

Progress and Challenges of Anti-VEGF Agents and Their Sustained-Release Strategies for Retinal Angiogenesis

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Abstract: Currently, the treatment for ocular neovascular diseases, including diabetic macular edema (DME) and age-related macular degeneration (AMD), mainly involves repeated intravitreal injection of anti-vascular endothelial growth factor (VEGF) drugs. Although it can preserve vision, repeated injections are an invasive treatment modality, leading to serious complications and reducing patient adherence to treatment. To reduce the frequency of administration, prolong the time of drug action, and avoid repeated intravitreal injections, the combination of sustained-release materials with anti-VEGF drug therapy has become an emphasis in ophthalmology. In this review, we highlight the current state of anti-VEGF technology, its challenges, and the sustained-release strategies under investigation or being used in clinical practice. Both continuous release and considerable therapeutic effects can be achieved by encapsulating anti-VEGF drugs in sustained-release materials to minimize the number of intravitreal injections. At present, two sustained-release materials are being tested in clinical research, and although basic research shows the strong therapeutic application prospects of extended-release drugs, its challenges mainly involve the discrepancy between the release rates in vitro and the efficiency of the drugs in vivo. Briefly, sustained release of anti-VEGF agents is an advantageous strategy for treating retinal angiogenesis.

Keywords: anti-vascular endothelial growth factor, retinal angiogenesis, anti-VEGF drugs, sustained-release strategies, ranibizumab, bevacizumab

Introduction

Currently, the most common blinding fundus diseases worldwide mainly include age-related macular degeneration (AMD),¹ diabetic retinopathy (DR),² retinal vein occlusions (RVO),³ and retinopathy of prematurity (ROP),⁴ which cause vision loss at almost all ages and impose a heavy socioeconomic burden. The common hallmark of these diseases is the formation of retinal neovascularization (RNV). The causes of pathological RNV mainly include local or total retinal ischemia, hypoxia, and persistent inflammatory states caused by different influencing factors, which further promote the secretion of proangiogenic cytokines by local cells, such as hypoxia-inducible factor-1 α (HIF-1 α),⁵ vascular endothelial growth factor (VEGF),^{5,6} and Platelet-derived growth factor (PDGF).⁷ The blood vessels of pathological RNV are more vulnerable to bleeding, causing different degrees of retinal hemorrhage and vitreous hemorrhage, and in severe cases, retinal cells proliferate and form a proliferative membrane, which can eventually lead to blindness.⁸ Therefore, the management of these diseases should focus on inhibiting the formation of RNV.

Among the proangiogenic cytokines, HIF-1 α as the key mediator of hypoxia is upregulated in the context of aberrant angiogenesis and low oxygen supply in body organs in health and disease.⁹ Increased expression of HIF-1 α correlates with the promotion of angiogenesis, tumor metastasis, and poor prognosis in multiple types of solid tumors, including

those of the colon and cervix. Upregulated HIF-1 α stimulates the development of new blood vessels, providing nutrients and oxygen to facilitate tumor growth and metabolism, clearing metabolic waste and CO₂.⁵ VEGF is a well-established HIF-1 target that plays essential roles in promoting angiogenesis.

VEGF is a highly specific endogenous pro-vascular endothelial cell growth factor. In normal physiologic conditions, VEGF secretion helps to reform endothelial cells in order to construct functional vessels without representing a leaky architecture. VEGF is a crucial target for the inhibition of RNV and the upregulation of VEGF is contributed to the increased vascular permeability, extracellular matrix degeneration, vascular endothelial cell migration and proliferation, and angiogenesis.¹⁰ There are three VEGRs, namely VEGFR-1, VEGFR-2 and VEGFR-3, among them VEGFR-1 and VEGFR-2 are contributed to the formation of new blood endothelial cells, and VEGFR-3 activity is contributed to the formation of new lymphatic endothelial cells.¹¹ Currently, blocking VEGF activity has become the main strategy for the treatment of neovascular ophthalmopathy. Many surveys have shown that anti-VEGF intravitreal injections can inhibit retinal neovascularization and vascular leakage and improve the visual outcomes in most of these conditions.^{12–14}

To reduce the public health burden of fundus neovascular disease, therapeutic approaches including pan retinal photocoagulation, anti-VEGF therapy, Intravitreal injection of steroid and vitrectomy are commonly used clinical strategies.^{15–17} Although intravitreal injection of anti-VEGF drugs can inhibit the further formation of retinal neovascularization to a certain extent, it requires one injection per month to achieve an effective concentration of the drug in the eye.¹⁸ Repeated intraocular injections cause a heavy economic and mental burden on patients, and with the increase in the number of injections, the risk of ocular and systemic diseases in patients also increases sharply.^{19,20} In addition, some patients showed insensitivity to treatment with anti-VEGF drugs as well as photoreceptor degeneration after long-term repeated treatment.²¹ Sustained-release drugs are manufactured to be more potent and slowly dissolved so to release small amounts of drugs into organs or tissues over an extended period.²² Scientific community is currently exploring controlled release technologies that can prolong the action time of anti-VEGF drugs in the eye. Both continuous release and considerable therapeutic effects can be achieved by encapsulating anti-VEGF drugs in sustained-release materials to minimize the number of intravitreal injections. In this review, we summarize the ocular fundus diseases characterized by pathological RNV, retrospect the development and current state of anti-VEGF agents. Notably, we highlight the sustained release strategies for anti-VEGF drugs under investigation or being used in clinical practice.

Ocular Neovascular Disease

Diabetes Mellitus (DM) and Diabetic Macular Edema (DME)

DM is one of the diseases with a rapidly increasing incidence worldwide and is a serious threat to human health. Currently, 300 million individuals have diabetes worldwide, and the incidence is rising rapidly. Over one-third of diabetic individuals will develop DR, and approximately 10% of these individuals will develop a vision-threatening disease.²³

DR falls into two broad categories: the earlier stage of non-proliferative diabetic retinopathy (NPDR) and the advanced stage of proliferative diabetic retinopathy (PDR). The classification of NPDR is based on clinical findings manifested by visible features, including microaneurysms, retinal hemorrhages, intraretinal microvascular abnormalities, and venous caliber changes, while PDR is characterized by the hallmark feature of pathological preretinal neovascularization.²³

An important additional categorization of DR is diabetic DME, which is an important manifestation of DR that occurs across all DR severity levels of both NPDR and PDR and represents the most common cause of vision loss in patients with DR.²⁴ DME arises from diabetes-induced breakdown of the blood-retinal barrier (BRB), with consequent vascular leakage of fluid and circulating proteins into the neural retina.^{23–25} The extravasation of fluid into the neural retina leads to abnormal retinal thickening and cystoid edema of the macula.²⁶

Retinal Vein Occlusions

RVO is the second most common retinal vascular disease after diabetic retinopathy.²⁷ RVO includes central retinal vein occlusion (CRVO), branch retinal vein occlusion (BRVO), and less commonly, hemi-retinal vein occlusion. BRVO is

four to six times more prevalent than CRVO.²⁸ In BRVO, there is occlusion of a branch of the retinal vein system, while in CRVO, the occlusion is located in the central retinal vein.²⁷ The estimated prevalence of CRVO worldwide is 2.5 million.²⁹ RVO is more prevalent in men than women and is more frequent in older age (over 65 years). RVO is associated with increasing age, systemic hypertension, cardiovascular disease, diabetes mellitus, hyperviscosity syndromes, and glaucoma.²⁸ The most common cause of progressive loss of vision from vein occlusions is macular edema.³⁰ VEGF is a cytokine released by hypoxic cells. It increases neovascularization and vascular permeability and leads to the development of macular edema in vein occlusions.^{31,32}

Age-Related Macular Degeneration

AMD is the leading cause of irreversible vision loss in individuals over 65 years of age. It is estimated that over 187 million people worldwide are affected by the disease.³³ It is characterized by progressive and chronic degeneration of the macula, a central region in the retina measuring 5.5 mm in diameter, responsible for high acuity vision.³⁴ There are two main types of AMD, namely non-neovascular or dry AMD and neovascular or wet AMD (nAMD). Early AMD is generally asymptomatic and is diagnosed by the detection of drusen deposits of a specific size and number. Approximately 20% of individuals diagnosed with AMD will develop a late-stage disease.³⁵ Of those who progress to the late stage, 70% will develop neovascular AMD, also known as exudative or wet AMD, and the remainder will progress toward geographic atrophy.^{35,36} Neovascular AMD is characterized by abnormal growth of choroidal blood vessels through Bruch's membrane, generally confined below the retinal pigment epithelium (RPE) and/or retina, although it can also penetrate beyond the subretinal space and within the retina, progressing to retinal angiomatous proliferation.³⁷ These "new" vessels are leaky and proliferative and may eventually lead to fibrosis and scarring, contributing to significant vision loss.^{35–38}

Retinopathy of Prematurity

ROP is a developmental retinal vaso-proliferative disease and a leading cause of visual impairment and blindness in children. It is characterized by the growth of abnormal vessels in the incompletely vascularized retina of preterm infants.³⁹ Globally, in 2010, an estimated 184,700 babies among 14.9 million preterm babies developed at least one stage of ROP, 20,000 of whom became blind (visual acuity < 20/400) or severely visually impaired (visual acuity from < 20/200 to \geq 20/400) from ROP, and of whom 12,300 others developed mild-moderate visual impairment (visual acuity from < 20/40 to \geq 20/200).⁴⁰ Retinal ischemia causes excessive production of VEGF, which results in neovascularization in severe ROP. Early gestational age at birth, low birth weight, and unregulated oxygen exposure are the main known risk factors for the development of ROP.⁴¹

Normal retinal vascularization begins at the optic nerve at 16 weeks of gestation and proceeds anteriorly, with full vascularization at 40 weeks of gestation. This orderly developmental process can be disrupted by the abrupt change from an intrauterine to an extrauterine environment in a preterm infant, leading to the development of ROP.⁴⁰

Therapies for Ocular Neovascular Disease

Intraocular treatment modalities for retinal neovascularization include laser photocoagulation, intravitreal injections of anti-VEGF and steroid agents, and vitreoretinal surgery. The laser results in the destruction of cells in the ischemic retina and produce proangiogenic factors such as VEGF, which are responsible for vasoproliferation.⁴² A laser is a destructive procedure that damages the peripheral retina and can result in visual field constriction.⁴³ Although the first-line therapy for most eyes with center-involved DME consists of anti-VEGF injections, intravitreal injections of steroids can also be effective.^{24–26} However, intravitreal steroid use is limited by more frequent ocular side effects, such as cataracts and glaucoma.

Since the availability of anti-VEGF agents, there has been a paradigm shift in the management and treatment of retinal neovascularization. Although initially used for the treatment of choroidal neovascularization in neovascular age-related macular degeneration, their application has rapidly spread to other indications as they outperform previously existing treatments. In the past decade, various clinical trials have repeatedly demonstrated superior visual acuity results and outcomes with anti-VEGF treatments.

Features of Anti-VEGF Agents

Anti-VEGF agents, with the effects of reducing RNV and choroidal neovascularization (CNV), and inhibiting vascular permeability, have revolutionized ocular neovascularization therapy. Currently, various anti-VEGF drugs are being used in ophthalmology, mainly including pegaptanib (Bausch & Lomb), bevacizumab (Genentech), ranibizumab (Novartis), aflibercept (Bayer), conbercept (Kanghong), brolucizumab (Novartis), abicipar-pegol (Allergan), faricimab (Roche), KSI-301 (Kodiak Sciences), and ziv-aflibercept (Zaltrap). According to their molecular modalities, these anti-VEGF agents can be divided into aptamers, monoclonal antibodies (mAbs), and recombinant fusion proteins. The timeline of development of anti-VEGF drugs being used in the field of ocular fundus disease is shown in Figure 1.

Pegaptanib

Pegaptanib (Bausch & Lomb, 50kDa) was the first drug to obtain Food and Drug Administration (FDA) approval for intravitreal administration. It is a pegylated aptamer that binds preferentially to the heparin-binding domain of the VEGF165 isoform.⁴⁴ Although proven to be efficient in inhibiting the neovascularization process,⁴⁵ its molecular features strongly limit its capacity to block VEGF-related pathways, thus making it a subsidiary therapeutic choice.

Bevacizumab

Bevacizumab (Genentech, San Francisco, CA) is a humanized monoclonal antibody of 148 kDa with activity against VEGF, which was first approved by the (FDA) for the treatment of metastatic colorectal cancer.⁴⁶ Bevacizumab's mechanisms of action is quite simple; as a pure anti-VEGF antibody, its main effect is to block the neovascular stimulus and VEGF-induced vascular permeability.⁴⁷ Furthermore, bevacizumab interacts with HIF-1 α , reducing its stimulating effect on VEGF production.⁴⁸

The use of intravitreal bevacizumab dramatically changed the treatment landscape of ocular diseases having VEGF as an etiological factor. Since 2015, bevacizumab has been shown to improve vision in addition to resolving subretinal fluid and decreasing macular edema in patients with neovascular AMD and RVO, respectively.⁴⁹ However, there is still no FDA approval of bevacizumab for any intraocular indication given the lack of large randomized clinical trial data. Nevertheless, off-label treatment is within the realm of the accepted “standard of care.”

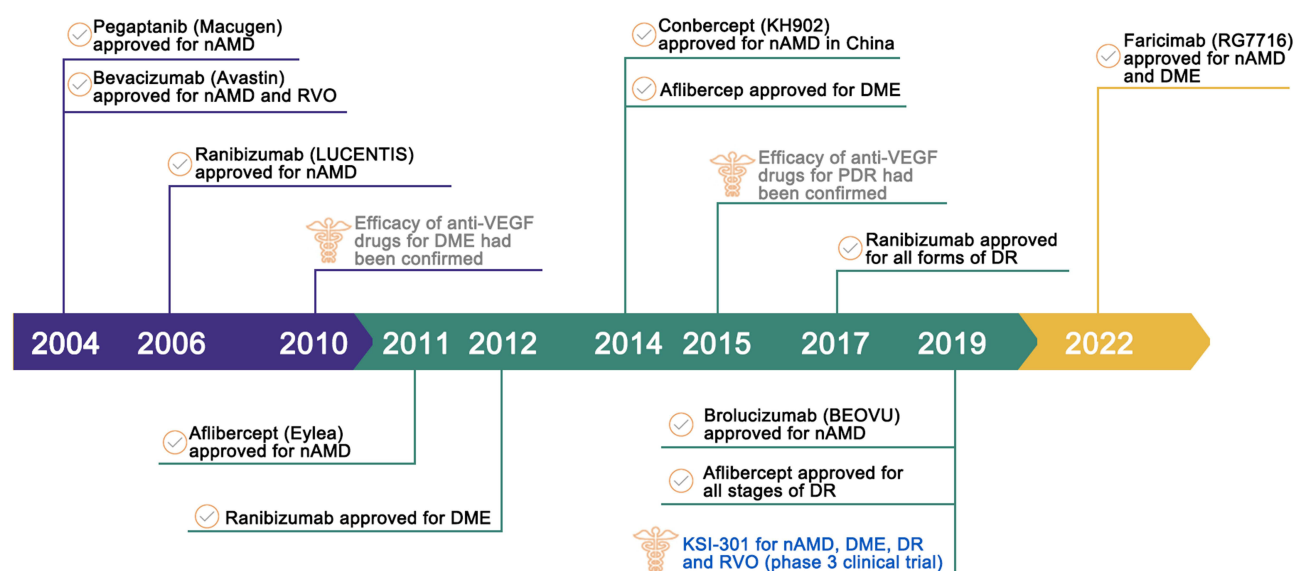


Figure 1 Development timeline of anti-VEGF agents applied in ocular fundus disease. Black: the time indicates the year of FDA approval for anti-VEGF drugs. Blue: the time represents the year of KSI-301 for phase 3 clinical trials. Grey: the time indicates the expanded ophthalmic application of anti-VEGF drugs.

Abbreviations: nAMD, neovascular age-related macular degeneration; RVO, retinal vein occlusion; DME, diabetic macular edema; PDR, proliferative diabetic retinopathy.

Ranibizumab

Ranibizumab (Novartis, Switzerland) is a 48 kDa recombinant humanized monoclonal antibody with affinity toward all isotypes of VEGF-A, inhibiting its biological activity. The lack of a fragment crystallizable (Fc) domain and its small molecule size might expand its affinity for more isoforms of VEGF-A (VEGF165, VEGF121, and VEGF110) and increase its diffusion within the retina and choroid.⁵⁰ Furthermore, since ranibizumab has only one binding site for VEGF, two molecules of ranibizumab bind to each VEGF dimer, with the ranibizumab/VEGF-A complex having a higher stability energy than bevacizumab⁵¹ and greater molecular affinity than bevacizumab and aflibercept.⁵² It was first approved in the United States for intraocular use in 2006 after two Phase 3 studies (MARINA and ANCHOR) demonstrating its safety and efficacy in limiting vision loss and disease progression in neovascular AMD.⁵³

Aflibercept

Aflibercept, also known as VEGF Trap-eye (Eylea[®], Regeneron, Rensselaer, NY, USA), is a 115 kDa recombinant humanized protein that acts as a soluble decoy receptor binding to VEGF-A, VEGF-B, and placental growth factor, thereby stopping the binding and activation of VEGF receptors.⁵⁴ Since its approval by the FDA for use in nAMD following the landmark VIEW1 and VIEW2 studies,⁵⁵ intravitreal aflibercept has been approved for a variety of other pathologies in 2011. There was also a suggestion that injections every 2 months with this VEGF trap may be equivalent to ranibizumab, the gold standard at the time.⁵⁶

Conbercept

Conbercept (Chengdu Kanghong Biotech Company, Sichuan, China) is currently the third most popular molecule belonging to the VEGF Trap family. It consists of a full human DNA sequence of 143 kDa, characterized by the fusion of the extracellular domain 2 of VEGFR1 and the extracellular domains 3 and 4 of VEGFR2 with the Fc portion of human IgG1.^{57,58} The pharmacokinetic profile of conbercept is quite similar to that of aflibercept; the main difference is the presence of a portion dedicated to VEGFR2, which was developed to potentially increase the efficacy and stability of conbercept and to produce a relative affinity for VEGF-A, VEGF-B, VEGF-C, and PGF.^{57,58}

Brolucizumab

Brolucizumab is a single-chain antibody fragment of 26 kDa, characterized by the absence of the Fc portion, and specifically developed to minimize its molecule size and improve its affinity for VEGF-A isoforms compared with the other molecules.⁵⁹ Brolucizumab has been recently approved for use in neovascular age-related macular degeneration, showing noninferiority and higher penetrance within the retina and choroid than the other anti-VEGF molecules.⁶⁰

Abicipar-Pegol

Abicipar-pegol belongs to the family of designed ankyrin repeat protein (DARPin) molecules, a class of molecules that can mimic antibodies and show a high affinity for the VEGF target. More specifically, abicipar-pegol is a recombinant protein of 34 kDa coupled to a polyethylene glycol fraction, binding all VEGF-A isoforms. Its affinity for VEGF-A was found to be comparable to that of aflibercept but remarkably higher than that of bevacizumab and ranibizumab.⁶¹

Faricimab

Faricimab is the second approved molecule belonging to the DARPIn class. The feature of this 150 kDa molecule is that it has two different targets. It can simultaneously and independently bind and neutralize both VEGF-A and Ang-2, the latter enabling interference with the Ang-1/Tie2 pathway.⁶² As mentioned above, the Ang-1/Tie2 pathway is a major pathogenic factor in the development of neovascularization and exudation.⁶³ As a result, faricimab offers an interesting multitarget approach.

KSI-301

KSI-301 (Kodiak Sciences Inc., Nasdaq: KOD, Palo Alto, California) is a new investigational intravitreal anti-VEGF drug of 950 kDa, with a biopolymer of 800 kDa. Its design optimized both its size and molar dose to increase its tissue availability, stability, biocompatibility, potency, and systemic clearance, which in turn are responsible for its durability and design as an optimal ocular PK.⁶⁴ KSI-301 blocks all isoforms of VEGF-A and is administered as an intravitreal injection. It has been shown to bind VEGF-A with high affinity (KD 6.75 pM), higher than its cognate receptors VEGFR1 and VEGFR2, and has a high bioavailability in both the retina and choroid/RPE.⁶⁵ The ocular tissue half-life has been demonstrated to be > 10.5 days in the retina and > 12.5 days in the choroid in rabbit models.⁶⁶

The clinical results (Phase 1a and Phase 1b) of KSI-301 have thus far shown excellent safety and greater efficacy in three retinal diseases: nAMD, DME, and RVO.⁶⁷ The ongoing phase-1b trial has also shown extraordinary biological durability, with most patients achieving treatment-free intervals of 6 months for nAMD, 6 months or longer for DME, and 4 months or longer for RVO.⁶⁸ However, these are interim results, and larger trials with long-term follow-up are required to assess the safety, efficacy, and durability of KSI-301 in all three retinal diseases.

Ziv-Aflibercept

Ziv-aflibercept (Zaltrap; Regeneron, Tarrytown, NY, and Bayer Health care, Leverkusen, Germany; 96 kDa) contains the same therapeutic molecule as Eylea but has a much higher osmolality (1000 mOsm/kg vs 300 mOsm/kg). It was approved in 2012 for the treatment of metastatic colorectal carcinoma.⁶⁹ It has recently been used as an intravitreal treatment for various chorioretinal vascular diseases. Several case series have demonstrated that intravitreal ziv-aflibercept is safe and effective as both short-term and long-term therapy.⁷⁰ Ziv-aflibercept has been used in several areas of the world because it seems to be as effective as other anti-VEGF drugs and is highly cost effective.⁷¹

Limitations of Anti-VEGF Therapy

The human eyeball is a spherical structure with a biological barrier that can protect it against foreign substances, but which limits the treatment routes, including through systemic and anterior segment administration. The presence of the BRB, composed of retinal capillary endothelial cells (inner BRB) and retinal pigment epithelial cells (outer BRB), requires a relatively higher initial drug concentration under a nonlocal administration mode.⁷² Therefore, the most common administration method of anti-VEGF drugs is intravitreal injection. Depending on the half-life of the drug and the severity of the disease, there are several main clinical treatment protocols: 1) monthly intravitreal injections for 3 months followed by an as-needed reinjection schedule; 2) monthly intravitreal injections for 3 months followed by an as-needed reinjection per month; and 3) monthly intravitreal injections for 3 months followed by an as-needed reinjection every 2 months.^{55,73} Some studies have shown that delaying or reducing the number of injections can lead to varying degrees of vision loss.^{55,73}

The use of anti-VEGF drugs has changed the outlook for patients with retinal vascular diseases. However, there are concerns regarding the dosage, timing of injection, and long-term functional outcomes of anti-VEGF treatments. Multiple intravitreal injections indicate a long-term duration of treatment and require good patient compliance. Moreover, repeated treatment places a certain economic and psychological burden on the patients, which may further reduce the patient compliance rate. In addition, repeated intravitreal injections can also cause ocular adverse effects, including persistent intraocular pressure (IOP) increases,⁷⁴ RPE tears,⁷⁵ and geographic atrophy.⁷⁶

Persistent Intraocular Pressure Increase

Serial injections of anti-VEGF agents (ranibizumab or bevacizumab) may lead to a persistent IOP increase.⁷⁴ Transient IOP fluctuations may also cause damage to the retinal nerve fiber layer. After a mean of 20 anti-VEGF injections (range, 8–40 injections), the mean IOP was 29.8 mm Hg (range, 22–58 mm Hg), compared with a baseline of 16.9 mm Hg (range, 14–21 mm Hg). Another study showed that in response to the significant acute and transient IOP increase 5 min after each intravitreal injection of anti-VEGF, Bruch's membrane opening (BMO) enlargement, cup widening and

deepening, and prelaminar tissue thinning were observed.⁷⁷ Compared with baseline values, significant BMO expansion and retinal nerve fiber layer thinning were observed in the third month. In eyes with more than six intravitreal injections, similar immediate post-injection changes, including IOP increase, prelaminar tissue thinning, and cup deepening, were observed at 1 year, while BMO expansion was not significant.⁷⁸

Retinal Pigment Epithelium Tears

The most common cause of an RPE tear is vascularized retinal pigment epithelial detachment (PED) in patients with exudative age-related macular degeneration. Although RPE tears can develop spontaneously in vascularized PEDs, the most recent cases have been associated with anti-VEGF injections.⁷⁵ The subretinal fluid within the PED applies hydrostatic pressure to the RPE and stretches it, and the PED increases as the hydrostatic pressure increases. Contraction of the choroidal neovascular membrane adds tractional forces to the RPE monolayer, especially in larger PEDs.⁷⁵

One observational study retrospectively evaluated the incidence of RPE tears in 72 consecutive patients (74 eyes) treated with intravitreal ranibizumab 0.5 mg for classic CNV. It was found by optical coherence tomography (OCT) and fundus examinations that the incidence of RPE tears in classical CNV patients treated with anti-VEGF was higher than that in untreated patients. The increased risk of an RPE tear after repeated injection of anti-VEGF therapy may be due to increased contraction of the choroidal neovascular membrane.⁷⁹ Another study also found that the first RPE tear occurred after an average of 4.5 ± 2.7 injections of anti-VEGF because of neovascular contraction of a type 1 neovascular complex, adherent to the posterior surface of the RPE and spanning a significant portion of the pigment epithelium detachment area. The second RPE tear occurred after an average of 7.1 ± 5.2 injections of anti-VEGF owing to constriction of the fibrous vessels induced by anti-VEGF therapy. Although a “double RPE tear” occurred, as in previous studies, the patient’s vision was not significantly affected because the macula was not involved.⁸⁰

Geographic Atrophy

Geographic atrophy secondary to AMD is a progressive and irreversible loss of photoreceptors, RPE cells, and choroidal capillaries. Geographic atrophy develops from intravitreal verruca pigmentation, followed by regression of the intravitreal verruca and pigmentation, and then RPE cell death. Some studies have found that geographic atrophy progresses faster in patients receiving anti-VEGF therapy.⁸¹

A retrospective review showed that the number of anti-VEGF injections was correlated with geographic atrophy progression, and each additional injection increased the probability of atrophy by 1.35 times. In this study, 72 eyes of 63 patients with exudative AMD were observed, all receiving ischemia-reperfusion injury (IRI) treatment only with a median number of injections of 6.⁷⁶ Fundus angiography and imaging examination showed that the cumulative number of anti-VEGF injections was significantly correlated with the progression of atrophy. The logistic regression model showed that each additional injection increased the probability of atrophy by 1.35 times. Similarly, in a treatment trial for exudative AMD, the number of anti-VEGF injections was also confirmed to be correlated with geographic atrophy progression.⁸²

Corneal Endothelial Injury and Limbal Mesenchymal Stem Cell (LMSC) Injury

Intravitreal injection is performed on the eye at approximately 3.5 mm to 4 mm behind the limbs, and subconjunctival drug regurgitation can be seen in this area. Anti-VEGF drugs in the aqueous humor are in direct contact with the corneal endothelium and are almost entirely cleared by aqueous circulation from the eyes. Therefore, the risk of corneal endothelial injury associated with repeated intravitreal injections should be considered.

Lass found by specular microscopy that repeated intravitreal aflibercept injection for 52 weeks had no apparent corneal endothelial toxicity in patients treated for neovascular age-related macular degeneration.⁸³ Currently, there is no definite conclusion about the mechanical effect of intravitreal injection or the long-term effects of anti-VEGF drugs on the cornea. However, additional clinical data is still needed to study the long-term risk because anti-VEGF agents are in direct contact with the corneal endothelium.⁸⁴

Studies have found that LMSCs have the potential to treat corneal injuries, and repeated intravitreal injection of anti-VEGF drugs may reduce LMSC reserves due to their toxic effects. There is a case report of a patient who developed iatrogenic limbal stem cell deficiency due to the intravitreal injection of bevacizumab seven times, indicating that repeated surgical trauma to the limbus and surrounding areas may damage the LMSCs.⁸⁵ Moreover, Ugur Acar also confirmed that administration of high doses (5 times and 10 times the clinical dose) or repeated standard doses of intravitreal anti-VEGF agents (aflibercept, ranibizumab, and bevacizumab) may adversely affect the proliferation indices and viability of compact bone derived-mesenchymal stem cells and LMSCs.⁸⁶

Structural Changes in Sclera

Repeated intravitreal anti-VEGF injection may lead to structural changes in the sclera. Zinkernagel assessed the effects of intravitreal anti-VEGF therapy on the scleral structure by spectral domain anterior segment OCT, and patients with surgical trauma from more than 30 intravitreal anti-VEGF injections were found to have a significantly thinner sclera.⁸⁷ These results suggest that alternating the injection site should be considered in patients requiring multiple intravitreal injections.

Choroid Vessel Changes

Studies have shown that anti-VEGF therapy reduces the diameter of the retinal and choroid vessels, reduces the thickness of the choroid, and damages the normal choroidal and retinal blood circulation. Anti-VEGF drugs (bevacizumab) exert two successive effects, including vasoconstriction of the retinal and choroidal blood vessels occurs very soon after injection (from minutes to hours) and is related to the link between VEGF and nitric oxide (NO), followed by a decrease in capillary density, which occurs from hours to a few days after injection.⁸⁸

Kim found via swept-source OCT that the thickness of the choroid was significantly reduced in patients treated with multiple intravitreal anti-VEGF injections over 3 months.⁸⁹ It was also observed that the choroidal blood flow and velocity decreased in patients with at least two intravitreal anti-VEGF injections (bevacizumab or ranibizumab), but such changes were reversible after drug withdrawal. However, if retinal ischemia is present at the same time, it may lead to serious consequences.⁹⁰

In addition, it was observed that anti-VEGF (aflibercept) treatment with three to five injections can effectively reduce the CNV lesion area but increases the vascular density of the CNV as observed by OCT angiography (OCTA), impacting the blood supply of the choroid.⁹¹ Therefore, for patients with repeated intravitreal injection of anti-VEGF drugs, the vascular morphology and distribution in the fundus should be monitored regularly.

Sustained-Release Devices of Anti-VEGF Agents for Retinal Angiogenesis Treatment

Systemic administration, periocular administration, and intravitreal injection of pharmaceuticals are possible routes of drug delivery to the posterior segment of the eye. Delivery of anti-VEGF agents to the posterior segment presents challenges that arise from the clearance pharmacokinetics and anatomy of the eye. The drug concentration in the eyes is considerably lower than the initial concentration after transfer to the blood circulation. The blood-retinal barrier also leads to low efficiency of the systemic administration route. In addition, the drug may have an adverse impact on other organs.⁹²

When using transscleral administration, the drug diffuses through the sclera and circulates to the target tissue of the posterior segment of the eye through the subconjunctival and retrobulbar area, near the posterior sclera or posterior Tenon capsule. Although this method is less invasive, various static barriers of the eye, such as the corneal epithelium, conjunctival epithelium, sclera, choroid, Bruch membrane, and retinal pigment epithelium, and dynamic barriers, including choroidal or conjunctival blood flow, hinder drug diffusion, increase drug loss, and reduce the treatment effects. In addition, the efficiency of drug delivery through the sclera is also affected by the properties of the drug itself, such as charge, hydrophilicity, and molecular weight.⁹³

The intravitreal drug delivery method directly delivers the drug to the posterior segment of the eye, effectively avoiding various barrier effects. Therefore, the current guidelines for the treatment of neovascular ophthalmopathy

mostly recommend the use of intravitreal injection of anti-VEGF drugs.^{94–97} However, as mentioned earlier, most anti-VEGF drugs have a short half-life, and the effective concentration can only be maintained for approximately 1 month.⁹⁸ Even the latest drug to hit the market, brotaruximab, is extended to only 3 months at an effective concentration in the eye.⁶⁰

To solve the problems of a short duration of action and the low effective concentration of drugs in the eye, researchers have focused on the application of sustained-release devices with anti-VEGF agents to meet the above requirements. The BRB helps to further maintain the intraocular concentration of the drug while minimizing systemic absorption and side effects; at the same time, the eye is also immune-exempt, which limits the occurrence of inflammatory responses associated with sustained-release drug carriers.⁹⁹

Currently, according to the research progress, sustained-release anti-VEGF drugs can be divided into a clinical trial phase and a basic investigation phase. According to the sustained release material, it can be divided into degradable materials and nondegradable materials. According to the delivery route of sustained-release drugs, it can be divided into intravitreal injection administration, scleral implantation, and subconjunctival injection administration. Several sustained-released materials and methods have been studied for anti-VEGF ocular delivery and are summarized in Table 1 and described in the following sections.

Sustained-Release Anti-VEGF Agents in the Clinical Trial Phase

Ranibizumab Sustained-Release Device

The Port Delivery System (PDS) with ranibizumab (Roche/Genentech), the most representative of the non-degradable implants, is currently in phase 3 clinical research for the continuous delivery of ranibizumab.^{100–103} The PDS is a permanent, refillable implant inserted through a small incision in the sclera and pars plana. A self-sealing septum in the center of the implant allows the drug to be replenished, thus ensuring that the PDS does not need to be removed from the eye. Ranibizumab moves down the concentration gradient from the implant reservoir by passive diffusion, continuously releasing ranibizumab into the vitreous over time through a porous metal release control element to control the rate of release.¹⁰⁰ The concentration of the drug in the implant gradually decreases over time, and when the concentration of ranibizumab in the PDS implant is 100 mg/mL initially, an effective drug concentration in the vitreous chamber can be achieved, persisting for at least six months.¹⁰¹

In a Phase 2 clinical trial, nAMD patients who received 100 mg/mL PDS showed comparable visual and anatomic outcomes but a reduced total number of ranibizumab intravitreal injections compared with those who received monthly intravitreal ranibizumab (0.5 mg) injections both at 9 month and 22 month, suggesting that PDS exhibited the potential to reduce the treatment burden in nAMD while maintaining vision.^{100,102} As a phase 3 non-inferiority and equivalence trial demonstrated that the adjusted mean change in best-corrected visual acuity (BCVA) score was +0.2 letters in the PDS arm and +0.5 letters in the monthly ranibizumab arm over 36 weeks. Among the patients enrolled, 98.4% did not receive supplemental treatment in the first 24-week interval. In addition, the PDS implants displayed fewer adverse events within 1 month of implantation.¹⁰³ Although the device can reduce the number of intravitreal injections, it still has the problems of poor drug stability and high complications such as endophthalmitis, which need additional larger samples and longer-term safety studies. Moreover, the PDS implants need to be surgically implanted and removed, which reduces the compliance of patients.

GB-102

GB-102 is a tyrosinase inhibitor developed by GrayBug Vision that targets VEGF-A and PDGF. GB-102, the injectable form of sunitinib, forms a depot in the inferior vitreous after being injected, which then gradually releases the drug and biodegrades over time.¹⁰⁴ Studies have shown that a single injection of GB-102 can achieve visual benefits in patients with nAMD after 6 months. In the phase 1/2a clinical study (ADAGIO; NCT03249740), 68% of patients at month 6 had received a single GB-102 injection, and their central subfield thickness (CST) was significantly reduced compared with that before treatment.^{104,105} In a subsequent phase 2b clinical study (ALTISSIMO; NCT03953079), CNV lesions secondary to a previously treated nAMD with the same initial dose of GB-102 every 6 months showed comparable

Table I Sustained-Release Devices of Anti-VEGF Agents for Retinal Angiogenesis Diseases

| Main Sustained-Release Device | Sustained-Release System | Active Molecule | Delivery Route | Degradable | Research Phase | Research Subject | Release Duration | Main Outcomes | Refs. |
|-------------------------------|-----------------------------------|-----------------|---------------------------------------|------------|---|------------------|---------------------------------------|---|-------|
| PDS | PDS | Ranibizumab | Inserted in the sclera and pars plana | No | Phase 2, multicenter, randomized, active treatment-controlled clinical trial | nAMD patients | 9 months | 1) PDS was well tolerated 2) PDS (100 mg/mL) showed visual and anatomic outcomes comparable with monthly intravitreal ranibizumab injections | [100] |
| PDS | PDS | Ranibizumab | Inserted in the sclera and pars plana | No | Phase 2, multicenter, randomized, active treatment-controlled clinical trial | nAMD patients | ~22 months | 1) PDS was well tolerated 2) PDS (100 mg/mL) showed visual and anatomic outcomes comparable with monthly intravitreal ranibizumab injections | [102] |
| PDS | PDS | Ranibizumab | Inserted in the sclera and pars plana | No | Phase 3, open-label, randomized, visual acuity assessor-masked noninferiority and equivalence trial | nAMD patients | 36~40 weeks | 1) Over 98% patients did not receive supplemental treatment in the first 24-week of PDS interval 2) PDS showed comparable BCVA outcomes over weeks 36 | [101] |
| GB-102 | Bioerodible polymer nanoparticles | Sunitinib | Intravitreal injection | Yes | Phase 1/2, multicenter clinical trial | nAMD patients | 6 months | A single dose of GB-102 injection reduced CST of 68% nAMD patients compared with that before treatment | [105] |
| GB-102 | Bioerodible polymer nanoparticles | Sunitinib | Intravitreal injection | Yes | Phase 2b, multicenter, visual examiner-masked, randomized active-controlled clinical trial | nAMD patients | 6 months | Every 6 months of GB-102 injection shown comparable BCVA changes and CST changes at month 10 and 12 | [106] |
| PLGA | NPinPMP | Bevacizumab | Intravitreal injection | Yes | Experiment | Rat | 4 months in vitro 2 months in vivo | 1) Bevacizumab was continuously released from NPinPMP for 4 months in vitro 2) Alexa-bevacizumab from NPinPMP could be detected for 2 months in rat vitreous | [113] |

| | | | | | | | | | |
|----------|--|----------------------------------|------------------------|-----|------------|----------------|--|--|-----------|
| PLGA | Albuminated-PLGA-nanoparticles | Bevacizumab | Intravitreal injection | Yes | Experiment | Rabbit | 8 weeks in vitro 56 days in vivo | 1) Sustained release of bevacizumab for 8 weeks in vitro 2) Nanoparticles with bevacizumab persistence in rabbit vitreous at day 56 | [114] |
| PLGA | PLGA-PEG-PLGA hydrogel | Bevacizumab | Intravitreal injection | Yes | Experiment | Rat Rabbit | 4 weeks in rat vitreous 42 days in rabbit vitreous | PLGA-PEG-PLGA hydrogel sustain released bevacizumab for 4 weeks in rat vitreous and for 42 days in rabbit vitreous | [115,116] |
| PLGA | PLGA/PCADK blend | Bevacizumab-dextran particles | Intravitreal injection | Yes | Experiment | Rabbit | 56 days in vivo | 1) PLGA/PCADK sustain released bevacizumab for 56 days in rabbit vitreous 2) PLGA/PCADK-bevacizumab inhibited the vessels proliferation in vivo | [117] |
| PLGA | PLGA | Ranibizumab biosimilar | Intravitreal injection | Yes | Experiment | HUVEC cells | 3 weeks in vitro | Ranibizumab biosimilar encapsulated PLGA continuously release d the drugs for 3 weeks and inhibited HUVEC cell proliferation and tube formation | [121] |
| Hydrogel | PolyActive™ hydrogel co-polymer | Dual anti-VEGF domain antibodies | Intravitreal injection | Yes | Experiment | Cynomolgus | Over 12 months in vitro 6 months in vivo | 1) Sustained release activity in vitro for more than 12 months 2) Anti-angiogenesis in Cynomolgus CNV model for 6 months | [130] |
| Hydrogel | Hyaluronic acid/dextran-based in situ hydrogel | Bevacizumab | Intravitreal injection | Yes | Experiment | Rabbit | 6 months in vivo | Continually released bevacizumab and maintained concentration up to 6 months | [131] |
| Hydrogel | Hyaluronic acid/dextran-based in situ hydrogel | Anti-VEGF antibody | Intravitreal injection | Yes | Experiment | Rhesus monkeys | At least 5 months | Maintained release in the monkey eye for at least 5 months and preventing the recurrent CNV | [132] |
| Chitosan | Chitosan-PLGA nanoparticles | Bevacizumab | - | Yes | Experiment | Goat sclera | Not provided | 1) Better permeability of bevacizumab across the sclera 2) Longer remaining time on the surface of sclera | [137] |

(Continued)

Table I (Continued).

| Main Sustained-Release Device | Sustained-Release System | Active Molecule | Delivery Route | Degradable | Research Phase | Research Subject | Release Duration | Main Outcomes | Refs. |
|----------------------------------|----------------------------------|--------------------|--------------------------------|------------|----------------|------------------|------------------------------------|--|-------|
| Chitosan | Chitosan-polyelectrolyte complex | Anti-VEGF antibody | - | Yes | Experiment | HUVEC cells | Over 30 days | 1) Sustained release for over 30 days 2) Inhibiting HUVEC cell proliferation and tube formation | [138] |
| Liposomes | Nanoscale liposomes | Bevacizumab | Intravitreal injection | Yes | Experiment | Rabbit | 42 days | Five times higher concentration of bevacizumab released from nanoscale liposomes in rabbit vitreous at day 42 | [139] |
| Liposomes | Bev-MVL | Bevacizumab | Intravitreal injection | Yes | Experiment | Rat | 28 days | Maintained therapeutic concentration of bevacizumab in rat vitreous fluids for 28 days | [140] |
| Light-responsive Nanoparticle | Light-responsive Nanoparticle | Nintedanib | Intravitreal injection | Yes | Experiment | Rat | 30 weeks in vitro 10 weeks in vivo | Inhibition of the rat CNV formation within 10 weeks | [144] |
| BioSilicon | BioSilicon | Bevacizumab | - | No | Experiment | - | 20 days | 1) The release of bevacizumab can be controlled by adjusting the pore size and surface area of BioSilicon 2) 80% bevacizumab can be released from BioSilicon with pores size of 100nm for 20 days | [145] |
| PEGDM/COL | PEGDM/COL | Ranibizumab | Episcleral implant | No | Experiment | Rat | 18 weeks | Ranizumab can penetrate the choroid and retina from PEGFM/COL through the rat sclera, inhibiting CNV formation for up to 18 weeks | [146] |
| Polymer solid lyophilized matrix | Polymer solid lyophilized matrix | Bevacizumab | Subconjunctival administration | Yes | Experiment | Rabbit | 12 weeks | Comparable effects in the rabbit vitreous up to 12 weeks | [147] |

Abbreviations: Refs, references; VEGF, vascular endothelial growth factor; PDS, Port Delivery System; nAMD, neovascular age-related macular degeneration; BCVA, best-corrected visual acuity; CST, central subfield thickness; PLGA, Poly (lactic acid-co-glycolic acid); NPInPMP, nanoparticles in porous microparticles; PLGA-PEG-PLGA, Poly (lactic acid-co-glycolic acid)-poly (ethylene glycol)-poly (lactic acid-co-glycolic acid); PLGA/PCADK, PLGA/poly cyclohexane-1,4-diyl acetone dimethylene ketal; HUVEC, Human umbilical vein endothelial cell; CNV, choroidal neovascularization; Bev-MVL, Bevacizumab-laden polycystic liposomes; PEGDM/COL, photopolymerized poly (ethyleneglycol) dimethacrylate that incorporated collagen microparticles.

BCVA changes and CST changes at months 10 and 12.¹⁰⁶ In addition, animal experiments showed that after a single intravitreal injection, the efficacy of GB-102 in inhibiting VEGF and PDGF could be sustained for up to half a year.¹⁰⁷

Non-degradable implants can achieve long-term release and show good biocompatibility, but when the drug is depleted, the material needs to be removed by secondary surgery, which increases the risks. Degradable anti-VEGF sustained-release drugs mainly use polymer materials to package anti-VEGF drugs or integrate with the drugs to achieve the purpose of sustained release. After the release of the anti-VEGF drug, the carrier material is gradually degraded in the eye without the need for secondary surgery. Furthermore, the release rate can be adjusted by changing its composition and molecular weight.

Sustained-Release Anti-VEGF Agents in the Experimental Phase

In recent years, due to the disadvantages of non-degradable slow-release drugs, more research focus has been placed on biodegradable slow-release drugs. Anti-VEGF sustained-release drugs mainly use polymer materials to package anti-VEGF drugs or integrate with the drugs to achieve the purpose of sustained release. After the release of the anti-VEGF drugs, the carrier materials will gradually degrade in the eye without the need for secondary surgery. The release rate can be adjusted by changing its composition and molecular weight.

Poly(lactic-Co-Glycolic Acid) (PLGA)

PLGA, one of the most widely studied polymers, is a co-polymer of polylactic acid (PLA) and polyglycolic acid (PGA).¹⁰⁸ PLGA is fully biodegradable and self-assembles into nanomicelles, and as one of the best-defined biomaterials for drug delivery. PLGA can be processed into virtually any shape and size, can encapsulate molecules of virtually any size, and is soluble in a variety of common solvents, including chlorinated solvents, tetrahydrofuran, acetone, and ethyl acetate.^{109,110} In water, PLGA is biodegraded by the hydrolysis of ester bonds.¹¹¹ Importantly, PLGA has been approved by the FDA for implant applications and is currently widely studied as a biodegradable controlled-release vehicle for drugs.¹¹² The above physicochemical properties make PLGA an excellent candidate for the intraocular sustained release of anti-VEGF drugs.

In 2013, Sarath K. Yandrapu et al applied supercritical infusion and pressure quenching technology to the preparation of bevacizumab-loaded nanoparticles in porous microparticles (NPinPMP). In vitro, bevacizumab is continuously released from NPinPMP for 4 months, maintaining its monomeric form, conformation, and activity. In addition, the concentration of bevacizumab in the vitreous returned to the baseline level at week 2 after intravitreal injection of bevacizumab in rats. Bevacizumab undergoes sustained release for up to 2 months from bevacizumab-loaded NPinPMP.¹¹³ Albuminated-PLGA-NPs were also used to contain bevacizumab and provided a sustained release of bevacizumab for approximately 2 months. Notably, the bevacizumab concentration in the vitreous of rabbits remained above 500 ng/mL, the minimum effective concentration that completely blocks VEGF-induced angiogenesis for at least 8 weeks.¹¹⁴ The payload of bevacizumab to PLGA-PEG-PLGA hydrogel or PLGA alone can also fulfill the sustained release of bevacizumab in the rat eye for 4 week¹¹⁵ and in the rabbit eye for 42 days.¹¹⁶ Jiaxin Liu et al found that a PLGA/polycyclohexane-1,4-diyl acetone dimethylene ketal (PLGA/PCADK) blend could be loaded with bevacizumab-dextran particles using solid-in-oil-in-water (S/O/W) emulsification, and it exhibited a release behavior of bevacizumab via an increasing total release over 50 days both in vitro and in vivo.¹¹⁷ Nevertheless, most investigations emphasized the modification and optimization of PLGA materials, aiming to manifest better penetration of the formulation and achieve a higher concentration and an extended release duration in the posterior ocular tissues.

To compare the function of bevacizumab-encapsulated PLGA and bevacizumab, XP Zhang et al used an alkaline burn-induced corneal neovascularization model and an oxygen-induced retinopathy model to demonstrate that bevacizumab-loaded PLGA improved the anti-angiogenesis capability for handling both corneal and retinal neovascularization.¹¹⁸ In recent years, considerable evidence has highlighted the critical role of immune inflammatory processes in the pathogenesis of retinal and choroidal neovascularization.¹¹⁹ Accordingly, dexamethasone, a widely used corticosteroid for treating diabetic retinopathy, ocular inflammation, and CNV, was loaded into PLGA/PEI nanoparticles (DPPNs), and bevacizumab was subsequently adsorbed onto the DPPNs (eBev-DPPNs) by static electricity. eBev-

DPPNs presented excellent antiangiogenic efficiency compared with dexamethasone or bevacizumab both in vitro and in a rabbit CNV model.¹²⁰

PLGA is currently the most broadly studied anti-VEGF agent sustained-release material, of which bevacizumab-loaded PLGA is the most widely investigated in retinal and choroidal neovascular diseases. There are also studies that combined ranibizumab biosimilar with PLGA particles. Release experiments in vitro showed that ranibizumab biosimilar could be continuously released from PLGA particles for several weeks, and the ability of ranibizumab-encapsulated PLGA to inhibit vessel proliferation was comparable to that of ranibizumab.¹²¹

Hydrogel-Based Drug Delivery Systems

Hydrogels are an ideal class of material for sustained-release drugs in addition to polymerizing implants and colloidal drug delivery systems.^{122–125} Hydrogels are composed of a three-dimensional network of hydrophilic polymer chains with high water retention capacity, the properties of which can be changed by monomers and crosslinkers.¹²⁶ Due to these properties of hydrogels, they have been successfully applied to soft contact lenses (SCLs), intraocular lenses, and artificial tear fluids, and the use of hydrogels as sustained-release carriers to deliver anti-VEGF drugs has also increased rapidly.¹²⁷ Similar to the challenges of PLGA designs, drug sustained-release hydrogels occupy only a small portion of the vitreous space and typically degrade over time and release the drug, without necessarily requiring excellent optical transparency. As a sustained-release drug system, hydrogels can effectively reduce the number of intraocular injections, thus helping to reduce the risk of endophthalmitis, retinal detachment, and lens injury caused by invasive procedures. In addition, the porous structure of the hydrogel guarantees that it can store higher doses of the drug than the free-drug form.^{127,128}

To be able to control and promote sustained drug release, researchers have developed a variety of sophisticated methods to optimize the formulation of hydrogels. In 2017, Xu et al used four poly (amidoamine) dendrimers to cross-link with PEG and dibenzocyclooctyne groups to promote azide-alkyne cycloaddition to form a hydrogel by strain.¹²⁹ In a study using primates, the researchers applied microparticles of PolyActiveTM hydrogel copolymer as an anti-VEGF delivery base, and two different anti-VEGF domain antibodies (dAb) attached to a human IgG1 Fc region served as dimeric anti-VEGF molecules. The efficacy results showed that this system had a lower IC₅₀ than aflibercept and maintained sustained release activity in vitro for more than 12 months. In vivo studies have shown that this delivery system can exert a protective effect in a cynomolgus CNV model for up to 6 months after a single intravitreal injection.¹³⁰

During the same period, Yu et al implemented catalyst-free chemical crosslinking between vinylsulfone-functionalized hyaluronic acid (HA-vs) and thiolated dextran (Dex-SH) to form an in-situ hydrogel. Mixed bevacizumab, HA-vs, and Dex-SH were injected into rabbit vitreous, and it continually released bevacizumab and maintained its concentration for up to 6 months, 107-fold higher than a bolus injection.¹³¹ Three years later, Yu's team used the same formula of this anti-VEGF controlled-release hydrogel system to prove that a relatively constant concentration of anti-VEGF drug can be maintained in the monkey eye for at least 5 months and that the dosage was sufficient to prevent recurrent CNV after a single injection.¹³²

Overall, hydrogel sustained-release vehicles can achieve longer anti-VEGF drug release than PLGA delivery systems, and their function has been demonstrated in CNV models of primates, although evidence from clinical studies is still lacking.

Chitosan

Chitosan is a natural cationic polymer with the characteristics of hydrophilia, biocompatibility, biodegradability, and mucoadhesivity.^{133–135} Chitosan has similar structural characteristics to specific parts of the extracellular matrix, enabling easier carry of drugs. In addition, chitosan presents a variety of shapes, including nanoparticles, thin films, and hydrogels. Chitosan covalently cross-linked hydrogels have a stable mesh structure and are an ideal biomedical drug carrier material.¹³⁶ Subconjunctival injections and subconjunctival delivery patterns are becoming potential methods for posterior ocular tissue therapy. Based on nanotechnology, chitosan-coated PLGA nanoparticles (NPs) of bevacizumab were prepared. Compared with the drug solution, chitosan-coated NPs demonstrated better permeability of bevacizumab across the sclera and a longer remaining time on the surface of the sclera, thereby improving the drug residence time and permeability. Therefore, bevacizumab-loaded chitosan-coated NPs can be used as candidate carriers for the subconjunctival injection of anti-VEGF drugs.¹³⁷ The chitosan polyelectrolyte complex (PEC) system, containing polysaccharides

alginate and chitosan, can produce a 30-day delivery of anti-VEGF antibodies and inhibit VEGF-induced endothelial cell proliferation and angiogenesis in vitro.¹³⁸

Liposomes

Liposomes are small vesicles made of lipid bilayers that can carry hydrophilic and lipophilic drugs simultaneously. Bevacizumab was encapsulated in nanoscale liposomes, and the concentration of free bevacizumab in the eyes receiving liposome bevacizumab was five times higher at 42 days compared to rabbit eyes with glass cavity injection of soluble bevacizumab.¹³⁹ There are also been studies of bevacizumab-laden polycystic liposomes (Bev-MVL) that more effectively maintain the therapeutic level of bevacizumab in vitreous fluids for a longer duration than bevacizumab solutions (Bev-S) for 28 days in rat CNV.¹⁴⁰

Light-Responsive Nanoparticle

Conventional anti-VEGF sustained-release drugs are released according to the sustained-release characteristics of the drug design after injection into the vitreous cavity, and their release cannot be controlled after injection. One class of drugs is prepared using triggered carriers that can be administered with high spatial and temporal resolution through external triggering. Taking advantage of the photoreceptor system of the eyeball, it is theoretically feasible to use light to control the drug release, and an injectable implant (ISFI) formed in situ in response to the light is a photosensitive liquid that can be injected into the vitreous body in a minimally invasive manner. Through noninvasive light irradiation of the cornea, it quickly changes its physical properties to form a transparent gel to achieve sustained release and improve the bioavailability of the drug.¹⁴¹

Studies have demonstrated the possibility of using light to control nanoparticles targeting cells and tumors. In studies using photosensitive nanoparticles to treat CNV, intravenous administration and intravitreal administration have both been reported,^{142,143} such as vitreous cavity photosensitive anti-VEGF nanoparticles, which stably retain encapsulated angiogenesis inhibitor (nintedanib) molecules in the vitreous and can trigger drug release for up to 30 weeks after injection by exposure to far ultraviolet light (UV) on demand, and animal experiments have also confirmed that it can inhibit the formation of rat CNV for up to 10 weeks.¹⁴⁴ The advantage is that light-responsive nanoparticles can trigger the on-demand release of the drug in a noninvasive manner in vitro, but there are also safety issues such as potential damage from the trigger source (far-ultraviolet radiation) to other tissues of the eye.

Others

BioSilicon, an oxidized porous silicon, was prepared with bevacizumab as its substrate. The release of bevacizumab can be controlled by adjusting the pore size and surface area of BioSilicon. Bevacizumab (80%) can be released from BioSilicon with a pore size of 100 nm for 20 days.¹⁴⁵

Photopolymerized poly (ethylene glycol) dimethacrylate incorporating collagen microparticles (PEGDM/COL) was developed as an episcleral implantable device for the sustained release of ranibizumab. By implanting the device through the sclera, ranibizumab can penetrate the choroid and retina through the rat sclera, inhibiting CNV formation for up to 18 weeks.¹⁴⁶

In addition, Burgalassi et al produced a polymer solid lyophilized matrix containing bevacizumab. A comparative study in rabbit eyes showed that the injection of lyophilized matrix under the conjunctiva could be used as a posterior ocular delivery system of bevacizumab, which is less invasive and has a longer drug release time.¹⁴⁷

Conclusion

In this review, we have summarized several retinal vascular diseases characterized by RNV as the main pathological feature and listed the current research status and the challenges of anti-VEGF drugs with FDA approval or those still in the research phases, as well as studies on the most common sustained-release materials of anti-VEGF drugs to address these challenges and dilemmas.

Although researchers are continuously trying to develop a variety of anti-neovascular therapies that are more efficient, have a longer half-life, and have safer and noninvasive delivery methods, the development and research of sustained-

release strategies for existing anti-VEGF drugs seem to be able to address the limitations of current anti-VEGF drug treatments faster and better. The advantages of sustained-release dosage forms of anti-VEGF agents include: 1) long-term therapeutic effects by continuous release of the medicine; 2) reduces frequency of dosing; 3) prolonged action of the medicaments; 4) control of drug therapy; 5) ability to modify the extent and rate of drug absorption; 5) improves patient compliance.

According to the recent publications, the most well-studied sustained-release materials include degradable and non-degradable materials. Degradable materials are now widely preferred by researchers because there is no need for secondary surgery to remove them. However, it's worth noticing that it is necessary to receive other intraocular injections of anti-VEGF agents if the degradable materials were degraded before completely controlling disease progression. The main advantage of non-degradable materials is that they can be repeatedly supplemented with drugs. The operation of supplementary drugs is simpler compared with intraocular injection, and as long as the implant material does not cause side effects on the eye, it can be placed there for a long time. Thereby ensuring maximum prolongation of the action time of anti-VEGF drugs. In addition, light-responsive nanoparticle also shows unique advantages for ocular applications, with the potential of triggering the on-demand release of the drug in a noninvasive manner. What can not be ignored is light-responsive strategies may cause the potential damage from far-ultraviolet radiation to tissues of the eye.

In terms of the anti-VEGF drugs carried in different forms, bevacizumab is the most commonly used drug, followed by ranibizumab, probably because these two drugs may have been used for the treatment of RNV diseases for a relatively longer time. With the ideal therapeutic effects of ranibizumab in ophthalmic research field, the combination of ranibizumab and sustained-released materials may provide convenience and benefits for patients.

The main concern for the application of sustained-released strategies is the effective duration. Most of the strategies can reach a release time of at least one and a half months, PDS shows a continuous release of anti-VEGF drugs for 1 year, and hydrogel-carried anti-VEGF drugs can also achieve sustainable release in the vitreous cavity of primates for six months to one year and can achieve therapeutic effects. Regarding to the research progress, the investigations on PDS and GB-102 are significantly ahead of other drugs and have entered the phase 2 and 3 clinical research phases.

Notably, most of the investigations are in the basic research stages. The use of PLGA has been reported more frequently, and hydrogel research has used primates as its main model to explore the effects of sustained-release anti-VEGF drugs. We should also note that some studies of sustained-release materials have focused more on demonstrating long-term drug release *in vitro* rather than evaluating the effects of anti-VEGF itself. Whether the mechanisms of sustained-released anti-VEGF drugs were changed needs more proofs.

The main problem facing the research is how to select and prepare the drug carrier, which can meet the requirements of stable combination with anti-VEGF drugs, long released periods, enhanced intraocular penetration and good biocompatibility. Furthermore, whether in basic or clinical studies, the abundance of anti-retinal angiogenesis drugs has shown surprising and promising outcomes. Studies on the administration of anti-VEGF with sustained-release materials are showing increasingly good progress. In the future, with advances in the development of nanotechnology, new materials, and the cooperation and complementarity among different research fields, we will achieve more efficient and safer therapeutic strategies for ocular fundus diseases.

Abbreviation

DME, diabetic macular edema; AMD, age-related macular degeneration; VEGF, vascular endothelial growth factor; RNV, retinal neovascularization; PDS, port delivery system; PLGA, polylactic-co-glycolic acid; DR, diabetic retinopathy; RVO, retinal vein obstruction; ROP, retinopathy of prematurity; HIF-1 α , hypoxia-inducible factor-1 α ; PDGF, platelet-derived growth factor; DM, diabetes mellitus; NPDR, nonproliferative diabetic retinopathy; PDR, proliferative diabetic retinopathy; DME, diabetic macular edema; BRB, blood-retinal barrier; CRVO, central retinal vein occlusion; BRVO, branch retinal vein occlusion; RPE, retinal pigment epithelium; CNV, choroidal neovascularization; FDA, Food and Drug Administration; nAMD, neovascular age-related macular degeneration; IOP, intraocular pressure; BMO, Bruch's membrane opening; PED, pigment epithelial detachment; IRI, ischaemia-reperfusion injury; LMSC, limbal mesenchymal stem cell; OCT, optical coherence tomography; OCTA, OCT angiography; BCVA, best-corrected visual

acuity; CST, central subfield thickness; PLA, polylactic acid; PGA, polyglycolic acid; NPInPMP, nanoparticles in porous microparticles; PLGA/PCADK, PLGA/polycyclohexane-1,4-diyl acetone dimethylene ketal; S/O/W, solid-in-oil-in-water; DPPNs, PLGA/PEI nanoparticles; SCLs, soft contact lenses; NPs, nanoparticles; ISFI, injectable implant; UV, ultraviolet light.

Data Sharing Statement

All data generated or analyzed during this study are included in this published article and are available from the corresponding author, Prof.Xiaorong Li (email address: xiaorli@163.com), on reasonable request.

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Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

Disclosure

The authors declare that they have no competing interests.

References

- Mitchell P, Liew G, Gopinath B, et al. Age-related macular degeneration. *Lancet*. 2018;392(10153):1147–1159. doi:10.1016/s0140-6736(18)31550-2
- Cheung N, Mitchell P, Wong TY. Diabetic retinopathy. *Lancet*. 2010;376(9735):124–136. doi:10.1016/s0140-6736(09)62124-3
- Tang Y, Cheng Y, Wang S, et al. Review: the development of risk factors and cytokines in retinal vein occlusion. *Front Med*. 2022;9:910600. doi:10.3389/fmed.2022.910600
- Dai C, Xiao J, Wang C, et al. Neurovascular abnormalities in retinopathy of prematurity and emerging therapies. *J Mol Med*. 2022;100(6):817–828. doi:10.1007/s00109-022-02195-2
- Huang YH, Kuo CH, Peng IC, et al. Recombinant thrombomodulin domain 1 rescues pathological angiogenesis by inhibition of hif-1 α -VEGF pathway. *Cell Mol Life Sci*. 2021;78(23):7681–7692. doi:10.1007/s00018-021-03950-3
- Uemura A, Fruttiger M, D'Amore PA, et al. Vegfr1 signaling in retinal angiogenesis and microinflammation. *Prog Retin Eye Res*. 2021;84:100954.
- Huang H. Pericyte-endothelial interactions in the retinal microvasculature. *Int J Mol Sci*. 2020;21(19). doi:10.3390/ijms21197413
- Huemer J, Khalid H, Wagner SK, et al. Phenotyping of retinal neovascularization in ischemic retinal vein occlusion using wide field OCT angiography. *Eye*. 2021;35(10):2812–2819. doi:10.1038/s41433-020-01317-9
- Mortezaee K. Hypoxia induces core-to-edge transition of progressive tumoral cells: a critical review on differential yet corroborative roles for hif-1 α and hif-2 α . *Life Sci*. 2020;242:117145. doi:10.1016/j.lfs.2019.117145
- Rattner A, Williams J, Nathans J. Roles of HIFs and VEGF in angiogenesis in the retina and brain. *J Clin Invest*. 2019;129(9):3807–3820. doi:10.1172/jci126655
- Majidpoor J, Mortezaee K. Angiogenesis as a hallmark of solid tumors - clinical perspectives. *Cell Oncol*. 2021;44(4):715–737. doi:10.1007/s13402-021-00602-3
- Campbell M, Doyle SL. Current perspectives on established and novel therapies for pathological neovascularization in retinal disease. *Biochem Pharmacol*. 2019;164:321–325. doi:10.1016/j.bcp.2019.04.029
- Jiang G, Han X, Qiao K, et al. Therapeutic effect of intravitreal anti-VEGF drugs on retinal neovascularization in diabetic retinopathy. *Minerva Med*. 2022. doi:10.23736/s0026-4806.22.07943-5
- Arrigo A, Aragona E, Bandello F. VEGF-targeting drugs for the treatment of retinal neovascularization in diabetic retinopathy. *Ann Med*. 2022;54(1):1089–1111. doi:10.1080/07853890.2022.2064541
- Tomita Y, Lee D, Tsubota K, et al. Updates on the current treatments for diabetic retinopathy and possibility of future oral therapy. *J Clin Med*. 2021;10(20). doi:10.3390/jcm10204666
- Lashay A, Riaz-Esfahani H, Mirghorbani M, et al. Intravitreal medications for retinal vein occlusion: systematic review and meta-analysis. *J Ophthalmic Vis Res*. 2019;14(3):336–366. doi:10.18502/jovr.v14i3.4791
- Hayreh SS. Photocoagulation for retinal vein occlusion. *Prog Retin Eye Res*. 2021;85:100964. doi:10.1016/j.preteyeres.2021.100964

18. Maxwell JD, Greig WR, Boyle JA, et al. Reiter's syndrome and psoriasis. *Scott Med J*. 1966;11(1):14–18. doi:10.1177/003693306601100103
19. Hirschke K, Bühler R, Apell HJ, et al. Inactivation of the Na, k-ATPase by radiation-induced free radicals. Evidence for a radical-chain mechanism. *FEBS Lett*. 1994;353(3):297–300. doi:10.1016/0014-5793(94)01067-6
20. Domańska-Janik K. Experimental hypoxia and some problems of oxygenic glucose metabolism in the central nervous system. *Neuropatol Pol*. 1972;10(1):17–43.
21. Brinkmann A, Winkelman K, Kückenmeister T, et al. Effect of long-term anti-VEGF treatment on viability and function of rpe cells. *Curr Eye Res*. 2022;47(1):127–134. doi:10.1080/02713683.2021.1931344
22. Zhao HY, Wu J, Zhu JJ, et al. Research advances in tissue engineering materials for sustained release of growth factors. *Biomed Res Int*. 2015;2015:808202. doi:10.1155/2015/808202
23. Stitt AW, Curtis TM, Chen M, et al. The progress in understanding and treatment of diabetic retinopathy. *Prog Retin Eye Res*. 2016;51:156–186. doi:10.1016/j.preteyeres.2015.08.001
24. Campochiaro PA, Brown DM, Pearson A, et al. Sustained delivery fluocinolone acetonide vitreous inserts provide benefit for at least 3 years in patients with diabetic macular edema. *Ophthalmology*. 2012;119(10):2125–2132. doi:10.1016/j.ophtha.2012.04.030
25. Zhang X, Zeng H, Bao S, et al. Diabetic macular edema: new concepts in patho-physiology and treatment. *Cell Biosci*. 2014;4:27. doi:10.1186/2045-3701-4-27
26. Boyer DS, Yoon YH, Belfort R Jr, et al. Three-year, randomized, sham-controlled trial of dexamethasone intravitreal implant in patients with diabetic macular edema. *Ophthalmology*. 2014;121(10):1904–1914. doi:10.1016/j.ophtha.2014.04.024
27. Hayreh SS, Zimmerman MB. Branch retinal vein occlusion: natural history of visual outcome. *JAMA Ophthalmol*. 2014;132(1):13–22. doi:10.1001/jamaophthalmol.2013.5515
28. Kolar P. Risk factors for central and branch retinal vein occlusion: a meta-analysis of published clinical data. *J Ophthalmol*. 2014;2014:724780. doi:10.1155/2014/724780
29. Yang LP, McKeage K. Intravitreal aflibercept (Eylea®): a review of its use in patients with macular oedema secondary to central retinal vein occlusion. *Drugs Aging*. 2014;31(5):395–404. doi:10.1007/s40266-014-0176-2
30. Sangroongruangsri S, Ratanapakorn T, Wu O, et al. Comparative efficacy of bevacizumab, ranibizumab, and aflibercept for treatment of macular edema secondary to retinal vein occlusion: a systematic review and network meta-analysis. *Expert Rev Clin Pharmacol*. 2018;11(9):903–916. doi:10.1080/17512433.2018.1507735
31. Rogers SL, McIntosh RL, Lim L, et al. Natural history of branch retinal vein occlusion: an evidence-based systematic review. *Ophthalmology*. 2010;117(6):1094–1101.e1095. doi:10.1016/j.ophtha.2010.01.058
32. Rhoades W, Dickson D, Nguyen QD, et al. Management of macular edema due to central retinal vein occlusion - The role of aflibercept. *Taiwan J Ophthalmol*. 2017;7(2):70–76. doi:10.4103/tjo.tjo_9_17
33. Rein DB, Wittenborn JS, Zhang X, et al. Forecasting age-related macular degeneration through the year 2050: the potential impact of new treatments. *Arch Ophthalmol*. 2009;127(4):533–540. doi:10.1001/archophthalmol.2009.58
34. Wong WL, Su X, Li X, et al. Global prevalence of age-related macular degeneration and disease burden projection for 2020 and 2040: a systematic review and meta-analysis. *Lancet Glob Health*. 2014;2(2):e106–e116. doi:10.1016/s2214-109x(13)70145-1
35. Ambati J, Fowler BJ. Mechanisms of age-related macular degeneration. *Neuron*. 2012;75(1):26–39. doi:10.1016/j.neuron.2012.06.018
36. Bhutto I, Luttly G. Understanding age-related macular degeneration (AMD): relationships between the photoreceptor/retinal pigment epithelium/bruch's membrane/choriocapillaris complex. *Mol Aspects Med*. 2012;33(4):295–317. doi:10.1016/j.mam.2012.04.005
37. Anguita R, Tasiopoulou A, Shahid S, et al. A review of aflibercept treatment for macular disease. *Ophthalmol Ther*. 2021;10(3):413–428. doi:10.1007/s40123-021-00354-1
38. Lim LS, Mitchell P, Seddon JM, et al. Age-related macular degeneration. *Lancet*. 2012;379(9827):1728–1738. doi:10.1016/s0140-6736(12)60282-7
39. Sen P, Agarwal AAK, Bhende P, et al. Treatment outcomes of combination of anti-vascular endothelial growth factor injection and laser photocoagulation in type 1 rop and apmp. *Int Ophthalmol*. 2022;42(1):95–101. doi:10.1007/s10792-021-02004-8
40. Blencowe H, Lawn JE, Vazquez T, et al. Preterm-associated visual impairment and estimates of retinopathy of prematurity at regional and global levels for 2010. *Pediatr Res*. 2013;74(Suppl1):35–49. doi:10.1038/pr.2013.205
41. Kim SJ, Port AD, Swan R, et al. Retinopathy of prematurity: a review of risk factors and their clinical significance. *Surv Ophthalmol*. 2018;63(5):618–637. doi:10.1016/j.survophthal
42. Mintz-Hittner HA, Kennedy KA, Chuang AZ. Efficacy of intravitreal bevacizumab for stage 3+ retinopathy of prematurity. *N Engl J Med*. 2011;364(7):603–615. doi:10.1056/NEJMoa1007374
43. Kychevalou A, Dorta P, Katz X. Zone I retinopathy of prematurity: clinical characteristics and treatment outcomes. *Retina*. 2006;26(7 Suppl):S11–S15. doi:10.1097/01.iae.0000244285.79004.e6
44. Lee JH, Canny MD, De Erkenez A, et al. A therapeutic aptamer inhibits angiogenesis by specifically targeting the heparin binding domain of vegf165. *Proc Natl Acad Sci U S A*. 2005;102(52):18902–18907. doi:10.1073/pnas.0509069102
45. Gragoudas ES, Adamis AP, Cunningham ET Jr, et al. Pegaptanib for neovascular age-related macular degeneration. *N Engl J Med*. 2004;351(27):2805–2816. doi:10.1056/NEJMoa042760
46. Ferrara N, Hillan KJ, Novotny W. Bevacizumab (avastin), a humanized anti-VEGF monoclonal antibody for cancer therapy. *Biochem Biophys Res Commun*. 2005;333(2):328–335. doi:10.1016/j.bbrc.2005.05.132
47. Kreisl TN, Kim L, Moore K, et al. Phase II trial of single-agent bevacizumab followed by bevacizumab plus irinotecan at tumor progression in recurrent glioblastoma. *J Clin Oncol*. 2009;27(5):740–745. doi:10.1200/jco.2008.16.3055
48. Spasic M, Chow F, Tu C, et al. Molecular characteristics and pathways of avastin for the treatment of glioblastoma multiforme. *Neurosurg Clin N Am*. 2012;23(3):417–427. doi:10.1016/j.nec.2012.05.002
49. Rosenfeld PJ, Moshfeghi AA, Puliafito CA. Optical coherence tomography findings after an intravitreal injection of bevacizumab (avastin) for neovascular age-related macular degeneration. *Ophthalmic Surg Lasers Imaging*. 2005;36(4):331–335.
50. Ferrara N, Damico L, Shams N, et al. Development of ranibizumab, an anti-vascular endothelial growth factor antigen binding fragment, as therapy for neovascular age-related macular degeneration. *Retina*. 2006;26(8):859–870. doi:10.1097/01.iae.0000242842

51. Platania CB, Di Paola L, Leggio GM, et al. Molecular features of interaction between VEGFA and anti-angiogenic drugs used in retinal diseases: a computational approach. *Front Pharmacol*. 2015;6:248. doi:10.3389/fphar.2015.00248
52. Yang J, Wang X, Fuh G, et al. Comparison of binding characteristics and in vitro activities of three inhibitors of vascular endothelial growth factor A. *Mol Pharm*. 2014;11(10):3421–3430. doi:10.1021/mp500160v
53. Rosenfeld PJ, Brown DM, Heier JS, et al. Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med*. 2006;355(14):1419–1431. doi:10.1056/NEJMoa054481
54. Pham B, Thomas SM, Lillie E, et al. Anti-vascular endothelial growth factor treatment for retinal conditions: a systematic review and meta-analysis. *BMJ Open*. 2019;9(5):e022031. doi:10.1136/bmjopen-2018-022031
55. Heier JS, Brown DM, Chong V, et al. Intravitreal aflibercept (VEGF trap-eye) in wet age-related macular degeneration. *Ophthalmology*. 2012;119(12):2537–2548. doi:10.1016/j.ophtha.2012.09.006
56. Arrigo A, Bandello F. Molecular features of classic retinal drugs, retinal therapeutic targets and emerging treatments. *Pharmaceutics*. 2021;13(7). doi:10.3390/pharmaceutics.13071102
57. Zhang M, Zhang J, Yan M, et al. Recombinant anti-vascular endothelial growth factor fusion protein efficiently suppresses choroidal neovascularization in monkeys. *Mol Vis*. 2008;14:37–49.
58. Suto K, Yamazaki Y, Morita T, et al. Crystal structures of novel vascular endothelial growth factors (VEGF) from snake venoms: insight into selective VEGF binding to kinase insert domain-containing receptor but not to fms-like tyrosine kinase-1. *J Biol Chem*. 2005;280(3):2126–2131. doi:10.1074/jbc.M411395200
59. Yannuzzi NA, Freund KB. Brolucizumab: evidence to date in the treatment of neovascular age-related macular degeneration. *Clin Ophthalmol*. 2019;13:1323–1329. doi:10.2147/ophth.s184706
60. Nguyen QD, Das A, Do DV, et al. Brolucizumab: evolution through preclinical and clinical studies and the implications for the management of neovascular age-related macular degeneration. *Ophthalmology*. 2020;127(7):963–976. doi:10.1016/j.ophtha.2019.12.031
61. Rodrigues GA, Mason M, Christie LA, et al. Functional characterization of abicipar-pegol, an anti-VEGF darpin therapeutic that potently inhibits angiogenesis and vascular permeability. *Invest Ophthalmol Vis Sci*. 2018;59(15):5836–5846. doi:10.1167/iov.18-25307
62. Nicolò M, Ferro Desideri L, Vagge A, et al. Faricimab: an investigational agent targeting the tie-2/angiopoietin pathway and VEGF-a for the treatment of retinal diseases. *Expert Opin Investig Drugs*. 2021;30(3):193–200. doi:10.1080/13543784.2021.1879791
63. Sahni J, Patel SS, Dugel PU, et al. Simultaneous inhibition of angiopoietin-2 and vascular endothelial growth factor-a with faricimab in diabetic macular edema: boulevard phase 2 randomized trial. *Ophthalmology*. 2019;126(8):1155–1170. doi:10.1016/j.ophtha.2019.03.023
64. Stern HD, Hussain RM. Ksi-301: an investigational anti-VEGF biopolymer conjugate for retinal diseases. *Expert Opin Investig Drugs*. 2022;31(5):443–449. doi:10.1080/13543784.2022.2052042
65. Chandrasekaran PR, Madanagopalan VG. Ksi-301: antibody biopolymer conjugate in retinal disorders. *Ther Adv Ophthalmol*. 2021;13:25158414211027708. doi:10.1177/25158414211027708
66. Liang H, Huang X, Ngo W, et al. Ksi-301: an anti-VEGF antibody biopolymer conjugate with extended half-life for treatment of neovascular retinal diseases. *Invest Ophthalmol Vis Sci*. 2018;59:211.
67. Patel S, Naor J, Qudrat A, et al. Phase 1 first-in-human study of ksi-301: a novel anti-VEGF antibody biopolymer conjugate with extended durability. *Invest Ophthalmol Vis Sci*. 2019;60:3670.
68. Diana V. Extended durability in exudative retinal diseases using the novel intravitreal anti-VEGF antibody biopolymer conjugate ksi-301: update from phase 1b study in patients with wamd, dme and rvo. In: *Angiogenesis, Exudation and Degeneration, 8 February 2020*. Kodiak sciences; 2020. <https://ir.kodiak.com/static-files/81d12c4b-cff4-4298-991c-a80e7be51ec5>.
69. Singh SR, Dogra A, Stewart M, et al. Intravitreal ziv-aflibercept: clinical effects and economic impact. *Asia Pac J Ophthalmol*. 2017;6(6):561–568. doi:10.22608/apo.2017263
70. Chhablani J, Narayanan R, Mathai A, et al. Short-term safety profile of intravitreal ziv-aflibercept. *Retina*. 2016;36(6):1126–1131. doi:10.1097/iae.0000000000000913
71. Ashraf M, El Kayal H, Souka AAR. Comparison between the short-term outcomes of bevacizumab and ziv-aflibercept in the treatment of primary diabetic macular oedema. *Acta Ophthalmol*. 2017;95(8):e803–e804. doi:10.1111/aos.13352
72. Fogli S, Del Re M, Rofi E, et al. Clinical pharmacology of intravitreal anti-VEGF drugs. *Eye*. 2018;32(6):1010–1020. doi:10.1038/s41433-018-0021-7
73. Schmidt-Erfurth U, Eldem B, Guymer R, et al. Efficacy and safety of monthly versus quarterly ranibizumab treatment in neovascular age-related macular degeneration: the excite study. *Ophthalmology*. 2011;118(5):831–839. doi:10.1016/j.ophtha.2010.09.004
74. Tseng JJ, Vance SK, Della Torre KE, et al. Sustained increased intraocular pressure related to intravitreal anti-vascular endothelial growth factor therapy for neovascular age-related macular degeneration. *J Glaucoma*. 2012;21(4):241–247. doi:10.1097/IJG.0b013e31820.d7d19
75. Ersoz MG, Karacorlu M, Arf S, et al. Retinal pigment epithelium tears: classification, pathogenesis, predictors, and management. *Surv Ophthalmol*. 2017;62(4):493–505. doi:10.1016/j.survophthal.2017.03.004
76. Lois N, McBain V, Abdelkader E, et al. Retinal pigment epithelial atrophy in patients with exudative age-related macular degeneration undergoing anti-vascular endothelial growth factor therapy. *Retina*. 2013;33(1):13–22. doi:10.1097/IAE.0b013e3182657fff
77. Gómez-Mariscal M, Puerto B, Muñoz-Negrete FJ, et al. Acute and chronic optic nerve head biomechanics and intraocular pressure changes in patients receiving multiple intravitreal injections of anti-VEGF. *Graefes Arch Clin Exp Ophthalmol*. 2019;257(10):2221–2231. doi:10.1007/s00417-019-04354-7
78. Filek R, Hooper P, Sheidow TG, et al. Two-year analysis of changes in the optic nerve and retina following anti-VEGF treatments in diabetic macular edema patients. *Clin Ophthalmol*. 2019;13:1087–1096. doi:10.2147/ophth.s199758
79. Konstantinidis L, Ambresin A, Zografos L, et al. Retinal pigment epithelium tears after intravitreal injection of ranibizumab for predominantly classic neovascular membranes secondary to age-related macular degeneration. *Acta Ophthalmol*. 2010;88(7):736–741. doi:10.1111/j.1755-3768.2009.01547.x
80. Mouallem A, Sarraf D, Chen X, et al. Double retinal pigment epithelium tears in neovascular age-related macular degeneration. *Retina*. 2016;36(11):2197–2204. doi:10.1097/iae.0000000000001062
81. Fleckenstein M, Mitchell P, Freund KB, et al. The progression of geographic atrophy secondary to age-related macular degeneration. *Ophthalmology*. 2018;125(3):369–390. doi:10.1016/j.ophtha.2017.08.038

82. Sophie R, Wang J, Campochiaro PA, et al. Risk of geographic atrophy in the comparison of age-related macular degeneration treatments trials. *Ophthalmology*. 2014;121(7):e34. doi:10.1016/j.ophtha.2013.12.043
83. Lass JH, Benetz BA, Menegay HJ, et al. Effects of repeated intravitreal aflibercept injection on the corneal endothelium in patients with age-related macular degeneration: outcomes from the re-view study. *Cornea*. 2018;37(5):596–601. doi:10.1097/ico.0000000000001535
84. Urban B, Szabowicz M, Bakunowicz-lazarczyk A. Effect of repeated intravitreal ranibizumab and aflibercept injections on the cornea in patients with age-related macular degeneration. *J Ophthalmol*. 2020;2020:4928905. doi:10.1155/2020/4928905
85. Capella MJ, Alvarez de toledo J, De la paz MF. Insuficiencia limbar secundaria a múltiples inyecciones intravítreas. [Limbal stem cell deficiency following multiple intravitreal injections]. *Arch Soc Esp Oftalmol*. 2011;86(3):89–92. Spanish. doi:10.1016/j.oftal.2010.11.018
86. Acar U, Erginturk Acar D, Alpaslan Pinarli F, et al. Effects of commonly used intravitreal anti-vascular endothelial growth factor drugs on mesenchymal stem cells derived from the limbus and ciliary body. *Clin Exp Ophthalmol*. 2016;44(7):587–596. doi:10.1111/ceo.12715
87. Zinkernagel MS, Schorno P, Ebnetter A, et al. Scleral thinning after repeated intravitreal injections of antivascular endothelial growth factor agents in the same quadrant. *Invest Ophthalmol Vis Sci*. 2015;56(3):1894–1900. doi:10.1167/iov.14-16204
88. Schraermeyer U, Julien S. Effects of bevacizumab in retina and choroid after intravitreal injection into monkey eyes. *Expert Opin Biol Ther*. 2013;13(2):157–167. doi:10.1517/14712598.2012.748741
89. Kim SW, Woo JE, Yoon YS, et al. Retinal and choroidal changes after anti vascular endothelial growth factor therapy for neovascular age-related macular degeneration. *Curr Pharm Des*. 2019;25(2):184–189. doi:10.2174/1381612825666190319165824
90. Bonnin P, Pournaras JA, Lazrak Z, et al. Ultrasound assessment of short-term ocular vascular effects of intravitreal injection of bevacizumab (avastin®) in neovascular age-related macular degeneration. *Acta Ophthalmol*. 2010;88(6):641–645. doi:10.1111/j.1755-3768.2009.01526.x
91. Wylęgała A, Wylęgała F, Wylęgała E. Aflibercept treatment leads to vascular abnormalization of the choroidal neovascularization. *J Healthc Eng*. 2018;2018:8595278. doi:10.1155/2018/8595278
92. Maurice D. Review: practical issues in intravitreal drug delivery. *J Ocul Pharmacol Ther*. 2001;17(4):393–401. doi:10.1089/108076801753162807
93. Nomoto H, Shiraga F, Kuno N, et al. Pharmacokinetics of bevacizumab after topical, subconjunctival, and intravitreal administration in rabbits. *Invest Ophthalmol Vis Sci*. 2009;50(10):4807–4813. doi:10.1167/iov.08-3148
94. Kertes PJ, Galic IJ, Greve M, et al. Canadian treat-and-extend analysis trial with ranibizumab in patients with neovascular age-related macular disease: one-year results of the randomized Canadian treat-and-extend analysis trial with ranibizumab study. *Ophthalmology*. 2019;126(6):841–848. doi:10.1016/j.ophtha.2019.01.013
95. Gillies M, Arnold J, Bhandari S, et al. Ten-year treatment outcomes of neovascular age-related macular degeneration from two regions. *Am J Ophthalmol*. 2020;210:116–124. doi:10.1016/j.ajo.2019.10.007
96. Dugel PU, Koh A, Ogura Y, et al. Hawk and harrier: phase 3, multicenter, randomized, double-masked trials of brolucizumab for neovascular age-related macular degeneration. *Ophthalmology*. 2020;127(1):72–84. doi:10.1016/j.ophtha.2019.04.017
97. Brynskov T, Munch IC, Larsen TM, et al. Real-world 10-year experiences with intravitreal treatment with ranibizumab and aflibercept for neovascular age-related macular degeneration. *Acta Ophthalmol*. 2020;98(2):132–138. doi:10.1111/aos.14183
98. Edington M, Connolly J, Chong NV. Pharmacokinetics of intravitreal anti-VEGF drugs in vitrectomized versus non-vitrectomized eyes. *Expert Opin Drug Metab Toxicol*. 2017;13(12):1217–1224. doi:10.1080/17425255.2017.1404987
99. Urtti A. Challenges and obstacles of ocular pharmacokinetics and drug delivery. *Adv Drug Deliv Rev*. 2006;58(11):1131–1135. doi:10.1016/j.addr.2006.07.027
100. Campochiaro PA, Marcus DM, Awh CC, et al. The port delivery system with ranibizumab for neovascular age-related macular degeneration: results from the randomized phase 2 ladder clinical trial. *Ophthalmology*. 2019;126(8):1141–1154. doi:10.1016/j.ophtha.2019.03.036
101. Sharma A, Kumar N, Parachuri N, et al. Ranibizumab port delivery system (rpds): realising long awaited dream of prolonged VEGF suppression. *Eye*. 2020;34(3):422–423. doi:10.1038/s41433-019-0479-y
102. Khanani AM, Callanan D, Dreyer R, et al. End-of-study results for the ladder phase 2 trial of the port delivery system with ranibizumab for neovascular age-related macular degeneration. *Ophthalmol Retina*. 2021;5(8):775–787. doi:10.1016/j.oret.2020.11.004
103. Holekamp NM, Campochiaro PA, Chang MA, et al. Archway randomized phase 3 trial of the port delivery system with ranibizumab for neovascular age-related macular degeneration. *Ophthalmology*. 2022;129(3):295–307. doi:10.1016/j.ophtha.2021.09.016
104. Boyer D. New developments in drug therapy for retinal disorders. In: *Hawaiian Eye & Retina Annual Meeting, January 21, 2019, kona, hawaii*; 2019.
105. Graybug vision presents top line results of phase 1/2a adagio study at Hawaiian eye & retina; 2019. Available from: <https://graybug.Com/graybug-vision-presents-top-line-results-of-phase-1-2a-adagio-study-at-hawaiian-eye-retina-2019/>. Accessed September 15, 2022.
106. Delaney-gesing A. Graybug vision releases preliminary topline results of phase 2b altissimo trial. *Ophthalmology times*; 2021. Available from: <https://www.Ophthalmologytimes.Com/view/graybug-vision-releases-preliminary-topline-results-of-phase-2b-altisimo-trial>. Accessed September 15, 2022.
107. Hussain RM, Shaikat BA, Ciulla LM, et al. Vascular endothelial growth factor antagonists: promising players in the treatment of neovascular age-related macular degeneration. *Drug Des Devel Ther*. 2021;15:2653–2665. doi:10.2147/dddt.s295223
108. Makadia HK, Siegel SJ. Poly lactic-co-glycolic acid (plga) as biodegradable controlled drug delivery carrier. *Polymers*. 2011;3(3):1377–1397. doi:10.3390/polym3031377
109. Uhrich KE, Cannizzaro SM, Langer RS, et al. Polymeric systems for controlled drug release. *Chem Rev*. 1999;99(11):3181–3198. doi:10.1021/cr940351u
110. Wu XS, Wang N. Synthesis, characterization, biodegradation, and drug delivery application of biodegradable lactic/glycolic acid polymers. Part ii: biodegradation. *J Biomater Sci Polym Ed*. 2001;12(1):21–34. doi:10.1163/156856201744425
111. Rapier CE, Shea KJ, Lee AP. Investigating plga microparticle swelling behavior reveals an interplay of expansive intermolecular forces. *Sci Rep*. 2021;11(1):14512. doi:10.1038/s41598-021-93785-6
112. Jain RA. The manufacturing techniques of various drug loaded biodegradable poly (lactide-co-glycolide) (plga) devices. *Biomaterials*. 2000;21(23):2475–2490. doi:10.1016/s0142-9612(00)00115-0
113. Yandrapu SK, Upadhyay AK, Petrash JM, et al. Nanoparticles in porous microparticles prepared by supercritical infusion and pressure quench technology for sustained delivery of bevacizumab. *Mol Pharm*. 2013;10(12):4676–4686. doi:10.1021/mp400487f
114. Varshochian R, Riazi-Esfahani M, Jeddi-Tehrani M, et al. Albuminated plga nanoparticles containing bevacizumab intended for ocular neovascularization treatment. *J Biomed Mater Res A*. 2015;103(10):3148–3156. doi:10.1002/jbm.a.35446

115. Xie B, Jin L, Luo Z, et al. An injectable thermosensitive polymeric hydrogel for sustained release of avastin® to treat posterior segment disease. *Int J Pharm.* 2015;490(1–2):375–383. doi:10.1016/j.ijpharm.2015.05.071
116. Ye Z, Ji YL, Ma X, et al. Pharmacokinetics and distributions of bevacizumab by intravitreal injection of bevacizumab-plga microspheres in rabbits. *Int J Ophthalmol.* 2015;8(4):653–658. doi:10.3980/j.issn.2222-3959.2015.04.02
117. Liu J, Li S, Li G, et al. Highly bioactive, bevacizumab-loaded, sustained-release plga/pcadk microspheres for intravitreal therapy in ocular diseases. *Int J Pharm.* 2019;563:228–236. doi:10.1016/j.ijpharm.2019.04.012
118. Zhang XP, Sun JG, Yao J, et al. Effect of nanoencapsulation using poly (lactide-co-glycolide) (plga) on anti-angiogenic activity of bevacizumab for ocular angiogenesis therapy. *Biomed Pharmacother.* 2018;107:1056–1063. doi:10.1016/j.biopha.2018.08.092
119. Hoffart L, Matonti F, Conrath J, et al. Inhibition of corneal neovascularization after alkali burn: comparison of different doses of bevacizumab in monotherapy or associated with dexamethasone. *Clin Exp Ophthalmol.* 2010;38(4):346–352. doi:10.1111/j.1442-9071.2010.02252.x
120. Liu J, Zhang X, Li G, et al. Anti-angiogenic activity of bevacizumab-bearing dexamethasone-loaded plga nanoparticles for potential intravitreal applications. *Int J Nanomedicine.* 2019;14:8819–8834. doi:10.2147/ijn.s217038
121. Tanetsugu Y, Tagami T, Terukina T, et al. Development of a sustainable release system for a ranibizumab biosimilar using poly (lactic-co-glycolic acid) biodegradable polymer-based microparticles as a platform. *Biol Pharm Bull.* 2017;40(2):145–150. doi:10.1248/bpb.b16-00437
122. Kirchhof S, Goepferich AM, Brandl FP. Hydrogels in ophthalmic applications. *Eur J Pharm Biopharm.* 2015;95:227–238. doi:10.1016/j.ejpb.2015.05.016
123. Yasin MN, Svirskis D, Seyfoddin A, et al. Implants for drug delivery to the posterior segment of the eye: a focus on stimuli-responsive and tunable release systems. *J Control Release.* 2014;196:208–221. doi:10.1016/j.jconrel.2014.09.030
124. Fanguiero JF, Andreani T, Egea MA, et al. Design of cationic lipid nanoparticles for ocular delivery: development, characterization and cytotoxicity. *Int J Pharm.* 2014;461(1–2):64–73. doi:10.1016/j.ijpharm.2013.11.025
125. Costa JR, Silva NC, Sarmento B, et al. Potential chitosan-coated alginate nanoparticles for ocular delivery of daptomycin. *Eur J Clin Microbiol Infect Dis.* 2015;34(6):1255–1262. doi:10.1007/s10096-015-2344-7
126. Lim HL, Hwang Y, Kar M, et al. Smart hydrogels as functional biomimetic systems. *Biomater Sci.* 2014;2(5):603–618. doi:10.1039/c3bm60288e
127. Cooper RC, Yang H. Hydrogel-based ocular drug delivery systems: emerging fabrication strategies, applications, and bench-to-bedside manufacturing considerations. *J Control Release.* 2019;306:29–39. doi:10.1016/j.jconrel.2019.05.034
128. Meyer CH, Krohne TU, Charbel Issa P, et al. Routes for drug delivery to the eye and retina: intravitreal injections. *Dev Ophthalmol.* 2016;55:63–70. doi:10.1159/000431143
129. Xu L, Cooper RC, Wang J, et al. Synthesis and application of injectable bioorthogonal dendrimer hydrogels for local drug delivery. *ACS Biomater Sci Eng.* 2017;3(8):1641–1653. doi:10.1021/acsbomaterials.7b00166
130. Adamson P, Wilde T, Dobrzynski E, et al. Single ocular injection of a sustained-release anti-VEGF delivers 6months pharmacokinetics and efficacy in a primate laser CNV model. *J Control Release.* 2016;244:1–13. doi:10.1016/j.jconrel.2016.10.026
131. Yu Y, Lau LC, Lo AC, et al. Injectable chemically crosslinked hydrogel for the controlled release of bevacizumab in vitreous: a 6-month in vivo study. *Transl Vis Sci Technol.* 2015;4(2):5. doi:10.1167/tvst.4.2.5
132. Yu Y, Lin X, Wang Q, et al. Long-term therapeutic effect in nonhuman primate eye from a single injection of anti-VEGF controlled release hydrogel. *Bioeng Transl Med.* 2019;4(2):e10128. doi:10.1002/btm.2.10128
133. Thanou M, Verhoef JC, Junginger HE. Oral drug absorption enhancement by chitosan and its derivatives. *Adv Drug Deliv Rev.* 2001;52(2):117–126. doi:10.1016/s0169-409x(01)00231-9
134. Bhattarai N, Gunn J, Zhang M. Chitosan-based hydrogels for controlled, localized drug delivery. *Adv Drug Deliv Rev.* 2010;62(1):83–99. doi:10.1016/j.addr.2009.07.019
135. Ways M, Lau TM, Khutoryanskiy VV. Chitosan and its derivatives for application in mucoadhesive drug delivery systems. *Polymers.* 2018;10(3). doi:10.3390/polym10030267
136. Peers S, Montembault A, Ladavière C. Chitosan hydrogels for sustained drug delivery. *J Control Release.* 2020;326:150–163. doi:10.1016/j.jconrel.2020.06.012
137. Pandit J, Sultana Y, Aqil M. Chitosan-coated plga nanoparticles of bevacizumab as novel drug delivery to target retina: optimization, characterization, and in vitro toxicity evaluation. *Artif Cells Nanomed Biotechnol.* 2017;45(7):1397–1407. doi:10.1080/21691401.2016.1243545
138. Fletcher NA, Krebs MD. Sustained delivery of anti-VEGF from injectable hydrogel systems provides a prolonged decrease of endothelial cell proliferation and angiogenesis in vitro. *RSC Adv.* 2018;8(16):8999–9005. doi:10.1039/c7ra13014g
139. Abrishami M, Zarei-Ghanavati S, Soroush D, et al. Preparation, characterization, and in vivo evaluation of nanoliposomes-encapsulated bevacizumab (avastin) for intravitreal administration. *Retina.* 2009;29(5):699–703. doi:10.1097/IAE.0b013e3181a2f42a
140. Mu H, Wang Y, Chu Y, et al. Multivesicular liposomes for sustained release of bevacizumab in treating laser-induced choroidal neovascularization. *Drug Deliv.* 2018;25(1):1372–1383. doi:10.1080/10717544.2018.1474967
141. Bisht R, Jaiswal JK, Chen YS, et al. Light-responsive in situ forming injectable implants for effective drug delivery to the posterior segment of the eye. *Expert Opin Drug Deliv.* 2016;13(7):953–962. doi:10.1517/17425247.2016.1163334
142. Dvir T, Banghart MR, Timko BP, et al. Photo-targeted nanoparticles. *Nano Lett.* 2010;10(1):250–254. doi:10.1021/nl903411s
143. Wang Y, Liu CH, Ji T, et al. Intravenous treatment of choroidal neovascularization by photo-targeted nanoparticles. *Nat Commun.* 2019;10(1):804. doi:10.1038/s41467-019-08690-4
144. Huu VA, Luo J, Zhu J, et al. Light-responsive nanoparticle depot to control release of a small molecule angiogenesis inhibitor in the posterior segment of the eye. *J Control Release.* 2015;200:71–77. doi:10.1016/j.jconrel.2015.01.001
145. Nadarassan DK. Sustained release of bevacizumab (avastin) from biosilicon. *Invest Ophthalmol Vis Sci.* 2014;55(13):1950.
146. Nagai N, Nezhad ZK, Daigaku R, et al. Transscleral sustained ranibizumab delivery using an episcleral implantable device: suppression of laser-induced choroidal neovascularization in rats. *Int J Pharm.* 2019;567:118458. doi:10.1016/j.ijpharm.2019.118458
147. Burgalassi S, Monti D, Nicosia N, et al. Freeze-dried matrices for ocular administration of bevacizumab: a comparison between subconjunctival and intravitreal administration in rabbits. *Drug Deliv Transl Res.* 2018;8(3):461–472. doi:10.1007/s13346-018-0520-x

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