

# Stem Cell Therapy in Inflammatory Bowel Disease: A Review of Achievements and Challenges

Cheng-Mei Tian<sup>1,2,\*</sup>, Yuan Zhang<sup>3,\*</sup>, Mei-Feng Yang<sup>4,\*</sup>, Hao-Ming Xu<sup>5</sup>, Min-Zheng Zhu<sup>6</sup>, Jun Yao<sup>1</sup>, Li-Sheng Wang<sup>1</sup>, Yu-Jie Liang<sup>7</sup>, De-Feng Li<sup>1</sup>

<sup>1</sup>Department of Gastroenterology, Shenzhen People's Hospital (The Second Clinical Medical College, Jinan University, The First Affiliated Hospital, Southern University of Science and Technology), Shenzhen, Guangdong, People's Republic of China; <sup>2</sup>Department of Emergency, Shenzhen People's Hospital (The Second Clinical Medical College, Jinan University, The First Affiliated Hospital, Southern University of Science and Technology), Shenzhen, Guangdong, People's Republic of China; <sup>3</sup>Department of Medical Administration, Huizhou Institute of Occupational Diseases Control and Prevention, Huizhou, Guangdong, People's Republic of China; <sup>4</sup>Department of Hematology, Yantian District People's Hospital, Shenzhen, Guangdong, People's Republic of China; <sup>5</sup>Department of Gastroenterology and Hepatology, Guangzhou Digestive Disease Center, Guangzhou First People's Hospital, School of Medicine, South China University of Technology, Guangzhou, Guangdong, People's Republic of China; <sup>6</sup>Department of Gastroenterology and Hepatology, The Second Affiliated Hospital, School of Medicine, South China University of Technology, Guangzhou, Guangdong, People's Republic of China; <sup>7</sup>Department of Child and Adolescent Psychiatry, Shenzhen Kangning Hospital, Shenzhen, Guangdong, People's Republic of China

\*These authors contributed equally to this work

Correspondence: Li-Sheng Wang; De-Feng Li, Department of Gastroenterology, Shenzhen People's Hospital (The Second Clinical Medical College, Jinan University, The First Affiliated Hospital, Southern University of Science and Technology), No. 1017, Dongmen North Road, Luohu District, Shenzhen, 518020, People's Republic of China, Tel +86 755 25533018, Email wanglsszrmy@163.com; ldf830712@163.com

**Abstract:** Inflammatory bowel disease (IBD), including Crohn's disease (CD) and ulcerative colitis (UC), is a group of chronic inflammatory diseases of the gastrointestinal tract. Repeated inflammation can lead to complications, such as intestinal fistula, obstruction, perforation, and bleeding. Unfortunately, achieving durable remission and mucosal healing (MH) with current treatments is difficult. Stem cells (SCs) have the potential to modulate immunity, suppress inflammation, and have anti-apoptotic and pro-angiogenic effects, making them an ideal therapeutic strategy to target chronic inflammation and intestinal damage in IBD. In recent years, hematopoietic stem cells (HSCs) and adult mesenchymal stem cells (MSCs) have shown efficacy in treating IBD. In addition, numerous clinical trials have evaluated the efficiency of MSCs in treating the disease. This review summarizes the current research progress on the safety and efficacy of SC-based therapy for IBD in both preclinical models and clinical trials. We discuss potential mechanisms of SC therapy, including tissue repair, paracrine effects, and the promotion of angiogenesis, immune regulation, and anti-inflammatory effects. We also summarize current SC engineering strategies aimed at enhancing the immunosuppressive and regenerative capabilities of SCs for treating intestinal diseases. Additionally, we highlight current limitations and future perspectives of SC-related therapy for IBD.

**Keywords:** stem cells, mesenchymal stromal cells, HSCs, immunosuppression, inflammatory bowel disease, ulcerative colitis, Crohn's disease

## Introduction

Inflammatory Bowel Disease (IBD) has a multifaceted etiology, including genetic susceptibility, immune dysregulation, external intestinal flora, and environmental factors.<sup>1</sup> Both internal and external causes can compromise the intestinal mucosal barrier, leading to chronic, non-specific inflammation, local structural changes, and intestinal dysfunction.<sup>2</sup> However, the extent and nature of the lesions are not limited to the intestinal mucosa and inflammation.<sup>3</sup> Repeated inflammation and microcirculation disorders can cause intestinal fistulae, stenosis, obstruction, perforation, gastrointestinal (GI) bleeding, sepsis, and other complications, increasing the risk of intestinal cell cancerization and death.<sup>1,4,5</sup> Although the overall morbidity of IBD remains at 0.5% in North America and Europe, its incidence is increasing yearly in Asia, Africa, and South America, resulting in a growing number of patients.<sup>4-7</sup>

## Graphic Abstract



Drugs used to treat IBD primarily target the systemic immune system, whether through non-targeted immunosuppressive agents or biological agents targeting TNF- $\alpha$ , integrin- $\alpha$  4 beta 7, interleukin-12 (IL-12), IL-23, and other factors. Their goal is to suppress excessive immune responses and active inflammatory responses.<sup>8,9</sup> However, inhibiting the inflammatory response alone is insufficient to completely cure IBD and achieve long-term remission and mucosal healing (MH).<sup>10,11</sup>

Stem cells (SCs) can differentiate, proliferate, and regulate various tissue cells through secretory functions.<sup>12</sup> Intestinal resident SCs, mainly intestinal stem cells (ISCs) and resident mesenchymal stem cells (MSCs), maintain the structure and function of intestinal structural and immune cells, preserving intestinal mucosal integrity and immune homeostasis.<sup>13</sup> Absence or abnormal activation of these SCs may cause mucosal and immune homeostasis disorders, which are the primary pathogenesis of IBD.<sup>13</sup> IBD is characterized by the loss of control of immune cells, damage to intestinal structural cells, and abnormalities of intestinal resident SCs.<sup>14,15</sup> Transplantation of SCs can regulate or rebuild immune cells, repair or supplement structural cells, such as intestinal epithelial cells (IECs), and potentially lead to a complete cure of IBD.<sup>16,17</sup> This review summarizes recent research on the use of SCs for treating IBD, providing targeted treatment recommendations and research strategies for IBD patients.

Research has shown that multiple types of SCs continuously generate and replace IECs, immune cells, and more, thereby maintaining the integrity of the intestinal epithelium and the homeostasis of intestinal immunity. The absence or abnormal activation of intestinal resident SCs can lead to intestinal epithelial dysfunction and immune dysfunction,

which are key pathogenic factors of IBD. Transplantation of SCs into the intestinal lesions of patients may offer a potential cure for IBD by repairing intestinal epithelial dysfunction and reestablishing immune homeostasis, resulting in MH and long-term remission.

## Commonly Used SC Types for IBD Treatment

Human cells can be classified into general cells and SCs based on their ability to proliferate and differentiate.<sup>18</sup> General cells, also called terminal differentiated cells, have a limited lifespan and lack the capacity to proliferate and differentiate.<sup>19</sup> In contrast, SCs have the potential for unlimited proliferation and differentiation and are regulated by their micro-environment, or niche, which allows them to differentiate into various cell types and maintain the structural and functional integrity of the organ.<sup>14</sup> SCs are further classified into totipotent, pluripotent, and unipotent SCs based on their differentiation potential, with totipotent SCs capable of self-renewal and differentiation into any cell type, pluripotent SCs capable of differentiating into multiple cell types, and unipotent SCs able to differentiate into only one cell type. Additionally, SCs can regulate human cells, acting as cell producers and managers in the body.<sup>15,17</sup> However, SCs *in vivo* are also affected by their local niche and can undergo structural changes, damage, depletion, cancerization, and even death,<sup>20</sup> which can affect their differentiation and secretion functions. SC therapy involves transplanting exogenous SCs into patients with impaired SC structure and function, supplementing, resetting, or transforming their own SCs to repair or replace damaged cells or tissues and cure diseases.<sup>21</sup> Various types of SCs, including hematopoietic stem cells (HSCs), MSCs, ISCs, and induced pluripotent stem cells (iPSCs), have been used for the treatment of IBD.<sup>12,22</sup> Please refer to [Table 1](#) for a list of commonly used SC types for IBD treatment.

### HSCs

Human HSCs possess potent proliferation and differentiation capabilities, enabling them to differentiate into blood and immune cells, as well as migrate to damaged tissues and differentiate into various “structural cells”, such as epithelial cells, endothelial cells (ECs), and fibroblasts.<sup>23</sup> Moreover, HSCs also have the ability to regulate blood, immune, and tissue cells.<sup>23</sup> Hematopoietic stem cell transplantation (HSCT) can fully restore the hematopoietic and immune systems and prevent abnormal blood and immune cell differentiation.<sup>24</sup> Although HSCT technology is well-established and has been used to treat hematological tumors, its application to IBD treatment is limited by its high immunogenicity, the need for a donor match, marrow clearance therapy, invasiveness, associated adverse effects, as well as a high recurrence rate. Additionally, immunosuppression is required, which further adds to its limitations.<sup>25–27</sup>

### MSCs

MSCs are pluripotent SCs that are widely distributed throughout the human body, including the bone marrow, umbilical cord, placenta, amniotic fluid, fat, and other tissues.<sup>28</sup> These cells have the ability to actively proliferate and differentiate in multiple directions,<sup>29</sup> including osteoblasts, chondrocytes, adipocytes, and cardiomyocytes.<sup>30</sup> Researchers have successfully induced bone marrow MSCs to differentiate into IECs by using fetal intestinal wall connective tissue.<sup>19</sup> In addition to their differentiation potential, MSCs possess strong immune regulation and tissue repair abilities, as well as targeted and directional migration to injured tissue sites. They also have the ability to promote SC implantation and provide hematopoietic support.<sup>29,31</sup>

Studies have found that intestinal resident mesenchymal stem cells (MtSCs) in patients with IBD exhibit abnormal differentiation or depletion, making it difficult to regulate immune cells and repair IECs.<sup>13</sup> Mesenchymal stem cell transplantation (MSCT), as a supplement or alternative to MtSCs, can regulate immune cells and repair IECs and has become the most widely used SC therapy for IBD.<sup>15</sup> The use of MSCs for IBD treatment offers several advantages. Firstly, MSCs have low immunogenicity and do not generally require matching or myeloablative treatment, allowing for immunosuppressive therapy to be given as needed after transplantation.<sup>19</sup> Secondly, MSCs can be easily obtained from multiple sources, such as umbilical cord blood, peripheral blood, or fat, and can be easily separated and cultured.<sup>14</sup> Thirdly, MSCs can be administered through various methods, such as intravenous, intraperitoneal, subcutaneous, or local injection, allowing for the best method to be selected based

**Table 1** For a List of Commonly Used SC Types for IBD Treatment

Advantages	Disadvantages
<b>HSCs</b>	
<p>HSCs have a high differentiation potential and can reconstruct the hematopoietic and immune systems.</p> <p>H SCT technology is mature and mainly used for the treatment of hematologic tumors, and has been found to be effective in IBD.</p>	<p>HSCs are highly immunogenic.</p> <p>H SCT requires search for a match, marrow clearance therapy and immunosuppression. There are invasive operations, many adverse effects and high relapse rate. Not an option for the treatment of IBD alone.</p>
<b>MSCs</b>	
<p>MSCs can be multidirectionally differentiated and have strong immunomodulatory and tissue repair capabilities.</p> <p>MSCs have low immunogenicity, MSC T does not require searching for a match and marrow clearance therapy, and there is no significant difference in efficacy and safety between allogeneic or autologous transplantation.</p> <p>The source is widely available, and can be derived from bone marrow, cord blood, peripheral blood or adipose tissue.</p> <p>Infusion is flexible, with the option of intravenous, intraperitoneal, subcutaneous, or local injection routes.</p>	<p>MSCs are heterogeneous and can change with the microenvironment, lose their immunomodulatory capacity, or even become pro-inflammatory cells.</p> <p>MSCs remain weakly immunogenic, potentially tumorigenic or pro-tumorigenic risk.</p> <p>MSCs are injected intravenously and are mostly sequestered in organs such as liver and lung, with very little homing to the site of IBD lesions (&lt; 1%).</p>
<b>BM-MSCs</b>	<p>Classical source, most studied, easy to extract and isolate.</p> <p>High differentiation potential and immunomodulatory ability.</p>
<b>hUC-MSC UCBSCs</b>	<p>The MSCs of the umbilical cord are abundant, less immunogenic, and have greater differentiation potential and regulatory immunity. It can be expanded in vitro and frozen for a long time.</p>
<b>PBSCs</b>	<p>Peripheral blood is easily accessible.</p>
<b>ASCs</b>	<p>Adequate source, easy to obtain, easy to isolate and easy to cultivate.</p> <p>Strong immunomodulatory and tissue repair ability, stable biological properties.</p>
<b>ISCs</b>	
<p>ISCs can differentiate into various types of IECs and resident immune cells, capable of repairing intestinal epithelium and regulating intestinal immune cells. ISCs are relatively new in research and have great potential to be cultivated into intestinal organoids, which are the best source of SCs for IBD treatment.</p>	<p>The extraction and culture of ISCs are relatively difficult. Relying on endoscopic manipulation and in vitro 3D cell culture techniques, the process is difficult and relatively complicated.</p>
<b>iPSCs</b>	
<p>It is easily accessible, malleable, and can be induced to form iISCs, iMSCs, and iESCs with similar functions as ISCs, MSCs, and ESCs. Preclinical studies clarify the therapeutic effects of iPSCs in IBD, and no clinical studies in IBD are available.</p>	<p>The preparation process is complex and requires precise control, with a high failure rate and uncertain safety.</p> <p>Induced SCs are not exactly the same as in vivo SCs in terms of gene expression, phenotypic characteristics, proliferation, and differentiation.</p>
<b>pESCs</b>	
<p>pESCs are totipotent SCs that have been successfully induced into ISCs. pESCs are still in preclinical studies and there are no clinical studies on IBD yet.</p>	<p>The preparation process is complex and requires precise control, with high failure rates and uncertain safety.</p> <p>Faced with the problem of differences between induced SCs and in vivo SCs.</p>

**Abbreviations:** ASCs, adipose mesenchymal stem cells; BM-MSCs, Bone marrow mesenchymal stem cells; HSCs, hematopoietic stem cells; HSCT, hematopoietic stem cells transplantation; IBD, Inflammatory bowel disease; iESCs, Induced embryonic SCs; iISCs, Induced intestinal stem cells; iMSCs, MSCs derived from iPSCs; iPSCs, induced pluripotent stem cells; ISCs, intestinal stem cells; MSCs, mesenchymal stem cells; PBSCs, Peripheral blood stem cells; pESCs, Parthenogenetic embryonic stem cells; IECs, intestinal epithelial cells; UCBSCs, Umbilical Cord Blood stem cells; hUC-MSC, Umbilical cord mesenchymal stem cells.

on the type of lesion, with the transplantation process being similar to that of ordinary drug injection.<sup>17</sup> Finally, both autologous and allogeneic MSCs can be used with no significant difference in efficacy and safety.<sup>17</sup>

However, it is worth noting that MSCs are heterogeneous and can change with the microenvironment, losing their immunomodulatory ability and even becoming pro-inflammatory cells.<sup>32</sup> There are many differences between MSCs from different tissue sources in the treatment of IBD (Table 1). There are potential pathogenic risks associated with the colonization of allogeneic MSCs in vivo, such as potential immunogenicity and potential tumorigenicity.<sup>19</sup> Attention should also be paid to the homing rate of MSCs, most of which stay in the liver, lung, and other organs after intravenous, intraperitoneal, and subcutaneous injection, with fewer than 1% homing to intestinal lesions. Therefore, MSCs mainly rely on paracrine signaling to regulate intestinal immunity and repair the intestinal mucosal barrier rather than through proliferation and differentiation.<sup>17</sup>

## ISCs

IECs have a short lifespan of 3–5 days and rely on the continuous division and replenishment of ISC located in the intestinal crypts to maintain the integrity of the intestinal epithelial barrier.<sup>33,34</sup> ISCs possess pluripotency and can differentiate into various IECs and intestinal resident immune cells, such as T cells, B cells, and dendritic cells (DCs).<sup>35,36</sup> In response to intestinal mucosal injury, ISCs can proliferate and differentiate to repair the injured mucosa and regulate resident immune cells in the intestine.<sup>37–39</sup> Dysfunction of IECs is a crucial factor in the development of IBD, and studies have found abnormal differentiation or insufficient regeneration of ISCs in IBD patients.<sup>38</sup> ISCs are an excellent source of SCs for the treatment of IBD due to their vital role in the maintenance of intestinal structure and function.<sup>40</sup> However, the extraction and in vitro culture of ISCs are challenging.<sup>40</sup> Recent advancements in 3D culture technology have led to the successful extraction, isolation, and in vitro culture of human ISCs by recreating the “niche” of ISCs in vitro, resulting in the development of “intestinal organoids” for in vitro cell research. The application of ISCs in animal models of IBD has shown initial success, providing hope for the application of ISCs in the treatment of IBD.<sup>41</sup> The extraction and in vitro culture of intestinal organoids from ISCs require complex procedures, including endoscopic input, to ensure implantation at the site of intestinal lesions.<sup>42</sup>

## iPSCs

Transcription factor genes, including Oct4, Sox, Myc, and Klf4, have been introduced into somatic cells, such as fibroblasts or blood cells, through retrovirus, leading to the generation of iPSCs.<sup>43</sup> These cells possess similar morphology, function, and gene expression to embryonic stem cells (ESCs) and can be induced into various cell types, including iESCs, iMSCs, or iISCs, exhibiting similar functions to ESCs, MSCs, or ISCs, as confirmed in vitro and animal experiments. iPSCs offer several advantages over ESCs, including the avoidance of ethical issues and immune rejection.<sup>44–46</sup> Spence et al have demonstrated the feasibility of using iPSCs to generate 3D intestinal tissues that closely resemble human intestinal tissues.<sup>47–49</sup> After a series of induction and culture, iPSCs are differentiated into IECs, indicating their ability to produce tissue-specific cells. However, the preparation process of iPSCs is complex and requires precise control, often resulting in a high failure rate and uncertain safety. Moreover, although iPSCs can be induced into iESCs, iMSCs, or iISCs, their gene expression, phenotypic characteristics, proliferation, differentiation, apoptosis, and aging may not be identical.<sup>48</sup>

## Parthenogenetic Embryonic Stem Cells (pESCs)

ESCs pose a high risk of cancer due to ethical restrictions, making it difficult to control and conduct clinical research on IBD. To overcome these issues, oocytes can be stimulated using physical and chemical methods to develop into parthenogenetic blastocysts, which can give rise to pluripotent stem cells known as pESCs.

These cells can be induced to differentiate into the three germ layers, and recent studies have shown successful induction of human pESCs into intestinal epithelial stem cells, bringing hope for the treatment of IBD.<sup>44,46</sup> However, the preparation process is complex, requiring precise control, and has a high failure rate with uncertain security.<sup>46</sup> Furthermore, ESCs or ISCs that have been differentiated in vitro may differ from their in vivo counterparts in terms of surface markers and functions. Therefore, their efficacy in clinical settings remains to be verified.<sup>50</sup>

## The Mechanisms of SC Therapy in IBD

After implantation, SCs have been proven to persist in the body for an extended period and demonstrate a lasting effect.<sup>12,21</sup> Each implanted SC represents a relatively independent life activity unit that can adapt well to the internal environment and serve as a producer and manager of human cells.<sup>22</sup> SCs act on IBD lesions by regulating immune cells and repairing the intestinal mucosal barrier, which directly combats chronic intestinal inflammation.<sup>51</sup> In addition to addressing inflammation, SCs can play a beneficial role in enhancing intestinal microecology, resolving microcirculation disorders, and preventing fibrosis and cancerization, among other benefits.<sup>15</sup>

### Regulating the Immune System

Implanted SCs affect their own SCs and immune cells through differentiation and secretion.<sup>52</sup> HSCT, following marrow clearing, replaces the body's own SCs, while MSCT and iPSC transplantation (ISCT) primarily serve as supplements and regulators of the body's own SCs. HSCT induces the differentiation of various types of immune cells, while MSCT and ISCT primarily regulate immune cells through the secretion of cytokines or vesicles.<sup>30,53</sup> For patients with IBD, MSCs primarily regulate immune cells in the intestinal tract by secreting cytokines or vesicles. They regulate the polarization of macrophages to the M2 type, inhibit the proliferation of helper T cells Th1 and Th17, promote the differentiation of regulatory T cells, and inhibit the maturation and differentiation of DC, ultimately inhibiting inflammation. However, there is limited evidence supporting the homing of MSCs to the intestinal mucosa, and it remains unclear whether they can differentiate into immune cells.<sup>54–56</sup>

#### T Lymphocytes

MSCs express indoleamine 2,3-dioxygenase (IDO), which can lead to tryptophan depletion and the production of tryptophan metabolites, promoting apoptosis of Th1 cells and inducing the differentiation and maturation of Th2 cells (Figure 1).<sup>57</sup> Additionally, MSCs inhibit the expression of pro-inflammatory factors, such as TNF- $\alpha$ , IFN- $\gamma$ , and IL-17, by Th1 cells and enhance the expression of anti-inflammatory factors, such as IL-10 and TGF- $\beta$ , by Th2 cells. They also promote the differentiation of Treg cells in CD4+T cells and inhibit the differentiation of Th1 and Th17 cells, thereby exerting an immunomodulatory role.<sup>58,59</sup> Moreover, MSCs can inhibit the signal transducer and activator of transcription 3 (STAT-3) pathway, reducing IL-17 expression and inhibiting the differentiation of T lymphocytes to Th17 cells while promoting differentiation to Treg cells by regulating the STAT3/STAT5 signaling pathway.<sup>60,61</sup>

#### Macrophages

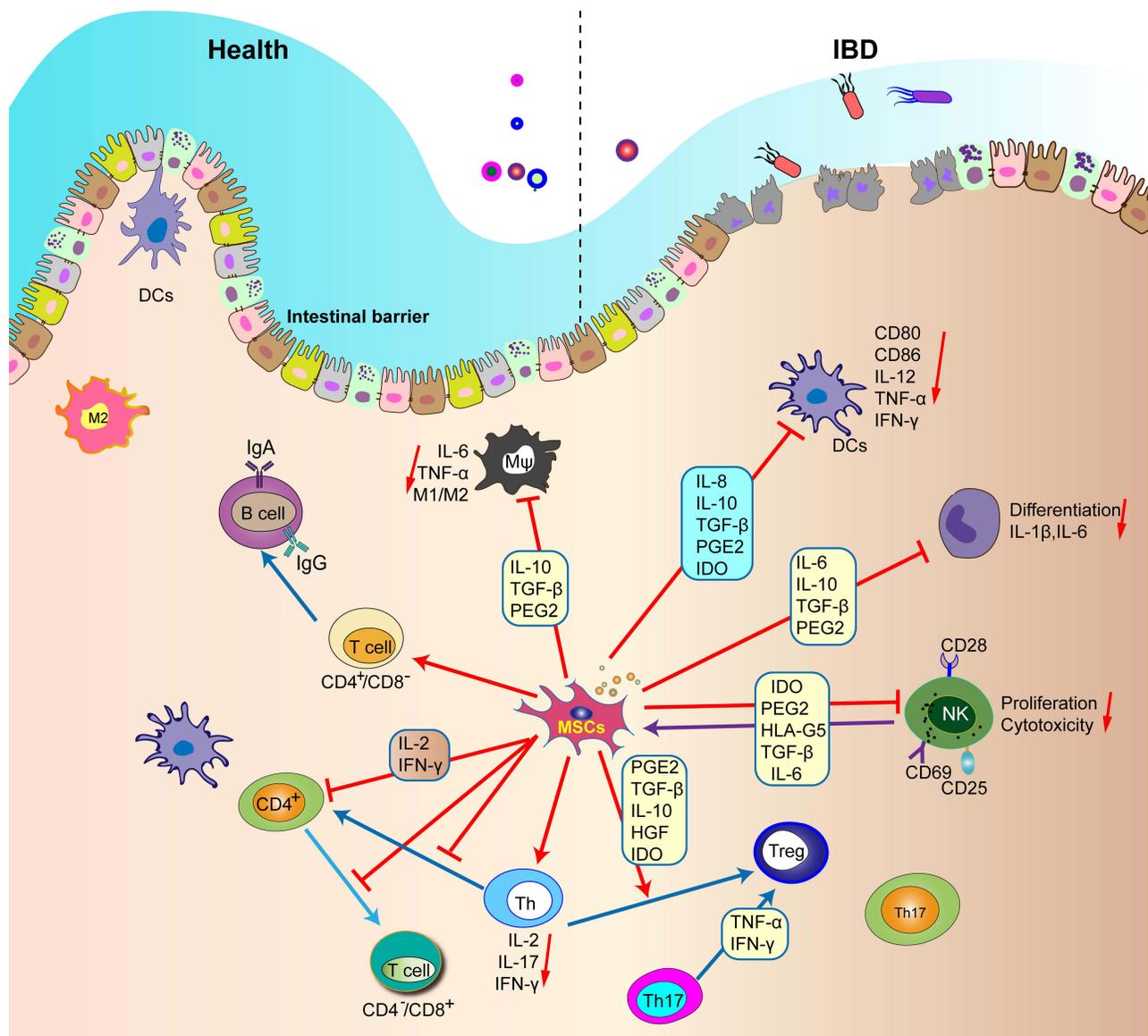
Inflammation-stimulated MSCs have been shown to secrete prostaglandin E2 (PGE2) and tumor necrosis factor  $\alpha$  stimulated gene-6 (TSG-6).<sup>62,63</sup> TSG-6 has been found to inhibit neutrophil migration, signal transduction of tissue-resident immune cells, and polarize macrophages into the M2 subtype (Figure 1). PGE2 can also bind to macrophage receptors, leading to a shift from the M1 subtype to the M2 subtype and inhibiting inflammation.<sup>64</sup>

#### DCs

Research has indicated that MSCs can down-regulate the expression of CD80, CD86, and IL-12 in DCs, thereby inhibiting DC maturation and activation. In addition, MSCs can inhibit the migration of mature DCs and weaken the antigen presentation ability of DCs.<sup>65</sup> Moreover, MSCs have the capacity to induce DCs to differentiate into tolerogenic dendritic cells (tDCs), which can effectively reduce inflammation by inhibiting effector T cells and promoting the activation of Treg cells (Figure 1).<sup>66</sup>

### Repair of Intestinal Mucosal Barrier

Studies have demonstrated that MSCs repair the intestinal mucosal barrier mainly by secreting cytokines or vesicles. Firstly, MSCs regulate the growth microenvironment of ISCs and promote their differentiation into IECs by secreting certain factors.<sup>67</sup> Secondly, MSCs secrete factors that promote the repair of IECs and reduce intestinal wall permeability.<sup>68</sup> Additionally, vesicles secreted by MSCs have been shown to inhibit apoptosis of IECs by reducing the cleavage of caspase-3, caspase-8, and caspase-9 in animal models of IBD.<sup>67</sup> Furthermore, MSCs can promote angiogenesis through secretion by promoting lymphatic endothelial cell (LEC) proliferation, migration, and



**Figure 1** MSCs regulate the function of all immune cells in the intestinal tract.

**Notes:** MSCs inhibit inflammatory M1 macrophages and promote their transformation to M2 phenotype. MSCs inhibit the maturation of DC and the secretion of tumor necrosis factor- $\alpha$ , interferon- $\gamma$  and IL-12 by DC. In addition, MSC can secrete PEG2, IL-10, TGF- $\beta$ , inducible, IDO and IL-6 to inhibit the proliferation of T cells and NK cells, thus reduce the production of inflammatory cytokines in Th1 and Th17, and inhibit immune response and inflammation.

**Abbreviations:** IDO, indoleamine 2,3-dioxygenase; DCs, dendritic cells; MSCs, Mesenchymal stem cells.

lymphangiogenesis. MSCs can also reduce oxidative stress damage in tissue cells by inhibiting the production of endogenous toxins, such as oxygen radicals.<sup>69–71</sup>

While MSCs can differentiate into IECs and myofibroblasts when homing to the intestinal mucosa, this method of repair is rare and not the primary repair mechanism.<sup>72,73</sup> Local injection of MSC preparations has been found to significantly increase the homing rate in IBD lesions and produce good local effects, which is the basis for the clinical application of local injection in the treatment of perianal fistula Crohn's disease (CD).<sup>12,19</sup>

## Improving Intestinal Microecology

The intestinal microbiota and host cells are interdependent and together constitute the intestinal microecology.<sup>74</sup> Their interaction results in a state of dynamic equilibrium, which is referred to as intestinal microecological homeostasis. It has become a consensus that the disorder of intestinal microecology is the primary pathogenesis of IBD. The maintenance of

intestinal microecological homeostasis is heavily reliant on intestinal immunity and mucosal barrier function. MSCs have an indirect impact on improving intestinal microecology through their effects on intestinal immunity and mucosal barrier function.<sup>75</sup> Furthermore, MSCs secrete various antimicrobial peptides, such as IL-10, PGE2, IDO, and IL-17, which have demonstrated antibacterial activity in multiple studies.<sup>76–78</sup>

## Promoting Angiogenesis

Microvascular dysfunction and damage to the endothelial barrier occur in IBD, which can impact colon tissue perfusion and healing.<sup>79</sup> MSCs secrete various angiogenic factors, including vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), transforming growth factor beta (TGF- $\beta$ ), and angiopoietin-1, which promote EC proliferation and the formation of new blood vessels.<sup>80,81</sup> Furthermore, MSCs can differentiate into vascular ECs and promote angiogenesis.<sup>82</sup> A study has also shown that MSCs can successfully differentiate into colon vascular ECs and promote angiogenesis.<sup>82</sup>

## Anti Fibrosis

Intestinal fibrosis is a common complication of IBD that often leads to intestinal stenosis and obstruction in patients with CD.<sup>83</sup> Research has identified the TGF- $\beta$  signaling pathway as the core mechanism driving intestinal fibrosis.<sup>84</sup> BM-MSCs have been shown to reverse the epithelial-mesenchymal transition (EMT) of TGF- $\beta$ 1-treated IEC-6 cells by carrying miR-200b, resulting in a significant reversal of intestinal fibrosis.<sup>85,86</sup> Furthermore, studies on placental MSCs have revealed that inhibiting Rho/MRTF/SRF expression can downregulate TGF- $\beta$ 1-induced fibroblast activation, reduce collagen deposition in the intestinal wall, and thus inhibit intestinal fibrosis.<sup>87,88</sup>

## SCs in Colorectal Cancer Cryotherapy

While the relationship between SCs and intestinal tumors is complex and controversial, many studies have shown that MSCs have certain anti-tumor properties.<sup>89,90</sup> For example, MSCs can migrate to the intestine after intraperitoneal injection and prevent the occurrence of colitis-related colorectal cancer by inhibiting tumor cell proliferation and inducing tumor cell apoptosis.<sup>91,92</sup> Additionally, some studies suggest that MSCs can modify the intestinal microbiome by increasing the number of anti-tumor bacteria, such as Parabacteroides and Staphylococcus, thus preventing tumor formation.<sup>93,94</sup>

## Therapeutic Effects of SCs in IBD

Numerous studies have explored the potential of SCs in treating IBD. To conduct a thorough review, we searched the PubMed and Web of Science Library databases using keywords, such as “stem cell therapy”, “mesenchymal stromal cells”, “HSCs”, “immunosuppression”, “inflammatory bowel disease”, “ulcerative colitis”, “Crohn’s disease”, and “perianal fistula CD”. We also searched the NIH clinical trial database (<https://ClinicalTrials.gov/>) for clinical trials of MSCs in the treatment of IBD. So far, 34 clinical trials involving MSCs have been registered and implemented for treating IBD.<sup>12,95</sup> From these studies, we carefully selected representative, complete, and high-quality studies and classified them into preclinical studies, completed clinical studies, and ongoing clinical studies. We summarized the operational process and research results, evaluated the feasibility, effectiveness, and safety, and objectively reflected on the research status.

## Preclinical Use of SCs for IBD Treatment

Preclinical research has been conducted on the application of various types of SCs, including HSCs, MSCs, ISC, iPSCs, and pESC, for treating IBD. These studies have evaluated the efficacy, feasibility, and safety of SC therapy in animal models of IBD and have gradually revealed the internal mechanisms underlying the therapy. These findings have shown us the potential application prospects and limitations of SC therapy for IBD.<sup>16,96</sup> However, further exploration and improvement of strategies are still necessary to lay the foundation for clinical research in this field.

## HSCs

HSCT has been widely used clinically for the treatment of hematologic diseases. However, due to its complexity, there have been few preclinical studies conducted on HSCT, and almost no studies have been conducted on animal models of IBD.<sup>25</sup>

## MSCs

A large number of preclinical studies have been conducted at the cellular level and on animal models of IBD regarding MSCs.<sup>19</sup> Most of these studies have confirmed the efficacy of MSCs in treating IBD, with desirable feasibility and safety.<sup>15</sup> Therefore, MSCs have become the primary choice for SC therapy for IBD. In several studies, MSCs from human umbilical cord blood, bone marrow, and adipose tissue are used to treat IBD animal models, achieving the goal of promoting symptom relief and pathological tissue repair.<sup>97–99</sup>

Qu et al have infused donor rats with BM-MSCs labeled with the fluorescent dye PKH26 into rats that have suffered indomethacin-induced GI tract injury. PKH26-positive BM-MSCs are subsequently found in the damaged mucosa, and the mucosal layer and crypt layer of the small intestine are thicker in BM-MSC-treated rats. This suggests that the therapeutic effect of MSCs is related to the regenerative repair of damaged tissues.<sup>100</sup>

Lian et al have established a mouse model of intestinal fibrosis by increasing the dose of TNBS by enema for 7 weeks, followed by introducing MSCs by enema for treatment. The results show that MSCs inhibit the expression of TGF- $\beta$  and phosphorylation of Smad2 and Smad3 after TNBS induction, thereby inhibiting EMT and reversing intestinal fibrosis.<sup>85</sup> Table 2 summarizes the preclinical studies on MSCs for the treatment of IBD completed in recent years.

**Table 2** Summarizes the Preclinical Studies on MSCs for the Treatment of IBD Completed in Recent Years

Author/ Time References	Animal Model	SCs Sources	SCs Route	SCs Dose (Cells)	Follow- Up Period	Primary Outcomes
Barnhoorn (2020) <sup>145</sup>	DSS- induced colitis	BM-MSC	IP	$2 \times 10^6$ cells	11 days	MSCs after in vivo aggregation show a favorable RNA expression profile for the treatment of colitis. MSCs spheroids showed high expression of Ki-67 and low levels of apoptotic marker cleaved caspase-3. Locally applied MSCs and MSCs spheroids are both able to ameliorate DSS-induced colitis and show similar clinical effects.
Barnhoorn (2020) <sup>145</sup>	DSS- induced colitis	BM-MSC	Endoscopic	$2 \times 10^6$ cells	4–6 days	Endoscopic injection can be a feasible and effective novel application route for MSCs therapy in patients with luminal IBD.
Chao (2016) <sup>146</sup>	TNBS- induced colitis	UCBSC	IP	$10 \times 10^6$ cells	14 days	The mortality in UCBSC-treated TNBS mice was 20% (55% in colitis model). IL-20 and TGF-Beta were significantly higher in UCBSC-treated mice ( $p = 0.04$ and $0.02$ respectively).
Cheng (2017) <sup>147</sup>	DSS- induced colitis	BM-MSC	IV	$5 \times 10^6$ cells	8 days	IL-25-MSCs treatment significantly attenuate the colon shortening ( $12 \pm 0.62$ cm); IL-25 could enhance immunomodulatory ability of MSCs via inhibiting Th17 immune response and promoting the regulation of Tregs cells.
de Aguiar (2018) <sup>148</sup>	DSS- induced colitis	ASCs	IP	$10 \times 10^6$ cells	7 days	ASCs-treated mice did not present severe reduction in colon length, and presented a reduced tissue damage score index (3). Significant reduction of IFN-gamma and TNF-Alpha, and reduction of IL-6 and MCP-1 protein levels. ASCs treatment reduced DCs and macrophages presence in the colon.
de Aguiar (2018) <sup>148</sup>	DSS- induced colitis	ASCs	IP	$2 \times 10^6$ cells	7 days	ASCs ameliorated the severity of DSS-induced colitis, reducing colitis pathological score and preventing colon shortening.

(Continued)

Table 2 (Continued).

Author/ Time References	Animal Model	SCs Sources	SCs Route	SCs Dose (Cells)	Follow- Up Period	Primary Outcomes
de la Portilla (2013) <sup>149</sup>	TNBS- induced colitis	ASCs	Local	60×10 <sup>6</sup> cells	24 weeks	First study which shows the homing migration of ASCs to areas of experimentally-induced colitis following rectal installation.
de la Portilla (2018) <sup>150</sup>	TNBS- induced colitis	ASCs	Local	2 × 10 <sup>6</sup> cells	10 days	There were no differences in component rectal wall thicknesses with a higher Hunter score in the treated group compared with the controls.
Fu (2017) <sup>151</sup>	TNBS- induced colitis	ASCs	IP	2 × 10 <sup>6</sup> cells	6 days	Decrease the weight loss and DAI score, MPO activity. Reduced levels of ROR and IL-17A; inhibited STAT3 phosphorylation, but increased STAT5 phosphorylation.
González- Rey (2009) <sup>152</sup>	DSS- induced chronic colitis	ASCs	IP	10×10 <sup>6</sup> cells	27 days	ASCs treatment protects against DSS-induced acute colitis as well as chronic severe colitis ( <i>p</i> 0.01 and <i>p</i> 0.001 respectively); reduces colonic inflammatory responses in DSS-induced chronic colitis ( <i>p</i> 0.001).
Gregoire (2018) <sup>153</sup>	Fistula Crohn's disease	ASCs	Local	(3–30) ×10 <sup>6</sup> cells	8 weeks	6/8 fistulas healed, 2/8 improved.
Heidari (2021) <sup>56</sup>	DSS- induced colitis	ASCs	IP	10×10 <sup>6</sup> cells	34 days	There was no significant difference in the survival rate among the study groups; however, there was a significant increase in terms of the colon length ( <i>p</i> 0.005). In the treated mice the level of mucosal damage was significantly lower ( <i>p</i> 0.005).
Heidari (2021) <sup>56</sup>	DSS- induced colitis	ASCs	IP	2 × 10 <sup>6</sup> cells	34 days	The regulatory effects of ASCs and their CM in inflammatory conditions because of colitis.
In Kap (2010) <sup>137</sup>	DSS- induced colitis	MSCs	IV	1 × 10 <sup>6</sup> cells	7 days	Anti-addressin Ab coating on MSCs increased cell delivery to inflamed colon and increased the efficacy of MSCs treatment of IBD.
Jianxia Hu (2016) <sup>154</sup>	Luminal Crohn's disease	UCBSC	IV	0.5×10 <sup>6</sup> cells	3 months	30/36 patients showed good response and diffuse and deep ulcer formation and severe inflammatory mucosa were improved markedly.
Lee (2016) <sup>155</sup>	DSS- induced colitis	BM-MSC	IV	10×10 <sup>6</sup> cells	33 days	IL-10 production was upregulated by about 10-fold in BM-MSC-treated mice and showed a preventive effect on weight loss.
Lee (2016) <sup>155</sup>	DSS- induced colitis	BM-MSC	IV	30×10 <sup>6</sup> cells	33 days	Infusion of BM-MSC at the onset of disease exerted preventive and rapid recovery effects.
Lee (2018) <sup>156</sup>	DSS- induced colitis	UCBSC	IP	2×10 <sup>6</sup> cells	12 days	The survival rate was further increased by co-treatment compared to UCBSC or MIS416 single treatments; colon lengths were significantly increased in cotreatment; colonic inflammation was more effectively resolved by co-treatment with MIS416 and UCBSC, and only co-treatment markedly decreased fibrosis and enhanced tissue regeneration.

(Continued)

Table 2 (Continued).

Author/ Time References	Animal Model	SCs Sources	SCs Route	SCs Dose (Cells)	Follow- Up Period	Primary Outcomes
Legaki (2016) <sup>147</sup>	DSS- induced colitis	ASCs	IP	1.5×10 <sup>6</sup> cells 200 µL / dose	7 days	CM treatment significantly decreased the extension and severity of the inflammation in comparison to the DSS-treated mice; the relative expression levels of IL-10 mRNA were significantly increased, and TGFβ1 was significantly higher (p 0.0001).
Mao (2017) <sup>157</sup>	DSS- induced colitis	UCBSC	IV	1.3×10 <sup>6</sup> cells	11 days	Exosomes from MSC have profound effects on alleviating DSS-induced IBD and may exert their impact through the modulation of IL-7 expression in macrophages.
Martín (2018) <sup>103</sup>	TNBS- induced colitis	ASCs	Local	10×10 <sup>6</sup> cells	11 days	Submucosal injection of human ASCs ameliorates the course of TNBS colitis in immunocompetent rats.
Martin Arranz (2018) <sup>158</sup>	TNBS- induced colitis	ASCs	Endoscopic	10× 0 <sup>6</sup> cells	11 days	The endoscopic score improved in the ASCs group by 47.1% ± 5.3% vs 21.8% ± 6.6% in the vehicle group.
Miyamoto (2017) <sup>159</sup>	TNBS- induced colitis	ASCs	IV and Local	1×10 <sup>6</sup> cells IV and 400 µL Local	7 days	ASCs transplantation significantly decreased the number of neutrophils, attenuated acute inflammation. In the TNBS-CM gel group ulcers were shallow and bleeding was not detected, therefore improved endoscopic score. In the gel group mRNA expression levels of TNF-Alpha, CXCL1, CCL2 and IL-6 were increased.
Molendijk (2015) <sup>160</sup>	Fistula Crohn's disease	BM-MS	Local	10,30, 90×10 <sup>6</sup> cells	6, 12, 24 weeks	At week twelve, 3 of 9 individual fistulas had healed in group 1 (33.3%), 6 of 7 had healed in group 2 (85.7%), 2 of 7 had healed in group 3 (28.6%), and 3 of 9 had healed in the placebo group (33.3%).
Pak (2018) <sup>161</sup>	DSS- induced colitis	BM-MS ASCs	Endoscopic	8×10 <sup>5</sup> , 1.1×10 <sup>6</sup> cells	1–3 days	The success rate was 37.60% for ASCs group and 35.20% for BM-MS group.
Panés (2016) <sup>109</sup>	Fistula Crohn's disease	ASCs	Local	120×10 <sup>6</sup> cells	24 weeks	Remission in the ITT (53 of 107 [50%] vs 36 of 105 [34%]; difference 15.2%, 97.5% CI 0.2–30.3; p = 0.024) C × 601 vs placebo.
Panés (2018) <sup>157</sup>	Fistula Crohn's disease	ASCs	Local	120×10 <sup>6</sup> cells	52 weeks	C × 601 achieved combined remission (56.3%) vs controls (38.6%).
Park (2018) <sup>109</sup>	DSS- induced colitis	ASCs	IP	10×10 <sup>6</sup> cells	20 days	The results suggest that PGE2, produced by co-culture of ASCs and THP-1, reduces M1 population, decreased the frequency of macrophage transition.
Park (2018) <sup>162</sup>	DSS- induced colitis	ASCs	IP	2 × 10 <sup>6</sup> cells	20 days	ASCs can suppress the inflammatory response by controlling the macrophage population, and ASCs may be therapeutically useful for the treatment of IBD.
Pouya (2018) <sup>163</sup>	DSS- induced colitis	MSCs	IP	500 µL, ×3	10 days	After infusion, colon inflammation was reduced and histopathological analysis showed a decrease in mucosal degeneration.
Song (2017) <sup>164</sup>	DSS- induced colitis	MSC- Exo UCBSC	IP	150µg 10×10 <sup>6</sup> cells;	36 days	MSC-Exo ameliorates the clinical parameters in DSS-induced colitis; the treated group showed significantly less MPO activity. The level of IL-17 was significantly decreased, whereas those of IL-10 and TGF-Beta1 were increased.

(Continued)

Table 2 (Continued).

Author/ Time References	Animal Model	SCs Sources	SCs Route	SCs Dose (Cells)	Follow-Up Period	Primary Outcomes
Song (2018) <sup>165</sup>	DSS-induced colitis	Canine ASCs	IP	2 × 10 <sup>6</sup> + TSG-6 siRNA	10 days	ASCs-secreted TSG-6 reduced inflammatory response and apoptosis in the colon; intraperitoneally infused ASCs did not migrate to the inflamed colon; increased M2 macrophages in the inflamed colon.
Soontarak (2018) <sup>48</sup>	DSS-induced colitis	iMSCs ASCs	IV	3 × 10 <sup>6</sup> cells	19 days	Colonic tissues from mice treated with either iMSCs or ASCs exhibited an overall reduction in transmural inflammation, with significantly less infiltration of inflammatory cells in the lamina propria, diminished mucosal ulceration and decreased mucosal collapse and granulation tissue formation.
Tanaka (2008) <sup>166</sup>	DSS-induced colitis	BM-MSC	IV	5 × 10 <sup>6</sup> cells	7 days	In the rectum of treated rats the mRNA expression of TNF-alpha and IL-1Beta was markedly decreased to (43.7 ± 25.5% p 0.05 and 14.5 ± 12% p 0.01 respectively), as well as COX-2 16.5 ± 15.2% (p 0.01).
Tanaka (2008) <sup>166</sup>	DSS-induced colitis	MSCs	IV	5 × 10 <sup>6</sup> cells	7 days	Exogenous MSCs accumulated in inflamed tissues and ameliorated DSS-induced colitis via a local anti-inflammatory action.
Wang (2016) <sup>130</sup>	DSS-induced colitis	BM-MSC	IP	0.5 × 10 <sup>6</sup> cells	10 days	Intraperitoneal injection is the best delivery way for MSCs: showed better mucosa recovery and higher cell engraftment at inflamed colon.
Wu (2018) <sup>167</sup>	DSS-induced colitis	UCBSC	IV	400µg UC-MSC	11 days	Exosomes from UCBSCs have profound effects on alleviating DSS-induced IBD and may exert their function by regulating the ubiquitin modification level.
Xu (2018) <sup>168</sup>	DSS-induced colitis	ERC	IV	3 × 10 <sup>6</sup> cells	10 days	ERC treatment significantly reduced the levels of TNF-Alpha, IL-1Beta and IL-6; ERC downregulated the expanded Th1 and Th17 cells in colitis, and elevated the proportion of Tregs in lymphocytes; ERC inhibited B-cell activation, differentiation and IgG production in colitis.
Yu (2017) <sup>169</sup>	DSS-induced colitis	T-MSCs	IP	20,40, × 10 <sup>6</sup> cells	30 days	Co-culture with T-MSCs clearly inhibited the PMA-stimulated proliferation of splenocytes by 60%; T-MSCs [×4] treated mice's survival rate was improved to that of the normal. T-MSCs [×2] injection also significantly improved the survival rate to 89% of the control. T-MSCs [×4] treatment inhibits DSS-induced colon shortening.

**Abbreviations:** AF-MSC, Amniotic fluid Mesenchymal stem cell; ASCs, adipose mesenchymal stem cells; BM-MSCs, Bone marrow mesenchymal stem cells; CCL2, Chemokine Ligand 2; CD, Crohn's disease; CXCL1, Chemokine Ligand 1; DAI, Disease activity index; DCs, Dendritic cells; DSS, Dextran Sulfate Sodium Salt; ERC, Endometrial regenerative cells; IBD, Inflammatory bowel disease; IFN- $\gamma$ , interferon- $\gamma$ ; IL-10, interleukin-10; IL-17A, interleukin-17A; IL-1 $\beta$ , interleukin-1 $\beta$ ; IL-20, interleukin-20; IL-25, interleukin-25; IL-6, interleukin-6; IL-7, interleukin-7; iMSCs, mesenchymal stem cells derived from induced pluripotent stem cells; IP, intraperitoneal injection; iPSCs, induced pluripotent stem cells; IV, intravenous injection; MCP-1, Monocyte Chemotactic Protein-1; MPO, myeloperoxidase; MSC-Exo, Mesenchymal stem cell exosomes; MSCs, mesenchymal stem cells; MSCT, mesenchymal stem cells transplantation; PFCD, perianal fistula Crohn's disease; PGE2, prostaglandin E2; PMA, THP-1 nuclear extract lysate; ROR, Retinoic acid related orphan receptor; SCs, stem cells; STAT3, signal transducer and activator of translation-3; TGF- $\beta$ , transforming growth factor- $\beta$ ; THP-1, Tohoku Hospital Pediatrics-1; T-MSCs, Tonsil MSCs; TNBS, 2,4,6-trinitro-Benzenesulfonic acid; TNF-alpha, tumor necrosis factor alpha; TSG-6, tumor necrosis factor  $\alpha$  stimulated gene-6; UCBSCs, Umbilical Cord Blood stem cells.

## ISCs

ISCs have been identified as a promising therapy for SCs, with their feasibility being affirmed through breakthrough technology. The efficacy of ISCs in treating IBD has also been preliminarily confirmed in preclinical studies. In recent years, the establishment of an in vitro ISC culture system and advancements in 3D culture technology have brought about promising prospects for patients with IBD or other refractory GI diseases.<sup>101</sup>

Fukuda et al are the first to isolate intestinal crypts (containing ISCs) from EGFP transgenic mice and form colonic-like organs after 1 week of 3D culture. The culture medium containing colonic organoids is subsequently injected into an

IBD mouse model induced by ethylene dinitril tetraacetic acid (EDTA) enema. One day after the injection, EGFP cells are found to be scattered on the surface of the recipient's colon, partially retaining the organoid structure. Two weeks later, EGFP cells are found to display a complex structure with some downward invagination to form a crypt-like structure. At week 4, the EGFP epithelium is stably sunk into the recipient's colon. This demonstrates that *in vitro* cultured mouse intestinal epithelial-like organs are able to reconstruct new epithelium in the recipient's colon.<sup>102</sup>

Shaker et al have cultured single SCs from the colon of mice to produce tiny organs of intestinal epithelial tissue, which are then transplanted into the damaged intestine of mice through an enema. The tiny organs attach to the damaged area and repair the mucosal damage. After 25 weeks of tracking, the tiny organs still show self-renewal ability, and the weight of the treated mice increases significantly.<sup>103</sup>

Recently, Watanabe et al have formed "intestinal organoids" from human colon fragments and mouse colon segments by 3D *in vitro* culture, respectively. They are injected into the intestine of a DSS-induced IBD model mouse with a thin tube, and the anus of the mouse is subsequently sutured. The transplanted intestinal organoid is confirmed to be attached to the damaged area of the mouse intestine by fluorescence detection, preserving the intestinal epithelial structure. Subsequently, the attached intestinal organoid is found to be able to expand in the mucosa of the mouse intestinal colon and reconstruct new intestinal epithelial tissue. Finally, as observed from the mouse colon specimens and tissue examination, the transplantation of intestinal-like organs significantly reduces the inflammatory response in the colon, and the intestinal damage is repaired.<sup>104</sup>

## iPSCs

Preclinical research on iPSCs has yielded promising results. In a mouse model of IBD, iMSCs are found to be as effective as adipose MSCs (ASCs), promoting intestinal angiogenesis and stimulating the proliferation of IECs and Lgr5+ ISCs.<sup>48</sup> In another study, iPSCs are induced from mouse skin fibroblasts and injected into mice with DSS-induced colitis. The mice that receive the iPSCs demonstrate a significant improvement in clinical symptom scores after just 1 day. After 72 hours, the colon specimens from these mice are significantly longer and heavier, with reduced transmural inflammation, mucosal ulceration, and infiltration of inflammatory cells in the intestinal mucosa.<sup>48</sup>

Spence et al have successfully generated definitive endoderm (DE) from human iPSCs by inducing them with Activin A, a nodal-related TGF- $\beta$  molecule. They then transfer the cells to a culture environment containing Wnt3a and FGF4 to create an "intestinal organoid" with a crypt structure of the intestinal epithelium containing various types of IECs and intestinal endocrine cells.<sup>49</sup>

## Completed Clinical Studies of SCs for IBD Treatment

Clinical research on IBD has primarily focused on MSCs and HSCs, with little attention given to ISCs and iPSCs.

### HSCT

The safety of HSCT for treating IBD has faced significant challenges due to its complex operation and many adverse reactions. As a result, HSCT is gradually being replaced by MSCT. While most completed clinical studies have achieved certain efficacy, some studies have shown no response to IBD treatment and have faced severe adverse reactions.<sup>26</sup> Burt et al have reported on 24 patients with refractory CD who are treated with autologous nonmyeloablative HSCT and immunosuppression using cyclophosphamide (CTX) and horse anti-thymocyte globulin (ATG), with no transplant-related deaths. The recurrence-free survival rates are 91%, 63%, 57%, 39%, and 19% at 1–5 years after transplantation, respectively. Nine patients have survived for more than 5 years without recurrence.<sup>105</sup> Cassinotti et al have performed autologous HSCT on four patients with refractory CD. In the third month, the primary endpoint of clinical remission is achieved in all patients. After a median follow-up of 16.5 months, all affected patients achieved complete fistula closure, and three out of four patients maintained both clinical and endoscopic remission even after the withdrawal of all drugs.<sup>106</sup> The ASTIC study (autologous stem cell international Crohn's disease trial) is a multicenter, randomized, Phase III intervention study. Twenty-three patients with refractory CD underwent autologous HSCT, while 22 received standard CD treatment (controls). The results show that only two out of 23 patients in the SC therapy group achieved complete clinical and endoscopic remission, whereas only one out of 22 patients in the control group achieved clinical and

endoscopic remission, with no statistical difference. The study has also found that all patients who received SC therapy experienced different degrees of complications, and infection is the most common.<sup>107</sup> However, when the evaluation criteria are lowered, the proportion of patients receiving HSCT treatment who stopped using immunosuppressive drugs is significantly higher than that of the control group, and the Crohn's disease activity index (CDAI) is also significantly lower than that of the control group. The quality of life and the remission of intestinal lesions under endoscopy are better than those of the control group, indicating that HSCT treatment still has a positive effect on these patients.<sup>107</sup>

## MSCs

Clinical studies on MSCs have primarily focused on patients with refractory perianal fistulous CD and refractory intestinal luminal IBD. MSCs can be sourced from various tissues, such as bone marrow, umbilical cord, and adipose tissue. Adipose tissue is a readily available and easy-to-culture source of MSCs that is increasingly used in clinical applications, and it now accounts for one-third of all MSC sources. The differentiation and secretory functions of MSCs are influenced by their growth environment, and by utilizing culture pretreatment and content modification, MSCs with specific functions can be generated. However, due to the heterogeneity of MSCs, there is still a lack of uniform research standards, leading to mixed results in clinical trials. The clinical applications of MSCs in IBD are broadly divided into two categories: local application in CD and intractable intraluminal IBD.<sup>29</sup>

## Box 1 MSCs for Local Application of CD

Local application of MSCs has been found to be effective in treating refractory perianal fistula in Crohn's disease (PFCD). The first clinical preparation of MSCs for PFCD, darvadstrocel/Cx601 (Takeda) (allogeneic adipose-derived stem cells), has already entered clinical treatment.<sup>108</sup> More optimized MSC preparations and derivatives are currently in clinical trial stages. In a review of 32 Phase I–III clinical trials of HSCT for CD complicated by anal fistula, it is found that more than half of the patients achieved complete remission, at least 2/3 of the patients responded to treatment, and no serious adverse effects were reported.<sup>19</sup> In a study conducted by Panés J, 212 patients with refractory, complex, and active perianal fistulas are included. Of these patients, 107 received allogeneic, expanded, and induced SCs (Cx601) treatment, while the remaining 105 were in the placebo group and received routine treatment. The intention-to-treat (ITT) analysis reveals that 53 individuals in the Cx601 group (50%) and 36 in the placebo group (34%) achieved combined remission. Among the patients who received Cx601 treatment, 18 (17%) of 103 experienced treatment-related adverse events compared to 30 (29%) of 103 in the placebo group. This study concludes that Cx601 is an effective and safe treatment for patients with refractory, complex, and active perianal fistulas.<sup>109</sup> Another study has included 40 patients with PFCD. Of these, 25 patients are in the darvadstrocel treatment group (an expanded allogeneic adipose-derived stem cells), while the remaining 15 are in the control group. Thirty-seven patients completed the 104-week follow-up, and seven treatment-emergent serious adverse events were reported. At week 104, clinical remission is reported in 14/25 (56%) patients in the darvadstrocel group and 6/15 (40%) patients in the control group.<sup>110</sup>

## Box 2 MSCs for Intractable Intraluminal IBD

For refractory intraluminal IBD, MSCs can be administered intravenously or intraperitoneally. This approach has shown promise in achieving relief of clinical symptoms and improvement of mucosal lesions while also being highly safe and cost-effective.<sup>15,19</sup>

In a recent randomized controlled trial (RCT), 82 patients with refractory intracavitary IBD were included, and 41 patients were randomly assigned to receive a total of four peripheral intravenous infusions of  $1 \times 10^6$  UC MSCs/kg once a week. After 12 months of treatment, the CDAI decreased by  $62.5 \pm 23.2$  in the UC-MSC group compared to  $23.6 \pm 12.4$  in the control group. Additionally, the Harvey-Bradshaw index (HBI) decreased by  $3.4 \pm 1.2$  in the UC-MSC group and by  $1.2 \pm 0.58$  in the control group. The corticosteroid dosage also decreased by  $4.2 \pm 0.84$  mg/day in the UC-MSC group and by  $1.2 \pm 0.35$  mg/day in the control group. These results suggest that UC-MSCs are effective in treating refractory intracavitary IBD.<sup>111</sup>

In a Phase II clinical study, 16 patients with intractable intraluminal CD and a CDAI score  $>250$  were included. Subjects received intravenous infusions of allogeneic MSCs ( $2 \times 10^6$  cells/kg body weight) weekly for 4 weeks. At day

42, among the 15 patients who completed the study, the mean CDAI score decreased from 370 to 203. Twelve patients had a clinical response, and seven patients had endoscopic improvement (47%) with a decrease in mean CD endoscopic index of severity (CDEIS) scores from 21.5 to 11.0.<sup>112</sup>

Knyazev et al have included 34 patients with refractory intracavitary CD, all of whom were treated with intravenous infusion of allogeneic BM-MSK preparations. Some patients (n=19) received azathioprine (AZA) combination therapy. The CDAI assessed before and 2, 6, and 12 months after transplantation was 337.6±17.1, 118.9±12.4, 110.3±11.1, and 99.9±10.8, respectively, with a significant decrease in CDAI after transplantation and comparable CDAI scores in both groups. Blood immunoassays show that the levels of pro-inflammatory factors, such as INF- $\gamma$ , TNF- $\alpha$ , and IL-12- $\beta$ , are significantly decreased and more pronounced in the combined ASA group.<sup>113</sup> Table 3 summarizes the clinical studies of MSCs in treating IBD completed in recent years.

**Table 3** Summarizes the Clinical Studies of MSCs in Treating IBD Completed in Recent Years

Author, Time and Country	Phase	Clinical Indication	Number	SC Sources Route	Adjuvant Therapy	Follow-Up Period	Primary Outcomes	Ref.
Oyama Y. 2005, USA	I	Refractory CD	12	Auto-HSCs IV	CTX and equine ATG	7–37 months	11 patients entered a sustained remission, only 1 patient has developed a recurrence of active CD, which occurred 15 months after HSCT.	[170]
Burt R. K. 2010, USA	I	Severe CD	24	Auto-HSCs IV	CTX and equine ATG	1–5 years	Eighteen of 24 patients are 5 or more years after transplantation.	[105]
Clerici M. 2011, Italy	N/A	Active moderate-severe CD	7	Auto-HSCs IV	CTX and rabbit ATG	1 year	Most of whom showed clinical and endoscopic complete remission and were maintained for one year.	[171]
Hasselblatt P. 2012, Germany	I/II	Refractory CD	12	Auto-PBSCs IV	High-dose CTX	0.5–10.3 years	5 patients achieved a clinical and endoscopic remission within 6 months after auto-PBSC. 7/9 relapses patients reduced maintenance dose of conventional drugs.	[172]
Hawkey C J. 2015, United Kingdom	III	Refractory CD	45	Auto-HSCs IV	High-dose CTX	1 year	Compared with conventional therapy, did not result in a statistically significant improvement in sustained disease remission at 1 year.	[107]
Kotlarz D. 2012, Germany	II	IBD with IL-10 gene defect	66	Allo-HSCs IV	Lemtrada F-ara-A treosulfan	2 years	Allo-HSCT was performed in 5 patients to induce sustained clinical remission.	[173]

(Continued)

Table 3 (Continued).

Author, Time and Country	Phase	Clinical Indication	Number	SC Sources Route	Adjuvant Therapy	Follow-Up Period	Primary Outcomes	Ref.
Liang J. 2012, China	N/A	Refractory IBD 4 CD, 3 UC	7	Allo-MSCs IV	Steroids and/or immunosuppressants	Mean 19 months	Diarrhea frequency and abdominal pain/cramps gradually improved in all the seven patients, and the index score decreases.	[174]
Forbes G. M. 2013 Australia	II	Luminal CD	16	Allo-MSCs Local	Corticosteroid, AZA	42 days	In a phase 2 study, administration of allogeneic MSCs reduced CDAI and CDEIS scores in patients with refractory luminal CD.	[112]
GarciaOlmo D. 2003, Spain	II	RFCD	1	ASCs Local	Olsalazine	3 months	Since the surgical procedure 3 month ago the patient has not experienced vaginal flatus or fecal incontinence through her vagina.	[175]
GarciaOlmo D. 2009, Spain	II	Complex PFCD	24	ASCs Local	Fibrin glue	1 year	Combination therapy appears to achieve higher rates of healing than fibrin glue alone.	[176]
Guadalajara H. 2011, Spain	II	PFCD	49	ASCs Local	Fibrin glue	3 years	A low proportion of the stem cell-treated patients with closure after the procedure. Remained free of recurrence after more than 3 years of follow-up.	[177]
Herreros M. D. 2012, Spain	III	Complex Anal fistula	200	ASCs Local	Fibrin glue	1 year	Achieving healing rates of approximately 40% at 6 months and of more than 50% at 1-year follow-up.	[178]
Dietz A. B. 2017 USA	I	Perianal CD	12	Auto-ASCs. Local	Nothing	<6 month	10/12 (83%) achieved combined remission at 24 weeks by fistula exam and MRI.	[179]
Dige A. 2019 Denmark	I	Perianal CD	21	Auto-ASCs. Repeatable Local	Nothing	<6 month	12/21 (57%) achieved clinical remission 6 months after last injection. 8/9 patient who underwent MRI had complete resolution at 6 months.	[180]
Lightner A. L. 2020 USA	I	Rectovaginal CD	5	Auto-ASCs. Local	Nothing	<6 month	3/5 (60%) with complete clinical response. 2/5 (40%) with partial clinical response. 0/5 patients with radiographic remission	[181]

(Continued)

Table 3 (Continued).

Author, Time and Country	Phase	Clinical Indication	Number	SC Sources Route	Adjuvant Therapy	Follow-Up Period	Primary Outcomes	Ref.
Nikolic M. 2021 Austria	I	Rectovaginal CD	4	Allo-ASCs. Local	Nothing	<6 month	Fistula closure and absence of drainage on exam at 6 months. 1/4 (25%) achieved clinical healing at 6 months.	[182]
Cho Y. B. 2015, Korea	II	Perianal CD	41	Auto-ASCs. Local	Nothing	>6 month	At 24 months, complete healing was observed in 21/26 (80.8%) patients in mPP group and 27/36 (75%) in mITT group.	[183]
Ciccocioppo R. 2015, Italy	I	Perianal CD	10	Auto-BM-ASCs. Local	Nothing	>6 month	Fistula relapse-free survival was 88% at 1 year, 50% at 2 years, and remaining at 37% for the remainder of the six-year follow-up. Mean CDAI score decreased from 300 to 150 at 6 years.	[184]
García-Olmo D. 2015, Spain	I	CD (3 with Perianal CD)	10	Auto-ASCs. Local / IV.	Nothing	>6 month	2/3 (66%) of CD patients achieved complete healing (re-epithelialization and absence of suppuration at 1 year) at 1 year.	[185]
Park K. J. 2016, Korea	I	Perianal CD	6	Auto-ASCs. Local	Nothing	>6 month	2/3 (66.7%) in group 1 and 1/3 (33.3%) in group 2 achieved clinical healing at 8 months.	[186]
García-Arranz M. 2016, Spain	I/II	Rectovaginal CD	10	Auto-ASCs. Local	Nothing	>6 month	Of 5 total patients who completed study, 3 achieved clinical healing (60%) (re-epithelialization of both vaginal and rectal sides and absence of drainage) at 52 weeks.	[187]
Panés J. 2018 Austria	III	Perianal CD	212	Auto-ASCs (107 Local) Placebo (105)	Nothing	>6 month	Using mITT, 58/103 (56.3%) of patients in treatment arm achieved combined remission (absence of external openings on exam and absence of collections > 2 cm on MRI) vs 39/101 (38.6%) in control arm at week 52.	[157]

(Continued)

Table 3 (Continued).

Author, Time and Country	Phase	Clinical Indication	Number	SC Sources Route	Adjuvant Therapy	Follow-Up Period	Primary Outcomes	Ref.
Wainstein C. 2018, Chile	I	Perianal CD	9	Auto-ASCs Local	Nothing	>6 month	8/9 (88.9%) patients with complete healing (absence of suppuration from the external fistula opening and complete epithelialization), 1/9 (11.1%) patients with partial healing at median follow-up of 31 months.	[188]
Knyazev O. V. 2018 Russia	II	Perianal CD	36	Auto-BM-MSCs Local	Nothing	>6 month	At 3 and 6 months, healing of 66.6% (8/12) in group 1, 60% (6/10) in group 2, and 7.14% (1/14) in group 3. At 1-year, healing of 58.3% (7/12) in group 1, 60% (6/10) in group 2, and 14.3% (2/14) in group 3.	[189]
Herreros M. D. 2019, Spain	I/II	CD (18 with perianal CD)	45	Auto-ASCs allo-ASCs Local / IV	Nothing	>6 month	55.5% of CD patients achieved healing (absence of suppuration). CD patients receiving SVF, auto-ASCs and allo-ASCs were cured by 40%, 66.6% and 55.5% respectively.	[190]
Barnhoorn M. C. 2020, The Netherlands	I	Perianal CD	21	Auto-BM-MSCs Local	Nothing	>6 month	Group 1: 3/4 (75%) healing at 4 years. Group 2: 4/4 (100%) healing at 4 years. Group 3: 2/5 (20%) healing at 4 years. Group 4: 0/3 (0%) healing at 4 years.	[145]
Zhou C. 2020, China	II	Perianal CD	22	Auto-ASCs Local	Nothing	>6 month	3 months: 10/11 (91%) in treatment arm vs 5/11 (45.5%) in placebo arm. 6 months: 8/11 (72.7%) in treatment arm vs 6/11 (54.5%) in placebo arm. 12 months: 7/11 (63.6%) in treatment arm vs 6/11 (54.5%) in placebo arm.	[191]
Laureti, S. 2020, Italy	II	Perianal CD	15	ASCs Lipogems® Local	Nothing	>6 month	10/15 (66.7%) with combined remission (closure of all external openings on exam and absence of collections > 3mm on MRI) at 24 months.	[192]

(Continued)

Table 3 (Continued).

Author, Time and Country	Phase	Clinical Indication	Number	SC Sources Route	Adjuvant Therapy	Follow-Up Period	Primary Outcomes	Ref.
Melmed G. Y. 2015, Italy	Ib/Ila	Luminal CD	50	PDA-001 IU, 4U IV	Nothing	>6 month	Decrease in CDAI by $\geq 100$ points and/or 25% from baseline at weeks 4 and 6. Phase I b: primary efficacy not reported Phase IIb: Placebo: 0/16 1U PDA-001: 5/15 (33%) 4U PDA-001: 5/13 (38.5%)	[193]
Dhere T. 2016, USA	I	Luminal CD	12	Auto- BM- MSCs IV	N/A	>6 month	5/11 (45.4%) achieved clinical response (decrease in CDAI by $\geq 100$ points at 2 weeks).	[194]
Hu J. 2016, China	I/II	With UC	70	UCBSCs IV	N/A	>6 month	29/34 (85.3%) with clinical response (decrease in total Mayo UC activity score of $\geq 3$ and $\geq 30\%$ from baseline) in group I vs 6/36 (16.7%) at 3 months.	[154]
Knyazev O. V. 2016 Russia	I	With UC	22	Auto-BM- MSCs IV	AZA	>6 month	Remission rate of 50% (6/12) in treatment group vs 10% (1/10) in control group at 3 years. Remission duration of 22 months in treatment group vs 20 months in control group at 3 years.	[195]
Gregoire C. 2018 Belgium	I/II	Luminal CD	13	Auto-BM- MSCs IV.	N/A	>6 month	2/13 (15.4%) with clinical response (decrease in CDAI by $\geq 100$ points) at 8 weeks.	[153]
Zhang J. 2018, China	I	Luminal CD	82	UCBSCs IV.	Corticosteroid	>6 month	Decrease in CDAI, HBI, and corticosteroid usage.	[111]
Knyazev O V. 2018 Russia	I/II	Luminal CD	34	BM-MSCs IV.	AZA	>6 month	Clinical remission (CDAI < 150) at 12 months.	[113]

**Abbreviations:** Allo, Allogeneic; ASCs, adipose mesenchymal stem cells; Auto, Autogenous; BM-MSCs, Bone marrow mesenchymal stem cells; CD, Crohn's disease; CDAI, Crohn's disease activity index; CDEIS, Crohn's Disease Endoscopic Index of Severity; HBI, Harvey-Bradshaw index; HSCT, hematopoietic stem cell transplantation; IBD, Inflammatory bowel disease; IL-10, interleukin-10; IP, intraperitoneal injection; IV, intravenous injection; mPP, Massively Parallel Processing; MRI, Magnetic resonance imaging; MSCs, mesenchymal stem cells; MSCT, mesenchymal stem cells transplantation; PBSCT, Peripheral blood stem cell transplantation; PDA-001, Human placenta-derived cells; PFCD, perianal fistula Crohn's disease; RFCD, rectovaginal fistula CD; SCs, stem cells; SVF, Stromal Vascular Fraction; UC, Ulcerative colitis; UCBSCs, Umbilical Cord Blood stem cells; CTX, Cyclophosphamide; ATG, antithymocyte globulin; F-ara-A, fludarabine; AZA, azathioprine; N/A, Not provided.

## Ongoing Clinical Studies of SCs for IBD Treatment

While there have been some completed clinical trials investigating the use of MSCs in the treatment of IBD, there are a number of limitations to these studies. For instance, there are few double-blind RCTs with relatively small sample sizes, and the standards used across these trials have not been uniform, which makes it difficult to draw clear and convincing conclusions. Although local injection of MSCs has shown promise in treating PFCD, intravenous injection of MSCs for refractory intestinal IBD has not yet produced the desired effect, and the efficacy of some trials remains uncertain.<sup>19</sup> In order to address these limitations, ongoing and future clinical trials should focus on improving treatment strategies and

standardizing protocols. Key technologies, such as optimizing the extraction and culture process of SCs, will help to ensure the best possible SC preparation.<sup>114</sup> It is important to clearly define the indications for SC therapy, determine the optimal route and dosage, and gradually expand the use of SC therapy for treating IBD. It is hoped that ongoing clinical trials investigating MSCs will provide clearer answers. The ongoing clinical trials investigating MSCs for IBD are summarized in Table 4.

The emergence of ISCs and iPSCs represents a promising avenue for future research, and clinical trials will be conducted gradually.<sup>115,116</sup> Recently, the research team at Tokyo Medical and Dental University has reported the

**Table 4** The Ongoing Clinical Trials Investigating MSCs for IBD

NCT Number	Sponsor Country	Phase	Clinical Indication	SC Sources	MSC Dose (Cells)	Administration Route	Status
03299413	Jordan	I/II	IBD	W. Jelly MSCs	$1.2 \times 10^9$ cells	iv.	Active
03115749	Montpellier	N/A	IBD	Intestinal MSCs	N/A	N/A	No recruiting
01914887	Spain	I/II	UC	Allogeneic ASCs	$6 \times 10^7$ cells	Colonoscope	Recruiting
01874015	Spain	I	CD	BM-MSCs	N/A	N/A	Recruiting
01157650	USA	I/ II	CD	Autologous MSCs	N/A	N/A	Completed
00294112	USA	II	CD	Adult human MSCs	$2, 8 \times 10^6$ cells	iv.	Completed
02677350	USA	I	CD	BM-MSCs	$2 \times 10^7$ cells	iv.	Withdrawn
02445547	China	I/II	CD	UCBSCs	$1 \times 10^6$ cell/kg	iv.	Completed
00543374	USA	III	CD	PROCHYMAL <sup>®</sup>	$0.6-1.2 \times 10^9$ cells	iv.	Completed
00482092	USA	III	CD	MSCs	$0.6-1.2 \times 10^9$ cells	iv.	Completed
01540292	Belgium	I/II	CD	MSCs	$1.5-2 \times 10^6$ cell/kg	iv.	Unknown status
04519671	USA	I/II	CD	BM-MSCs	$7.5 \times 10^7$ cells	iv.	Recruiting
04519684	USA	I/II	CD	BM-MSCs	$7.5 \times 10^7$ cells	iv.	Recruiting
01144962	USA	I/II	PFCD	BM-MSCs	$1-9 \times 10^7$ cells	Local injection	Completed
04519697	Nether-land	I/II	PFCD	MSCs	$7.5 \times 10^7$ cells	Local injection	Recruiting
04073472	USA	I	PFCD	BM-MSCs	$6 \times 10^7$ cells	Local injection	No recruiting
04548583	USA	I/II	CD	BM-MSCs	$1.5-3 \times 10^8$ cells	Endoscopic	Recruiting
03183661	China	I	CD	Allogenic-ASCs	$2, 8 \times 10^6$ cell/ kg	iv.	Enrolling
01221428	Austria	I/II	UC	UCBSCs	$2 \times 10^7$ cells	iv.	Unknown
01541579	Austria	III	PFCD	ASCs	$1.2 \times 10^8$ cells	Local injection	Completed
04312113	USA	I	UC	Autologous-ASCs	$1.5-3 \times 10^7$ cells	Intraarterial	Recruiting
04543994	USA	I/II	UC	BM-MSCs	$1.5, 3 \times 10^8$ cells	Endoscopic	Recruiting
01233960	USA	III	UC	MSCs	$2 \times 10^8$ cells	Intraarterial	Completed

(Continued)

Table 4 (Continued).

NCT Number	Sponsor Country	Phase	Clinical Indication	SC Sources	MSC Dose (Cells)	Administration Route	Status
03609905	China	I/II	UC	ASCs	$5 \times 10^7$ cells	Colonoscope	Recruiting
03901235	Belgium	I/II	N/A	MSCs	N/A	Local injection	Recruiting
02442037	China	I/II	UC	UCBSCs	$1 \times 10^6$ cell/kg	iv.	Recruiting
02580617	Korea	I	CD	ASCs	$0.15-1 \times 10^8$ cells	N/A	Recruiting
01510431	USA	N/A	CD	MSCs	$2 \times 10^8$ cells	N/A	Not available
02403232	Italy	II	CD	ASCs	N/A	N/A	Recruiting
02403232	Korea	I/II	CD	UCBSCs	0.5, $1 \times 10^8$ cells	iv.	Unknown status
02926300	Korea	I/II	CD	SCs	Not provided.	N/A	Recruiting
03220243	USA	I	PFCD	MSCs	$2 \times 10^7$ cells	Local injection	Completed
01915927	USA	I	PFCD	MSCs	$2 \times 10^7$ cells	Local injection	Completed
03449069	USA	I	PFCD	MSCs	$2 \times 10^7$ cells	Local injection	Recruiting

**Abbreviations:** ASCs, adipose mesenchymal stem cells; BM-MSCs, Bone marrow mesenchymal stem cells; CD, Crohn's disease; IBD, Inflammatory bowel disease; iv., intravenous injection; MSCs, mesenchymal stem cells; PFCD, perianal fistula Crohn's disease; PROCHYMAL<sup>®</sup>, Human Adult stem cells; SCs, stem cells; UC, Ulcerative colitis; UCBSCs, Umbilical Cord Blood stem cells; USA, United States, N/A, Not provided, WJ, Jelly, Wharton Jelly.

successful transplantation of the world's first "organoid" into the intestinal mucosa of a refractory UC patient, marking the beginning of the organoid SC era. The team collected normal intestinal mucosa (including ISCs) from the patient under endoscopy, cultured it in vitro for approximately 1 month, and constructed an "organoid" with a diameter of 0.1–0.2 mm. The "organoid" is then transplanted to the intestinal lesion site under endoscopy and fixed in place with a degradable film. According to reports, the patient's intestinal symptoms have improved, and they are in good condition.

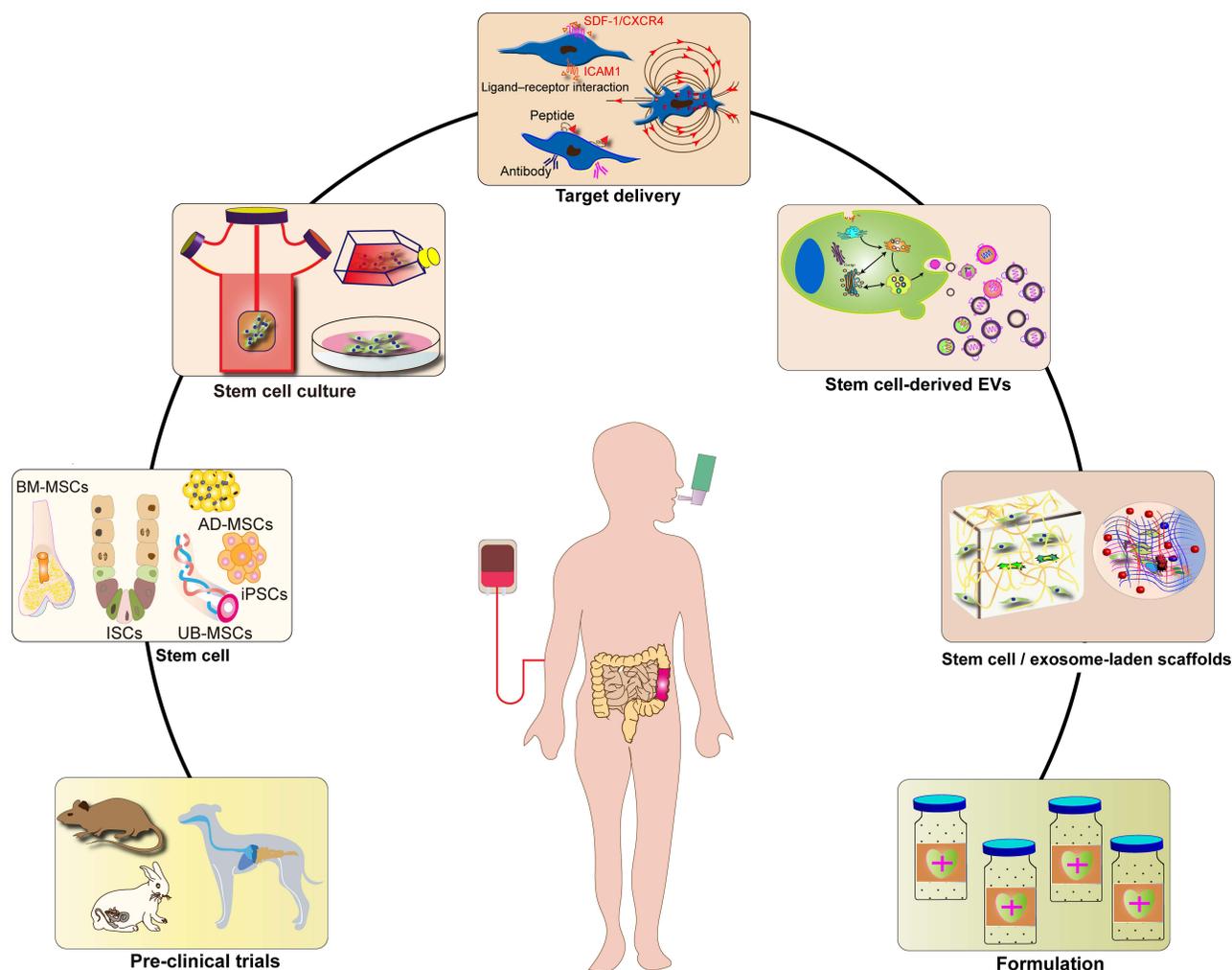
## Improvement Strategies of SC-Based IBD Therapies

Clinical trials have provided valuable insights into the potential applications and efficacy of SC therapy for IBD. However, it is important to acknowledge the limitations of current SC therapy, including its efficacy, technology, and safety.<sup>117</sup> When administered intravenously or intraperitoneally, only a small number of SCs home to intestinal lesions, with their effect primarily dependent on their secretory function. The secretion of SCs is also greatly influenced by their "niche" environment. Obtaining a large number of high-quality, homogeneous SC preparations remains technically challenging. Safety concerns associated with SC therapy include microcirculatory and coagulation dysfunction, immune rejection, and the risk of tumor formation.<sup>96,114</sup>

To enhance the use of SC therapy in treating IBD, several modifications have been proposed, such as changes to the culture conditions and cultivation methods for SCs, modification of SC contents and genetic genes, and extraction of SC derivatives. These strategies have the potential to optimize SC therapy and improve its efficacy and safety in treating IBD.<sup>114</sup> Figure 2 illustrates some of these improvement strategies for SC-based IBD therapies.

### Box 1 Changing SC Culture for IBD 3D Culture Method

Cells can migrate and grow in 3D culture vectors made from different materials, forming 3D cell-vector complexes.<sup>114,118</sup> The 3D culture environment provides a better simulation of the in vivo SC growth environment and can more effectively regulate cell proliferation and differentiation. This allows for the cultivation of more cell types and the formation of tissue structures similar to those found in vivo.<sup>119</sup> For instance, human ISCs can be cultured in 3D to generate intestinal



**Figure 2** Stem cell-based therapy strategies for inflammatory bowel diseases.

epithelial organs, which can be used as IBD disease models. The introduction of human ISCs through the anus can also improve the targeting and colonization rate of SCs, further increasing the therapeutic potential.<sup>104</sup>

In another example, MSCs are cultured in 3D microsphere scaffolds constructed from poly(lactic-co-glycolic acid) (PLGA), resulting in improved viability of the MSCs. The addition of a broad-spectrum caspase inhibitor (IDN6556) to the 3D medium further increases the cell viability of MSCs, which is validated by intraperitoneal injection into an IBD mouse model.<sup>120</sup>

### Changing Culture Conditions

The cultivation of SCs not only requires media but also an environment that mimics their *in vivo* growth. The proliferation and differentiation of SCs can be induced purposefully by adjusting the chemical, physical, or microbial environment of the culture medium, which enhances the therapeutic effects of SCs in IBD.<sup>90,121</sup> For instance, in one study, various exosome-free fetal bovine sera (exosome-free FBS) were added to the medium of MSCs, resulting in enhanced immunomodulatory effects of MSCs in an animal model of IBD.<sup>122,123</sup> The addition of drugs or biomolecules, such as aspirin, B fibroblast growth factor (FGF), all-trans retinoic acid (ATRA), activin A, and FGF2, to the culture medium has also been reported to enhance the beneficial effects of MSC treatment.<sup>124,125</sup> Furthermore, the addition of pro-inflammatory cytokines, such as IFN- $\gamma$ , TNF- $\alpha$ , and IL-1 $\beta$ , to the culture medium has been shown to increase the immunosuppressive function of MSCs.<sup>126,127</sup>

## Box 2 Modification of SCs for IBD

### Genetic Modification

To increase the function and efficacy of SCs, specific functional genes can be introduced into them through gene transfer viral vectors. For instance, IL-10 can be genetically delivered to MSCs, resulting in MSCs with IL-10 overexpression that enhance immunosuppression and tissue repair.<sup>128,129</sup> Additionally, genes expressing IL-37b can be delivered to MSCs, and MSCs expressing IL-37b can inhibit the production of pro-inflammatory cytokines and chemokines, as well as suppress tissue migration and infiltration of neutrophils.<sup>130</sup> In mouse models of IBD, both MSCs expressing IL-35 and MSCs expressing IL-25 have been shown to increase the therapeutic effect of MSC therapy.<sup>131,132</sup> The therapeutic effect of MSCs can also be increased by gene transduction to produce miR-181a overexpressing MSCs, where the secreted miR-181a can regulate the development of T and B cells.<sup>133</sup> Furthermore, there are increasing attempts to apply CRISPR-Cas9-mediated gene modifications to the therapeutic field of MSCs, which can enhance the ability of MSCs to regulate immunity.<sup>134,135</sup>

### Adding Surface Markers

Surface markers play a crucial role in the recognition and targeting of SCs. By introducing cellular markers to SCs or exosomes, the recognition of IBD lesion sites can be enhanced, and the targeting of SCs can be improved, thereby increasing the homing ability of SCs or exosomes to specific sites. For instance, in one study, MSCs were incubated with antibodies directed against vascular cell adhesion molecules (VCAM1), which resulted in an increased therapeutic effect of VCAM1-treated MSCs as they were found to migrate more effectively to the injured colon.<sup>136,137</sup> In another study, pretreating MSCs with stromal-derived factor 1 (SDF-1) induced an increased expression of CXCR4 chemokine receptors, allowing more MSCs to colonize the injured colon more rapidly, thereby enhancing their therapeutic effect.<sup>138</sup> Additionally, the use of intercellular adhesion molecule 1 (ICAM1) overexpressing MSCs in an IBD mouse model is designed to direct infusion of ICAM1-MSCs to inflamed intestinal tissues and spleens, thereby increasing their therapeutic efficacy.<sup>139</sup>

## Box 3 SCs Derivatives for IBD

The use of MSC-derived extracellular vesicles (EVs), particularly exosomes, as a substitute for MSC therapy has been validated in animal experiments, and clinical trials have reported comparable therapeutic outcomes.<sup>98,140</sup> Mechanistic studies have revealed that MSCs act as immune modulators or repairers of the intestinal mucosal barrier by targeting immune cells or structural cells at sites of intestinal inflammation, mainly through paracrine EVs that carry a large number of bioactive molecules.<sup>141</sup> The extraction of MSC-derived EVs is an attractive strategy for MSC-based therapy. EVs offer the advantages of cell-free therapy, reducing the potential immune rejection of MSCs and the risk of tumor formation. Furthermore, EVs have the ability to cross multiple physiological barriers in the body, act as carrier vesicles, and better preserve the formulation.<sup>142,143</sup> The study conducted by Yang et al involved the intraperitoneal injection of exosome preparations derived from the supernatant of human UC-MSC cultures into IBD model mice. Through carrying TSG-6, MSC-derived EVs repair the mucosal barrier and regulate intestinal immunity, significantly improving IBD symptoms and reducing mortality.<sup>144</sup>

## Conclusions and Future Perspectives

Although current treatments for IBD can provide temporary relief from clinical symptoms, achieving sustained remission and MH remains a challenge. SCs have the potential to directly improve chronic inflammation in the intestine by modulating immune cells and repairing the intestinal mucosal barrier. Furthermore, SCs can indirectly improve intestinal inflammatory damage and prevent IBD complications by positively impacting intestinal microecology, microcirculatory disorders, fibrosis, and carcinogenesis. The use of SCs in IBD treatment is expected to achieve sustained remission and MH. Preclinical studies of SCs in IBD models are increasing, and various SCs, such as MSCs, HSCs, ISCs, and iPSCs, have demonstrated positive efficacy and stable safety in IBD animal models, making them suitable for further clinical studies. Although HSCT can significantly improve IBD in clinical studies, its safety and cost-effectiveness are low, and it is not a viable option for the treatment of IBD alone. However, most clinical trials of MSCs applied to refractory IBD

have reported positive efficacy and can achieve durable remission, especially in patients with refractory PFCD, confirming their safety and effectiveness. Additionally, clinical trials involving MSCs-derived EVs are underway. With the advancement of key technologies, SCs, such as ISCs and iPSCs, are gradually being applied in clinical studies for IBD treatment. Intestinal organoids derived from SCs in 3D culture are also being investigated in clinical research on IBD.<sup>104</sup>

However, significant progress is still needed to achieve safe, efficient, and standardized use of SC therapy for IBD. Firstly, achieving homogeneity in SCs is crucial. SCs are influenced by tissue sources and culture environments, leading to significant heterogeneity. Currently, there are no uniform standards and control comparisons in the tissue source and culture of SCs, making it challenging to achieve homogeneity in screening IBD patients. This heterogeneity affects the consistency of IBD clinical studies, leading to significant individual differences in study results and obstacles to sharing and applying results, ultimately limiting clinical translation. Secondly, the potential hazards of SC therapy must be considered. Compared to chemical drugs or biological agents, SCs are complex, and much is still unknown about their structure and function. Current observations of the effects of SCs *in vivo* are limited to particular diseases, and the full extent of their effects *in vivo* is not yet fully understood. These potential therapeutic risks limit clinical translation. Thirdly, standardization of SC therapy is necessary. Currently, there are no uniform standards regarding infusion mode, infusion dose, and time interval of SCs, and clear indications and contraindications have yet to be established.<sup>19</sup>

Currently, studies on SCs in IBD are primarily focused on the preclinical phase, with clinical applications limited to localized fistulas and refractory IBD. However, as the technology for SCs applications continues to mature, SCs are poised to become a routine “weapon” in clinical treatment and can be applied to general IBD patients in the future. Inflammation, injury, and neoplastic lesions of the digestive tract can all be targets for SC treatment.

## Abbreviations

ASCs, adipose MSCs; ASTIC study, autologous stem cell international Crohn’s disease trial; ATRA, all-trans retinoic acid; BM-MSCs, Bone marrow MSCs; CD, Crohn’s disease; CDAI, Crohn’s disease activity index; CRISPR-Cas9, Clustered Regularly Interspersed Short Palindromic Repeats-Cas9; CXCR4, CXC chemokine receptor 4; DCs, Dendritic cells; ECs, endothelial cells; EMT, epithelial-mesenchymal transition; ESCs, embryonic SCs; EV, extracellular vesicle; FGF, fibroblast growth factor; HSCs, hematopoietic stem cells; HSCT, HSCs transplantation; IBD, Inflammatory bowel disease; ICAM1, intercellular adhesion molecule 1; ICAM1-MSCs, intercellular adhesion molecule 1-overexpressing MSCs; IDO, indoleamine 2,3-dioxygenase; IECs, intestinal epithelial cells; IFN- $\gamma$ , interferon- $\gamma$ ; IL-10, interleukin-10; IL-12, interleukin-12; IL-17, interleukin-17; IL-1 $\beta$ , interleukin-1 $\beta$ ; IL-23, interleukin-23; IL-37b, interleukin-37b; iMSCs, MSCs derived from iPSCs; iPSCs, IPSCs; ISCs, ISCs; LEC, lymphatic endothelial cell; Lgr5+, leucine rich repeat containing G protein coupled receptor 5; LIF, leukemia inhibitory factor; MNM, modified neuronal medium; MSCs, Mesenchymal SCs; MSCT, MSCs transplantation; MtSCs, intestinal resident MSCs; PBSCs, Peripheral blood SCs; PDGF, platelet-derived growth factor; pESCs, Parthenogenetic embryonic SCs; PFCD, perianal fistula Crohn’s disease; PGE2, prostaglandin E2; PLGA, poly lactic-co-glycolic acid; SCs, stem cells; SDF-1, stromal derived factor 1; STAT-3, signal transducer and activator of translation-3; tDC, tolerogenic dendritic cell; TGF- $\beta$ , transforming growth factor- $\beta$ ; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ ; TSG-6, tumor necrosis factor  $\alpha$  stimulated gene-6; UC, Ulcerative colitis; UCBSCs, Umbilical Cord Blood SCs; VCAM1, vascular cell adhesion molecule 1; VEGF, vascular endothelial growth factor.

## Acknowledgments

The authors thank prof. Yu-Qiang Nie for editing the manuscript.

## Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

## Funding

This work was supported by Science and Technology Innovation Committee of Shenzhen (No. JCYJ20200109150700942, No. JCYJ20150403101028164, No. JCYC20170307100911479, No. JCYJ20190807145617113 and No. JCYJ20210324113802006), the Guangdong Basic and Applied Basic Research Foundation (No. 2020A1515011581) and National Natural Science Foundation of China (No. 81800489).

## Disclosure

The authors report no conflicts of interest in this work.

## References

1. Ramos GP, Papadakis KA. Mechanisms of disease: inflammatory bowel diseases. *Mayo Clin Proc.* 2019;94(1):155–165. doi:10.1016/j.mayocp.2018.09.013
2. Lightner AL. Stem cell therapy for inflammatory bowel disease. *Clin Transl Gastroenterol.* 2017;8(3):e82. doi:10.1038/ctg.2017.7
3. Guan Q. A comprehensive review and update on the pathogenesis of inflammatory bowel disease. *J Immunol Res.* 2019;2019:7247238. doi:10.1155/2019/7247238
4. Kaplan GG, Windsor JW. The four epidemiological stages in the global evolution of inflammatory bowel disease. *Nat Rev Gastroenterol Hepatol.* 2021;18(1):56–66. doi:10.1038/s41575-020-00360-x
5. Coward S, Clement F, Benchimol EI, et al. Past and future burden of inflammatory bowel diseases based on modeling of population-based data. *Gastroenterology.* 2019;156(5):1345–1353.e1344. doi:10.1053/j.gastro.2019.01.002
6. Ng SC, Shi HY, Hamidi N, et al. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet.* 2017;390(10114):2769–2778. doi:10.1016/s0140-6736(17)32448-0
7. Kaplan GG. The global burden of IBD: from 2015 to 2025. *Nat Rev Gastroenterol Hepatol.* 2015;12(12):720–727. doi:10.1038/nrgastro.2015.150
8. Rogler G, Vavricka S, Schoepfer A, Lakatos PL. Mucosal healing and deep remission: what does it mean? *World J Gastroenterol.* 2013;19:7552–7560. doi:10.3748/wjg.v19.i43.7552
9. Hommes D, Colombel JF, Emery P, Greco M, Sandborn WJ. Changing Crohn's disease management: need for new goals and indices to prevent disability and improve quality of life. *J Crohns Colitis.* 2012;6 Suppl 2:S224–S234. doi:10.1016/s1873-9946(12)60502-9
10. Abraham BP, Ahmed T, Ali T. Inflammatory bowel disease: pathophysiology and current therapeutic approaches. *Handb Exp Pharmacol.* 2017;239:115–146. doi:10.1007/164\_2016\_122
11. Wright EK, Ding NS, Niewiadomski O. Management of inflammatory bowel disease. *Med J Aust.* 2018;209(7):318–323. doi:10.5694/mja17.01001
12. Zhang HM, Yuan S, Meng H, et al. Stem cell-based therapies for inflammatory bowel disease. *Int J Mol Sci.* 2022;23. doi:10.3390/ijms23158494
13. Grim C, Noble R, Uribe G, et al. Impairment of tissue-resident mesenchymal stem cells in chronic ulcerative colitis and crohn's disease. *J Crohns Colitis.* 2021;15(8):1362–1375. doi:10.1093/ecco-jcc/ijab001
14. Huang Y, Wu Q, Tam PKH. Immunomodulatory mechanisms of mesenchymal stem cells and their potential clinical applications. *Int J Mol Sci.* 2022;23. doi:10.3390/ijms231710023
15. Eiro N, Fraile M, González-Jubete A, González LO, Vizoso FJ. Mesenchymal (stem) stromal cells based as new therapeutic alternative in inflammatory bowel disease: basic mechanisms, experimental and clinical evidence, and challenges. *Int J Mol Sci.* 2022;23(16):8905. doi:10.3390/ijms23168905
16. Mishra R, Dhawan P, Srivastava AS, Singh AB. Inflammatory bowel disease: therapeutic limitations and prospective of the stem cell therapy. *World J Stem Cells.* 2020;12(10):1050–1066. doi:10.4252/wjsc.v12.i10.1050
17. Che Z, Ye Z, Zhang X, et al. Mesenchymal stem/stromal cells in the pathogenesis and regenerative therapy of inflammatory bowel diseases. *Front Immunol.* 2022;13:952071. doi:10.3389/fimmu.2022.952071
18. Artinger KB, Watanabe M. Introduction to “stem cells” special issue. *Birth Defects Res.* 2022;114:921–925. doi:10.1002/bdr2.2061
19. Ko JZ-H, Johnson S, Dave M. Efficacy and safety of mesenchymal stem/stromal cell therapy for inflammatory bowel diseases: an up-to-date systematic review. *Biomolecules.* 2021;11(1):82. doi:10.3390/biom11010082
20. Sherman SE, Agudo J. Immune-mediated specific depletion of intestinal stem cells. *Methods Mol Biol.* 2020;2171:25–39. doi:10.1007/978-1-0716-0747-3\_2
21. Kolios G, Moodley Y. Introduction to stem cells and regenerative medicine. *Respiration.* 2013;85(1):3–10. doi:10.1159/000345615
22. Biehl JK, Russell B. Introduction to stem cell therapy. *J Cardiovasc Nurs.* 2009;24(2):98–105. doi:10.1097/JCN.0b013e318197a6a5
23. da Silva Meirelles L, Chagastelles PC, Nardi NB. Mesenchymal stem cells reside in virtually all post-natal organs and tissues. *J Cell Sci.* 2006;119(11):2204–2213. doi:10.1242/jcs.02932
24. Goodell M. Introduction to a review series on hematopoietic stem cells. *Blood.* 2015;125(17):2587. doi:10.1182/blood-2015-03-615005
25. DiNicola CA, Zand A, Hommes DW. Autologous hematopoietic stem cells for refractory Crohn's disease. *Expert Opin Biol Ther.* 2017;17(5):555–564. doi:10.1080/14712598.2017.1305355
26. Ruiz MA, Junior RLK, Piron-Ruiz L, et al. Medical, ethical, and legal aspects of hematopoietic stem cell transplantation for Crohn's disease in Brazil. *World J Stem Cells.* 2020;12(10):1113–1123. doi:10.4252/wjsc.v12.i10.1113
27. Balassa K, Danby R, Rocha V. Haematopoietic stem cell transplants: principles and indications. *Br J Hosp Med.* 2019;80(1):33–39. doi:10.12968/hmed.2019.80.1.33
28. Rodríguez-Fuentes DE, Fernández-Garza LE, Samia-Meza JA, et al. Mesenchymal stem cells current clinical applications: a systematic review. *Arch Med Res.* 2021;52(1):93–101. doi:10.1016/j.arcmed.2020.08.006

29. Zhang X, Wang S, Ding X, Guo J, Tian Z. Potential methods for improving the efficacy of mesenchymal stem cells in the treatment of inflammatory bowel diseases. *Scand J Immunol.* 2020;92(3):e12897. doi:10.1111/sji.12897
30. Mao F, Wu Y, Tang X, et al. Exosomes derived from human umbilical cord mesenchymal stem cells relieve inflammatory bowel disease in mice. *Biomed Res Int.* 2017;2017:5356760. doi:10.1155/2017/5356760
31. Sarsenova M, Kim Y, Raziyeva K, et al. Recent advances to enhance the immunomodulatory potential of mesenchymal stem cells. *Front Immunol.* 2022;13:1010399. doi:10.3389/fimmu.2022.1010399
32. de Wolf C, van de Bovenkamp M, Hoefnagel M. Regulatory perspective on in vitro potency assays for human mesenchymal stromal cells used in immunotherapy. *Cytotherapy.* 2017;19(7):784–797. doi:10.1016/j.jcyt.2017.03.076
33. Umar S. Intestinal stem cells. *Curr Gastroenterol Rep.* 2010;12(5):340–348. doi:10.1007/s11894-010-0130-3
34. Nalapareddy K, Zheng Y, Geiger H. Aging of intestinal stem cells. *Stem Cell Rep.* 2022;17(4):734–740. doi:10.1016/j.stemcr.2022.02.003
35. Liu J, Walker NM, Cook MT, Ootani A, Clarke LL. Functional Cfr in crypt epithelium of organotypic enteroid cultures from murine small intestine. *Am J Physiol Cell Physiol.* 2012;302(10):C1492–C1503. doi:10.1152/ajpcell.00392.2011
36. Barker N, van de Wetering M, Clevers H. The intestinal stem cell. *Genes Dev.* 2008;22(14):1856–1864. doi:10.1101/gad.1674008
37. Gehart H, Clevers H. Tales from the crypt: new insights into intestinal stem cells. *Nat Rev Gastroenterol Hepatol.* 2019;16(1):19–34. doi:10.1038/s41575-018-0081-y
38. Neal MD, Richardson WM, Sodhi CP, Russo A, Hackam DJ. Intestinal stem cells and their roles during mucosal injury and repair. *J Surg Res.* 2011;167(1):1–8. doi:10.1016/j.jss.2010.04.037
39. Guiu J, Hannezo E, Yui S, et al. Tracing the origin of adult intestinal stem cells. *Nature.* 2019;570(7759):107–111. doi:10.1038/s41586-019-1212-5
40. Tan DW, Barker N. Intestinal stem cells and their defining niche. *Curr Top Dev Biol.* 2014;107:77–107. doi:10.1016/b978-0-12-416022-4.00003-2
41. Jasper H. Intestinal stem cell aging: origins and interventions. *Annu Rev Physiol.* 2020;82(1):203–226. doi:10.1146/annurev-physiol-021119-034359
42. Sugimoto S, Sato T. Establishment of 3D intestinal organoid cultures from intestinal stem cells. *Methods Mol Biol.* 2017;1612:97–105. doi:10.1007/978-1-4939-7021-6\_7
43. Liang R, Xiao X, Luo L, et al. Efficient definitive endoderm differentiation from human parthenogenetic embryonic stem cells induced by activin A and Wnt3a. *Ann Clin Lab Sci.* 2020;50(4):468–473.
44. Liang R, Wang Z, Kong X, et al. Differentiation of human parthenogenetic embryonic stem cells into functional hepatocyte-like cells. *Organogenesis.* 2020;16(4):137–148. doi:10.1080/15476278.2020.1848237
45. Gonzalez R, Garitaonandia I, Crain A, et al. Proof of concept studies exploring the safety and functional activity of human parthenogenetic-derived neural stem cells for the treatment of Parkinson's disease. *Cell Transplant.* 2015;24(4):681–690. doi:10.3727/096368915x687769
46. Schickl H, Braun M, Dabrock P. Ways out of the patenting prohibition? Human parthenogenetic and induced pluripotent stem cells. *Bioethics.* 2017;31(5):409–417. doi:10.1111/bioe.12334
47. Ferreira AF, Calin GA, Picanço-Castro V, et al. Hematopoietic stem cells from induced pluripotent stem cells – considering the role of microRNA as a cell differentiation regulator. *J Cell Sci.* 2018;131(4). doi:10.1242/jcs.203018
48. Soontarak S, Chow L, Johnson V, et al. Mesenchymal Stem Cells (MSC) Derived from Induced Pluripotent Stem Cells (iPSC) equivalent to adipose-derived MSC in promoting intestinal healing and microbiome normalization in mouse inflammatory bowel disease model. *Stem Cells Transl Med.* 2018;7(6):456–467. doi:10.1002/sctm.17-0305
49. Spence JR, Mayhew CN, Rankin SA, et al. Directed differentiation of human pluripotent stem cells into intestinal tissue in vitro. *Nature.* 2011;470(7332):105–109. doi:10.1038/nature09691
50. Sagi I, Chia G, Golan-Lev T, et al. Derivation and differentiation of haploid human embryonic stem cells. *Nature.* 2016;532(7597):107–111. doi:10.1038/nature17408
51. Lu Y, Xu Y, Zhang S, et al. Human gingiva-derived mesenchymal stem cells alleviate inflammatory bowel disease via IL-10 signalling-dependent modulation of immune cells. *Scand J Immunol.* 2019;90(3):e12751. doi:10.1111/sji.12751
52. Otsuka R, Wada H, Murata T, Seino K-I. Immune reaction and regulation in transplantation based on pluripotent stem cell technology. *Inflamm Regen.* 2020;40(1):12. doi:10.1186/s41232-020-00125-8
53. Cao L, Xu H, Wang G, et al. Extracellular vesicles derived from bone marrow mesenchymal stem cells attenuate dextran sodium sulfate-induced ulcerative colitis by promoting M2 macrophage polarization. *Int Immunopharmacol.* 2019;72:264–274. doi:10.1016/j.intimp.2019.04.020
54. Wang J, Pei B, Yan J, et al. hucMSC-derived exosomes alleviate the deterioration of colitis via the miR-146a/SUMO1 axis. *Mol Pharm.* 2022;19(2):484–493. doi:10.1021/acs.molpharmaceut.1c00450
55. Jiang -X-X, Zhang Y, Liu B, et al. Human mesenchymal stem cells inhibit differentiation and function of monocyte-derived dendritic cells. *Blood.* 2005;105(10):4120–4126. doi:10.1182/blood-2004-02-0586
56. Heidari N, Abbasi-Kenarsari H, Namaki S, et al. Adipose-derived mesenchymal stem cell-secreted exosome alleviates dextran sulfate sodium-induced acute colitis by treg cell induction and inflammatory cytokine reduction. *J Cell Physiol.* 2021;236(8):5906–5920. doi:10.1002/jcp.30275
57. Mounayar M, Kefaloyianni E, Smith B, et al. PI3ka and STAT1 interplay regulates human mesenchymal stem cell immune polarization. *Stem Cells.* 2015;33(6):1892–1901. doi:10.1002/stem.1986
58. Qin Z, Wang P-Y, Wan -J-J, et al. MicroRNA124-IL6R mediates the effect of nicotine in inflammatory bowel disease by shifting Th1/Th2 balance toward Th1. *Front Immunol.* 2020;11:235. doi:10.3389/fimmu.2020.00235
59. Terraza-Aguirre C, Campos-Mora M, Elizondo-Vega R, et al. Mechanisms behind the immunoregulatory dialogue between mesenchymal stem cells and Th17 cells. *Cells.* 2020;9(7):1660. doi:10.3390/cells9071660
60. Reis M, Mavin E, Nicholson L, et al. Mesenchymal stromal cell-derived extracellular vesicles attenuate dendritic cell maturation and function. *Front Immunol.* 2018;9:2538. doi:10.3389/fimmu.2018.02538
61. Qian X, An N, Ren Y, et al. Immunosuppressive effects of mesenchymal stem cells-derived exosomes. *Stem Cell Rev Rep.* 2021;17(2):411–427. doi:10.1007/s12015-020-10040-7

62. Song W-J, Li Q, Ryu M-O, et al. Canine adipose tissue-derived mesenchymal stem cells pre-treated with TNF-alpha enhance immunomodulatory effects in inflammatory bowel disease in mice. *Res Vet Sci.* 2019;125:176–184. doi:10.1016/j.rvsc.2019.06.012
63. Li Y, Ma K, Zhang L, Xu H, Zhang N. Human umbilical cord blood derived-mesenchymal stem cells alleviate dextran sulfate sodium-induced colitis by increasing regulatory T cells in mice. *Front Cell Dev Biol.* 2020;8:604021. doi:10.3389/fcell.2020.604021
64. Cao X, Duan L, Hou H, et al. IGF-1C hydrogel improves the therapeutic effects of MSCs on colitis in mice through PGE 2 -mediated M2 macrophage polarization. *Theranostics.* 2020;10(17):7697–7709. doi:10.7150/thno.45434
65. García JR, Quirós M, Han WM, et al. IFN- $\gamma$ -tethered hydrogels enhance mesenchymal stem cell-based immunomodulation and promote tissue repair. *Biomaterials.* 2019;220:119403. doi:10.1016/j.biomaterials.2019.119403
66. Wang Y, Chen X, Cao W, Shi Y. Plasticity of mesenchymal stem cells in immunomodulation: pathological and therapeutic implications. *Nat Immunol.* 2014;15(11):1009–1016. doi:10.1038/ni.3002
67. Yang J, Liu XX, Fan H, et al. Extracellular vesicles derived from bone marrow mesenchymal stem cells protect against experimental colitis via attenuating colon inflammation, oxidative stress and apoptosis. *PLoS One.* 2015;10(10):e0140551. doi:10.3389/fimmu.2022.952071
68. Rager TM, Olson JK, Zhou Y, Wang Y, Besner GE. Exosomes secreted from bone marrow-derived mesenchymal stem cells protect the intestines from experimental necrotizing enterocolitis. *J Pediatr Surg.* 2016;51(6):942–947. doi:10.1016/j.jpedsurg.2016.02.061
69. Liang X, Zhang L, Wang S, Han Q, Zhao RC. Exosomes secreted by mesenchymal stem cells promote endothelial cell angiogenesis by transferring miR-125a. *J Cell Sci.* 2016;129(11):2182–2189. doi:10.1242/jcs.170373
70. Wang X, Wang H, Cao J, Ye C. Exosomes from adipose-derived stem cells promotes VEGF-C-dependent lymphangiogenesis by regulating miRNA-132/TGF- $\beta$  pathway. *Cell Physiol Biochem.* 2018;49:160–171. doi:10.1159/000492851
71. Guillén MI, Platas J, Pérez Del Caz MD, Mirabet V, Alcaraz MJ. Paracrine anti-inflammatory effects of adipose tissue-derived mesenchymal stem cells in human monocytes. *Front Physiol.* 2018;9:661. doi:10.3389/fphys.2018.00661
72. Komori M, Tsuji S, Tsujii M, et al. Involvement of bone marrow-derived cells in healing of experimental colitis in rats. *Wound Repair Regen.* 2005;13:109–118. doi:10.1111/j.1067-1927.2005.130114.x
73. Hayashi Y, Tsuji S, Tsujii M, et al. Topical implantation of mesenchymal stem cells has beneficial effects on healing of experimental colitis in rats. *J Pharmacol Exp Ther.* 2008;326:523–531. doi:10.1124/jpet.108.137083
74. Ahlawat S, Kumar P, Mohan H, Goyal S, Sharma KK. Inflammatory bowel disease: tri-directional relationship between microbiota, immune system and intestinal epithelium. *Crit Rev Microbiol.* 2021;47:254–273. doi:10.1080/1040841x.2021.1876631
75. Bailey CC, Zhong G, Huang IC, Farzan M. IFITM-family proteins: the cell's first line of antiviral defense. *Annu Rev Virol.* 2014;1:261–283. doi:10.1146/annurev-virology-031413-085537
76. Yagi H, Chen AF, Hirsch D, et al. Antimicrobial activity of mesenchymal stem cells against Staphylococcus aureus. *Stem Cell Res Ther.* 2020;11:293. doi:10.1186/s13287-020-01807-3
77. Alcayaga-Miranda F, Cuenca J, Martin A, et al. Combination therapy of menstrual derived mesenchymal stem cells and antibiotics ameliorates survival in sepsis. *Stem Cell Res Ther.* 2015;6:199. doi:10.1186/s13287-015-0192-0
78. Yang R, Liu Y, Kelk P, et al. A subset of IL-17(+) mesenchymal stem cells possesses anti-Candida albicans effect. *Cell Res.* 2013;23:107–121. doi:10.1038/cr.2012.179
79. Danese S, Sans M, de la Motte C, et al. Angiogenesis as a novel component of inflammatory bowel disease pathogenesis. *Gastroenterology.* 2006;130(7):2060–2073. doi:10.1053/j.gastro.2006.03.054
80. Manieri NA, Mack MR, Himmelrich MD, et al. Mucosally transplanted mesenchymal stem cells stimulate intestinal healing by promoting angiogenesis. *J Clin Invest.* 2015;125:3606–3618. doi:10.1172/jci81423
81. Tao H, Han Z, Han ZC, Li Z. Proangiogenic features of mesenchymal stem cells and their therapeutic applications. *Stem Cells Int.* 2016;2016:1314709. doi:10.1155/2016/1314709
82. Wang C, Li Y, Yang M, et al. Efficient differentiation of bone marrow mesenchymal stem cells into endothelial cells in vitro. *Eur J Vasc Endovasc Surg.* 2018;55(2):257–265. doi:10.1016/j.ejvs.2017.10.012
83. Wang Y, Huang B, Jin T, et al. Intestinal fibrosis in inflammatory bowel disease and the prospects of mesenchymal stem cell therapy. *Front Immunol.* 2022;13:835005. doi:10.3389/fimmu.2022.835005
84. Lovisa S, Genovese G, Danese S. Role of epithelial-to-mesenchymal transition in inflammatory bowel disease. *J Crohns Colitis.* 2019;13(5):659–668. doi:10.1093/ecco-jcc/jyy201
85. Lian L, Huang Q, Zhang L, et al. Anti-fibrogenic potential of mesenchymal stromal cells in treating fibrosis in crohn's disease. *Dig Dis Sci.* 2018;63(7):1821–1834. doi:10.1007/s10620-018-5082-8
86. Choi YJ, Koo JB, Kim HY, et al. Umbilical cord/placenta-derived mesenchymal stem cells inhibit fibrogenic activation in human intestinal myofibroblasts via inhibition of myocardin-related transcription factor A. *Stem Cell Res Ther.* 2019;10(1):291. doi:10.1186/s13287-019-1385-8
87. Srivastava AS, Feng Z, Mishra R, et al. Embryonic stem cells ameliorate piroxicam-induced colitis in IL10 $^{-/-}$  KO mice. *Biochem Biophys Res Commun.* 2007;361(4):953–959. doi:10.1016/j.bbrc.2007.07.139
88. Murry CE, Keller G. Differentiation of embryonic stem cells to clinically relevant populations: lessons from embryonic development. *Cell.* 2008;132(4):661–680. doi:10.1016/j.cell.2008.02.008
89. Wang M, Cai J, Huang F, et al. Pre-treatment of human umbilical cord-derived mesenchymal stem cells with interleukin-6 abolishes their growth-promoting effect on gastric cancer cells. *Int J Mol Med.* 2015;35(2):367–375. doi:10.3892/ijmm.2014.2019
90. Zheng X-B, He X-W, Zhang L-J, et al. Bone marrow-derived CXCR4-overexpressing MSCs display increased homing to intestine and ameliorate colitis-associated tumorigenesis in mice. *Gastroenterol Rep.* 2019;7(2):127–138. doi:10.1093/gastro/goy017
91. He R, Han C, Li Y, Qian W, Hou X. Cancer-preventive role of bone marrow-derived mesenchymal stem cells on colitis-associated colorectal cancer: roles of gut microbiota involved. *Front Cell Dev Biol.* 2021;9:642948. doi:10.3389/fcell.2021.642948
92. Koh GY, Kane A, Lee K, et al. Parabacteroides distasonis attenuates toll-like receptor 4 signaling and Akt activation and blocks colon tumor formation in high-fat diet-fed azoxymethane-treated mice. *Int J Cancer.* 2018;143(7):1797–1805. doi:10.1002/ijc.31559
93. Zhang X, Hu X, Rao X. Apoptosis induced by Staphylococcus aureus toxins. *Microbiol Res.* 2017;205:19–24. doi:10.1016/j.micres.2017.08.006
94. Nakatsuji T, Chen TH, Butcher AM, et al. A commensal strain of Staphylococcus epidermidis protects against skin neoplasia. *Science Adv.* 2018;4(2):eaao4502. doi:10.1126/sciadv.aao4502

95. Wang LT, Liu KJ, Sytwu HK, Yen ML, Yen BL. Advances in mesenchymal stem cell therapy for immune and inflammatory diseases: use of cell-free products and human pluripotent stem cell-derived mesenchymal stem cells. *Stem Cells Transl Med.* 2021;10:1288–1303. doi:10.1002/sctm.21-0021
96. Shi MY, Liu L, Yang FY. Strategies to improve the effect of mesenchymal stem cell therapy on inflammatory bowel disease. *World J Stem Cells.* 2022;14:684–699. doi:10.4252/wjsc.v14.i9.684
97. Banerjee A, Bizzaro D, Burra P, et al. Umbilical cord mesenchymal stem cells modulate dextran sulfate sodium induced acute colitis in immunodeficient mice. *Stem Cell Res Ther.* 2015;6(1):79. doi:10.1186/s13287-015-0073-6
98. Robinson AM, Miller S, Payne N, et al. Neuroprotective potential of mesenchymal stem cell-based therapy in acute stages of TNBS-induced colitis in guinea-pigs. *PLoS One.* 2015;10(9):e0139023. doi:10.1371/journal.pone.0139023
99. Xie M, Qin H, Luo Q, et al. Comparison of adipose-derived and bone marrow mesenchymal stromal cells in a murine model of crohn's disease. *Dig Dis Sci.* 2017;62(1):115–123. doi:10.1007/s10620-016-4166-6
100. Qu B, Xin G-R, Zhao L-X, et al. Testing stem cell therapy in a rat model of inflammatory bowel disease: role of bone marrow stem cells and stem cell factor in mucosal regeneration. *PLoS One.* 2014;9(10):e107891. doi:10.1371/journal.pone.0107891
101. Watanabe M. Retraction: adult tissue stem cell therapy for gastrointestinal diseases by Mamoru Watanabe. *J Gastroenterol Hepatol.* 2018;33(1):329. doi:10.1111/jgh.12555
102. Fukuda M, Mizutani T, Mochizuki W, et al. Small intestinal stem cell identity is maintained with functional Paneth cells in heterotopically grafted epithelium onto the colon. *Genes Dev.* 2014;28(16):1752–1757. doi:10.1101/gad.245233.114
103. Shaker A, Rubin DC. Stem cells: one step closer to gut repair. *Nature.* 2012;485(7397):181–182. doi:10.1038/485181a
104. Watanabe S, Kobayashi S, Ogasawara N, et al. Transplantation of intestinal organoids into a mouse model of colitis. *Nat Protoc.* 2022;17(3):649–671. doi:10.1038/s41596-021-00658-3
105. Burt RK, Craig RM, Milanetti F, et al. Autologous nonmyeloablative hematopoietic stem cell transplantation in patients with severe anti-TNF refractory Crohn disease: long-term follow-up. *Blood.* 2010;116(26):6123–6132. doi:10.1182/blood-2010-06-292391
106. Cassinotti A, Annaloro C, Ardizzone S, et al. Autologous haematopoietic stem cell transplantation without CD34+ cell selection in refractory Crohn's disease. *Gut.* 2008;57(2):211–217. doi:10.1136/gut.2007.128694
107. Hawkey CJ, Allez M, Clark MM, et al. Autologous hematopoietic stem cell transplantation for refractory Crohn Disease: a randomized clinical trial. *JAMA.* 2015;314(23):2524–2534. doi:10.1001/jama.2015.16700
108. Bislenghi G, Wolthuis A, Van Assche G, et al. Cx601 (darvadstrocel) for the treatment of perianal fistulizing Crohn's disease. *Expert Opin Biol Ther.* 2019;19(7):607–616. doi:10.1080/14712598.2019.1623876
109. Panes J, Garcia-Olmo D, Van Assche G, et al. Expanded allogeneic adipose-derived mesenchymal stem cells (Cx601) for complex perianal fistulas in Crohn's disease: a Phase 3 randomised, double-blind controlled trial. *Lancet.* 2016;388(10051):1281–1290. doi:10.1016/S0140-6736(16)31203-X
110. Garcia-Olmo D, Garcia-Arranz M, Herreros D, et al. A Phase I clinical trial of the treatment of Crohn's Fistula by adipose mesenchymal stem cell transplantation. *Dis Colon Rectum.* 2005;48(7):1416–1423. doi:10.1007/s10350-005-0052-6
111. Zhang J, Lv S, Liu X, Song B, Shi L. Umbilical cord mesenchymal stem cell treatment for Crohn's disease: a randomized controlled clinical trial. *Gut Liver.* 2018;12(1):73–78. doi:10.5009/gnl17035
112. Forbes GM, Sturm MJ, Leong RW, et al. A Phase 2 study of allogeneic mesenchymal stromal cells for luminal Crohn's disease refractory to biologic therapy. *Clin Gastroenterol Hepatol.* 2014;12(1):64–71. doi:10.1016/j.cgh.2013.06.021
113. Knyazev OV, Kagrananova AV, Fadeeva NA, et al. Mesenchymal stromal cells of bone marrow and azathioprine in Crohn's disease therapy. *Ter Arkh.* 2018;90(2):47–52. doi:10.26442/terarkh201890247-52
114. Seo Y, Kang M-J, Kim H-S. Strategies to potentiate paracrine therapeutic efficacy of mesenchymal stem cells in inflammatory diseases. *Int J Mol Sci.* 2021;22(7):3397. doi:10.3390/ijms22073397
115. Lucafò M, Muzzo A, Marcuzzi M, et al. Patient-derived organoids for therapy personalization in inflammatory bowel diseases. *World J Gastroenterol.* 2022;28(24):2636–2653. doi:10.3748/wjg.v28.i24.2636
116. Arjmand B, Rabbani Z, Soveysi F, et al. Advancement of organoid technology in regenerative medicine. *Regen Eng Transl Med.* 2022;1–14. doi:10.1007/s40883-022-00271-0
117. Lopez-Santalla M, Garin MI. Improving the efficacy of mesenchymal stem/stromal-based therapy for treatment of inflammatory bowel diseases. *Biomedicines.* 2021;9(11):1507. doi:10.3390/biomedicines9111507
118. Petrenko Y, Syková E, Kubinová Š. The therapeutic potential of three-dimensional multipotent mesenchymal stromal cell spheroids. *Stem Cell Res Ther.* 2017;8(1):94. doi:10.1186/s13287-017-0558-6
119. Chen Y, Shu Z, Qian K, Wang J, Zhu H. Harnessing the properties of biomaterial to enhance the immunomodulation of mesenchymal stem cells. *Tissue Eng Part B Rev.* 2019;25(6):492–499. doi:10.1089/ten.TEB.2019.0131
120. Hanson SE, King SN, Kim J, et al. The effect of mesenchymal stromal cell–hyaluronic acid hydrogel constructs on immunophenotype of macrophages. *Tissue Eng Part A.* 2011;17(19–20):2463–2471. doi:10.1089/ten.TEA.2010.0716
121. Li H, Shen S, Fu H, et al. Immunomodulatory functions of mesenchymal stem cells in tissue engineering. *Stem Cells Int.* 2019;2019:9671206. doi:10.1155/2019/9671206
122. Kang JY, Oh M-K, Joo H, et al. Xeno-free condition enhances therapeutic functions of human Wharton's Jelly-derived mesenchymal stem cells against experimental colitis by upregulated indoleamine 2,3-dioxygenase activity. *J Clin Med.* 2020;9(9):2913. doi:10.3390/jcm9092913
123. Wu X, Wu D, Mu Y, Zhao Y, Ma Z. Serum-free medium enhances the therapeutic effects of umbilical cord mesenchymal stromal cells on a murine model for acute colitis. *Front Bioeng Biotechnol.* 2020;8:586. doi:10.3389/fbioe.2020.00586
124. Liu Y, Xiao F, Hu X, et al. Recovery and maintenance of NESTIN expression in umbilical cord-MSC using a novel culture medium. *AMB Express.* 2020;10(1):132. doi:10.1186/s13568-020-01067-7
125. Salmenkari H, Laitinen A, Forsgård RA, et al. The use of unlicensed bone marrow-derived platelet lysate-expanded mesenchymal stromal cells in colitis: a pre-clinical study. *Cytotherapy.* 2019;21(2):175–188. doi:10.1016/j.jcyt.2018.11.011
126. Shin T-H, Ahn J-S, Oh S-J, et al. TNF- $\alpha$  priming elicits robust immunomodulatory potential of human tonsil-derived mesenchymal stem cells to alleviate murine colitis. *Biomedicines.* 2020;8(12):561. doi:10.3390/biomedicines8120561

127. Yu D, Zhao Y, Wang H, et al. IL-1 $\beta$  pre-stimulation enhances the therapeutic effects of endometrial regenerative cells on experimental colitis. *Stem Cell Res Ther.* 2021;12(1):324. doi:10.1186/s13287-021-02392-9
128. Payne NL, Dantanarayana A, Sun G, et al. Early intervention with gene-modified mesenchymal stem cells overexpressing interleukin-4 enhances anti-inflammatory responses and functional recovery in experimental autoimmune demyelination. *Cell Adh Migr.* 2012;6(3):179–189. doi:10.4161/cam.20341
129. Song SY, Hong J, Go S, et al. Interleukin-4 gene transfection and spheroid formation potentiate therapeutic efficacy of mesenchymal stem cells for osteoarthritis. *Adv Healthcare Mater.* 2020;9(5):e1901612. doi:10.1002/adhm.201901612
130. Wang W-Q, Dong K, Zhou L, et al. IL-37b gene transfer enhances the therapeutic efficacy of mesenchymal stromal cells in DSS-induced colitis mice. *Acta Pharmacol Sin.* 2015;36(11):1377–1387. doi:10.1038/aps.2015.51
131. Nan Z, Fan H, Tang Q, et al. Dual expression of CXCR4 and IL-35 enhances the therapeutic effects of BMSCs on TNBS-induced colitis in rats through expansion of tregs and suppression of Th17 cells. *Biochem Biophys Res Commun.* 2018;499(4):727–734. doi:10.1016/j.bbrc.2018.03.043
132. Fu Y, Ni J, Chen J, et al. Dual-functionalized MSCs that express CX3CR1 and IL-25 exhibit enhanced therapeutic effects on inflammatory bowel disease. *Mol Ther.* 2020;28(4):1214–1228. doi:10.1016/j.ymthe.2020.01.020
133. Liu L, Wang Y, Fan H, et al. MicroRNA-181a regulates local immune balance by inhibiting proliferation and immunosuppressive properties of mesenchymal stem cells. *Stem Cells.* 2012;30(8):1756–1770. doi:10.1002/stem.1156
134. Lu Q, Chen R, Du S, et al. Activation of the cGAS-STING pathway combined with CRISPR-Cas9 gene editing triggering long-term immunotherapy. *Biomaterials.* 2022;291:121871. doi:10.1016/j.biomaterials.2022.121871
135. Crippa S, Conti A, Vavassori V, et al. Mesenchymal stromal cells improve the transplantation outcome of CRISPR-Cas9 gene-edited human HSPCs. *Mol Ther.* 2022;30(10):3333. doi:10.1016/j.ymthe.2022.09.005
136. Chen Q, Li Y, Chen Z, Du H, Wan J. Anti-VCAM 1 antibody-coated mesenchymal stromal cells attenuate experimental colitis via immunomodulation. *Med Sci Monit.* 2019;25:4457–4468. doi:10.12659/msm.914238
137. Ko IK, Kim B-G, Awadallah A, et al. Targeting improves MSC treatment of inflammatory bowel disease. *Mol Ther.* 2010;18(7):1365–1372. doi:10.1038/mt.2010.54
138. Li X, Lan X, Zhao Y, et al. SDF-1/CXCR4 axis enhances the immunomodulation of human endometrial regenerative cells in alleviating experimental colitis. *Stem Cell Res Ther.* 2019;10(1):204. doi:10.1186/s13287-019-1298-6
139. Li X, Wang Q, Ding L, et al. Intercellular adhesion molecule-1 enhances the therapeutic effects of MSCs in a dextran sulfate sodium-induced colitis models by promoting MSCs homing to murine colons and spleens. *Stem Cell Res Ther.* 2019;10(1):267. doi:10.1186/s13287-019-1384-9
140. Robinson AM, Sakkal S, Park A, et al. Mesenchymal stem cells and conditioned medium avert enteric neuropathy and colon dysfunction in guinea pig TNBS-induced colitis. *Am J Physiol Gastrointest Liver Physiol.* 2014;307(11):G1115–G1129. doi:10.1152/ajpgi.00174.2014
141. Heidari M, Pouya S, Baghaei K, et al. The immunomodulatory effects of adipose-derived mesenchymal stem cells and mesenchymal stem cells-conditioned medium in chronic colitis. *J Cell Physiol.* 2018;233:8754–8766. doi:10.1002/jcp.26765
142. Lykov AP, Poveshchenko OV, Bondarenko NA, et al. Therapeutic potential of biomedical cell product in DSS-induced inflammation in the small intestine of C57Bl/6J mice. *Bull Exp Biol Med.* 2018;165:576–580. doi:10.1007/s10517-018-4216-5
143. Lee KE, Jung S-A, Joo Y-H, et al. The efficacy of conditioned medium released by tonsil-derived mesenchymal stem cells in a chronic murine colitis model. *PLoS One.* 2019;14(12):e0225739. doi:10.1371/journal.pone.0225739
144. Yang S, Liang X, Song J, et al. A novel therapeutic approach for inflammatory bowel disease by exosomes derived from human umbilical cord mesenchymal stem cells to repair intestinal barrier via TSG-6. *Stem Cell Res Ther.* 2021;12:315. doi:10.1186/s13287-021-02404-8
145. Barnhoorn MC, Wasser MN, Roelofs H, et al. Long-term evaluation of allogeneic bone marrow-derived mesenchymal stromal cell therapy for Crohn's disease perianal fistulas. *J Crohns Colitis.* 2020;14:64–70. doi:10.1093/ecco-jcc/jjz116
146. Chao K, Zhang S, Qiu Y, et al. Human umbilical cord-derived mesenchymal stem cells protect against experimental colitis via CD5(+) B regulatory cells. *Stem Cell Res Ther.* 2016;7:109. doi:10.1186/s13287-016-0376-2
147. Legaki E, Roubelakis MG, Theodoropoulos GE, et al. Therapeutic potential of secreted molecules derived from human amniotic fluid mesenchymal stem/stroma cells in a mice model of colitis. *Stem Cell Rev Rep.* 2016;12(5):604–612. doi:10.1007/s12015-016-9677-1
148. de Aguiar CF, Castoldi A, Andrade-Oliveira V, et al. Mesenchymal stromal cells modulate gut inflammation in experimental colitis. *Inflammopharmacology.* 2018;26(1):251–260. doi:10.1007/s10787-017-0404-6
149. de la Portilla F, Alba F, García-Olmo D, et al. Expanded allogeneic adipose-derived stem cells (eASCs) for the treatment of complex perianal fistula in Crohn's disease: results from a multicenter phase I/IIa clinical trial. *Int J Colorectal Dis.* 2013;28:313–323. doi:10.1007/s00384-012-1581-9
150. de la Portilla F, Yuste Y, Pereira S, et al. Local mesenchymal stem cell therapy in experimentally induced colitis in the rat. *Int J Stem Cell.* 2018;11(1):39–47. doi:10.15283/ijsc17074
151. Fu ZW, Zhang ZY, Ge HY. Mesenteric injection of adipose-derived mesenchymal stem cells relieves experimentally-induced colitis in rats by regulating Th17/Treg cell balance. *Am J Transl Res.* 2018;10:54–66.
152. Gonzalez-Rey E, Anderson P, Gonzalez MA, et al. Human adult stem cells derived from adipose tissue protect against experimental colitis and sepsis. *Gut.* 2009;58(7):929–939. doi:10.1136/gut.2008.168534
153. Gregoire C, Briquet A, Pirenne C, et al. Allogeneic mesenchymal stromal cells for refractory luminal Crohn's disease: a Phase I–II study. *Dig Liver Dis.* 2018;50(11):1251–1255. doi:10.1016/j.dld.2018.08.015
154. Hu J, Zhao G, Zhang L, et al. Safety and therapeutic effect of mesenchymal stem cell infusion on moderate to severe ulcerative colitis. *Exp Ther Med.* 2016;12(5):2983–2989. doi:10.3892/etm.2016.3724
155. Lee HJ, Oh S-H, Jang HW, et al. Long-term effects of bone marrow-derived mesenchymal stem cells in dextran sulfate sodium-induced murine chronic colitis. *Gut Liver.* 2016;10(3):412–419. doi:10.5009/gnl15229
156. Lee B-C, Shin N, Lee JY, et al. MIS416 enhances therapeutic functions of human umbilical cord blood-derived mesenchymal stem cells against experimental colitis by modulating systemic immune milieu. *Front Immunol.* 2018;9:1078. doi:10.3389/fimmu.2018.01078
157. Panés J, García-Olmo D, Van Assche G, et al. Long-term efficacy and safety of stem cell therapy (Cx601) for complex perianal fistulas in patients with Crohn's disease. *Gastroenterology.* 2018;154(5):1334–1342.e1334. doi:10.1053/j.gastro.2017.12.020

158. Martín Arranz E, Martín Arranz MD, Robredo T, et al. Endoscopic submucosal injection of adipose-derived mesenchymal stem cells ameliorates TNBS-induced colitis in rats and prevents stenosis. *Stem Cell Res Ther.* 2018;9(1):95. doi:10.1186/s13287-018-0837-x
159. Miyamoto S, Ohnishi S, Onishi R, et al. Therapeutic effects of human amnion-derived mesenchymal stem cell transplantation and conditioned medium enema in rats with trinitrobenzene sulfonic acid-induced colitis. *Am J Transl Res.* 2017;9(3):940–952.
160. Molendijk I, Bonsing BA, Roelofs H, et al. Allogeneic bone marrow-derived mesenchymal stromal cells promote healing of refractory perianal fistulas in patients with Crohn's disease. *Gastroenterology.* 2015;149(4):918–927.e916. doi:10.1053/j.gastro.2015.06.014
161. Pak S, Hwang SW, Shim IK, et al. Endoscopic transplantation of mesenchymal stem cell sheets in experimental colitis in rats. *Sci Rep.* 2018;8(1):11314. doi:10.1038/s41598-018-29617-x
162. Park S-R, Kim J-W, Jun H-S, et al. Stem cell secretome and its effect on cellular mechanisms relevant to wound healing. *Mol Ther.* 2018;26(2):606–617. doi:10.1016/j.ymthe.2017.09.023
163. Pouya S, Heidari M, Baghaei K, et al. Study the effects of mesenchymal stem cell conditioned medium injection in mouse model of acute colitis. *Int Immunopharmacol.* 2018;54:86–94. doi:10.1016/j.intimp.2017.11.001
164. Song J-Y, Kang HJ, Hong JS, et al. Umbilical cord-derived mesenchymal stem cell extracts reduce colitis in mice by re-polarizing intestinal macrophages. *Sci Rep.* 2017;7(1):9412. doi:10.1038/s41598-017-09827-5
165. Song W-J, Li Q, Ryu M-O, et al. TSG-6 released from intraperitoneally injected canine adipose tissue-derived mesenchymal stem cells ameliorate inflammatory bowel disease by inducing M2 macrophage switch in mice. *Stem Cell Res Ther.* 2018;9(1):91. doi:10.1186/s13287-018-0841-1
166. Tanaka F, Tominaga K, Ochi M, et al. Exogenous administration of mesenchymal stem cells ameliorates dextran sulfate sodium-induced colitis via anti-inflammatory action in damaged tissue in rats. *Life Sci.* 2008;83(23–24):771–779. doi:10.1016/j.lfs.2008.09.016
167. Wu Y, Qiu W, Xu X, et al. Exosomes derived from human umbilical cord mesenchymal stem cells alleviate inflammatory bowel disease in mice through ubiquitination. *Am J Transl Res.* 2018;10(7):2026–2036.
168. Xu X, Wang Y, Zhang B, et al. Treatment of experimental colitis by endometrial regenerative cells through regulation of B lymphocytes in mice. *Stem Cell Res Ther.* 2018;9(1):146. doi:10.1186/s13287-018-0874-5
169. Yu Y, Song EM, Lee KE, et al. Therapeutic potential of tonsil-derived mesenchymal stem cells in dextran sulfate sodium-induced experimental murine colitis. *PLoS One.* 2017;12(8):e0183141. doi:10.1371/journal.pone.0183141
170. Oyama Y, Craig RM, Traynor AE, et al. Autologous hematopoietic stem cell transplantation in patients with refractory Crohn's disease. *Gastroenterology.* 2005;128(3):552–563. doi:10.1053/j.gastro.2004.11.051
171. Clerici M, Cassinotti A, Onida F, et al. Immunomodulatory effects of unselected haematopoietic stem cells autotransplantation in refractory Crohn's disease. *Dig Liver Dis.* 2011;43(12):946–952. doi:10.1016/j.dld.2011.07.021
172. Hasselblatt P, Drognitz K, Potthoff K, et al. Remission of refractory Crohn's disease by high-dose cyclophosphamide and autologous peripheral blood stem cell transplantation. *Aliment Pharmacol Ther.* 2012;36(8):725–735. doi:10.1111/apt.12032
173. Kotlarz D, Beier R, Murugan D, et al. Loss of interleukin-10 signaling and infantile inflammatory bowel disease: implications for diagnosis and therapy. *Gastroenterology.* 2012;143(2):347–355. doi:10.1053/j.gastro.2012.04.045
174. Liang J, Zhang H, Wang D, et al. Allogeneic mesenchymal stem cell transplantation in seven patients with refractory inflammatory bowel disease. *Gut.* 2012;61(3):468–469. doi:10.1136/gutjnl-2011-300083
175. Garca-Olmo D, García-Arranz M, García LG, et al. Autologous stem cell transplantation for treatment of rectovaginal fistula in perianal Crohn's disease: a new cell-based therapy. *Int J Colorectal Dis.* 2003;18(5):451–454. doi:10.1007/s00384-003-0490-3
176. Garcia-Olmo D, Herreros D, Pascual I, et al. Expanded adipose-derived stem cells for the treatment of complex perianal fistula: a phase II clinical trial. *Dis Colon Rectum.* 2009;52(1):79–86. doi:10.1007/DCR.0b013e3181973487
177. Guadalajara H, Herreros D, De-La-Quintana P, et al. Long-term follow-up of patients undergoing adipose-derived adult stem cell administration to treat complex perianal fistulas. *Int J Colorectal Dis.* 2012;27(5):595–600. doi:10.1007/s00384-011-1350-1
178. Herreros MD, Garcia-Arranz M, Guadalajara H, De-La-Quintana P, Garcia-Olmo D. Autologous expanded adipose-derived stem cells for the treatment of complex cryptoglandular perianal fistulas: a phase III randomized clinical trial (FATT 1: fistula advanced therapy trial 1) and long-term evaluation. *Dis Colon Rectum.* 2012;55(7):762–772. doi:10.1097/DCR.0b013e318255364a
179. Dietz AB, Dozois EJ, Fletcher JG, et al. Autologous mesenchymal stem cells, applied in a bioabsorbable matrix, for treatment of perianal fistulas in patients with Crohn's disease. *Gastroenterology.* 2017;153(1):59–62.e52. doi:10.1053/j.gastro.2017.04.001
180. Dige A, Hougaard HT, Agnholt J, et al. Efficacy of injection of freshly collected autologous adipose tissue into perianal fistulas in patients with Crohn's disease. *Gastroenterology.* 2019;156(8):2208–2216.e2201. doi:10.1053/j.gastro.2019.02.005
181. Lightner AL, Dozois EJ, Dietz AB, et al. Matrix-delivered autologous mesenchymal stem cell therapy for refractory rectovaginal Crohn's fistulas. *Inflamm Bowel Dis.* 2020;26(5):670–677. doi:10.1093/ibd/izz215
182. Nikolic M, Stift A, Reinisch W, et al. Allogeneic expanded adipose-derived stem cells in the treatment of rectovaginal fistulas in Crohn's disease. *Colorectal Dis.* 2021;23(1):153–158. doi:10.1111/codi.15324
183. Cho YB, Park KJ, Yoon SN, et al. Long-term results of adipose-derived stem cell therapy for the treatment of Crohn's fistula. *Stem Cells Transl Med.* 2015;4(5):532–537. doi:10.5966/sctm.2014-0199
184. Ciccocioppo R, Gallia A, Sgarella A, et al. Long-term follow-up of Crohn disease fistulas after local injections of bone marrow-derived mesenchymal stem cells. *Mayo Clin Proc.* 2015;90(6):747–755. doi:10.1016/j.mayocp.2015.03.023
185. Garcia-Olmo D, Guadalajara H, Rubio-Perez I, et al. Recurrent anal fistulae: limited surgery supported by stem cells. *World J Gastroenterol.* 2015;21(11):3330–3336. doi:10.3748/wjg.v21.i11.3330
186. Park KJ, Ryoo S-B, Kim JS, et al. Allogeneic adipose-derived stem cells for the treatment of perianal fistula in Crohn's disease: a pilot clinical trial. *Colorectal Dis.* 2016;18(5):468–476. doi:10.1111/codi.13223
187. Garcia-Arranz M, Herreros MD, González-Gómez C, et al. Treatment of Crohn's-related rectovaginal fistula with allogeneic expanded-adipose derived stem cells: a phase I-IIa clinical trial. *Stem Cells Transl Med.* 2016;5(11):1441–1446. doi:10.5966/sctm.2015-0356
188. Wainstein C, Quera R, Fluxá D, et al. Stem cell therapy in refractory perineal Crohn's disease: long-term follow-up. *Colorectal Dis.* 2018;20(3):O68–O75. doi:10.1111/codi.14002
189. Knyazev OV, Fadeeva NA, Kagamanova AV, et al. Stem cell therapy for perianal Crohn's disease. *Ter Arkh.* 2018;90(3):60–66. doi:10.26442/terarkh201890360-66

190. Herreros MD, Garcia-Olmo D, Guadalajara H, et al. Stem cell therapy: a compassionate use program in perianal fistula. *Stem Cells Int.* 2019;2019:6132340. doi:10.1155/2019/6132340
191. Zhou C, Li M, Zhang Y, et al. Autologous adipose-derived stem cells for the treatment of Crohn's fistula-in-ano: an open-label, controlled trial. *Stem Cell Res Ther.* 2020;11(1):124. doi:10.1186/s13287-020-01636-4
192. Laureti S, Gionchetti P, Cappelli A, et al. Refractory complex Crohn's perianal fistulas: a role for autologous microfragmented adipose tissue injection. *Inflamm Bowel Dis.* 2020;26(2):321–330. doi:10.1093/ibd/izz051
193. Melmed GY, Pandak WM, Casey K, et al. Human placenta-derived cells (PDA-001) for the treatment of moderate-to-severe Crohn's disease. *Inflamm Bowel Dis.* 2015;21(8):1809–1816. doi:10.1097/mib.0000000000000441
194. Dhere T, Copland I, Garcia M, et al. The safety of autologous and metabolically fit bone marrow mesenchymal stromal cells in medically refractory Crohn's disease - a Phase I trial with three doses. *Aliment Pharmacol Ther.* 2016;44(5):471–481. doi:10.1111/apt.13717
195. Кныазев ОВ, Парфенов АИ, Коноплянский АГ, Болдырева ОН. Мезенхимальные стволовые клетки в комбинированном лечении язвенного колита [Use of mesenchymal stem cells in the combination therapy of ulcerative colitis]. *Ter Arkh.* 2016;88(2):44–48. Russian. doi:10.17116/terarkh201688244-48

Journal of Inflammation Research

Dovepress

## Publish your work in this journal

The Journal of Inflammation Research is an international, peer-reviewed open-access journal that welcomes laboratory and clinical findings on the molecular basis, cell biology and pharmacology of inflammation including original research, reviews, symposium reports, hypothesis formation and commentaries on: acute/chronic inflammation; mediators of inflammation; cellular processes; molecular mechanisms; pharmacology and novel anti-inflammatory drugs; clinical conditions involving inflammation. The manuscript management system is completely online and includes a very quick and fair peer-review system. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/journal-of-inflammation-research-journal>