ORIGINAL RESEARCH

SLC39A1 Overexpression is Associated with Immune Infiltration in Hepatocellular Carcinoma and Promotes Its Malignant Progression

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Background: Solute carrier family 39 member 1 (SLC39A1) has been identified as a zinc ion transport protein that possesses oncogenic properties in various types of cancers. However, its potential function in hepatocellular carcinoma (HCC) remains unknown. This study aimed to investigate the expression profile and potential mechanisms of SLC39A1 in HCC.

Methods: SLC39A1 expression was analyzed using multiple databases. The clinical significance and associated biological pathways of SLC39A1 were investigated using bioinformatics analysis. Potential correlations between SLC39A1 expression and tumor immunity in HCC were also evaluated using single-sample gene set enrichment analysis (GSEA). Sixty paired HCC samples were used to verify the expression pattern of SLC39A1. In vitro studies were performed to investigate the oncogenic effects of SLC39A1 in HCC. Western blot analysis was conducted to further investigate the possible involved signaling pathways.

Results: The overexpression of SLC39A1 in HCC was determined by bioinformatics analysis and was confirmed in tissues from our center. SLC39A1 overexpression was also significantly correlated with worse prognosis, advanced TNM stage, and histological grade. GSEA analysis demonstrated that SLC39A1 overexpression was involved in various tumor-related pathways, such as the cell cycle and Wnt signaling pathway. SLC39A1 knockdown repressed the proliferation, invasion, and migration abilities of HCC cells. Furthermore, SLC39A1 knockdown decreased the expression of the tumor progression-related proteins (eg, cyclin D1 and MMP2) and Wnt signaling pathway-related proteins (eg, Wnt3A and β -catenin). In addition, SLC39A1 overexpression may be associated with impaired tumor immunity in HCC, as evidenced by the increased infiltration of Th2 cells and reduced infiltration of cytotoxic cells. **Conclusion:** These findings preliminarily suggested the crucial effect of SLC39A1 overexpression on HCC tumor progression and immunosuppression, suggesting its potential as a novel prognostic and therapeutic target in HCC.

Keywords: SLC39A1, hepatocellular carcinoma, prognosis, Wnt signaling pathway, immune infiltration

Introduction

The predominant malignant liver tumor, HCC, is one of the most common causes of cancer-related mortality globally.¹ Despite the rapid development of curative therapies such as partial hepatectomy, liver transplantation, radiofrequency ablation, and targeted therapy, the overall survival of HCC patients is still poor due to tumor heterogeneity and the specific tumor microenvironment.² Based on some immune checkpoints, specifically expressed on T cells, scientists have provided some novel therapies for HCC patients. For instance, patients receiving atezolizumab plus bevacizumab had longer progression-free survival compared with those receiving sorafenib.³ Another study showed that HCC patients receiving the combination therapy of sintilimab and a bevacizumab biosimilar (IBI305) had a longer median progression-free survival than those receiving sorafenib.⁴ However, approximately 75% of HCC patients show a weak response to

immunotherapies, and the reason remains unknown.⁵ Therefore, a comprehensive investigation of the underlying molecular mechanisms involved in HCC progression and the associated immune microenvironment is increasingly needed for developing more effective therapeutic strategies.

Solute carrier family 39 members (SLC39As) have been identified as zinc metal ion transporting proteins, and their dysregulation has been proven to induce cellular zinc dyshomeostasis and exhibit various oncogenic properties in multiple malignancies, such as lung, cervical, and pancreatic cancers.^{6–11} For instance, SLC39A4 accelerates epithelial mesenchymal transition (EMT), and induces resistance to cisplatin therapy in lung cancer cells. It also induces phosphorylation of focal adhesion kinase (FAK) and paxillin, thus promoting tumor progression in pancreatic cancer cells.^{9,10} Conversely, SLC39A8 effectively inhibits the development of clear cell renal cell carcinoma by blocking EMT progression, whereas knockdown of SLC39A7 attenuates the metastatic ability of cervical cancer.^{11,12} As the first member of SLC39As, SLC39A1 was mainly expressed on the plasma membrane and involved in the zinc absorption activity.^{7,13} Oncogenic studies about SLC39A1 have been conducted in several malignancies, such as glioma and prostate cancer. Wang et al demonstrated that SLC39A1 overexpression promotes tumor progression and correlates with unfavorable clinical outcomes in glioma.¹⁴ In addition, downregulation of SLC39A1 impairs the accumulation of zinc in tumor cells, thereby promoting the malignant phenotype of prostate cancer.^{15,16} However, little is known about the regulatory mechanism of SLC39A1 in HCC progress.

In the current study, the expression levels of SLC39A1 and its clinical significance in HCC were comprehensively investigated using multiple bioinformatics analyses tools. In addition, the oncological role of SLC39A1 in HCC was determined through functional enrichment analysis and in vitro studies.

Materials and Methods

Public Databases Analyses

The Oncomine database (http://www.oncomine.org) is an oncogene database that provides different data sets for metaanalysis. We used the Oncomine database to evaluate the gene expression of SLC39As.¹⁷ TIMER database (https:// cistrome.shinyapps.io/timer/) was used to assess the expression levels of SLC39A1 in pan-cancer.^{18,19} HCCDB, an HCC expression atlas database, collects 15 HCC expression datasets (approximately 4000 samples) to be used publicly for analysis. We used HCCDB to analyze the expression pattern of SLC39A1 in different HCC datasets.²⁰ The sequencing data of mRNA and the clinical data of HCC (including 50 normal and 374 tumor tissues) were collected from The Cancer Genome Atlas (TCGA) for further analysis. MethSurv (https://biit.cs.ut.ee/methsurv/), an interactive portal for the evaluation of prognosis based on DNA methylation status, was used to analyze the DNA methylation sites of SLC39A1 and its prognostic value in HCC.²¹ Finally, the open access portal, cBioportal database (http://cbioportal. org), which provides comprehensive cancer genomics datasets analysis, was used to analyze the altered expression of SLC39A1 and its effect on the prognosis of HCC patients.²²

Correlation Analysis of SLC39A1 Expression and Clinicopathological Features in HCC

The relationship between the clinicopathological characteristics of HCC patients and SLC39A1 expression was analyzed using Student's *t*-test or the Wilcoxon signed-rank sum test using the ggplot2 package. Survinier and survival packages were used for survival and Cox analyses, and the time ROC package was used to draw the receiver operating characteristic curve (ROC).

Functional Enrichment Analysis

The DESeq2 package was used to identify the differentially expressed genes (DEGs) in the two groups of patients having high and low SLC39A1 expression patterns.²³ Genes with absolute fold changes (FC) larger than 1.5 and adjusted P value < 0.05 were screened for further analysis. The ClusterProfiler package was used for Gene Ontology (GO) enrichment analysis of SLC39A1-related DEGs.²⁴ The functions with adjusted P values < 0.05, and FDR < 0.2 indicated significance. GSEA was used to investigate the biological pathways mediated by high SLC39A1 expression.²⁵ The

enriched pathways with adjusted *P* value < 0.05, and FDR < 0.25 indicated significance after 1000 permutations. The R package ClusterProfiler was used for statistical analysis and graphical plotting.

Immune Infiltration Analysis

Single-sample gene set enrichment analysis (ssGSEA) from the gene set variation analysis (GSVA) package was used to quantify the relative infiltration levels of 24 immune cells.²⁶ The Spearman correlation analysis and Wilcoxon rank sum test were used to assess the immune infiltration associated with SLC39A1 expression in HCC.

Clinical Specimens

Fresh-frozen HCC tissues and paired normal adjacent tissues of 60 HCC patients were collected after hepatectomy at the Sun Yat-sen Memorial Hospital between 2016 and 2019. None of the patients received any therapy such as chemotherapy or radiotherapy prior to surgery. After surgical resection, all tissues were immediately stored at -80 °C or liquid nitrogen until RNA extraction. This study was approved by the Medical Ethics Committee of Sun Yat-sen Memorial Hospital, and written informed consent was obtained from all participants before participation in the study. The clinical information of the participants is summarized in <u>Supplementary Table S1</u>.

Cell Culture and Plasmid Transfection

Human Huh7 cells, obtained from the Cell Bank of the Chinese Academy of Sciences (Shanghai, China), were cultured in Dulbecco's modified Eagle's medium (DMEM; Gibco BRL, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS; Procell Life Science & Technology Co., Ltd., Wuhan, China) in an incubator with 5% CO₂ at 37 °C. Plasmids containing the negative control sequence (shNC) or the SLC39A1 knockdown sequence (sh#1-3) were obtained from Kidan Biosciences (Guangzhou, China). Briefly, cells were seeded in 6-well plates, cultured to reach an appropriate confluence, and then transfected with different plasmids using Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. After 48 h of incubation, transfected cells were harvested to be used for quantitative real-time polymerase chain reaction (qRT-PCR), cell function experiments, and Western blot analysis. SLC39A1 knockdown sequences were as follows: sh#1, 5'-GGGTATGGACAGGTCATAAACCTCGAGGTTTATGACCTGTCCATACCC-3'; sh#2, 5'-CCTGTTGGTCCTAGCCCATTTCTCGAGAAATGGGCTAGGACCAACAGG-3'; sh#3, 5'-GCAGATCACACTGGC TTACAACTCGAGTTGTAAGCCAGTGTGATCTGC-3'.

RNA Extraction and qRT-PCR

Total RNA was isolated from tissues and cell lines using the EZ-press RNA Purification Kit (EZBioscience, Roseville, USA) and then was reverse-transcribed to cDNA with Evo M-MLV RT Premix for qPCR (Accurate Biotechnology Co., Ltd., Hunan, China). qRT-PCR was performed to evaluate the SLC39A1 expression levels in cell lines and tissues using the $2^{-\Delta\Delta CT}$ method. The following primers were used: SLC39A1, 5'-GAACAAGAGATGGTCAAGTC-3' (forward), and 5'-ATGTGAGCCTGTCCTTATG-3' (reverse); actin, 5'-CTACCTCATGAAGATCCTCACCGA-3' (forward), and 5'-TTCTCCTTAATGTCACGCACGATT-3' (reverse).

Western Blot

Radioimmunoprecipitation assay (RIPA) lysis buffer (CWBIO, Beijing, China) with phosphatase and protease inhibitors (CWBIO, Beijing, China) was used to lyse cultured cells, followed by centrifugation at 14,000 rpm for 30 min at 4°C. A bicinchoninic acid assay (CWBIO, Beijing, China) was used to measure protein concentrations. After proteins were separated on SDS–PAGE gels (EpiZyme, Shanghai, China), the measured proteins were transferred onto PVDF membranes (Roche Applied Science, Germany). Appropriate antibodies were then used to carry out immunoblot analysis of all related proteins. Finally, bands were visualized using an Omni-ECLTM Femto Light Chemiluminescence Kit (EpiZyme, Shanghai, China) with a G: BOX Chemi XT4 imager (Syngene, UK). The antibodies for the relative proteins mentioned in this study are listed in <u>Supplementary Table S2</u>.

Cell Counting Kit-8 (CCK-8) Assay

The Cell Counting Kit-8 (APExBIO, Houston, Texas, USA) was used to examine the cellular viability. Briefly, Huh7 cells transfected with different plasmids (shNC, sh#1, sh#2, and sh#3) were seeded in 96-well plates at a density of 2×10^3 cells/well. At days 1–6, 10 µL CCK-8 reagent were mixed with 100 µL DMEM, and the mixed solution was added to each well and re-incubated for 2 h, followed by measurement of the absorbance at a wavelength of 450 nm using a microplate reader (Thermo MK3, Thermo Fisher Scientific, USA).

5-Ethynyl-20-Deoxyuridine (EdU) Assay

EdU Imaging Kits (Cy3) (APExBIO, Houston, Texas, USA) were used to evaluate the cellular proliferation. Briefly, Huh7 cells were seeded into 24-well plates at a density of 7×10^4 cells/well and incubated for 24 h. After re-incubation with the EdU reagent for 2h, the cells were fixed and stained for 30 min. Nucleic acid was stained with Hoechst 33342. Representative images were obtained using a fluorescence microscope.

Transwell Assay

Transfected cells were seeded into each upper transwell chamber (Corning, NY, USA) with or without Matrigel (Corning, NY, USA) at a density of 5×10^4 cells/well and incubated in 200 µL serum-free DMEM. The lower chambers contained 500 µL DMEM supplemented with 100 µL FBS. Then, cells from the upper compartments were removed after 48 h, and the migrated cells were fixed with paraformaldehyde for 15 min, followed by staining with 0.3% crystal violet for 10 min at room temperature. Cell counting was performed using a microscope.

Statistical Analysis

All experimental data were analyzed using SPSS software (version 25; Chicago, IL, USA) and GraphPad Prism software (version 8.0; San Diego, CA, USA). All bioinformatics statistical analyses were conducted using R software version 4.0.5 (https://www.r-project.org/). Statistical significance was defined as P < 0.05, unless otherwise specified.

Results

SLC39A1 Was Upregulated in HCC

SLC39As expression in HCC was evaluated using the Oncomine database. Results indicated that SLC39A1 expression was notably upregulated in HCC tissues compared with that in adjacent normal tissues (Figure 1A). SLC39A1 pancancer expression was investigated using the TIMER database. As shown in Figure 1B, SLC39A1 expression levels were markedly elevated in most cancers, including stomach adenocarcinoma, hepatocellular carcinoma, cholangiocarcinoma, esophageal carcinoma, rectum adenocarcinoma, colon adenocarcinoma, glioblastoma multiforme, bladder urothelial carcinoma, and breast invasive carcinoma. However, SLC39A1 was markedly downregulated in pheochromocytoma, paraganglioma, thyroid carcinoma, prostate adenocarcinoma, and kidney chromophobe. To further verify the over-expression of SLC39A1 in HCC, we assessed the mRNA levels of SLC39A1 in different HCC cohorts from the HCCDB database, which showed that SLC39A1 expression levels in HCC were also markedly upregulated compared to that in adjacent normal tissues (Figure 1C).

SLC39A1 Overexpression Was Associated with Unfavorable Clinical Outcomes in HCC

As shown in Table 1, high SLC39A1 expression was markedly correlated with poor clinicopathological characteristics such as TNM stage, histological grade, and alpha fetoprotein (AFP) levels. HCC patients with a higher TNM stage (P = 0.014), advanced histologic grades (P < 0.001), and higher AFP levels (P < 0.001) had higher expression levels of SLC39A1 (Figure 2A–C). In addition, Kaplan–Meier survival analysis demonstrated that HCC patients with lower SLC39A1 expression had a superior overall survival (OS), progression-free survival and disease specific survival time when compared with those having higher SLC39A1 expression (Figure 2D–F). ROC analysis indicated that the area under curves (AUC) of SLC39A1 expression for 1-, 3-, and 5-year OS were 0.753, 0.636, and 0.646, respectively, suggesting a reliable prognostic



Figure I SLC39A1 is over-expressed in Hepatocellular Carcinoma (HCC). (A) The different expression of SLC39A family between normal and tumor in HCC from Oncomine database. (B) Dynamic expression of SLC39A1 in pan-cancer. (C) Overexpression of SLC39A1 in HCC was identified in HCCDB database. *P < 0.05, **P < 0.01, ***P < 0.001.

Table I	Correlation	Between	SLC39A1	Expression and	d Clinicopathological	Features of HCC	2 Patients from T	CGA
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Parameters		SLC39A1 Expression		P value
		Low	High	
TNM stage (n=350)	Stage I	99 (28.3%)	74 (21.1%)	0.029*
	Stage II	43 (12.3%)	44 (12.6%)	
	Stage III	34 (9.7%)	51 (14.6%)	
	Stage IV	I (0.3%)	4 (1.1%)	
Gender (n=374)	Female	56 (15.0%)	65 (17.4%)	0.377
	Male	131 (35.0%)	122 (32.6%)	
Age (n=373)	≤60	82 (22.0%)	95 (25.5%)	0.196
	>60	105 (28.2%)	91 (24.4%)	
Histologic grade (n=369)	GI	36 (9.8%)	19 (5.1%)	< 0.001*
	G2	99 (26.8%)	79 (21.4%)	
	G3	45 (12.2%)	79 (21.4%)	
	G4	5 (1.4%)	7 (1.9%)	
Adjacent hepatic tissue inflammation (n=237)	None	70 (29.5%)	48 (20.3%)	0.303
	Mild	50 (21.1%)	51 (21.5%)	
	Severe	(4.6%)	7 (3.0%)	
AFP (ng/mL) (n=280)	≤400	127 (45.4%)	88 (31.4%)	< 0.001*
	>400	18 (6.4%)	47 (16.8%)	
Albumin (g/dl) (n=369)	<3.5	40 (13.3%)	29 (9.7%)	0.537
	≥3.5	122 (40.7%)	109 (36.3%)	
Prothrombin time (n=297)	≤4	110 (37.0%)	98 (33.0%)	0.828
	>4	49 (16.5%)	40 (13.5%)	
Child-Pugh grade (n=241)	А	119 (49.4%)	100 (41.5%)	1.000
	В	12 (5.0%)	9 (3.7%)	
	С	I (0.4%)	0 (0.0%)	
Fibrosis ishak score (n=215)	0	47 (21.9%)	28 (13.0%)	0.261
	1/2	15 (7.0%)	16 (7.4%)	
	3/4	12 (5.6%)	16 (7.4%)	
	5/6	45 (20.9%)	36 (16.7%)	
Vascular invasion (n=318)	No	119 (37.4%)	89 (28.0%)	0.060
	Yes	50 (15.7%)	60 (18.9%)	

Notes: Statistical significance was calculated by the chi-square test and Fisher's extract test. *P<0.05.

predictive discrimination for SLC39A1 (Figure 2G). Univariate and multivariate Cox analyses indicated that SLC39A1 expression was an independent unfavorable prognostic biomarker for HCC patients (Figure 3A and B).

Prognostic Value of SLC39A1 DNA Methylation and Genetic Alteration in HCC

As shown in Figure 4A, nine CpG methylated sites were found in SLC39A1, among which four were identified as unfavorable prognostic factors (cg16414271, HR = 1.735, P = 0.0047; cg10624480, HR = 1.556, P = 0.033; cg14747580, HR = 1.542, P = 0.026; and cg11411904, HR = 1.509, P = 0.049), and one was identified as a favorable prognostic factor (cg09079613, HR = 0.505, P = 0.00068; Figure 4B–F). In addition, Spearman correlation analysis demonstrated that the four CpGs of SLC39A1 were negatively correlated with SLC39A1 expression (cg11073883: Cor = -0.214, P < 0.001; cg08623548: Cor = -0.271, P < 0.001; cg09079613: Cor = -0.290, P < 0.001; cg0546481: Cor = -0.143, P = 0.006; Figure S1a–d). We further investigated the prognostic value of SLC39A1 genetic alterations in HCC. Results indicated that approximately 13% of HCC patients had SLC39A1 genetic alterations, most of which were amplified. The most common nucleotide change was C > T transversion (Figure 4G). SLC39A1 genetic alterations in HCC patients resulted in a poor OS (P < 0.05; Figure 4H).



Figure 2 Overexpression of SLC39A1 correlates with poor survival and unfavorable clinicopathological characteristics in Hepatocellular Carcinoma (HCC). (A-C) SLC39A1 expression of HCC was significantly associated with TNM stage, Histologic grade, and alpha fetoprotein (AFP) level. (D-F) Kaplan-Meier analysis showed HCC patients with high expression of SLC39A1 had poor overall survival (D), progression-free survival (E) and disease specific survival (F) than those with low expression in TCGA. (G) ROC (Receiver operator characteristic curve) analysis showed that SLC39A1 was a reliable prognostic predictor in patients with HCC. *P < 0.05, ***P < 0.001.

Functional Enrichment Analysis of SLC39A1 Expression in HCC

To further elucidate the potential mechanism of SLC39A1 in HCC, GO enrichment analysis and GSEA were conducted. GO enrichment analysis results indicated that SLC39A1 might have a regulatory effect on various cellular components and molecular functions, including the catenin complex, transmembrane transporter complex, serine-type peptidase activity, DNA-binding transcriptional activator activity, RNA polymerase II-specific, metal ion transmembrane

	Characteristics	Hazard Ratio(95% CI)		P value
н	istologic grade (G3&G4 vs. G1&G2)	1.091 (0.761-1.564)	He-H	0.636
	AFP(ng/ml) (>400 vs. <=400)	1.075 (0.658-1.759)	⊢ <mark>,</mark>	0.772
	Age (>60 vs. <=60)	1.205 (0.850-1.708)	┝╁●──┥	0.295
	Residual tumor (R1&R2 vs. R0)	1.604 (0.812-3.169)	⊢ <mark>⊢</mark> •−−−−−	0.174
TNM stag	ge (Stage III&Stage IV vs. Stage I&Stage II)	2.504 (1.727-3.631)	¦	— <0.001
	Child-Pugh grade (B&C vs. A)	1.643 (0.811-3.330)	⊢ <mark>⊢</mark> ● −−−−−	- 0.168
	Albumin(g/dl) (>=3.5 vs. <3.5)	0.897 (0.549-1.464)	⊢●¦→→	0.662
	Prothrombin time (>4 vs. <=4)	1.335 (0.881-2.023)	⊦ <mark>¦_●</mark> −−−4	0.174
	Vascular invasion (Yes vs. No)	1.344 (0.887–2.035)	⊢	0.163
	SLC39A1 (High vs. Low)	1.868 (1.312-2.658)	¦ ⊨ _ i	<0.001
			1 1 1 1 2 3	
	Characteristics	Hazard Ratio (95% CI)		P value
		2 204 (4 648, 2 478)		L <0.00
I NM stage (Stage III&Stage IV vs. Stage I&Stage II)	2.394 (1.648-3.478)		— <0.00 ⁷
	SLC39A1 (High vs. Low)	1.692 (1.164–2.458)	¦ ⊢_ ♦1	0.006
			1.0 1.5 2.0 2.5 3.	.0 3.5

Figure 3 Univariate and multivariate regression cox analysis of SLC39A1. (A) Univariate cox regression analysis was performed to confirmed high expression of SLC39A1 was associated with overall survival (OS) of HCC patients. (B) Multivariate cox regression analysis was performed to confirmed high expression of SLC39A1 was independent risk factor for OS of HCC patients.

transporter activity, and receptor ligand activity. As predicted, biological processes (BP) associated with zinc regulation were significantly enriched in the high SLC39A1 group, including cellular zinc ion homeostasis, cellular response to zinc ions, zinc ion homeostasis, and response to zinc ions (Figure 5A). In addition, GSEA demonstrated that SLC39A1 overexpression may be involved in many tumor progression-related pathways, such as the Wnt signaling pathway, MAPK signaling pathway, cell cycle, DNA replication, NOD-like receptor signaling pathway, and TGF- β signaling pathway (Figure 5B–G). These results suggest that SLC39A1 overexpression promotes HCC tumorigenesis and metastasis.

Correlation Between Immune Infiltration and SLC39A1 Expression

We performed ssGSEA to investigate the potential association between SLC39A1 expression levels and the infiltration levels of 24 types of immune cells in HCC. Results revealed that Th2 cells were positively associated with SLC39A1 expression (Cor > 0.3, P < 0.05), while cytotoxic cells were negatively associated with SLC39A1 expression (Cor < -0.3, P < 0.05; Figure 6A–C). Tumors having higher SLC39A1 expression exhibited higher infiltration levels of Th2 cells but lower infiltration levels of cytotoxic cells (Figure 6D and E). These findings indicate that SLC39A1 overexpression may be involved in the immune suppression of HCC.

Validation of SLC39A1 Overexpression in HCC in the Study's Cohort

Given that SLC39A1 is upregulated in multiple HCC databases, qRT-PCR experiments and genomic sequencing were performed to validate the overexpression of SLC39A1 in tumor tissues. As shown in Figure 7A, SLC39A1 mRNA levels were significantly overexpressed in 60 HCC tissues compared to their paired adjacent non-tumorous tissues (P < 0.0001). The 60 patients were then stratified into three groups; over-expression, down-expression, and no-change under the thresholds of P < 0.05, and FC > 1.5 or < 0.67. Among the 60 HCC samples from our center, 38 (63%) exhibited



Figure 4 Prognostic value of SLC39A1 DNA methylation and genetic alteration in HCC. (A) The DNA methylation clustered expression of SLC39A1. (B–F) Overall survival (OS) curve of cg09079613 (B), cg16414271 (C), cg10624480 (D), cg14747580 (E) and cg11411904 (F) in HCC patients. (G) The overview of genetic alteration in SLC39A1. (H) OS curve between altered and unaltered group of SLC39A1.



Figure 5 Gene Ontology (GO) enrichment analysis and Gene set enrichment analysis (GSEA) of SLC39A1 in HCC. (**A**) GO enrichment analysis of SLC39A1 related genes in HCC. (**B–G**) GSEA showed that the group with high expression of SLC39A1 was significantly associated with many tumor progression-related pathways, such as Wnt signaling pathway (**B**). Cell cycle (**C**), DNA replication (**D**), TGF- β signaling pathway (**E**), NOD like receptor signaling pathway (**F**) and MAPK signaling pathway (**G**).



Figure 6 The expression of SLC39A1 was associated with the immune infiltration in the tumor microenvironment. (A) Correlation between the relative abundance of 24 immune cells and SLC39A1 expression. (B–E) Box plot and correlation diagrams showing the difference of Th2 and cytotoxic cells infiltration level between SLC39A1-high and low group. *** P < 0.001.

SLC39A1 upregulation in tumor tissues, while six (10%) exhibited SLC39A1 downregulation in tumor tissues (Figure 7B and C). Furthermore, genomic sequencing of 20 pairs of HCC samples showed that SLC39A1was significantly overexpressed in tumor tissues than in paired adjacent non-tumorous tissues (P < 0.0001; Figure 7D).

SLC39A1 Promotes HCC Cellular Proliferation, Migration, and Invasion via the Wnt/ β -Catenin Pathway

To further determine the underlying biological mechanism of SLC39A1 in HCC, Huh7 cells were transfected with sh-SLC39A1 plasmid. A significant reduction in SLC39A1 expression was observed in the shRNA group (P < 0.0001) (Figure 8A). Using CCK-8 and EdU assays, knockdown of SLC39A1 significantly suppressed the Huh7 cellular proliferation (Figure 8B and C). Transwell assays demonstrated that the cellular migration and invasion abilities were significantly inhibited in the sh-SLC39A1 group compared to those in the shNC group (Figure 8D). In addition, upon knocking down SLC39A1, a significant reduction was observed in the protein levels of cyclin D1, N-cadherin, and MMP2, which contribute to tumor proliferation, migration, and invasion (Figure 8E). Furthermore, Wnt/ β -catenin signaling pathway-related proteins, including Wnt3A, phosphorylation-GSK3 β (p-GSK3 β), and β -catenin, were also markedly decreased in the sh-SLC39A1 groups (Figure 8F). These findings preliminarily illustrate that cellular proliferation, migration, and invasion of HCC are markedly suppressed by SLC39A1 knockdown via the Wnt/ β -catenin pathway.

Discussion

Multiple zinc transporters in the SLC39A family have been demonstrated to mediate the dysregulation of zinc accumulation in tumor cells and are associated with the malignant phenotype of multiple cancers, such as gastric, lung, and breast cancers.^{27–29} The current study revealed that SLC39A1, one of the SLC39A family members, exerts a pro-tumorigenic effect in HCC. Results also showed that SLC39A1 overexpression was notably observed in HCC



Figure 7 The overexpression of SLC39A1 in HCC patients. (A) The differences in the expression of SLC39A1 in 60 tumor and paired normal tissues. (B) The Fold Changes (FC) of SLC39A1 expression in 60 HCC patients. (C) Distribution of three groups (High, Low and No change) in 60 HCC patients. (D) The SLC39A1 expression results of genomic sequence in 20 HCC patients. ****P < 0.0001.

tissues and correlated with advanced TNM stage, histologic differentiation, AFP levels, and poor prognosis of HCC patients. This suggests an unfavorable predictive prognostic role of SLC39A1 in HCC.

GSEA analysis demonstrated that various oncogenic BP were mediated by SLC39A1 overexpression, such as cell cycle, DNA replication and Wnt signaling pathway, which contribute to tumor proliferation and metastasis.^{30–35} This was also evidenced by the fact that SLC39A1 downregulation notably repressed HCC cellular proliferation, migration, and invasion. Furthermore, SLC39A1 knockdown also decreased the expression of cyclin D1, N-cadherin, and MMP2, which are critical for the cellular proliferation and metastasis of the tumor.^{36–41} Of note, downregulation of some members of Wnt signaling pathway was also observed upon knocking down SLC39A1, indicating the potential role of SLC39A1 in the Wnt signaling pathway. Taken together, these findings suggest a critical pro-tumor effect of SLC39A1 overexpression on HCC tumor proliferation and metastasis. However, in-depth experimental studies are needed to elucidate the underlying mechanisms of SLC39A1 in hepatocellular carcinogenesis.

Reduced cellular zinc content affects the balance between Th1 and Th2 cytokine shifts and the activation of many immune cells.^{42,43} This study also demonstrated the specific effect of SCL39A1 on the tumor immune infiltration of HCC. Higher SLC39A1 expression was significantly associated with higher infiltration of Th2 cells but lower infiltration of cytotoxic cells. Th2 cells have been widely considered as tumor-promoting immune cells, leading to unfavorable clinical outcomes in HCC.^{44,45} Higher levels of Th2 cytokines (eg, interleukin [IL]-4 and IL-10) were observed in metastatic HCC, whereas Th1 cytokines (eg, interferon [IFN]- γ and IL-1) were notably decreased.^{46,47} IL-4 and IL-10 have been reported to restrain IFN- γ production and impair the cytotoxic cell-mediated anti-tumor immunity,^{48–51} suggesting that an increased shift from Th1 cells to Th2 cells promotes tumor progression and immunosuppression. In addition, aberrant Wnt signaling pathway in the tumor microenvironment also impairs the cytotoxic T cell infiltration and



shNC sh#1 sh#2 sh#3

Figure 8 Effects of SLC39A1 on HCC proliferation and metastasis. (A) The change of SLC39A1 expression after transfected sh-SLC39A1. (B and C) CCK-8 assay and EdU assay demonstrated that transfection with sh-SLC39A1 plasmid inhibited the proliferation of Huh7 cell. Scale bar = 100μ m. (D) Transwell assays indicated that migration and invasion ability of Huh7 cell was reduced after transfecting with sh-SLC39A1 plasmid. Scale bar = 100μ m. (E) The downregulation of N-cadherin, MMP2 and CyclinD1 in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected with sh-SLC39A1 plasmid was detected by Western blot. (F) The downregulation of Wnt3A, p-GSK3 β and β -catenin in Huh7 cell transfected w

anti-tumor function, inducing therapeutic resistance to immune checkpoint inhibitors.^{33,52–54} Therefore, blocking the Wnt signaling pathway through targeting SLC39A1 may be a promising therapeutic strategy to improve the current HCC immunotherapeutic approaches. Further studies should be performed to elucidate the underlying mechanism of SLC39A1 overexpression in immune cell infiltration in HCC.

The results described in this study may provide new insights into the malignant phenotype and the modulation of immune infiltration in HCC. However, this study had several limitations. First, the findings of this study were dependent on the quality of the data retrieved from the publicly-available databases. Second, further in vivo studies are required to explore and validate the molecular mechanisms of SLC39A1 in HCC.

Conclusions

In conclusion, this study identified SLC39A1 as an unfavorable prognostic biomarker for HCC. SLC39A1 overexpression may promote tumor progression and pro-tumor immunity via the Wnt/ β -catenin signaling pathway, which may be a novel therapeutic approach in HCC.

Data Sharing Statement

The original contributions presented in the study are included in the article and <u>Supplementary Materials</u>. Further inquiries can be directed to the corresponding authors, Yajin Chen, cyj0509@126.com and Changzhen Shang, shang-cz_sysu@163.com.

Ethical Standards

The study was reviewed and approved by the Ethics Committee of Sun Yat-sen Memorial Hospital. The patients/ participants provided their written informed consent to participate in this study. The study was completed in accordance with the Declaration of Helsinki.

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Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work. Xiaowu Ma and Hongkai Zhuang contributed equally to this manuscript. Co-corresponding authors: Yajin Chen and Changzhen Shang.

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Disclosure

All authors declare that they have no conflicts of interest for this work.

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