Effect of Telmisartan and Quercetin in 5 Fluorouracil-Induced Renal Toxicity in Rats

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Purpose: The present study was designed to evaluate the possible synergistic effects of telmisartan and quercetin in 5 fluorouracil (5-FU) induced nephrotoxicity in rats.

Methodology: Forty male rats were randomly divided into five groups: The negative control group, the positive control group that received 5-FU, the telmisartan group, receiving 10 mg/kg, the quercetin group, receiving 80 mg/kg, and the combination of telmisartan and quercetin group. All the treatments were given orally for 14 days. A single intraperitoneal injection of 5-FU (150 mg/kg) on day 13 of the experiment was given except for the negative control group. On the 15th day after scarification, approximately 5 mL of blood was collected and used for measurement of CBC, urea, creatinine, and uric acid. The kidneys were used for histopathological examination and for the measurement of kidney injury molecule-1 (KIM-1), neutrophil gelatinase-associated lipocalin (NGAL), Cystatin C (Cys-C), and total antioxidant capacity (TAOC).

Results: The combination therapy significantly attenuated the levels of tissue KIM-1, NGAL, Cys-C, and serum uric acid as well as blood inflammatory markers, Neutrophil/Lymphocyte (NLR), Monocyte/Lymphocyte (MLR), and Platelets/Lymphocyte ratios (PLR), and restored the TAOC. The histopathological findings greatly support the biochemical tests.

Conclusion: The results strongly suggest the renoprotective effects of telmisartan and quercetin in combination against the nephrotoxic effect of 5-FU through decreasing the levels of KIM-1, NGAL, and cys-C, and the novel inflammatory markers of kidney injury like NLP, MLR, and PLR, as well as decreasing uric acid and restoring the TAOC. The proposed mechanism could be the additive inhibitory effect on RAS provided by both telmisartan and quercetin.

Keywords: nephrotoxicity, telmisartan, quercetin, inflammatory markers, KIM-1, NGAL

Introduction

Renal impairment may take the form of acute or chronic. Epidemiological studies show that drug-induced nephrotoxicity is the third most common cause of acute kidney disease,1 and it is one of the major contributing factors to chronic kidney disease and end-stage renal disease.2

The pivotal roles of the kidney including the secretion, reabsorption, filtration, bioactivation, and excretion of most of the substances that enter the biological system, render it to be a good target for many medications to induce their toxicity. The increased and sustained exposure to numerous medicines and drug metabolites can cause harm to the functional unit of the kidney, including the vascular element, tubules, and the glomeruli.3 5-FU, the uracil nucleotide analog, is one of the cytotoxic medications used in various types of cancers such as stomach, colorectal, and pancreatic cancers in addition to cervical and breast cancer.4 The toxicities of 5-FU appear in several organs such as the kidney, liver, CNS, and the heart.9 Many mechanisms have been proposed for the nephrotoxic effect of 5-FU; DNA damage and nontargeting apoptotic cell death via increasing caspase-3 activity in the kidneys are the most deleterious effect of 5-FU.10,11 Additionally, attenuating the kidney’s ability to scavenge and neutralize free radicals, particularly ROS, predisposes it to different types of oxidative stress assaults, which play a key role in triggering 5-FU-induced renal damage.5,12 Furthermore, triggering inflammatory response is another proposed mechanism of 5-FU-induced nephrotoxicity because
It has been shown to increase activities of proinflammatory mediators such as tumor necrosis factor-alpha, interleukin-1, and interleukin-6 in the organs of rats treated with 5-FU.\textsuperscript{10,11}

The local effect of the renin-angiotensin system (RAS) has taken pronounced attention, since the components of this system have been found in various tissues, suggesting their role in cellular growth and differentiation.\textsuperscript{13} Prorenin uptake and intrarenal activation of the kidney RAS is thought to be the cause of renal injury and microvascular alterations in the kidney.\textsuperscript{14} Angiotensin II may cause pressure-induced renal injury via its ability to induce systemic and glomerular hypertension or cause ischemia-induced renal injury secondary to intrarenal vasoconstriction and decreased renal blood flow.\textsuperscript{15} Targeting this system is one of the strategies for protecting renal damage. The suppression of RAS by angiotensin receptor blockers (ARBs) has been shown to have renoprotective benefits.\textsuperscript{16} Telmisartan is one of the ARBs, characterized by the longest half-life that extends to 24 hours and a high affinity to AT1 receptor when compared to other older ARBs.\textsuperscript{17,18}

Combining ARBs with natural products like curcumin and resveratrol has been proven to be more successful in lowering metabolic disturbances, inflammation, insulin resistance (IR), hypertension, and a variety of other diabetic problems.\textsuperscript{19,20}

Flavonoids found in medicinal plants, vegetables, and fruits have the potential to be a novel medication source. In this regard, quercetin ($\text{C}_{15}\text{H}_{10}\text{O}_{7}$) is a well-known flavonoid with antihypertensive$^{21}$ and renoprotective properties.\textsuperscript{22} Adjuvant use of quercetin with other medications like sitagliptin has proven to exert cardioprotective effects$^{23}$ and protect against testicular damage induced by doxorubicin.\textsuperscript{24} Another study showed that the concomitant use of quercetin with valsartan was effective in enhancing intestinal absorption and attenuating valsartan outflow.\textsuperscript{25} All these effects, together with the approved regulatory effect on RAS offered by quercetin,\textsuperscript{26} suggest the use of quercetin with a drug like telmisartan for the possible synergistic renoprotective effects induced by 5-FU. Accordingly, the present study was designed to evaluate the effect of telmisartan and quercetin alone and in combination in a rat model of nephrotoxicity induced by 5-FU.
Materials and Methods

Experimental Animals
Male Wistar albino rats weighing 150–230 g were purchased from the animal house of the University of Tikrit. The rats were kept in the animal house of the College of Pharmacy/University of Sulaimani in a well-ventilated plastic cage, in normal conditions (temperature 25±2°C) and light–dark cycles. They were fed a standard pellet diet and had free access to water. The rats were acclimatized for 2 weeks before the beginning of the experiment. The experimental protocols met the Guidelines for Animal Experimentation and were approved by the ethical committee of the University of Sulaimani (Certificate no.6 on the date 14/11/2021) following the institutional Animal Ethics Committee. The study was performed by the Canadian Council on Animal Care (CCAC) guidelines.

Study Design
Forty fully-grown adult male rats were randomly divided into five groups, comprising eight animals each as follows:

- **Group 1**: The negative control group received only 1 mL distilled water (D.W) by oral gavage for 14 days.
- **Group 2**: The positive control group received 1 mL D.W by oral gavage for 14 days and a single intraperitoneal injection of 5-fluorouracil (150 mg/kg) on the 13th day of the experiment.
- **Group 3**: The Telmisartan group received telmisartan (10 mg/kg) by oral gavage for 14 days with the protocol of 5-FU on the 13th day of the experiment.
- **Group 4**: The Quercetin group received quercetin (80 mg/kg) by oral gavage for 14 days with the protocol of 5-FU on the 13th day of the experiment.
- **Group 5**: The combination group received both telmisartan (10 mg/kg) and quercetin (80 mg/kg) by oral gavage for 14 days with the protocol of 5-FU on the 13th day of the experiment.

All institutional and national guidelines for the care and use of laboratory animals were followed. The doses of quercetin, telmisartan, and 5-FU were chosen depending on previous studies.

Measurement of the Weight and Relative Organ Weight of the Rats
The rats utilized in the experiment were weighed before starting the treatment and on scarification day, using a weight measurement scale. The kidneys were removed carefully, cleaned of all other tissue, and weighed. Relative organ weight was measured using the following equation:

Relative organ weight = Organ weight (g) x [1,000/Body weight (g)].

Tissue Homogenization and Blood Sample Collection
On the 15th day of the experiment the animals were anesthetized using chloroform. After euthanizing the animals, approximately 5 mL of blood was collected by cardiac puncture, the blood was placed in a tube and allowed to clot, then centrifuged at 3,000 rpm for 20 minutes and the separated serum directly went through laboratory testing.

Both kidneys were harvested: one of the kidneys was washed with ice-cold normal saline and put in 25 mL of 10% formaldehdy for later histopathological examination. The other kidney was removed and washed with ice-cold phosphate buffer saline PBS solution, then the whole kidney was weighed, dissected, and homogenized with every 1 g of the kidney, 9 mL of PBS was added according to the instruction from bioassay technology Elisa kits, homogenization was done with an homogenizer machine. The homogenized samples were immediately centrifuged at 5,000 g for 5 min in a cold centrifuge and the supernatant fluids were collected and stored at −80°C for later kidney biomarker analysis.

Measurement of Biochemical Markers
Determination of Renal Function, CRP, and CBC
Serum levels of blood urea nitrogen (BUN), creatinine (Cr), uric acid, and C-reactive protein (CRP) determined using Cobas c 311 analyzers for Clinical Chemistry and Homogeneous Immunology (HIA) and CBC were measured using an automated hematology analyzer.
Kidney-Specific Biomarker Analysis

The supernatant of kidney tissue homogenate was used using ELISA kits (Bioassay technology laboratory, Shanghai, China) for measuring KIM-1 (Cat.No E0549Ra), NGAL (Cat.No E0762Ra), Cys-C (Cat.No E0145Ra), and TAOC (Cat NoSH0242).

Histopathological Protocol

Initially, animals were euthanized in a humane practice, continuously, after animal sacrificing necropsy started by collecting tissue samples for histopathological preparation. Briefly, kidney samples were cut and immobilized into tissue cassettes and then fixed with 10% buffered formaldehyde solution for about 48 hours. Afterward, the sections were dehydrated by passing through a series of ascending concentrations of ethanol alcohol (50%, 60%, 70%, 80%, 90%, and 100%), followed by three phases of xylene clearance. Next, the processed kidney tissues were impregnated and embedded in melted paraffin casts using an automated wax embedder. Paraffinized tissues were sectioned to 6 µm using a semi-automated rotary microtome. Then after, tissue sections were fixed on glass slides and dried using a hot plate tissue holder. Later on, glass slides with their mounted tissue sections were deparaffinized and cleaned with two xylene changes then dried in a hot oven at 50°C for 15 minutes. Finally, tissue sections were stained with Harris’s hematoxylin and eosin solution, cleaned with xylene and cover slipped, then viewed under a bright field light microscope.

Quantitative Lesion Scoring

Lesion scoring was evaluated quantitatively via image analyzer software (AmScope, 3.7) using an ocular lens camera (MD500, 2019) fixed with a bright field light microscope (NOVEL XSZ-N107T, China). In kidney sections, renal tubular cellular degeneration was estimated and measured in the percentage of calculated cell numbers from randomly selected different fields, whereas vascular congestion was assessed in micrometers and statistically evaluated as the mean percentage. While inflammatory cells were counted in 10 randomly chosen fields under high power magnification (100X), the mean average was calculated statistically as a percentage. Lastly, necrotic cells with their pyknotic nuclei were counted from randomly selected fields, then evaluated as the calculated average mean percentage. In conclusion, the mean percentage of all calculated values was expressed as following the lesion scoring-grading system (score 0–10% as no lesions; score 10–25% as mild; score 25–50% as moderate; score 50–75% as severe; score 75–100 as critical).

Statistical Analysis

The statistical analysis was performed using GraphPad Prism8. The values of the measured parameters were expressed as mean ± standard deviation (SD). For the comparisons between different groups, one-way analysis of variance (ANOVA), followed by Tukey’s multiple comparisons test were performed. The results were considered statistically significant when the p-value was less than 0.05.

Results

Effect of Telmisartan and Quercetin Alone or in Combination on Total Weight and Relative Weight of the Rats

In the present study a significant reduction was seen in the positive control group in comparison with the negative control group, (p=0.0003). Meanwhile, telmisartan and quercetin each alone and in combination groups significantly increased the weight of the rats when compared to the positive control group (p=0.0028, 0.0057, and 0.0014, respectively) (Figure 1A). Regarding the relative weight; no significant changes were detected in any of the treatment groups (Figure 1B).

Effect of Telmisartan and Quercetin Alone or in Combination on Serum Levels of Urea, Creatinine, and Uric Acid

The use of 5-FU produced a significant increase in the serum level of urea in comparison with the negative control group (p=0.0008). Only quercetin alone produced a significant reduction when compared with the positive control group (p=0.015), (Figure 2A). Regarding the effect on serum creatinine, only telmisartan alone showed a significant reduction...
when compared to the positive control group ($p=0.024$) (Figure 2B). Serum uric acid significantly increased in the 5-FU treated group in comparison with the negative control ($p=0.018$). Both telmisartan and the combination group significantly decreased the level when compared with the positive control ($p=0.0023$ and $0.0013$, respectively) (Figure 2C).

**Effect of Telmisartan and Quercetin Alone or in Combination on CRP, Neutrophil/Lymphocyte (NLR), Monocyte/Lymphocyte (MLR), and Platelets/Lymphocyte Ratio (PLR)**

C-reactive protein has been increased significantly ($p=0.003$) in the positive control group when compared to the negative control group, and all the treatment groups; telmisartan, quercetin, and the combination group resulted in significant attenuation of CRP level ($p=0.001$, $0.007$, and $0.001$, respectively) (Figure 3A). Meanwhile, significant elevation was observed in each NLR, MLR, and PLR in 5-FU treated group when compared with the negative control ($p=0.012$, $0.005$, $0.0013$, respectively).
and 0.0001, respectively). The combination treatment was able to decrease NLR and MLR significantly in comparison with the positive control group (\( p = 0.01 \) and 0.01, respectively) (Figure 3B and C). Concerning the PLR; all the treatment groups; telmisartan, quercetin, and the combination group, resulted in significant attenuation of PLR (\( p = 0.02, 0.006, \) and 0.001, respectively) (Figures 3D).

**Effect of Telmisartan and Quercetin Alone or in Combination KIM-1, NGAL, Cys-C, and TAOC Levels in Kidney Tissue**

The use of 5-FU produced a significant increase in the level of KIM-1 in comparison with the negative control group (\( p < 0.0001 \)). All the treatment groups including telmisartan, quercetin alone, and in combination were able to produce a significant reduction in the KIM-1 value in the kidneys tissue when compared to the positive control group (\( p = 0.0034, 0.0066, \) and 0.0004, respectively) (Figure 4A). Concerning NGAL level; 5-FU produced a significant increase when
compared with the negative control group ($p=0.01$). Meanwhile, a significant decrease has been observed with telmisartan ($p=0.008$) and quercetin ($p=0.009$), with a significant reduction in the combination group when compared to the positive control group ($p=0.006$) (Figure 4B). The tissue level of cystatin-C has increased significantly in the positive control group when compared to the negative control group ($p=0.0001$). Telmisartan alone decreased the level of Cys-C significantly ($p=0.0022$). Quercetin alone and the combination group were also able to decrease its level significantly ($p=0.0009$, and $0.0002$, respectively) (Figure 4C). Regarding TAOC level in the tissue, a significant decline in its level has been achieved by the use of 5-FU when compared to the negative control group ($p=0.046$). Only the combination group was able to increase the level of TAOC significantly ($p=0.047$) (Figure 4D).

**Histopathology Findings**

Generally speaking, the quantitative morphometric assay and lesion scoring of kidney sections are illustrated in Table 1. Interestingly, animals in treatment group 5 (G5) which is a combination of telmisartan 10 mg/kg and quercetin 80 mg/kg, show enormous improvement in the abnormal morphological changes induced by a single dose of 5-FU 150 mg/kg, which is evident by a significant ($p<0.05$) reduction in the percentage of acute cellular swelling (hydropic degeneration).
Table 1 | Histological Quantitative Evaluation of Kidney Sections

<table>
<thead>
<tr>
<th>Experimental Groups</th>
<th>Cellular Degeneration* (Mean %)**</th>
<th>Necrotic Cells* (Mean %)**</th>
<th>Inflammatory Cells* (Mean %)**</th>
<th>Vascular Congestion (Mean %)**</th>
<th>Lesion Scoring (0−100%)</th>
<th>Lesion Grading</th>
</tr>
</thead>
<tbody>
<tr>
<td>N=8</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(G1) NCG</td>
<td>8.11%a</td>
<td>3.41%a</td>
<td>4.78%a</td>
<td>5.39%a</td>
<td>0−10%</td>
<td>No lesion</td>
</tr>
<tr>
<td>(G2) PCG (5-FU)</td>
<td>94.54%a</td>
<td>89.26%a</td>
<td>86.35%a</td>
<td>78.92%a</td>
<td>75−100%</td>
<td>Severe</td>
</tr>
<tr>
<td>(G3) TEL+(5-FU)</td>
<td>72.86%d</td>
<td>70.59%d</td>
<td>64.73%d</td>
<td>65.53%d</td>
<td>50−75%</td>
<td>Moderate</td>
</tr>
<tr>
<td>(G4) QUR+(5-FU)</td>
<td>74.62%d</td>
<td>72.39%d</td>
<td>67.35%d</td>
<td>62.78%d</td>
<td>50−75%</td>
<td>Moderate</td>
</tr>
<tr>
<td>(G5) TEL+QUR+(5-FU)</td>
<td>58.22%d</td>
<td>57.43%d</td>
<td>52.66%d</td>
<td>49.16%d</td>
<td>50−75%</td>
<td>Moderate</td>
</tr>
</tbody>
</table>

Notes: *Tubules cellular degeneration, necrotic cells, and inflammatory cells were estimated in (%) of calculated cell numbers. Vascular congestion is estimated in (µm) surface area. **Each value represents the mean percentage (n=8). Statistical comparison among groups: Mean values with different letters have significant differences at p<0.05. G1: Negative control group (D.W); G2: Positive control group (D.W) and a single dose of 5-fluorouracil (5-FU) 150 mg/kg; G3: Telmisartan 10 mg/kg and a single dose of 5-fluorouracil (5-FU) 150 mg/kg; G4: Quercetin 80 mg/kg and a single dose of 5-fluorouracil (5-FU) 150 mg/kg; G5: Telmisartan 10 mg/kg and quercetin 80 mg/kg then a single dose of 5-fluorouracil (5-FU) 150 mg/kg. (%) of calculated cells keys: a = 0−10%, b= 10−25%, c= 25−50%, d= 50−75%, e=75−100%.

Discussion

The use of herbal medicine as an adjuvant with other medications has widely increased nowadays, and the results are promising in the management of many diseases such as hypertension, diabetes mellitus, and rheumatoid arthritis. Alternative medicines have also shown great effectiveness when used as add-ons with cancer chemotherapeutic agents to ameliorate the toxicities associated with their use that directly contribute to the limited effectiveness. In the current study, combining quercetin with telmisartan aimed to attenuate the toxic effects produced by 5-FU; this effect appeared to maintain the normal growth of the animals during the period of the study. A study revealed an increase in the levels of anabolic growth factors as well as protection against muscle damage in subjects who used quercetin for 2 weeks. Other studies showed the protective effect of quercetin against 5-FU-induced toxicities. Furthermore, another study conducted on the nephroprotective effect of telmisartan in diabetic rats showed noticeable maintenance of renal functions. In the present study, telmisartan significantly decreased the level of serum creatinine and this finding was in tune with another study. Meanwhile, quercetin maintained a normal function of the kidney by decreasing serum levels of urea. Another biomarker of kidney injury is the elevation of uric acid level. Uric acid buildup in the renal interstitium can draw in inflammatory cells, lead to persistent interstitial inflammation, and result in fibrosis. The significant increased which occurred in the level of uric acid by 5-FU was reversed by each of telmisartan and the combination group. High uric acid concentrations can encourage the generation of reactive oxygen species and oxidative stress in renal tubular epithelial cells, which can lead to a number of clinical consequences. A study has demonstrated the ability of quercetin to inhibit the activity of xanthine oxidoreductase and eventually decrease the production of uric acid. Additionally, telmisartan has been proven to attenuate the level of uric acid and its deleterious consequences.

Inflammatory reaction plays a critical role in the pathogenesis of kidney damage and the condition is known as nephritis. An intricate web of interactions between renal parenchymal cells and local immune cells, such macrophages and dendritic cells, as well as the enlistment of circulating monocytes, lymphocytes, and neutrophils, constitutes inflammation, a condition closely related to renal illness. Monocytes are crucial for tissue remodeling, inflammation, and innate and adaptive immunity. Potential substitute indicators for inflammation include neutrophils and
The results of the current study clearly revealed significant elevation in the levels of CRP, NLR, MLR, and PLR in the 5-FU treated group. These inflammatory biomarkers are considered as novel predictors of acute kidney injury, and the combination therapy produced a maximum anti-inflammatory effect in this regard. This could be attributed to the documented anti-inflammatory effect of telmisartan and quercetin. This combination was also effective in attenuating the levels of KIM-1, NGAL, and cystatin C. KIM-1 is a membrane glycoprotein that serves as a new biomarker for kidney injury. The level of KIM-1 usually increases during toxic damage to the kidney. Another biomarker of renal function is cystatin-C. Kidney damage produced by 5FU markedly increased the level of cystatin C, and the combination groups were able to decrease its level. The synergistic effect of quercetin and telmisartan was also notified in maintaining the total antioxidant status in the present work. The effectiveness of telmisartan could be attributed to increased blood flow to the kidneys and antioxidant, anti-inflammatory, and anti-apoptotic effects.

**Figure 5** Photomicrograph of kidney from groups: (G1): Negative control group received (D.W), display no prominent lesions, apparent by distinctive renal glomeruli (RG), in addition, renal tubules (RT) show no significant lesions except for mild and non-significant cellular swelling. (G2): Positive control group received (D.W) and 5-fluorouracil (5-FU) 150 mg/kg. Renal tubular epithelia show significant and severe vacuolar swelling (yellow arrows) evident by pale enlarged cytoplasm, together with the presence of a profound number of necrotic cells manifested by pyknotic nuclei and acidophilic cytoplasm. The section reveals interstitial infiltration of inflammatory cells (IF), in addition to pyknotic nuclei within the renal glomerular endothelial cells (G3): Telmisartan 10 mg/kg and a single dose of 5-fluorouracil (5-FU) 150 mg/kg, express moderate cellular swelling within the renal tubular epithelia (yellow arrows) in addition to the presence of some vacuolar change inside the glomerular endothelial cells (RG). Moreover, the section shows mild eosinophilic hyaline cast (HC) within the tubular lumen. (G4): Quercetin (80 mg/kg) and a single dose of 5-FU 150 mg/kg, display significant cellular swelling within the lining tubular epithelial and glomerular cells (yellow arrows), together with the incidence of pyknotic nuclei in the renal tubular epithelia (RT). In addition, the section also shows obvious glomerular atrophy (GA) which is simultaneous with minor dilation of Bowman’s capsule. (G5): Telmisartan 10 mg/kg and quercetin (80 mg/kg) with a single dose of 5-FU 150 mg/kg, displays moderate cellular swelling in the renal tubular epithelial cells (RT), with the slight deposition of deep pinkish proteinaceous materials within the tubular lumina (yellow arrows). Additionally, there is some mild vacuolar degeneration (VD) within the endothelial lining of renal glomerular capillaries (RG). H&E. Scale bar: 4 mm.
antiapoptotic effects. Quercetin, on the other hand, has been shown in other studies to improve endothelial function by activating nitric oxide synthase, and through the interference with RAS via downregulating the expression of the Ag II 1a receptor. Additionally, the combination therapy was successful in maintaining the antioxidant capacity. The antioxidant capacity of quercetin is very well documented previously, and the nephroprotective effect of telmisartan is partially attributed to its ability to attenuate renal oxidative stress. Many studies proved the superiority of telmisartan as nephroprotective in comparison with other antihypertensive agents via protecting the renovascular function and improving atherosclerosis. Meanwhile a large body of evidence demonstrated the renoprotective effects of quercetin through anti-inflammatory activity and attenuation of apoptosis and fibrosis. The histopathological finding of the current study greatly supports the biochemical test; where a pronounced improvement has been observed in the combination therapy via clear improvement in the lesion scoring, tissue degeneration, and necrosis.

Conclusion

The results of the current study strongly demonstrate the renoprotective effects of the combination of telmisartan and quercetin against the cytotoxic effect of 5-FU via attenuating the levels of kidney damage specific markers such as KIM-1, NGAL, and cystatin C, and the novel inflammatory markers of kidney injury like NLR, MLR, and PLR as well as restoring total antioxidant capacity. Furthermore, the combination therapy also ameliorated tissue degeneration and lesion scoring in the histopathological finding. The proposed mechanism could be the additive inhibitory effect on RAS provided by both telmisartan and quercetin. These finding suggest the use of this combination as add-on therapy with 5-FU to minimize the toxic effects afforded by the later one.

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Disclosure

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