Study on Changes in Immune Function After Microwave Ablation of Papillary Thyroid Microcarcinoma

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Background: In recent years, papillary thyroid microcarcinoma (PTMC) has been a main cause of the high incidence of thyroid carcinoma. No existing study has reported whether microwave ablation (MWA) affects patients’ immunity. Therefore, this study explored the effects of MWA treatment on the immune functions of patients with PTMC.

Methods: This study included 50 patients diagnosed with PTMC who received MWA treatment under ultrasound guidance at the ultrasound department of our hospital from January 2019 to October 2020. Changes in immune function after MWA treatment in PTMC patients were detected by T lymphocyte subsets and cytokines secreted by T helper cells.

Results: At 1 day and 2 weeks after MWA treatment, the proportions of CD3+, CD4+ and CD4+/CD8+ T lymphocyte subsets and the levels of the cytokines interleukin (IL)-2 and interferon (IFN)-γ in the peripheral blood of the patients were significantly higher than those before MWA treatment (P<0.05). The levels of CD8+ T lymphocytes, tumour necrosis factor (TNF)-α, IL-4, IL-6, IL-10 and IL-17A were not significantly different from those before MWA treatment (P>0.05). One month after MWA treatment, the proportions of CD3+, CD4+, CD8+ and CD4+/CD8+ T lymphocytes and the levels of the cytokines IL-2, IL-4, IL-6, IL-10, IFN-γ, TNF-α and IL-17A were not significantly different from those before MWA treatment (P>0.05).

Conclusion: The immune functions of patients with PTMC are temporarily enhanced after MWA treatment, which has important clinical significance for patients’ anti-PTMC ability.

Keywords: microwave ablation, papillary thyroid microcarcinoma, T lymphocyte, cytokine
Materials and Methods

Study Subjects

Fifty patients diagnosed with PTMC who received MWA treatment under ultrasound guidance at the ultrasound department of our hospital from January 2019 to October 2020 were enrolled as the research subjects, including 8 males and 42 females aged 18–60 (41.65±9.92) years old. The average size of the lesions was 83.57±77.23 mm³, and the average ablation time was 21.13±6.43 seconds at an ablation power of 25 W. All of the patients and their families signed informed consent forms, and this study was approved by the ethics committee of our hospital.

The inclusion criteria were as follows: (1) papillary carcinoma preoperatively diagnosed by needle cytology or histopathology; (2) a single lesion without extrathyroid invasion or cervical lymph node metastasis; (3) a maximum lesion diameter ≤10 mm; (4) normal thyroid function before MWA treatment and no history of other thyroid diseases (such as acute or chronic thyroiditis); and (5) ineligibility for or refusal of open thyroid surgery. The exclusion criteria were as follows: (1) multiple lesions; (2) extrathyroid invasion, cervical lymph node metastasis or distant metastasis; (3) pregnancy or breastfeeding; (4) other serious systemic or immune-related diseases; (5) severe liver and kidney or cardiopulmonary insufficiency, and (6) severe coagulation dysfunction.

Instruments

(1) The MWA machinery used in this study was an ECO-100C MWA treatment instrument (Nanjing Yigao Microwave System Engineering Co., Ltd.). The ablation needle used was an ECO-100AI3 disposable MWA needle, and the microwave emission frequency for ablation was 2450 MHz. (2) The ultrasound system used in this study was a Mindray Resona 7-colour ultrasound diagnostic instrument with a probe frequency of 7–15 MHz. (3) The ultrasound contrast agent used in this study was 1.8 mg/mL sulfur hexafluoride (Bracco, Italy). The ultrasound contrast agent was dissolved in 5 mL of normal saline and shaken well, and 2.0 mL of the suspension was quickly injected through the patient’s median cubital vein, followed by a bolus of 5 mL of saline. Contrast-enhanced ultrasound was performed before, immediately after and at the follow-up after the MWA treatment. (4) An FC 500MCL flow cytometer (Beckman Coulter, USA) was used to determine the proportions of CD3⁺, CD4⁺, CD8⁺ and CD4⁺/CD8⁺ T lymphocytes in the peripheral blood. A BD FACSaria II flow cytometer (BD, USA) was used to detect cytokine levels in the peripheral blood.

Methods

(1) Preoperative preparation: All of the patients underwent a routine ultrasound examination to determine the tumour size, shape, number, location, echo, peripheral blood flow distribution and adjacent vital organs and the presence of suspicious lymph nodes (Figure 1A). The supine position was used to allow full exposure of the neck. All patients underwent ECG monitoring, oxygen inhalation, and intravenous line placement. Routine disinfection and draping were performed before MWA, and 1% lidocaine hydrochloride was percutaneously injected for local infiltration anaesthesia.

(2) Liquid insulation zone: For lesions close to important organs or tissues, such as the carotid artery, trachea, oesophagus or recurrent laryngeal nerve, an insulation solution was repeatedly injected before ablation treatment. A 20-mL syringe was used to inject saline into the space around the tissue for insulation, forming a liquid insulation zone at least 5 mm in width7 (Figure 1B).

(3) Skin opening: A needle equipped with a 20-mL syringe was used to open the skin at the puncture point to create a channel for insertion of the microwave antenna through the skin.

(4) MWA: The MWA instrument was turned on, and cold circulation was checked. After the microwave antenna was inserted into the target area through the skin opening, multisection fixation ablation mode was used in combination with expanded ablation at an ablation power of 25 W. To achieve radical treatment, the ablation range should be expanded by at least 2 mm into the normal thyroid tissue beyond the tumour (Figure 1C).

(5) Immediately after MWA treatment, contrast-enhanced ultrasound was performed to determine the ablation scope, and whether the filling defect area completely covered and exceeded the range of the tumour before MWA treatment was evaluated. If the ablation range was not satisfactory, ablation was repeated (Figure 1D).

(6) After MWA treatment, to prevent bleeding, an ice pack was placed on the ablation area for 30 minutes.
Detection Indicators

Venous blood samples were collected from the patients before MWA treatment and 1 day, 2 weeks and 1 month after MWA treatment to detect the proportions of T lymphocyte subsets (CD3+ , CD4+ , CD8+ and CD4+/CD8+) and cytokine levels (interleukin (IL)-2, interferon (IFN)-γ, tumour necrosis factor (TNF)-α, IL-4, IL-6, IL-10 and IL-17A) in the peripheral blood.

Statistical Analysis

SPSS 26.0 software was used for data processing. Normally distributed measurement data are presented as the means ± standard deviations, and nonnormally distributed data are presented as the medians and interquartile ranges. Intragroup comparisons were performed by a paired t-test or rank-sum tests of related samples. P<0.05 indicated a statistically significant difference.

Results

Complications

During MWA treatment, 1 of the 50 patients developed hoarseness and recovered completely within 1 hour after MWA treatment, and on the next day, laryngoscopy showed that the movement ranges of both vocal folds were normal. Two of the 50 patients developed radial pain in the area behind the ear on the same side as the ablation, which resolved within 30 minutes after MWA treatment.

Effectiveness of MWA Treatment

Contrast-enhanced ultrasound performed 1 day after MWA showed that the lesion area was completely covered by the ablation range and was free of contrast agent filling. The volume of the ablation area gradually decreased at 2 weeks and 1 month after MWA.

Figure 1 Ultrasound-guided microwave ablation (MWA) of papillary thyroid microcarcinoma (PTMC). The open arrow shows the PTMC lesion. (A) Two-dimensional ultrasound showing the PTMC lesion in the left lobe of the thyroid next to the trachea, with a hypoechoic shadow and a clear boundary. (B) Under ultrasound guidance, normal saline was injected between the thyroid and the trachea to form an insulation zone, which appears as an anechoic strip (indicated by the triangle). (C) Under ultrasound guidance, an MWA needle (indicated by the closed arrow) was inserted into the PTMC lesion percutaneously along the predetermined needle channel. (D) Contrast-enhanced ultrasound performed after ablation shows that the ablation range (indicated by the diamond arrow) completely covers the PTMC lesion.
Changes in T Lymphocyte Subsets and Cytokines

Changes in T lymphocyte subsets in the peripheral blood of the patients before and after MWA treatment: One day and 2 weeks after MWA treatment, the proportions of $CD3^+$, $CD4^+$ and $CD4^+/CD8^+$ T lymphocytes in the peripheral blood of the patients were significantly higher than those before MWA treatment ($P<0.05$), while the proportion of $CD8^+$ T lymphocytes was not significantly different from that before MWA treatment ($P>0.05$). One month after MWA treatment, the proportions of $CD3^+$, $CD4^+$, $CD8^+$ and $CD4^+/CD8^+$ T lymphocytes in the peripheral blood were not significantly different from those before MWA treatment ($P>0.05$) (Table 1). The changes in T lymphocyte subsets in the peripheral blood of the patients before and after MWA treatments are shown in Figure 2A.

Changes in cytokine levels in the peripheral blood of the patients before and after MWA treatment: One day and 2 weeks after MWA treatment, the levels of IL-2 and IFN-$\gamma$ secreted by T helper 1 (Th1) cells were significantly higher than those before MWA treatment ($P<0.05$), while the TNF-$\alpha$ concentration was not significantly different from that before MWA treatment ($P>0.05$). Moreover, the levels of IL-4, IL-6 and IL-10 secreted by T helper 2 (Th2) cells were not significantly different from those measured before MWA treatment ($P>0.05$), and the level of IL-17A secreted by T-helper 17 (Th17) cells was not significantly different from that measured before MWA treatment ($P>0.05$).

One month after MWA treatment, the levels of IL-2, IL-4, IL-6, IL-10, IFN-$\gamma$, TNF-$\alpha$ and IL-17A were not significantly different from those measured before MWA treatment ($P>0.05$) (Table 2). The changes in the cytokine levels in the peripheral blood of the patients before and after MWA treatment are shown in Figure 2B.

<table>
<thead>
<tr>
<th></th>
<th>Before MWA</th>
<th>1 Day After MWA</th>
<th>2 Weeks After MWA</th>
<th>1 Month After MWA</th>
</tr>
</thead>
<tbody>
<tr>
<td>$CD3^+$</td>
<td>71.22±8.13</td>
<td>74.06±7.31*</td>
<td>74.23±6.68*</td>
<td>72.39±6.19</td>
</tr>
<tr>
<td>$CD4^+$</td>
<td>42.41±7.80</td>
<td>46.16±8.03*</td>
<td>45.09±7.64*</td>
<td>41.99±7.49</td>
</tr>
<tr>
<td>$CD8^+$</td>
<td>24.60±6.89</td>
<td>23.92±6.58</td>
<td>23.93±6.55</td>
<td>25.19±6.42</td>
</tr>
<tr>
<td>$CD4^+/CD8^+$</td>
<td>1.93±0.93</td>
<td>2.15±0.97*</td>
<td>2.11±0.95*</td>
<td>1.81±0.72</td>
</tr>
</tbody>
</table>

**Note:** *Compared with before MWA treatment $P<0.05$.

**Abbreviation:** MWA, Microwave ablation.

**Table 1 Changes in T Lymphocyte Subsets ($\bar{x} \pm s$, n=50, %)**

**Figure 2** The effects of MWA treatment on the immune function of patients with PTMC. (A) Changes in T lymphocyte subsets in the peripheral blood of patients before and after MWA treatment. (B) Changes in cytokine levels in the peripheral blood of the patients before and after MWA treatment.
Discussion

With the flourishing development of ablation techniques, MWA has gradually become a novel method for the treatment of thyroid diseases. The efficacy of MWA for the treatment of benign thyroid nodules and recurrent thyroid cancer has been widely recognized. The use of MWA for PTMC is becoming standardized and is being extensively applied. The expert group in China has issued a number of consecutive expert consensuses with the aims of affirming the efficacy of MWA for the treatment of PTMC, standardizing the indications and contraindications of MWA for PTMC and promoting the application of MWA for PTMC treatment. Previous studies have reported changes in the immune capacity after MWA treatment for liver cancer, showing that the proportions of CD4+ and CD4+/CD8+ T lymphocytes in the peripheral blood increased after ablation, suggesting that MWA treatment for primary liver cancer not only inactivates tumour cells but also increases the number of immune cells available to exert antitumor functions. Some scholars have studied changes in serum Th1- and Th2-type cytokines in patients with liver cancer after radiofrequency ablation and found that Th1-type cytokines (IL-2 and TNF-α) increased in peripheral blood after ablation, while Th2-type cytokines (IL-4 and IL-10) decreased, suggesting that radiofrequency ablation can improve immune functions and enhance antitumor abilities. However, no report on changes in immune functions after MWA for PTMC are available. Therefore, this study analysed changes in T lymphocyte subsets (CD3+, CD4+, CD8+ and CD4+/CD8+) and cytokine concentrations (IL-2, IL-4, IL-6, IL-10, TNF-α, IFN-γ and IL-17A) to explore the effects of MWA on immune function in PTMC patients.

No serious complications occurred after MWA treatment in the 50 patients included in this study. The range of the contrast-enhanced ultrasound completely covered the lesion on the first day after MWA treatment, and the volume of the ablation area gradually decreased at 2 weeks and 1 month after MWA treatment, indicating that MWA is a safe and effective treatment method.

T lymphocyte subsets (CD3+, CD4+, CD8+ and CD4+/CD8+) are the main indicators of cellular immune functions, all of which play an important role in immune regulation by playing an antiviral role and regulating the functions of the immune system. This study found that 1 day and 2 weeks after MWA treatment, the proportions of CD3+, CD4+ and CD4+/CD8+ T lymphocytes in the patients’ peripheral blood increased significantly, suggesting enhancement of the patients’ immune functions, which is consistent with the results of domestic and international scholars who have studied the use of MWA for liver cancer. The change in the number of CD3+ T lymphocytes can be used to evaluate the overall immune functional status of tumour patients. An increase in the number of CD3+ T lymphocytes indicates an increase in the total number of mature T lymphocytes and an enhanced biological effect. This study found that the number of CD3+ T lymphocytes in the peripheral blood of the patients increased 1 day and 2 weeks after MWA treatment, indicating that MWA treatment has a positive regulatory effect on T lymphocytes. CD4+ T lymphocytes are the most important hub cells in the regulation of immune responses. These cells can stimulate the expression of cytokines by a variety of immune cells and Th cells and mainly play an important role in antitumor immunity. In this study, the CD4+ T lymphocyte count in the peripheral blood of the patients at 1 day and 2 weeks after MWA treatment increased significantly, indicating that MWA promotes the secretion of immune cells. CD8+ T lymphocytes have a specific immunosuppressive effect as part of the

### Table 2 Changes in Cytokine Levels Before and After MWA Treatment (n=50, Pg/Ml)

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>Before MWA</th>
<th>1 Day After MWA</th>
<th>2 Weeks After MWA</th>
<th>1 Month After MWA</th>
</tr>
</thead>
<tbody>
<tr>
<td>IL-2</td>
<td>0.98(0.41,1.31)</td>
<td>1.28(0.60,1.81)*</td>
<td>1.09(0.58,1.47)*</td>
<td>0.99(0.34,1.32)</td>
</tr>
<tr>
<td>IL-4</td>
<td>0.45(0.20,1.32)</td>
<td>0.65(0.41,1.09)</td>
<td>0.56(0.34,0.85)</td>
<td>0.42(0.24,0.65)</td>
</tr>
<tr>
<td>IL-6</td>
<td>2.11(1.23,3.20)</td>
<td>2.23(1.85,3.07)</td>
<td>2.3(1.67,2.98)</td>
<td>2.34(1.17,3.08)</td>
</tr>
<tr>
<td>IL-10</td>
<td>1.77(1.16,2.17)</td>
<td>1.91(1.25,2.41)</td>
<td>1.72(1.36,2.25)</td>
<td>1.65(1.01,2.18)</td>
</tr>
<tr>
<td>TNF-α</td>
<td>0.54(0.31,0.98)</td>
<td>0.65(0.42,0.96)</td>
<td>0.54(0.40,0.85)</td>
<td>0.53(0.15,0.94)</td>
</tr>
<tr>
<td>IFN-γ</td>
<td>0.47(0.23,0.69)</td>
<td>0.90(0.40,2.96)*</td>
<td>0.89(0.46,1.63)*</td>
<td>0.58(0.30,0.73)</td>
</tr>
<tr>
<td>IL-17A</td>
<td>0.52(0.13,2.25)</td>
<td>0.85(0.13,3.77)</td>
<td>0.93(0.22,2.81)</td>
<td>0.68(0.05,1.71)</td>
</tr>
</tbody>
</table>

**Note:** Compared with before MWA treatment P<0.05.

**Abbreviation:** MWA, Microwave ablation.
immune response and play a negative regulatory role by releasing inhibitory cytokines. The results of this study showed no significant changes in CD8\(^+\) T lymphocytes before and after MWA treatment, which may indicate that MWA treatment does not act by regulating CD8\(^+\) T lymphocytes. The proportion of CD4\(^+\)/CD8\(^+\) T lymphocytes is a sensitive indicator of immune function disorders of the human body and reflects the overall status of immune function. The results of this study showed that the proportion of CD4\(^+\)/CD8\(^+\) T lymphocytes in the peripheral blood increased significantly 1 day and 2 weeks after MWA treatment, indicating that the overall immune function of the body was enhanced. The various changes in the T lymphocyte subsets indicate that MWA treatment not only inactivates tumour cells but also increases the number of immune cells released by the body.\(^{14}\)

We also studied major changes in cytokines secreted by Th1 cells (IL-2, INF-γ and TNF-α), Th2 cells (IL-4, IL-6 and IL-10) and Th17 cells (IL-17A). In this study, IL-2 and IFN-γ levels increased at 1 day and 2 weeks after MWA treatment, indicating that the body’s antitumor immune effect increased and that the immune ability increased in a short period after MWA. However, the levels of these factors at 1 month after MWA were similar to the preoperative levels; a possible explanation is that as the inactivated tumour tissue is gradually absorbed by the body, the secretion of various cytokines gradually reaches a dynamic balance.

The results of this study showed that the proportions of CD3\(^+\), CD4\(^+\) and CD4\(^+\)/CD8\(^+\) T lymphocytes in the peripheral blood of patients after MWA were significantly increased and that the levels of IL-2 and INF-γ after MWA were significantly higher than those before MWA, indicating that MWA for PTMC is conducive to short-term improvement in patients’ immune function.

At present, the mechanism of immune regulation after MWA treatment of PTMC is still not clear. According to a similar study,\(^{15}\) the possible regulatory mechanisms are as follows:

1. Reduction in the tumour burden: MWA employs microwave needle thermoablation to cause coagulation and necrosis of tumour cells to completely inactivate them, thereby reducing the tumour burden, decreasing the secretion of immunosuppressive factors and relieving the immunosuppressive effect of cancer cells on the body. (2) Thermal effect and membrane structure theory: cancer cells undergo degeneration and necrosis after exposure to the heating stimulus of the MWA needle and remain in the body as antigens. The antigenic tumour determinants on the surfaces of cancer cells are fully exposed, which increases the antigenicity of the tumour cells and facilitates antibody binding to the tumour cells. When these antigens are taken up by antigen-presenting cells, they induce T lymphocytes to exert immune responses, thereby improving the body’s immune functions. (3) Immune responses of cytokines and humoural factors: A previous study showed that thermal therapy can increase the activity of lymphocytes and enhance the body’s immunity by increasing the secretion of cytokines. Tumour cells reduce IL-2 levels in the body by releasing sIL-2R to competitively bind to the IL-2 secreted by Th1 cells, thus exerting an inhibitory effect on the immune system. (4) Inflammatory response and immune stimulation: pathological examinations of tumour tissue after heat treatment revealed a large number of lymphocytes surrounding the necrotic foci, suggesting that the inflammatory response can eliminate tumour cells,\(^{16}\) and through this mechanism, IFN-γ promotes the expression of inflammatory factors to exert an immune effect.

5. Immune response of heat shock protein: MWA mainly converts microwave energy into heat energy, which then kills cancer cells as the temperature increases. When cancer cells are heated, the body is believed\(^{17}\) to undergo a stress response and can synthesize a stress protein, heat shock protein (HSP), which protects the body. After T lymphocytes recognize heat shock proteins, they participate in cellular immunity to initiate a specific immune response.

The limitations of this study are that only changes in various T lymphocyte subsets and cytokines at 1 day, 2 weeks and 1 month after MWA were compared with those before MWA, and no long-term follow-up values were analysed. Therefore, our next step is to further explore the long-term effects of MWA on the immune function of the body. Additionally, this study had a small sample size, and future studies require a large sample size for in-depth research.

An inclusion criterion in this study was the absence of cervical lymph node metastasis. However, neck lymph node dissection is still a hot topic in surgery.\(^{18}\) The main objective of this study was to evaluate changes in T lymphocyte subsets and cytokines in patients’ peripheral blood. In future research, we will select more meaningful markers for analysis, including changes in markers such as calcitonin or carcinoembryonic antigen before and after MWA treatment. During our research, we found that many patients with thyroid cancer have Hashimoto’s thyroiditis, and some scholars...
have reported that Hashimoto’s thyroiditis has a good prognostic effect on papillary thyroid carcinoma.\textsuperscript{19} Therefore, the influence of Hashimoto’s thyroiditis on the immune function of PTMC patients after ablation is also an important topic.

In summary, MWA for PTMC is safe and effective. The CD3\textsuperscript{+}, CD4\textsuperscript{+} and CD4\textsuperscript{+}/CD8\textsuperscript{+} T lymphocytes and the cytokines IL-2 and IFN-\(\gamma\) in the peripheral blood of the patients increased at 1 day and 2 weeks after MWA treatment, indicating that patients’ immune functions are enhanced in the short term, which has important clinical significance for anti-PTMC activity.

**Institutional Review Board Statement**
The study was conducted in accordance with the Declaration of Helsinki and approved by the Ethics Committee of China-Japan Union Hospital of Jilin University (protocol code 2019082112 and date of approval: 21 August 2019).

**Informed Consent Statement**
Informed consent was obtained from all subjects involved in the study.

**Author Contributions**
All authors made significant contributions to the work reported, whether the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; provided final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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**Disclosure**
The authors declare no conflicts of interest in this work.

**References**


