Survivin-T34A: molecular mechanism and therapeutic potential

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Abstract: The inhibitor of apoptosis protein survivin’s threonine 34 to alanine (T34A) mutation abolishes a phosphorylation site for p34(cdc2)–cyclin B1, resulting in initiation of the mitochondrial apoptotic pathway in cancer cells; however, it has little known direct effects on normal cells. The possibility that targeting survivin in this way may provide a novel approach for selective cancer gene therapy has yet to be fully evaluated. Although a flurry of work was undertaken in the late 1990s and early 2000s, only minor advances on this mutant have recently taken place. We recently described that cells generated to express a stable form of the mutant protein released this survivin-T34A to the conditioned medium. When this conditioned medium was collected and deposited on naive tumor cells, conditioned medium T34A was as effective as some chemotherapeutics in the induction of tumor cell apoptosis, and when combined with other forms of genotoxic stressors potentiated their killing effects. We hope with this review to revitalize the T34A field, as there is still much that needs to be investigated. In addition to determining the therapeutic dose and the duration of drug therapy required at the disease site, a better understanding of other key factors is also important. These include knowledge of target cell populations, cell-surface receptors, changes that occur in the target tissue at the molecular and cellular level with progression of the disease, and the mechanism and site of therapeutic action.

Keywords: survivin, T34A, apoptosis, proliferation, therapy

Survivin is important in cancer

Apoptosis is a physiological cell suicide program that is critical for the development and maintenance of healthy tissues.1 The evasion of apoptosis, as identified by Hanahan and Weinberg, is a primary characteristic of human cancer2 with proteins involved in its control of significant biological interest, as they could prove to be important targets for therapy development. The inhibitor of apoptosis (IAP) family consists of proteins with a proven ability to inhibit apoptosis, although much of how this works mechanistically is still to be revealed. Survivin is an important IAP because its expression is among the most tumor specific of all human gene products.3,4 It has been classified as a bifunctional protein, as it can inhibit apoptosis and cell cycle.5 More specifically, survivin’s promoter exhibits M-phase-inducible transactivation and, in HeLa cells, both survivin protein and mRNA are upregulated at the G2/M-phase of the cell cycle.5 Survivin expression is enhanced during embryonic and fetal development but not in terminally differentiated normal tissue.6 Its expression has also been seen in virtually every type of human cancer,7 making survivin an attractive protein in the study of cancer pathophysiology, drug discovery, and medical diagnosis. Our laboratory results have recently shown that cancer cells release survivin into the extracellular space8
phosphorylated in small 50–150 nm vesicles called exosomes. This is significant, as it may reveal new mechanisms about how cancer cells communicate with one another and how they affect the tumor microenvironment. We hope that this discovery may provide other options for targeted therapy. However, this discovery begs for studies about why and how survivin is packaged within exosomes, how survivin from exosomes is utilized by the recipient cells, and what are the effects of exosomal survivin on normal human cells.

Survivin physically associates with and is phosphorylated on its threonine 34 (Thr34) by the cyclin-dependent kinase p34(cdc2)–cyclin B1 protein complex. Loss of Thr34 phosphorylation results in dissociation of the caspase-9-survivin protein complex and caspase-dependent apoptosis. Initially, Thr34 through site-directed mutagenesis was changed to Ala (T34A) as a nonphosphorylated mimic of survivin. When replication-deficient adenoviruses encoding this survivin-T34A mutant were produced and used to infect cancer cell lines, spontaneous apoptosis resulted. In contrast, the viability of normal human cells, including major organs, experienced enhanced protection against genotoxic stress as well as exhibited an increased replicative and invasive potential. Alternatively, cancer cells grown in conditioned medium from T34A cells began to apoptose through a caspase-2- and caspase-9-dependent pathway that was further enhanced by the addition of chemo- and radiotherapeutic modalities. We believe that this extracellular survivin-T34A could be used as a possible ‘white knight’ to survivin’s ‘dark family’. The following sections detail T34A’s journey as a novel potential cancer therapeutic. The aforementioned studies have already provided impetus for further studies on T34A, but much is still unknown. The advancement of future T34A studies may provide a more comprehensive understanding of cancer and the potential of using T34A to battle it.

**T34A induces apoptosis**

What makes this survivin-T34A mutant important as a potential therapeutic is that little or no toxicity has resulted from its treatment in vitro or in vivo. T34A-treated mice showed significantly longer survival rates compared with control mice as well as having no pathologic changes in the major organs. More specifically, tumor cells treated in vitro with T34A exhibited reduced cell survival that was associated with morphological features of apoptosis. Studies of mouse xenographs treated with survivin-T34A concluded with significant tumor size reduction and increased animal survival rates except in mice subcutaneously injected with the IGROV1 ovarian carcinoma cell line, where there were no significant differences between T34A-infected tumors and controls. These results may be explained, however, as IGROV1 cells do not express significant levels of endogenous survivin and T34A, as shown in normal survivin-nonexpressing cells, does not induce apoptosis. It was not until IGROV1 cells were infected with adenovirus vector-expressing caspase-3 (Ad-caspase-3) or coinfectected with Ad-caspase-3 and Ad-T34A that tumor size was reduced and survival rates extended. It is of much interest that the adenoviruses used in these studies upregulated survivin mRNA, as shown by reverse transcription-polymerase chain reaction, and concomitantly resulted in a half-fold reduction in another IAP, XIAP (X-linked inhibitor of apoptosis protein). Furthermore, in melanoma cells that have been engineered to express an inducible survivin-T34A, XIAP and the IAP Livin were cleaved upon T34A induction. These reductions may in part be responsible for the more favorable outcomes observed in these studies. In contrast, it has been postulated that the transfection of cells may affect endogenous levels of XIAP, but there was no effect on XIAP protein levels following transfections using virus vector controls or Ad-survivin or Ad-T34A treatment. There were also...
no changes in other IAPs or members of the Bcl-2 family.20 Although survivin-T34A may not affect endogenous levels of other IAPs, a survivin-IAP (XIAP, cIAP1, and cIAP2) complex has been confirmed, as well as a suggestion for direct interaction between survivin and XIAP through their BIR (baculovirus IAP repeat) domains.30 Furthermore, evidence that T34A can only kill cells expressing survivin has been shown in its inability to kill normal human cells treated in vitro11,31 and in vivo.11,15 However, T34A has shown significant killing effects in normal endothelial cells that express high levels of endogenous survivin.19 Whether this is the result of some interaction between endogenous survivin, other IAPs and the T34A mutant, or a competition between them for protein interactors is currently unknown.

Mechanistic examination of T34A-induced apoptosis revealed mitochondrial events that preceded caspase activation, including depolarization and the release of cytochrome c, AIF (apoptosis-inducing factor), and Smac/DIABLO (second mitochondria-derived activator of caspase/direct inhibitor of apoptosis-binding protein with low isoelectric point).11,19,20,32 Also, T34A treatment of melanoma cells has been shown to involve Apaf-1, Mcl-1, and PARP (poly [ADP-ribose] polymerase) cleavage.19,20,25 Stress-induced cytochrome c release results in apoptosis formation and caspase activation,13 thus, several groups have reported that caspase-3 may be important to T34A-induced apoptosis.11,18-21,26,34 A significant increase in apoptosis was reported in ovarian carcinoma cells treated with T34A and caspase-3 in vivo and in vitro.18 We, however, when working with cervical carcinoma HeLa cells, were unable to show cleavage of caspase-3 or caspase-8, though cleavage of caspase-9 and caspase-2 was recorded.8 Further experiments revealed that T34A induced a reduction in the proforms of caspase-2, caspase-7, and caspase-9; a cleavage of PARP; and an induction of mitochondrial depolarization. Treatment with caspase inhibitors Z-DEVD-FMK and Z-VAD-FMK has been shown to counter the effect of T34A treatment, suggesting that T34A-induced apoptosis may act through the caspase-dependent pathway.20 Additionally, the earliest proapoptotic event observed in T34A-treated cells was nuclear translocation of mitochondrial AIF, which triggers both mitochondrial-associated apoptosis and caspase-independent DNA fragmentation.20 Taken together, survivin-T34A targets the intrinsic20 or mitochondria-associated apoptotic pathway to most probably induce caspase-dependent and independent apoptosis (Figure 1). However, with the exception of one study,36 survivin has not been shown to directly inhibit caspases, and whether survivin-T34A disrupts caspase activity has yet to be fully elucidated. We postulate that survivin-T34A-induced apoptosis begins further upstream in the intrinsic apoptotic pathway initiating mitochondrial depolarization as well as inhibition of IAPs known to directly inhibit caspases (eg, XIAP-caspase-3).

The tumor suppressor gene, p53, has been widely studied and is known to induce apoptosis.37,38 As a transcription factor, p53 is activated by a variety of genotoxic and cytotoxic stresses. Upon activation, p53 prevents the proliferation of genetically compromised cells by regulating the expression of a battery of genes that initiate cell cycle arrest, apoptosis, and DNA repair (Figure 2).39 Transcriptionally, p53 activates Mdm2, which in turn targets p53 for ubiquitin-proteasome-dependent degradation. In response to stress, a decrease in Mdm2 protein levels and/or its activity and the interaction between Mdm2 and p53 lead to p53 stabilization.40 The relationship between survivin and p53 may be important, as survivin regulates cell division and inhibits apoptosis, whereas p53 inhibits cell cycle progression and induces apoptosis.21 Survivin was recently shown to be transcriptionally repressed by wild-type p53, and when overexpressed in cells sensitive to p53-dependent cell death markedly inhibited apoptosis induced by ultraviolet light.41

When cancer cells that had been stably transfected with survivin-T34A were treated with the DNA damage-inducing Adriamycin, p53 protein levels were significantly elevated compared with DNA damage alone or when combined with stably transfected survivin.21 Increased levels of p53 mRNA preceded the elevation in p53 protein levels seen with T34A-treated cells, suggesting that p53 may be directly upregulated by survivin-T34A. Survivin disruption by survivin-T34A also resulted in the degradation of Mdm2. Proteasome inhibition and caspase-3 inhibition blocked this survivin-T34A-associated decrease in Mdm2, resulting in the stabilization of p53. Survivin-T34A was not shown to affect mRNA levels of Mdm2.21 Furthermore, the p53 homologs TAp63, ΔNp63, TAp73, and ΔNp73 were analyzed for changes following survivin-T34A treatment. The response of these p53 homologs is important because TAp63 and TAp73 both transactivate similar genes as p53, whereas the ΔNp63 and ΔNp73 function as antagonists to p53 and the TA homologues.42-44 ΔNp63, TAp73, and ΔNp73 were all decreased in response to survivin-T34A, whereas TAp63 was not detectable. These findings suggest that p53 transcription may be differentially regulated by survivin-T34A.21 Taken together, the ability of survivin-T34A to perturb the p53/Mdm2 feedback loop, resulting in increasing levels of p53 and p53’s ability to then transcriptionally repress survivin, may prove to be just
the therapeutic strategy needed for targeting p53-associated stress-induced repair pathways in cancer.

**Survivin Thr³⁴ plays a role in cell cycle regulation**

Survivin is a structurally unique IAP family protein that is expressed in mitosis in a cell cycle-dependent fashion and localized to components of the mitotic apparatus. Survivin has been described as bifunctional, in that it is involved in both inhibition of apoptosis and control of cell division. In order to study the effect survivin phosphorylation at Thr³⁴ had on cell division, the nonphosphorylatable T34A form of survivin was compared with T34E, which acts as a phosphomimetic. As a phosphomimetic, T34E assumes the role of a phosphorylated Thr³⁴ and therefore mimics an activated survivin. T34A-treated cells grew two-fold faster than control cells, whereas T34E cells grew 2.5-fold slower. Depletion studies showed that only T34A was normally distributed and able to support cell proliferation, whereas T34E was not.
Survivin has been known to be upregulated in G2/M of the cell cycle, but T34A’s ability to induce apoptosis is G2/M independent. The cell cycle profile of T34A-treated cells was normal, whereas T34E-treated cells had a significantly increased G2/M population, suggesting that cell division failed. Furthermore, cancer cells that exhibited endogenous levels of functional survivin when treated with T34A showed significant mitotic index inhibition, as well as a reduction in colony formation. Ultimately, when endogenous survivin was depleted, T34A restored survivin’s mitotic function, whereas T34E was mitotically incompetent. We therefore conclude that phosphorylated Thr inhibits mitosis, but nonphosphorylated survivin is important for mitosis.

T34A has been shown to decrease the proportion of cells in S-phase following S-phase arrest. Evidence for survivin-T34A regulation of chromosomal passenger complexes and central spindle checkpoint during mitosis has been generated. In this work, survivin-T34A was also shown to be able to restore the spindle checkpoint defect following mitosis arrest, unlike the other non-wild-type forms, as well as rescue the cells from cytokinesis failure. These findings strongly suggest that survivin-T34A is a functional complement for endogenous survivin. Because survivin-T34A functionally behaves as survivin, it can localize appropriately during mitosis and thus correctly colocalize Aurora B at the centromeres and midzone, allowing proper mitosis regulation. Further experiments show that survivin-T34A correctly localizes BubR1 to the kinetochores and can restore localization of endogenous Borealin/Dasp B to both centromeres and central spindles. Taken together, survivin-T34A allows for the enhanced proliferative effects of survivin, whereas survivin-T34E does not. This suggests that the nonphosphorylated survivin Thr (T34A) accounts for cellular proliferation as opposed to the phosphorylated survivin Thr (T34E). Also, contrasting survivin Thr for its cytoprotective activity revealed that T34E is responsible for the antiapoptotic characteristics, whereas T34A allows for caspase activation and induced apoptosis.

**T34A opposes classical cancer characteristics**

Angiogenesis is crucial for cell survival and function, and without it tumor cells are not able to obtain oxygen and nutrients. Targeting tumor angiogenesis is a relatively new therapeutic strategy, and it is believed that if the tumor’s oxygen and nutrient supply is limited, tumor growth and development will be inhibited and may result in apoptosis. Survivin-T34A treatment of tumor-bearing mice resulted in inhibited tumor-induced angiogenesis and increased apoptosis. Tumor sections taken from human breast cancer transplants in mice were stained with anti-CD31 antibody to determine microvessel density. Survivin-T34A-treated groups showed significantly reduced microvessel density compared with controls. In vivo results showed T34A-induced apoptotic characteristics in murine endothelial cells as well as decreased new vessel formation when compared with control mice. Furthermore, experiments using CT26 colorectal cancer cells showed similar results. It is still unknown how survivin-T34A treatment reduces tissue staining for the angiogenesis marker CD31. The elucidation of this mechanism may provide insight into this tumor-acquired capability. Survivin has been shown to be upregulated by vascular endothelial growth factor (VEGF), which is an important promoter of angiogenesis. Perhaps survivin regulation by VEGF promotes angiogenesis by protecting developing vessels from apoptosis. Therefore, survivin-T34A inhibits VEGF’s promotion of angiogenesis, which may be mediated through loss of p53, activation of phosphatidylinositol 3′-kinase, or phosphorylation of signal transducers and activators of transcription.

Once a tumor mass becomes constricted, cells acquire the ability to invade surrounding tissue and metastasize to distant sites, which accounts for 90% of human cancer deaths. The inhibition of survivin using adenoviruses (Ad-T34A) or short hairpin RNA dramatically inhibited invasiveness of prostate cancer cells in the in vitro invasion assay and spontaneous metastasis in the Dunning prostate cancer in vivo model. Treatment of subcutaneous breast cancer tumors in mice...
using T34A plasmids or Ad-T34A inhibited metastasis of breast cancer cells from the primary tumor site to the lungs and peritoneum, respectively. Specifically, control treatment groups were observed to have a greater number of metastatic nodules and structural destruction of pulmonary alveoli compared with the T34A treatment group in transplanted mice. In light of the role that metastasis plays in cancer death, T34A’s metastasis-inhibitory role is important, as current therapeutics offer limited capabilities.

Cancerous lesions promote tumor growth, motility, invasion, and angiogenesis via oncogene-driven immunosuppressive leukocyte infiltrates. T34A has been shown to enhance T-lymphocyte activity against autologous tumor cells. In addition, T-cells derived from mouse xenographs exhibited higher cytotoxicity against target tumor cells in T34A-treated mice. Further studies revealed that adoptive transfer of CD8+ T-lymphocytes, and not their CD4+ T-lymphocyte counterparts, isolated from the spleen of T34A-treated mice exhibited a decrease in tumor volume. Perhaps the treatment of tumor cells with T34A or with immune cells that have been primed by survivin-T34A association will redress the immunosuppression associated with cancer.

Translational approaches and obstacles
Given survivin’s role in chemotherapy and radiation resistance, results using T34A would suggest that its role is to promote sensitivity to chemotherapy and radiotherapy. In the first study to investigate T34A’s ability to enhance cancer cell chemosensitivity, transfected survivin-T34A-induced apoptosis was as effective as Taxol alone, whereas transfected survivin-T34A cells enhanced Taxol-induced apoptosis. Subsequent studies involving other chemotherapeutics have also revealed enhanced apoptosis in T34A-expressing cancer cells and xenograph models (Flutamide, Paclitaxel). Survivin-T34A combined with Adriamycin exhibited no added effect over Adriamycin alone. This study was followed by studies designed to characterize phosphorylated survivin Thr’s protective role. Following Adriamycin treatment of tumor cells, p34(cdc2) was coimmunoprecipitated with survivin. These findings suggest that p34(cdc2) phosphorylates survivin on Thr following Adriamycin treatment thus protects the cells against the chemotherapeutic. These results were correlated with elevated MPM-2 mitotic phosphopeptope expression. Although survivin-T34A combined with Adriamycin exhibited no added effect over Adriamycin alone, it may still be possible to synergistically enhance Adriamycin’s effect with pretreatment of survivin-T34A. The same study further showed that increased doses of Flavopiridol treatment were associated with suppression of survivin Thr phosphorylation.

Human glioblastoma multiforme (GBM) cells were treated with adenoviral vectors containing T34A or survivin in vivo and in vitro. No change was seen in normal cells treated with either survivin form, but radiosensitivity was exhibited in T34A-treated cells compared with survivin-treated or control cells. Several other studies have confirmed similar results, suggesting that survivin only affects the radiosensitivity of tumors cells and that radioresistance can be reduced by T34A treatment. Interestingly, when cancer cells were irradiated and then treated with T34A, radiosensitivity was enhanced even more. Enhanced radiation sensitivity may be explained by cleavage and activation of caspase-3 in T34A-treated cells. Caspase inhibitors, except those for caspase-9, have been able to reduce apoptotic phenotypes in X-irradiation of T34A-treated cells. DNA double strand breakage was greater in T34A-treated cells, suggesting that T34A may interfere with the cells’ ability for DNA repair postirradiation.

Conclusion
Survivin-T34A has a rich history, having first been utilized as a mimic of nonphosphorylated survivin and now as a possible cancer therapeutic. It has potential in molecular therapy because of its minimal toxicity and its ability to induce apoptosis and immune modulation while reducing angiogenesis, metastasis, and cell cycle progression in survivin-expressing cells (Figure 3). With the discovery that a T34A-conditioned medium taken from cells that overexpress and release survivin-T34A to the extracellular space is able to kill cancer cells in vitro, we hope to further enhance the scientific data banks and pave new paths for clinical modalities and regimens. For this to occur, however, additional testing of this agent on normal tissues, specifically testing its ‘myelosuppressive effects’, will have to be completed. Also, additional studies to synergistically enhance chemo- and radiotherapies, whether in combination with or through survivin-T34A pretreatment, must be accomplished. The knowledge that will make this possible includes the elucidation of survivin-T34A target cell populations, cell-surface receptors, changes that occur in the target tissue at the molecular and cellular level, the presence or development of resistance that could limit its use, the mechanism of therapeutic action, and its site of action. As survivin and p53 are important opposing regulators of cancer, further research on their interactions and signaling
pathways will likely provide specific clues for understanding this complicated relationship.

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