Non-invasive and transdermal measurement of blood uric acid level in human by electroporation and reverse iontophoresis

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Abstract: The aim of this study was to find out the optimum combination of electroporation (EP) and reverse iontophoresis (RI) for noninvasive and transdermal determination of blood uric acid level in humans. EP is the use of high-voltage electric pulse to create nano-channels on the stratum corneum, temporarily and reversibly. RI is the use of small current to facilitate both charged and uncharged molecule transportation across the skin. It is believed that the combination of these two techniques has additional benefits on the molecules’ extraction across the human skin. In vitro studies using porcine skin and diffusion cell have indicated that the optimum mode for transdermal uric acid extraction is the combination of RI with symmetrical biphasic direct current (current density = 0.3 mA/cm²; phase duration = 180 s) and EP with 10 pulses per second (voltage = 100 V/cm²; pulse width = 1 ms). This optimum mode was applied to six human subjects. Uric acid was successfully extracted through the subjects’ skin into the collection solution. A good correlation (r² = 0.88) between the subject’s blood uric acid level and uric acid concentrations in collection solutions was observed. The results suggest that it may be possible to noninvasively and transdermally determine blood uric acid levels.

Keywords: Reverse iontophoresis, electroporation, uric acid, monitoring, noninvasive, transdermal

Introduction

Gout is one of the most common forms of arthritis, affecting more than 700,000 adults in the UK and nearly three million adults in the USA, and accounting for almost four million outpatient visits every year, with a substantial economic burden. Excessive amounts of uric acid, known as hyperuricemia, is one of most frequent metabolism disturbances. Hyperuricemia and high mean serum uric acid concentrations as risk factors for gout have been well documented in the past decades. Gouty arthritis, a rapid onset of joint inflammation, is a complication of hyperuricemia and is precipitated by deposits of uric acid crystals in the synovial fluid and synovial lining. Recent research has highlighted that patients with hyperuricemia and gout are at increased risk of morbidity and mortality related to cardiovascular disease. Therefore, blood uric acid monitoring is very important.

The routine method to determine blood uric acid level is achieved by blood sampling. However, this method is invasive, painful, and inconvenient. To the best of our knowledge, there is no noninvasive method to determine the blood uric acid level. Therefore, reverse iontophoresis (RI) and electroporation (EP), noninvasive techniques, were used in this study to transdermally and noninvasively extract uric acid in humans.
RI refers to the passage of a low level of current through the skin to promote the transport of both charged and neutral molecules. The main mechanisms that contribute to RI are the electro migration of charged species to the electrode of opposite polarity, electroosmosis of neutral molecules to the cathode or anode, or a combination of both. RI across the skin has been investigated as a noninvasive method for clinical monitoring. Its first clinical application was the transdermal extraction of glucose. EP is the use of an intense electric pulse to make the cell membrane transiently porous and then permeable to exogenous molecules present in the surrounding media. EP is widely used for the loading of cells with drugs and the delivery of drugs into the cells of living tissues. Recently, EP has been used for transcutaneous sampling of molecules, like glucose. Several papers have reported the use of combined RI and EP to enhance the effect of extracting metabolites, but no papers have been found for the use of combined RI and EP for the extraction of uric acid and this was the novelty of this study. The aim of this study was to find out the optimum combination of EP and RI on noninvasive and transdermal determination of blood uric acid level in humans.

**Materials and methods**

**Reagents and solutions**

All reagents used in this study were commercially available and used without further purification: phosphate buffered saline and uric acid were purchased from Sigma Chemical Company (St. Louis, MO) and uric acid assay kit (Catalog number: K608–100) from BioVision (Mountain View, CA). De-ionized water, purified by a Millipore Milli-Q UFplus System (Bedford, MA) was used to prepare all solutions.

**Equipment**

The diffusion cell for in vitro studies was the same as previously described (Figure 1). The diffusion cell for human studies was newly-designed in this study, with a 5 mm diameter for each electrode chamber and the two electrode chambers were 11 mm apart (Figure 2). RI and EP devices were also newly-designed in this study. The RI device has the accuracy of ± 0.01 µA on delivering current and ±0.01 ms on timing. The EP device has the accuracy of ±0.1 V on delivering voltage and ±0.01 ms on timing. A Bio-Rad 680 microplate reader (Bio-Rad, Hemel Hempstead, UK) was used for all colorimetric analysis.

**Preparation of silver–silver chloride electrodes**

Silver–silver chloride (Ag/AgCl) electrodes were used in both in vitro and human studies. The Ag/AgCl electrodes were prepared by chloridizing 99.99% pure silver wire, 1 mm in diameter and 25 mm in length (Aldrich Chemical Company Inc, Milwaukee, WI), immersed in 0.1 M HCl solution (Pt-cathode) for 90 minutes at an applied current of 314 µA.

**In vitro studies**

All experiments were conducted using diffusion cells, in which both electrode chambers were located on the same surface side of a porcine skin (250 µm) obtained by...
measurement of blood uric acid

the electrode chambers and stored in microcentrifuge tubes at 4°C for later quantification of uric acid. The subject’s blood uric acid level was also measured before experiments at the central laboratory of the Puli Christian Hospital.

Determination of the extracted uric acid

Colorimetric assay was used to determine the amount of uric acid extracted through the porcine or human skin during the in vitro and human studies. An excellent linear relationship ($r^2 > 0.9$) between uric acid concentrations and its relative absorbance was found, allowing the uric acid concentration to be calculated simply by linear regression.

Statistical analysis

One-way ANOVA was used to determine whether there were significant differences between EP parameters within a RI parameter, and between RI parameters within an EP parameter for the uric acid extraction. Post hoc comparisons were made with independent samples $t$-tests. A linear regression test was used to determine the relationship between the subject’s blood uric acid level and uric acid concentrations in collection solutions. All statistical analyses were carried out using SPSS software (SPSS Inc., Chicago, IL), with the level of statistical significance set at $P < 0.05$.

Results and discussion

In vitro studies

In this study, 195 combinations of RI and EP were chosen for the transdermal uric acid extraction in order to find out the optimum combination of EP and RI on noninvasive and transdermal determination of blood uric acid level in humans.

Uric acid is a negatively-charged molecule and its molecular weight is about 168 Da. Because of its small size and negative charge, uric acid was expected to be transdermally extractable. Electromigration was expected to be the dominant physical factor for the uric acid extraction.

As shown in Figure 4, the application of RI, EP, or the combination of both could facilitate a higher uric acid extraction as compared to diffusion alone. It was found that the uric acid extraction increased as the phase duration of the symmetrical biphasic direct current (PDSBdc) increased. At 50 V/cm², 100 V/cm², and 150 V/cm², uric acid extraction using the 180 s PDSBdc was found to be significantly higher ($P < 0.001$ in all cases) than that of protocols utilizing 0 s, 15 s, and 30 s PDSBdc conditions. Although an independent samples $t$-test revealed no significant difference ($P = 0.617$) between uric acid extraction using the 60 s and 180 s PDSBdc,
it was observed that the longer the PDSBdc, the better the uric acid extraction was. Therefore, 180 s PDSBdc was recommended as the optimum setting. On the other hand, the combination of RI and EP could facilitate a higher uric acid extraction as compared to RI or EP alone. This was because EP could create reversible nanochannels on the skin\textsuperscript{10,11} and this could further facilitate the uric acid extraction as compared with RI alone. It was also found that the higher the voltage of the EP, the better the uric acid extraction was. This meant that 150 V/cm\textsuperscript{2} EP was better than 50 V/cm\textsuperscript{2} EP ($P < 0.001$ in all cases of 15 s, 30 s, 60 s, and 180 s PDSBdc), as 150 V/cm\textsuperscript{2} EP had relatively more energy to effectively create more nanochannels. On the other hand, there was no significant difference ($P = 0.413$) between the 100 V/cm\textsuperscript{2} EP and 150 V/cm\textsuperscript{2} EP on the uric acid extraction. 100 V/cm\textsuperscript{2} EP was therefore recommended as the optimum setting. Also, no significant difference ($P = 0.552$) was found between 1 ms and 3 ms pulse width of the EP on the uric acid extraction. As such, a 1 ms pulse width of the EP was recommended as the optimum setting because 1 ms pulse width was commonly used in clinic. Moreover, EP with 10 pulses per second facilitated significantly more uric acid extraction ($P < 0.05$) as compared with EP with 5 pulses per second. Therefore, EP with 10 pulses per second was recommended as the optimum setting. In a nutshell, the optimum combination of RI and EP on uric acid extraction was as follows:

### Table 1 Experimental protocol for the in vitro studies

<table>
<thead>
<tr>
<th>Uric acid extraction</th>
<th>Reverse iontophoresis parameters</th>
<th>Symmetrical biphasic dc (0.3 mA/cm\textsuperscript{2})</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No Voltage</td>
<td>Voltage 50 V/cm\textsuperscript{2}</td>
</tr>
<tr>
<td></td>
<td>No Voltage</td>
<td>Voltage 100 V/cm\textsuperscript{2}</td>
</tr>
<tr>
<td></td>
<td>No Voltage</td>
<td>Voltage 150 V/cm\textsuperscript{2}</td>
</tr>
<tr>
<td></td>
<td>0 ms Phase duration</td>
<td>1 ms</td>
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<tr>
<td></td>
<td>15 s Phase duration</td>
<td>3 ms</td>
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<td></td>
<td>30 s Phase duration</td>
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<tr>
<td></td>
<td>60 s Phase duration</td>
<td>3 ms</td>
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<tr>
<td></td>
<td>180 s Phase duration</td>
<td>1 ms</td>
</tr>
</tbody>
</table>

**Notes:** One hundred ninety five different combinations of reverse iontophoresis and electroporation parameters were selected for the in vitro transdermal uric acid extraction, in order to determine the optimum combination for uric acid extraction.
RI: symmetrical biphasic dc with a current density of 0.3 mA/cm² and a phase duration of 180 s
EP: 100 V/cm² electric pulse with a pulse width of 1 ms and 10 pulses per second.

**Human studies**

No erythema was observed on any subject’s skin after the application of RI and EP. In addition, no subject reported pain or even unpleasant feeling during or after the experiment. Therefore, this human study demonstrated that RI and EP could provide a safe, painless, and non invasive extraction of uric acid in humans.

In this human study, the combination of RI and EP significantly promoted transdermal uric acid extraction, but there are no other human studies for data comparison. As shown in Figure 5, a good correlation \( r^2 = 0.88 \) between the subject’s blood uric acid level and uric acid concentrations in collection solutions was observed. This meant that the subject’s blood uric acid level could be predicted by the uric acid concentrations in collection solutions using the linear regres-

**Figure 3** Experimental protocol showing the time sequence for the application of electroporation, reverse iontophoresis, and sample collection.

**Figure 4** In vitro studies of the transdermal and noninvasive extraction of uric acid by different combinations of reverse iontophoresis (RI) and electroporation (EP). For the RI setting, it was a symmetrical biphasic dc with the current density of 0.3 mA/cm². The legend shows the electroporation setting where PW and P/s are the pulse width and pulse per second of the electroporation, respectively.
sion equation. Therefore, a transdermal and noninvasive method for blood uric acid monitoring was established.

Despite promising early results, the correlation between the subject’s blood uric acid level and uric acid concentration in the collection solution was still not too high. Hence, it is necessary to test this method using larger subject groups in order to achieve a better correlation between the subject’s blood uric acid level and uric acid concentration in the collection solution. On the other hand, subjects with a large difference of blood uric acid level are expected to improve the correlation as well.

Conclusion

Uric acid could be transdermally and noninvasively extracted by the combination of RI and EP. The optimum mode was the combination of the RI with symmetrical biphasic dc (current density = 0.3 mA/cm²; phase duration = 180 s) and EP with 10 pulses per second (voltage = 100 V/cm²; pulse width = 1 ms). A good correlation between the subject’s blood uric acid level and uric acid concentrations in collection solutions was observed. Therefore, a transdermal and noninvasive method for blood uric acid monitoring was established.

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Disclosure

The authors report no conflicts of interest in this work.

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