis, and inhibiting ferroptosis can alleviate these conditions. Consequently, ferroptosis has emerged as a potential therapeutic target for newborn disorders. However, most research conducted so far has been limited to animal and cell studies, necessitating further clinical investigations to validate these findings.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Disclosure

The authors declare no conflicts of interest in this work.

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