Bothrops erythromelas snake venom induces a proinflammatory response in mice splenocytes

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Abstract: Snake venom is a complex biological mixture used for immobilization and killing of prey for alimentation. Many effects are inflicted by this venom, such as coagulation, necrosis, bleeding, inflammation, and shock. This study aimed to evaluate the inflammatory activity promoted by Bothrops erythromelas and Crotalus durissus cascavella snake venom. It was observed that both B. erythromelas and C. d. cascavella venom induced higher interferon-gamma and interleukin-6 production. Nitric oxide (NO) was significantly produced only by B. erythromelas venom, which also showed a higher rate of cell death induction when compared with C. d. cascavella. Results showed that B. erythromelas and C. d. cascavella venom induced distinct response in vitro through cytokines and NO production. However, B. erythromelas induces a proinflammatory response and a higher rate of cell death in relation to C. d. cascavella venom.

Keywords: snake venom, Bothrops, nitric oxide, necrosis, Crotalus

Introduction
Snake envenomation is a health problem mainly in rural areas of tropical and subtropical countries.1,2 In Brazil, 20,000 bites by venomous snakes are reported each year, approximately 90% of which are inflicted by the genus Bothrops.3 People bitten by Bothrops species manifest local and systemic manifestations like edema, pain, ecchymosis, blisters, myonecrosis, gingival bleeding, hematuria, epistaxis, hemorrhage, and leukocyte infiltration.4,5

The genus Crotalus contains several species of snakes, such as Crotalus durissus cascavella (usually found in scrublands of the Brazilian northeast), which is responsible for approximately 1500 cases of snakebite annually.6,7 Crotalus species envenomation is caused by a complex mixture of biologically active substances, such as toxins, enzymes, and peptides.8 Clinical manifestations induced by Crotalus venom in South America are neurotoxicity, myotoxicity, and renal damage. The venom also has antithrombotic, platelet-aggregating activities and an analgesic effect and can cause acute renal failure.9–11

Many studies show that the venom of different snakes induces specific immunomodulatory responses in vitro and in vivo.12–14 These responses are divided between T-helper 1 (Th1) and Th2 cells and are mediated by specific cytokines that determine effective functions of immune system compounds.15 Different cytokines are involved in both Th1 and Th2 responses. Interleukin-2 (IL-2) has multiple, sometimes opposing, functions during an inflammatory response and is a potent inducer of T-cell proliferation and Th1 and Th2 effector T-cell differentiation.16 IL-6 is a critical factor for hematopoiesis through regulation of the entry of hematopoietic stem cells
into the cell cycle, proliferation of cells committed to the myeloid and lymphoid lineage, and maturation of B-cells into antibody-producing cells.\(^{17}\) IL-10 is an important immunoregulatory cytokine that influences innate and adaptive immune responses.\(^{18}\) Interferon-gamma (IFN-\(\gamma\)) is a cytokine secreted by activated T and natural killer cells and regulates host defense, inflammation, and autoimmunity. The balance of these cytokines determines which response is predominant, whether Th1 or Th2.\(^{19}\)

Ophidian envenomation induces physiological and immunological consequences. To investigate the immuno-modulatory response induced by both Bothrops erythromelas and C. d. cascavella Brazilian snake venom, the present study analyzed nitric oxide (NO), IL-2, IL-6, IL-10, and IFN-\(\gamma\) production on stimulated mice splenocyte cultures. Cytotoxic assays and cell viability tests were also performed to evaluate, when possible, the damage induced by the venom of both snakes. We believe that these preliminary studies involving immune response in mice splenocytes may promote a better understanding of Bothrops and Crotalus envenomations.

Materials and methods

Animals

Male BALB/c mice (aged 6 to 8 weeks) were raised at the animal facilities of the Oswaldo Cruz Foundation (Rio de Janeiro, Brazil) and maintained at the animal facilities of the Aggeu Magalhães Research Center of the Oswaldo Cruz Foundation in Recife, Brazil. All mice were treated and sacrificed in accordance with the Oswaldo Cruz Foundation Commission for Experiments with Laboratory Animals (Ministry of Health, Brazil, 0266/05).

Preparation of B. erythromelas and C. d. cascavella venom

Male and female specimens of B. erythromelas and C. d. cascavella snakes were milked, and their venom was maintained at 4°C until use. The protein concentration of the venom was analyzed by the Lowry test.\(^{20}\)

Preparation of splenocytes

Splenocytes were obtained according to Pereira et al.\(^{21}\) After euthanizing each animal with carbon dioxide (CO\(_2\)) gas, the spleen was removed aseptically and placed in a Falcon tube containing RPMI 1640 with fetal calf serum (complete medium). In a vertical flow, each spleen was transferred to a Petri dish, where it was macerated. The cell suspensions obtained were transferred to Falcon tubes containing approximately 10 mL of incomplete medium per spleen, and centrifuged at 4°C, 200 \(\times\) g for 5 minutes. After discarding the supernatant, distilled water was added to the sediment to promote lysis of red blood cells. The supernatant, containing no cellular debris, was collected and centrifuged at 4°C, 200 \(\times\) g for 5 minutes. The sediment (containing cells) was resuspended in complete RPMI 1640. An aliquot of each cell suspension was separated and diluted in trypan blue to be quantified in a Neubauer chamber, and cell viability was determined.

In vitro cytotoxicity assays

The cytotoxicity of the venom was determined using BALB/c mice splenocytes (6 \(\times\) 10\(^5\) cells/well) cultured in 96-well plates in RPMI 1640 media (Sigma-Aldrich Chemical Co, St Louis, MO, USA) supplemented with 10% fetal bovine serum (FCS; Cultilab, Campinas, SP, Brazil) and 50 \(\mu\)g/mL of gentamycin (Novafarma, Anápolis, GO, Brazil). Each venom was evaluated at six concentrations (1, 5, 10, 25, 50, and 100 \(\mu\)g/mL) in triplicate on two independent assays. Cultures were incubated in the presence of \(^3\)H-thymidine (Amersham Biosciences) (1 \(\mu\)Ci/well) for 24 hours at 37°C and 5% CO\(_2\). After this period, the content of the plate was harvested to determine the \(^3\)H-thymidine ([\(^3\)H]Tdr) incorporation using a beta-radiation counter (\(\beta\)-matrix 9600, Packard). The toxicity of the venom was determined by comparing the percentage of \(^3\)H-thymidine incorporation (as an indicator of cell viability) of venom-treated wells in relation to untreated wells. Saponin (0.05%), a known cytotoxic compound, was used as a positive control. Concanavalin A (Con A) and Phytohemagglutinin (PHA) were used as references for immunological assays. Nontoxic concentrations were defined as those causing a reduction of \(^3\)H-thymidine incorporation below 30% in relation to untreated controls.

Measurement of cytokine levels in splenocyte supernatants

Splenocytes were cultured in 24-well plates (TPP) at a density of 10\(^6\) cells/well. Cytokines were quantified in 24, 48, 72 hours, and 6 day supernatants from cultures stimulated with B. erythromelas snake venom (100, 10, and 1 \(\mu\)g/mL), C. d. cascavella snake venom (100, 10, and 1 \(\mu\)g/mL), Con A (2.5 \(\mu\)g/mL), PHA (5 \(\mu\)g/mL), or maintained only in culture medium (control). The levels of IL-2, IL-6, IL-10, and IFN-\(\gamma\) were measured by sandwich ELISA, according to the manufacturer’s suggested protocols. The monoclonal antibodies used were from Kit OptEIA (BD Biosciences), having previously been titered. Plates with 96 wells (Nalge Nunc International Corporation) were sensitized with specific
anticytokine antibodies (according to the manufacturer’s instructions) and incubated “overnight” at 4°C. Cytokine standards were added after serial dilution from their initial concentrations (800, 10000, 8000, and 8000 pg/mL for IL-2, IL-6, IL-10, and IFN-γ, respectively). After washes, 50 µL of all samples and standards were added in duplicate and the plates incubated for 2 hours at room temperature. Subsequently, the specific antibodies were combined with biotin (according to the manufacturer’s instructions) and incubated for 1 hour 30 minutes at room temperature. Revealer solution containing 2.2-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) diaminium salt was added. The reaction was blocked with 1 M sulfuric acid, and the reading was carried out in a spectrophotometer (Bio-Rad 3550, Hercules, CA, USA) at 415 nm. Sample concentrations were calculated in the linear region of the titration curve of cytokine standards, and final concentrations were expressed in pg/mL, using the Microplate Manager Version 4.0 software (Bio-Rad Laboratories).

### Analysis of cell viability by annexin V-FITC and propidium iodide staining

Splenocytes were treated with *B. erythromelas* venom (100, 10, and 1 µg/mL), *C. d. cascavella* venom (100, 10, and 1 µg/mL), Con A (2.5 µg/mL), and PHA (5 µg/mL). These treated cells were maintained in culture in 24-well plates (TPP) for 24 hours to analyze their cell viability. Untreated cells (control) were used as a negative control. Following this, lymphocytes were centrifuged at 4°C, 450 × g for 10 minutes. After discarding the supernatant, 1 mL of PBS 1X was added to the sediment, and the mixture was centrifuged at 4°C, 450 × g for 10 minutes. After discarding the supernatant, the pellet was resuspended in a binding buffer (10 mM HEPES [pH 7.4], 150 mM NaCl, 5 mM KCl, 1 mM MgCl₂, and 1.8 mM CaCl₂), and annexin V conjugated with fluorescein isothiocyanate (FITC) (1:500) and propidium iodide (PI, 20 µg/mL; 10⁶ cells) was added to each labeled cytometer tube. Flow cytometry was performed on a FACSCalibur (Becton Dickinson Biosciences, Mountain View, CA, USA) and analyzed using CellQuest Pro software (Becton Dickinson). Results analysis was performed on graphs by dot plot. Double negatives (annexin-FITC+/PI⁻) were considered viable cells. Annexin-FITC+/PI⁻/spleenocytes in the early stage of apoptosis. Double positive annexin-FITC+/PI⁺ were considered to be spleen cells in the late stage of apoptosis and only PI⁺ cells were considered necrotic.

### In vitro nitrite analysis

Splenocytes were used to evaluate the concentration of nitrite after treatment with *B. erythromelas* venom (100, 10, and 1 µg/mL), *C. d. cascavella* venom (100, 10, and 1 µg/mL), Con A (2.5 µg/mL), PHA (5 µg/mL), or maintained only in culture medium (control) after 24, 48, 72 h, and 6 days of incubation. Culture media were carefully collected for subsequent measurement by the colorimetric Griess method (Ding et al.). NO concentration was estimated by the standard curve (3.12–100 µmol/mL), and the reading was carried out in a spectrophotometer (Bio-Rad 3550) at 490 nm.

### Statistical analysis

Data were analyzed using nonparametric tests. Differences among groups were analyzed by one-way analysis of variance (ANOVA) followed by either Tukey’s t-test or a Kruskal–Wallis nonparametric test. All results were expressed as mean values of groups ± standard deviation of four independent experiments per group and were analyzed considering the value of *P* < 0.05 as statistically significant.

### Results

The cytotoxicity threshold was expressed as the highest concentration tested that was not cytotoxic for the splenocytes. Saponin was used as a positive control. Results showed that Saponin (0.05%), which is known for its cytotoxicity, demonstrated a higher inhibition. Con A and PHA, used as references for immunological assays, did not show toxic activity at 50 to 1 µg/mL concentrations for Con A and at neither concentration for PHA. Both *B. erythromelas* and *C. d. cascavella* venom was cytotoxic above 1 µg/mL concentration, indicating the higher toxic effect induced by the snake venom in splenocyte cultures (Table 1). Because of the higher toxic effect induced by the venom of both snakes,

### Table 1 Cytotoxic effect induced by *Bothrops erythromelas* and *Crotalus durissus cascavella* venom

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Lectin concentrations (µg/mL)</th>
<th>Inhibition (%)</th>
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<tr>
<td></td>
<td>100</td>
<td>50</td>
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<tr>
<td><em>B. erythromelas</em></td>
<td>62</td>
<td>55</td>
</tr>
<tr>
<td><em>C. d. cascavella</em></td>
<td>61</td>
<td>56</td>
</tr>
<tr>
<td>Con A</td>
<td>58</td>
<td>–</td>
</tr>
<tr>
<td>PHA</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Saponin</td>
<td>94</td>
<td>91</td>
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Notes: *Percent of cellular proliferation inhibition. (-) = Noncytotoxic concentrations were defined as those causing a reduction of [³H]-thymidine incorporation below 30% in relation to untreated controls.
all immunomodulatory assays in this study (ELISA, nitrite detection, and flow cytometry) were performed using a nontoxic dose, ie, 1 µg/mL.

**Snake venom promoted different values to IFN-γ production**

Immunological investigations of splenocytes stimulated in vitro with *B. erythromelas* snake venom were performed comparing this group with splenocytes stimulated with *C. d. cascadavella* snake venom. Because of their immunological properties, Con A and PHA mitogens were used as positive controls, and unstimulated cells (cells + medium) were used as a negative control. Four experimental times, ie, 24, 48, 72 hours, and 6 days, were used in our assays. In 24 hours, Con A (2149 ± 311) showed higher and statistically significant values in relation to the control (*P* ≤ 0.05). However, at the same time, *B. erythromelas* venom (1422 ± 480 and 1214 ± 380 at 100 and 1 µg/mL concentrations, respectively) and *C. d. cascadavella* (1234 ± 348 at 1 µg/mL concentration) induced higher, but not statistically significant, IFN-γ production. At 48 hours, only *C. d. cascadavella* (1374 ± 288 at 1 µg/mL) showed statistically significant values in relation to the control (*P* ≤ 0.05). The time of greatest IFN-γ production for all stimuli in vitro was 72 hours (Figure 1A–E). At this time, we observed that both *B. erythromelas* and *C. d. cascadavella* (at 100 µg/mL) showed significant values (Figure 1D–E). Finally, after 6 days of assay, only Con A (6032 ± 2545) and *C. d. cascadavella* (3090 ± 601 at 100 µg/mL) showed significant IFN-γ production (*P* ≤ 0.05). As shown in Figure 1, IFN-γ levels in supernatant cultures showed peak values at 72 hours.

![Figure 1](https://www.dovepress.com/)

**Notes:** Splenocytes were incubated under distinct experimental conditions, including: A) the absence of exogenous stimuli – control culture (□); B) in the presence of PHA (□); C) in the presence of Con A (□); D) in the presence of distinct concentrations (--- 100 µg/mL, --- 10 µg/mL, and ---- 1 µg/mL) of *B. erythromela* (□); or E) *C. d. cascadavella* venoms (□). Data are expressed as mean levels of IFN-γ (pg/mL) ± standard error. Rectangles highlight the data points at 72 hours of incubation, identified as the first statistically significant (*P* < 0.05) time point for IFN-γ production as compared with 24 and 48 hours of incubation (*) and therefore selected for further analysis. Horizontal bars represent the average of four independent experiments per group.
of assay. Thus, 72 hours was the time chosen to analyze other cytokines in this study, such as IL-2, IL-6, IL-10, and NO.

Profile of cytokine release and comparative analysis at 72 hours of assay
Other cytokines, such as IL-2, IL-6, IL-10, and NO, were measured in our study. IL-2 was not produced at significant levels for any treatment with snake venom at all experimental times. Only Con A showed statistically significant induction of production of this cytokine at 24, 48, and 72 hours (628 ± 42, 768 ± 13, and 151 ± 1.9 and P < 0.05, respectively) (data not shown). Similar to IFN-γ, IL-6 was also produced in statistically significant values by B. erythromelas (13,235 ± 1,850 at 1 µg/mL) and by C. d. cascavella (11,606 ± 1,038 at 10 µg/mL) at 72 hours of assay. The same result was observed for IL-10 and NO production at 72 hours of assay (Figure 2B and C). IL-10 was detected in splenocytes stimulated with B. erythromelas venom (100 and 10 µg/mL concentrations) and C. d. cascavella venom (1 µg/mL) when compared with the control (Figure 2B). Although NO production showed higher levels, only B. erythromelas venom (1 µg/mL) showed higher values (P ≤ 0.05) in relation to the control at 72 hours of assay (Figure 2C). In addition, we could also see that although there were no differences among venom stimulus in relation to IFN-γ production (Figure 2A), we observed a dichotomy in the synthesis of NO and IL-10 at 72 hours of assay. In fact, when we increased doses of venom from B. erythromelas, a gradual increase in IL-10 production and a decrease in NO release occurred (Figure 2B and C). The opposite effect was observed, ie, an increased C. d. cascavella dose induced a lower IL-10 production and a higher NO release (Figure 2B and C). These observations are very important for understanding venom molecular differences and signaling mechanisms in mice splenocytes.

Analysis of IFN-γ/IL-10 balance between snake venom
An imbalance in the ratio between proinflammatory and anti-inflammatory events is possible. To analyze this characteristic, we assessed the balance of the synthesis of IFN-IL-10 at 72 hours of in vitro stimuli. Figure 3 shows that although no significant difference was observed in this ratio, high doses of B. erythromelas venom (100 µg/mL) were able to promote an immunoregulatory effect, characterized by an inverse relationship in the synthesis of IFN-γ and IL-10.

Profile of cell viability indicated higher necrosis induced by B. erythromelas venom
After the cytotoxicity analysis, we investigated possible cell damage induced by snake venom in mice splenocytes. For this analysis we used apoptosis, late apoptosis, and necrosis parameters to indicate the induction of death and cellular damage promoted by the venom of both snakes (Figure 4A–E). The absence of exogenous stimuli was used as control. Figure 4A shows the cell viability status of BALB/c mice splenocytes following 24 hours of in vitro stimuli with snake venom. The data analysis indicated that B. erythromelas (at 100, 10, and 1 µg/mL concentrations) induced higher late apoptosis and necrosis (at 100 and 10 µg/mL) (Figure 4C and D). On the other hand, C. d. cascavella venom only showed higher necrosis induction at 1 µg/mL concentration (Figure 4D). Comparison between the venom of both snakes also showed higher late apoptosis induced by B. erythromelas

![Figure 2](https://www.dovepress.com/)

**Figure 2** Comparative analysis of IFN-γ, NO, and IL-10 production by BALB/c mice splenocytes following 72 hours of in vitro stimuli with snake venom.

**Notes:** Splenocytes were incubated under distinct experimental conditions, including the absence of exogenous stimuli – control culture (C); in the presence of PHA (PH); in the presence of Con A (Con A); in the presence of distinct concentrations (100 µg/mL, 10 µg/mL, and 1 µg/mL) of B. erythromelas; or C. durissus venom. The results are expressed as mean levels of IFN-γ, NO, and IL-10 (µg/mL) ± standard error. Significant differences at P < 0.05 are highlighted by ↑↓ as compared with control cultures and by connecting lines for intragroup comparisons. Dashed rectangles highlight the venom concentration (1 µg/mL) that induced a dichotomous profile between B. erythromelas (proinflammatory) and C. d. cascavella (regulatory). NO = not done. Horizontal bars represent the average of four independent experiments per group.
Luna et al

Snakes of the genera Bothrops and Crotalus are responsible for the majority of the ophidian accidents in Latin America and, specifically, Brazil. In humans and experimental animals, Bothrops envenomation induces systemic hemorrhage and blood incoagulability in addition to intense tissue damage at the site of injection. Symptoms are due to the presence of a variety of toxins in the venom. On the other hand, Crotalus envenomation does not induce a significant inflammatory reaction at the site of the bite but exerts severe systemic neuro-, nephro-, hepato-, and myotoxic effects.

Many studies have investigated immunomodulatory effects induced by the venom of different snakes, and these immunological alterations were observed as both cellular and humoral responses. The aim of this study was the same, but before analyzing immunological aspects and the stimulation profile exerted by B. erythromelas and C. d. cascavella snake venom, it was necessary to investigate of cytotoxicity levels promoted by the venom of both snakes against mice splenocytes.

We used tritiate thymidine ([3H]TdR) in our assays, given that thymidine acts through the incorporation of

![Figure 3](https://www.dovepress.com/)

Figure 3 Analysis of IFN-γ/IL-10 balance produced by BALB/c mice splenocytes following 72 hours of in vitro stimuli with snake venom. Notes: Top panel represents the IFN-γ/IL-10 ratio, considering the distinct experimental conditions, including absence of exogenous stimuli – control culture (□); presence of Con A (□); presence of distinct concentrations (100 µg/mL, 10 µg/mL, and 1 µg/mL) of B. erythromelas (□), or C. d. cascavella venom (□). The results are expressed as mean IFN-γ/IL-10 ratio ± standard error. Bottom panels represent correlation analysis between IFN-γ (pg/mL) and IL-10 (pg/mL) following in vitro stimuli with B. erythromelas (□) or C. d. cascavella (□) at distinct concentrations (100 µg/mL, 10 µg/mL, and 1 µg/mL). Correlation indices (r and P-values) are provided in the figure. Significance was considered at P < 0.05 and highlighted in bold. Horizontal bars represent the average of four independent experiments per group.

(at 1 µg/mL concentration) in relation to C. d. cascavella venom (Figure 4C).

**Discussion**

Snakes of the genera Bothrops and Crotalus are responsible for the majority of the ophidian accidents in Latin America and, specifically, Brazil. In humans and experimental animals, Bothrops envenomation induces systemic hemorrhage and blood incoagulability in addition to intense tissue damage at the site of injection. Symptoms are due to the presence of a variety of toxins in the venom. On the other hand, Crotalus envenomation does not induce a significant inflammatory reaction at the site of the bite but exerts severe systemic neuro-, nephro-, hepato-, and myotoxic effects.

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tритион в ДНК клеток и это наиболее часто используемый метод для оценки специфичности влияния на продукцию интерферона. У нас были получены результаты, демонстрирующие, что укусы гадюки вызывают более выраженные цитотоксические эффекты при экспозиции высокими дозами (100 μg/mL). Ломонте и соавт.

Габрово, инвестигуя Bothrops asper гадюку, установил, что цитотоксичность в макрофагах, включая мембрану, приводит к лизису. Вместе с тем, гаути и соавт. показали, что фракция PLA2 укуса Bothrops marajoensis вызывает цитотоксичность в макрофагах, включая мембрану, приводя к лизису. В дополнение, Crotalus durissus terrificus укус вызывает цитотоксические эффекты в спленоцитах в vitro при дозе 5 μg/mL.

Многие исследования описывают участие цитокинов в заболевании, включая гадюку, реснитца, пауков и пауков.

В целях доказательства влияния укуса гадюки, мы исследовали профиль IFN-γ, IL-2, IL-6, IL-10, и NO производства в спленоцитах при введении гадюки B. erythromelas и C. d. cascavella в vitro. В наших исследованиях, мы установили, что лучшее время для анализа цитотоксичности было 72 часа. В целях доказательства влияния укуса гадюки, мы исследовали цитотоксичность при введении IFN-γ, IL-6, IL-10, и NO производства, но цитотоксичность укусов двух гадюк была значимо ниже, чем укус C. d. cascavella, а укус B. erythromelas вызвал статистически значимое увеличение NO. Дальнейшие исследования по данной теме показывают возможное молекулярное взаимодействие гадюки и спленоцитов.
differences and signaling mechanisms induced by the venom of both snakes.

Similar results were demonstrated by Rangel-Santos et al. These authors affirm that splenocytes treated with *Crotalus d. terrificus* venom did not show IL-2 and IL-10 cytokine release. In fact, other studies have demonstrated, through in vivo and in vitro assays, the same profile of cytokine release observed in our study. Escocard et al showed IL-6 production induced by crude venom of *Bothrops atrox*; Petricevich et al investigating the venom of *Bothrops asper* and *B. jararaca*, demonstrated that the venom of both snakes induced prominent elevations on the production of IL-6, IL-10, and IFN-γ; and Lomonte et al showed that injection of *Bothrops asper* venom into mouse paws also invoked increased levels of IL-6 in the serum.

Higher NO release observed in splenocytes stimulated in vivo with *B. erythromelas* venom may be a consequence of characteristics induced by this venom in host tissue, such as damage, prominent necrosis, hemorrhage, and edema, which are characteristics of *Bothrops* envenomation. Furthermore, experimental studies showed that *B. asper* venom induces phagocytosis and triggers microbicidal functions of peritoneal leukocytes in vivo with a consequent increase in the production of hydrogen peroxide and NO by macrophages, followed by the generation of toxic peroxynitrites. In fact, NO is also produced during the host response evoked by *B. asper* venom. This venom induced the synthesis of NO following intramuscular and intraperitoneal injections in mice, mainly through induction of expression of the inducible nitric oxide synthase (iNOS) promoted by higher IFN-γ release. In addition, a protective role of NO was demonstrated against the lethal activity of this venom. Barros et al also showed similar results, indicating that low IFN-γ production was followed by undetected NO levels in supernatant cultures. These studies showed similar proinflammatory responses as achieved in our assays.

Distinct actions of both *B. erythromelas* and *C. d. cascavela* venom shown in this study may be explained by the different toxin composition and clinical manifestations presented by this venom in envenomated humans and mice, as previously explained. Other *Bothrops* genus species, such as *B. asper*, *B. jararaca*, and *B. atrox*, show similar induction of histamine, bradykinin, eicosanoids, prostaglandin, and cytokine production involved in *Bothrops* envenomations. Beyond, similar to our data, other studies affirm the higher Th1 response induced by *Bothrops* envenomations promoted in association with a higher proinflammatory response observed on ophidian envenomation. Although *Crotalus* envenomations induce similar hemorrhagic and tissue-damaging effects observed in *Bothrops* envenomations, the production of prostaglandins (potent proinflammatory mediators) induced by *Crotalus* venom is still unclear. Beyond that, some studies have shown the anti-inflammatory profile of this venom.

Although many stimuli can induce immunomodulatory responses, eg, snake venom, it is necessary to verify whether this response is beneficial for the stimulated immune cell, because some studies affirm that specific stimuli (biological or chemical) can induce damage, such as apoptosis or necrosis, on target cells through excessive free radical accumulation. In our cell viability assay, *B. erythromelas* venom induced higher inhibition of cell damage than *C. d. cascavela* venom. Teixeira et al also reported similar results showing depletion of neutrophils in mice treated with *B. asper* venom and amytotoxic PLA2 fraction. In addition, Angulo and Lomonte, analyzing the mechanism of action of myotoxin II, a phospholipase A2 of *B. asper* venom, showed that this toxin can induce necrotic cell death through degenerative events related to Ca2+ influx. Studies analyzing *Crotalus* venom effects were performed with isolated fraction (toxins) of this venom, and it has been shown that these toxins cause a reduction in the resting potential of the membrane and an increase in membrane conductance. Rangel-Santos et al showed in their assays that *Crotalus d. terrificus* venom induced higher inhibition of the cellular proliferative response, in cultures stimulated by Con A, in both crude venom and crotoxin (isolated fraction). However, their cell viability test did not show significant results. In our study we used total *Crotalus* venom, and we suggest that this methodology may be key to promoting low cell damage in comparison with *B. erythromelas*. Many studies have investigated the cell damage induced by this viperid snake venom, correlating the role of venom toxins with inflammatory responses. Diaz et al, treating human endothelial cells with BaP1, observed that this venom fraction induces apoptosis through caspase-8 activation. In addition, Gallagher et al observed in their study that fibroblasts treated with jararhagin activated an apoptotic cascade through death receptor (extrinsic) apoptosis pathways.

Snake venom is very complex, containing hundreds of biologically active compounds associated with the inflammatory process. In this study we observed that *B. erythromelas* and *C. d. cascavela* venom induced a marked immunomodulatory response in vitro through cytokines and NO production. However, it is evident that *B. erythromelas* promotes a proinflammatory profile and that *C. d. cascavela* venom has the opposite effect, ie, is anti-inflammatory.
We suggest that these data can enhance our understanding of immunological consequences induced by ophidian envenomation, especially concerning South American species, and may promote a better understanding for future therapeutic strategy by anti-ophidic venom treatment.

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Disclosure

No conflicts of interest were declared in relation to this paper.

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