

# Human serum albumin nanoparticles as an efficient noscapine drug delivery system for potential use in breast cancer: preparation and *in vitro* analysis

Safaa Sebak  
Maryam Mirzaei  
Meenakshi Malhotra  
Arun Kulamarva  
Satya Prakash

Biomedical Technology and Cell  
Therapy Research Laboratory,  
Department of Biomedical  
Engineering, Faculty of Medicine,  
McGill University, Montreal, Quebec,  
Canada

**Abstract:** Drug delivery systems such as nanoparticles can provide enhanced efficacy for anticancer agents. Noscapine, a widely used cough suppressant for decades has recently been shown to cause significant inhibition and regression of tumor volumes without any detectable toxicity in cells or tissues. Nanoparticles made of human serum albumin (HSA) represent promising strategy for targeted drug delivery to tumor cells by enhancing the drug's bioavailability and distribution, and reducing the body's response towards drug resistance. In the present study, we report for the first time the incorporation and delivery of noscapine-loaded HSA nanoparticles to tumor cells. The nanoparticles were designed and optimized to achieve a particle size in the range of 150–300 nm with a drug-loading efficiency of 85%–96%. The nanoparticles were evaluated *in vitro* for their anticancer activity and efficacy on breast cancer cells.

**Keywords:** HSA, encapsulation, microcapsule, nanomedicine, nanotechnology, tumor volumes

## Introduction

Most commonly used chemotherapeutic drugs such as paclitaxel interact with tubulin, the major protein of mitotic spindles, causing growth arrest in metaphase. These agents induce the polymerization of tubulin and stop mitosis of rapidly dividing cells; thereby leading to apoptosis. Despite impressive results, available chemotherapeutic drugs have severe symptoms associated with them including myelosuppression.<sup>1</sup> Also as a result of repeated and prolonged administration of chemotherapeutic agents, drug resistance can occur.<sup>1,2</sup> In addition, these drugs are toxic to healthy tissues, and have poor bioavailability that results in the need of extended intravenous infusions and the use of toxic solubility agents.<sup>1,3</sup>

Noscapine, a phthalide isoquinoline alkaloid derived from opium, has been used as an oral antitussive agent and has shown very few toxic effects in animals and humans. It is a naturally occurring tubulin-binding agent currently undergoing clinical trials for anticancer therapy. Noscapine, which lacks sedative, analgesic, and euphoric properties, has been used for decades as a cough suppressant. Noscapine showed little or no toxicity on kidney, liver, heart, bone marrow, spleen, and small intestine at tumor suppressive doses.<sup>4</sup> Noscapine was also shown to cause growth arrest of tumor cells during mitosis and induces apoptosis<sup>5–7</sup> and can therefore be used as an alternative to current chemotherapeutic drugs.<sup>8–10</sup> It was recently reported that noscapine was effective in reducing the growth of lymphoma and increasing the survival of tumor-bearing mice when administered in their drinking water.<sup>7,11–15</sup> However, current drug delivery methods of high concentration of noscapine are inadequate.

Correspondence: Satya Prakash  
Faculty of Medicine, McGill University,  
3775 University Street, Montreal,  
H3A2B4, Quebec, Canada  
Tel +1 514 398 3676  
Fax +1 514 398 7461  
Email satya.prakash@mcgill.ca

To increase the concentration of noscapine at the tumor site, we have encapsulated noscapine into colloidal drug nanocarriers made of human serum albumin (HSA). Nanoparticles of minimal size can enhance the efficacy of drug metabolism with minimal side effects since they allow for the possibility of site-specific targeted delivery.<sup>16–18</sup> Nanoparticles also offer benefits to carry functional groups (amino and carboxylic groups) that can be used for surface modifications. The emergence of nanoscaled devices as drug delivery systems has been remarkable in such a short span of time and has surpassed conventional methods of drug delivery. Nanoparticles help to increase the stability of drugs and possess useful controlled release properties. HSA-based nanoparticles can be well tolerated without any serious side effects, which is supported by clinical studies with registered HSA-based particle formulations such as Albutex.<sup>18–21</sup> Albumin has been shown to be biodegradable, nontoxic, easy to purify, and soluble in water, allowing ease of delivery by injection and thus an ideal candidate for nanoparticle preparation. Incorporation of suitable drugs in nanoparticles has been shown to protect the pharmacological active substances from degradation during storage as well as from early degradation/inactivation after injection.<sup>22–24</sup>

Many different possibilities have been discussed and tested to reduce the toxicity and increase antitumor efficacy of anticancer drugs using nanodevices.<sup>25–27</sup> Encapsulating noscapine in nanoparticles will help to increase its efficacy and lowers any side effects. In this paper we report for the first time encapsulation of noscapine into HSA nanoparticles with a described protocol on optimization of drug loading and preparation of nanoparticles and their effectiveness on breast cancer cell line (SK-BR-3).

## Materials and methods

### Materials

Human serum albumin (HSA fraction V, purity 96%–99%), noscapine hydrochloride and 8% glutaraldehyde, were purchased from Sigma Aldrich (Oakville, Ontario, Canada). For cell culture, fetal bovine serum (FBS), trypsin, McCoy's 5a medium, penicillin/streptomycin, and the SK-BR-3 cell line were purchased from American Type Culture Collection (Ontario, Canada). Cell-line was cultured according to supplier's instructions. All other reagents were purchased from Fisher (Ontario, Canada).

### Preparation of noscapine nanoparticles

Drug-free HSA nanoparticles and noscapine-loaded nanoparticles crosslinked with glutaraldehyde were prepared

using a pH-coacervation method.<sup>28,29</sup> 100 mg of HSA was dissolved in 2 mL of water or NaCl. Noscapine at a concentration of 5–30 mg/mL was incubated with solution for 4–8 h at room temperature.<sup>30</sup> The pH was adjusted to 8 by the addition of 1 M NaOH. Nanoparticles were formed by adding 8 mL of ethanol drop-wise at a constant rate of 1 mL/min under constant magnetic stirring. The particles were stabilized by crosslinking with 100  $\mu$ L of 8% glutaraldehyde solution. The crosslinking was performed for at least 24 h under constant magnetic stirring at room temperature.<sup>30,31</sup> Drug loading was evaluated following an indirect method of collecting the supernatant of purified particles. The quantity of unloaded free drug present in the supernatant was measured by spectrophotometer that lead to the quantification of the percentage of drug loaded into the nanoparticles.

### Purification of HSA nanoparticles

The resulting albumin nanoparticles were purified by three cycles of ultra-centrifugation (20,000 g, 30 min) followed by redispersion of pellet in water to original volume. Each redispersion step was performed in an ultrasonication bath (Branson 2510; Branson, Danbury, CT) for 15 min.

### Particle size and zeta potential of nanoparticles

The size of noscapine albumin nanoparticles was determined by photon correlation spectroscopy (PCS) using a high performance particle size analyzer (Malvern Instruments, Westborough, MA). The samples were diluted with distilled water and measured at 25°C at a scattering angle of 90°. Size distribution was characterized by a polydispersity index (PI) and the zeta potential was measured with the technique of electrophoretic laser Doppler anemometry, using a Zeta Potential Analyzer (Brookhaven Instruments, Holtsville, NY). Morphological characteristics were examined using scanning electron microscope (Hitachi S-4700 FE-SEM; Hitachi, Tokyo, Japan). In order to determine the stability of the nanocarriers when dispersed in aqueous solution, they were stored at room temperature for 5 days while being regularly monitored for size and zeta potential each day. The size and shape of the nanoparticles were also examined by scanning electron microscopy (SEM).

### Drug release profile of noscapine-loaded HSA nanoparticles

Determination of drug release of noscapine was performed by dispersing 10 mg of noscapine-loaded nanoparticles

in 10 mL phosphate-buffered saline (PBS; pH 7.4) under constant shaking at 200 rpm/min at 37°C. The samples were centrifuged and the amount of free nescapine in the supernatant was determined at predefined time intervals using a UV spectrophotometer Victor<sup>3</sup> V 1420 Multilabel Counter (Perkin Elmer, Boston, MA) at 310 nm. This analysis was performed three times for each sample.

## Cell culture

SK-BR-3 cells were grown in McCoy's 5a medium with 10% FBS and 1% penicillin/streptomycin. Cells were cultured in a humidified incubator containing 5% CO<sub>2</sub> at 37°C.

## *In vitro* cell viability of nescapine-loaded HSA nanoparticles

The determination of cell viability is a common assay to evaluate the *in vitro* cytotoxicity of biomaterials. The MTS assay is a quantitative and rapid colorimetric method for measuring the viability of cells. The cytotoxicity study of the nanoparticles was examined *in vitro* on the breast cancer cell-line SK-BR-3; the cell lines were cultured in 96-well plates at an initial concentrations of 5000 cells/well in fresh medium supplemented with 10% FBS. The cell proliferation was determined by cell counting after trypsinization and trypan blue staining. After 24 h of culture, cells were adherent and the medium was replaced by nescapine-loaded nanoparticles at a final concentration of 50 µg/mL. The media was removed after 24 h and replaced with fresh medium supplemented with 10% FBS. The cell viability and cytotoxicity were determined using MTS cell proliferation kit. At predetermined time intervals, 20 µl of MTT was added in each well and incubated for 4 h in a humidified incubator containing 5% CO<sub>2</sub> at 37°C and absorbance was measured at 490 nm using a Victor<sup>3</sup>V 1420 Multilabel Counter spectrophotometer (Perkin Elmer).<sup>32–34</sup>

## Results and discussion

### Preparation and visualization of nescapine nanoparticles

The objective of the present study was to design HSA nanoparticles for nescapine delivery and establish a standard protocol for their preparation. HSA nanoparticles containing nescapine were prepared by coacervation method, in which ethanol was used as a dissolving agent, followed by crosslinking using glutaraldehyde.<sup>29</sup> The pH-coacervation method has widely been used in the encapsulation of both water-soluble and insoluble drugs. Initially we investigated the effect of the process conditions such as the aqueous HSA concentration, the rotation speed of the magnetic stirrer, the

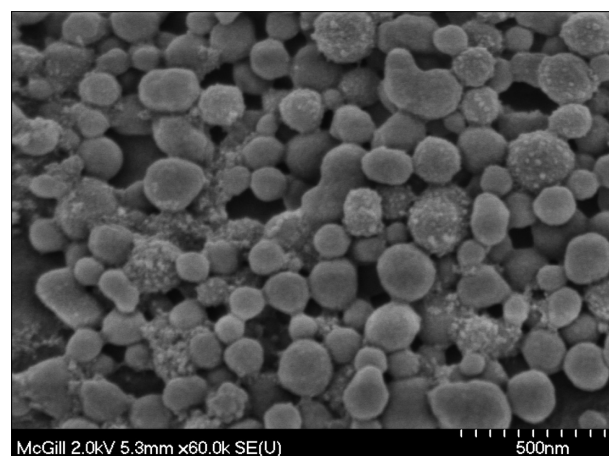
pH of the solution prior to ethanol addition, and the rate of ethanol addition. After experimentation and optimization a final preparation method was prepared. Uniform particles with narrower size distribution were achieved at higher rotation speed of the magnetic stirrer with ethanol, dropped at a constant flow rate using a peristaltic pump. With the modified pH-coacervation technique reported in this study, we achieved nescapine-loaded HSA nanoparticles of diameters between 150 and 300 nm and narrow size distribution determined by the PI of < 0.4. The SEM micrographs reveal morphological aspects of nanoparticles with a spherical shape and uniform size distribution in the desired range of nanoparticle sizes (Figure 1).

### Degree of crosslinking of nescapine-loaded nanoparticles

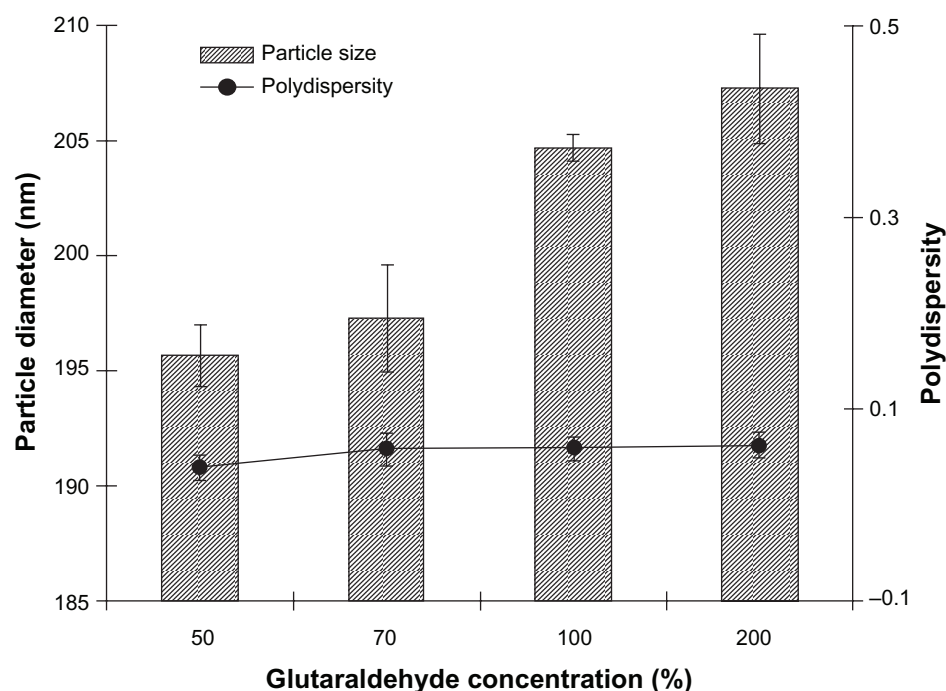
Crosslinking of nanoparticles with a crosslinking agent can limit their degradation rate and hydration potential, thereby attaining slow-release kinetics.<sup>35,36</sup> The influence of the crosslinking process (glutaraldehyde concentration, 50%–200%) on the size of albumin nanoparticles was investigated. This process plays a major role in the stability and drug release of albumin nanoparticles. Figure 2 shows the effect of different glutaraldehyde concentrations on particle diameter and PI of nescapine-loaded nanoparticles. The results reveal that the concentration of glutaraldehyde had little or no effect on particle size or polydispersity.

### Influence of pH and crosslinking on size and zeta potential of HSA nanoparticles

It was found that the pH of the nescapine-HSA solution prepared prior to ethanol addition was a crucial factor in



**Figure 1** Scanning electron micrograph of glutaraldehyde crosslinked nanoparticles prepared at pH 8, 60k resolution. The nanoparticles were found to be of uniform size and narrow size distribution.



**Figure 2** Effect of glutaraldehyde concentration on diameter and polydispersity index of noscapine (5 mg/mL) loaded human serum albumin (HSA) nanoparticles prepared with 10 mM NaCl solution and 100 mg of HSA protein at pH 8.

affecting the particle size of the resulting nanoparticles (Figure 3). At pH 5–7, particles were of larger diameter with high polydispersity indices. The particle size was greatest at pH 7 at approximately 2500 nm. At pH greater than 7 the particle diameter was reduced and at pH 8 the particles had optimal size and narrow size distribution at approximately 175–200 nm. The nanoparticles prepared at higher pH of 8–8.2, when viewed under SEM, revealed uniform and spherical particles.

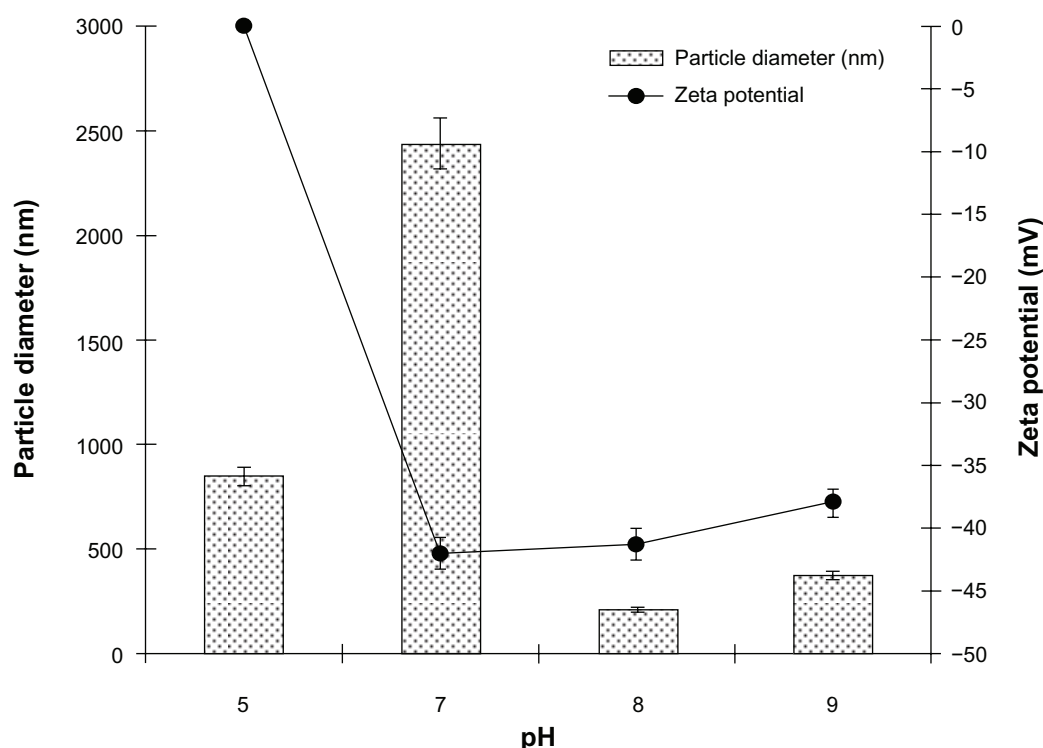
The electrical behavior of the noscapine-loaded HSA nanoparticles was also evaluated at different pH (Figure 3). Higher surface charge was attained on HSA nanoparticles by increasing the pH of the HSA solution. It was observed that with increasing pH > 7, the surface charge of prepared nanoparticles was reduced; particles prepared at pH 8 had their surface charge reduced to  $-47\text{mV}$ . It was noted that preparation of particles at higher pH also leads to repulsion among the HSA molecules due to steric effect and lead to aggregation during particle formation.

The stability of the nanoparticles was also investigated. Noscapine-loaded nanoparticles prepared at pH 8.2 were stored for 5 days in water at  $4^{\circ}\text{C}$  and at predefined times the samples were analyzed with regard to size (Figure 4). The particle size increased slightly to 190 nm, but particles were found to be stable after 5 days.

## Investigation of noscapine delivery by nanoparticles

Noscapine was adsorbed to HSA in solution prior to nanoparticle preparation. The pH was adjusted to 8.2 at different noscapine concentrations ranging from 5 mg/mL to 30 mg/mL of HSA. All preparations were stabilized with glutaraldehyde. Figure 5 illustrates the encapsulation efficiency of HSA nanoparticles at different drug concentrations. Maximum encapsulation was attained at noscapine concentrations of 5 mg/mL with an efficiency of 97%. Encapsulation efficiency was shown to gradually decrease with increasing drug concentration. The lowest encapsulation efficiency was seen at noscapine concentrations of 25 mg/mL and 30 mg/mL suggesting that at higher concentrations of noscapine more incubation time is probably required as well as greater concentration of HSA is needed for efficient encapsulation.

The drug release profiles of noscapine-loaded HSA nanoparticles were also investigated. Noscapine albumin nanoparticles prepared under experimental conditions described previously were tested *in vitro* and released at  $37^{\circ}\text{C}$  in PBS at pH 7.4. It was found that with increasing drug concentrations the particle size and polydispersity increased dramatically. Figure 6 shows the *in vitro* release of cumulative amounts of noscapine from albumin nanoparticles



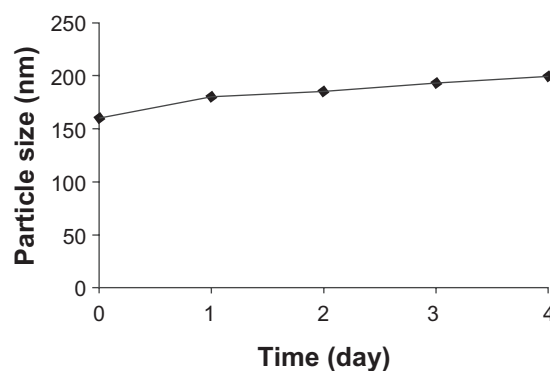
**Figure 3** Effect of pH on diameter and polydispersity index of noscapine (5 mg/mL) loaded human serum albumin (HSA) nanoparticles prepared with 10 mM NaCl solution and 100 mg of HSA protein.

as a function of time. For noscapine concentrations of 5 mg/mL, the initial drug released was around 10% which was considered a biphasic way of release; it is characterized by an initial rapid release period followed by a step of slower release. For noscapine-loaded nanoparticles with noscapine concentrations of 5 mg/mL and 10 mg/mL, the release profile was slow after 72 h, with less than 30% of the drug being released. While at concentrations of 15 mg/mL of noscapine more than 60% of the drug had been released after 72 h. The release profile seemed moderately slow.

### Viability of HSA nanoparticles loaded with noscapine in a SK-BR-3 cell line

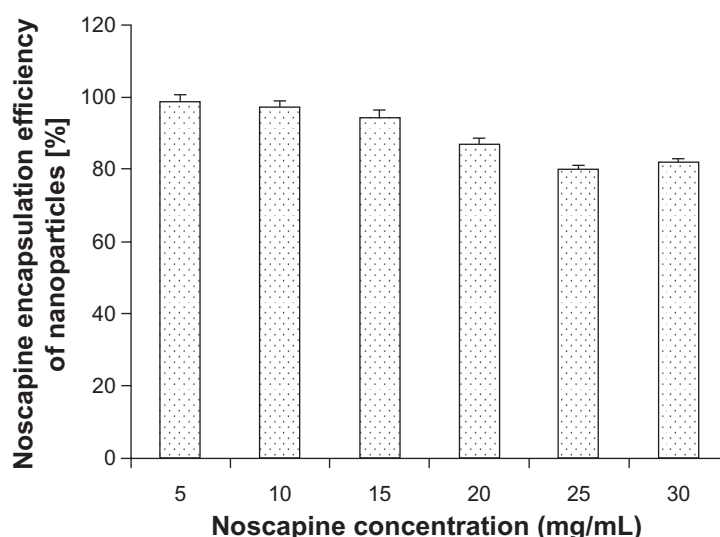
HER-2 positive SK-BR-3 breast cancer cells were used to examine the efficacy of noscapine prepared nanoparticles. The cell lines were cultured in 96-well plates at an initial concentration of 5000 cells/well in fresh medium. After 24 h of culture, cells were adherent, and medium was replaced with prepared noscapine nanoparticles and drug-free HSA nanoparticles. We analyzed the cell viability and cytotoxicity of noscapine-loaded nanoparticles prepared at noscapine concentrations of 5, 10, and 15 mg/mL, and compared them with drug-free nanoparticles. Both

drug-loaded and drug-free nanoparticles were added in SK-BR-3 breast cancer cell lines over time periods of 24, 48, 72, 96, and 144 h (Figure 7). As expected the efficacy of the particles depended on the concentration of drug-loaded. Noscapine nanoparticles with concentrations of 5 mg/mL had the lowest efficacy with cell viability of 60% after 144 hrs. The cell viability was less than 35% for noscapine nanoparticles prepared with noscapine concentrations of 10 and 15 mg/mL after 144 h. The drug-free nanoparticles had



**Figure 4** Stability of noscapine (5 mg/mL) loaded human serum albumin (HSA) nanoparticles over 5 days. Particle size was monitored. Noscapine-loaded HSA nanoparticles prepared with 10 mM NaCl solution and 100 mg of HSA protein at pH 8.





**Figure 5** Noscapine encapsulation of human serum albumin nanoparticles (50 mg/mL) in dependence on noscapine concentration.

little effect on the cell viability of the cells, and after 48 h the cell viability of cells with drug-free nanoparticles was approximately 93%. Gradually, the viability of cells treated with drug-free nanoparticles slightly decreased to about 80% after 144 h. Hence noscapine-loaded nanoparticles could potentially be used in the treatment of tumor cells.

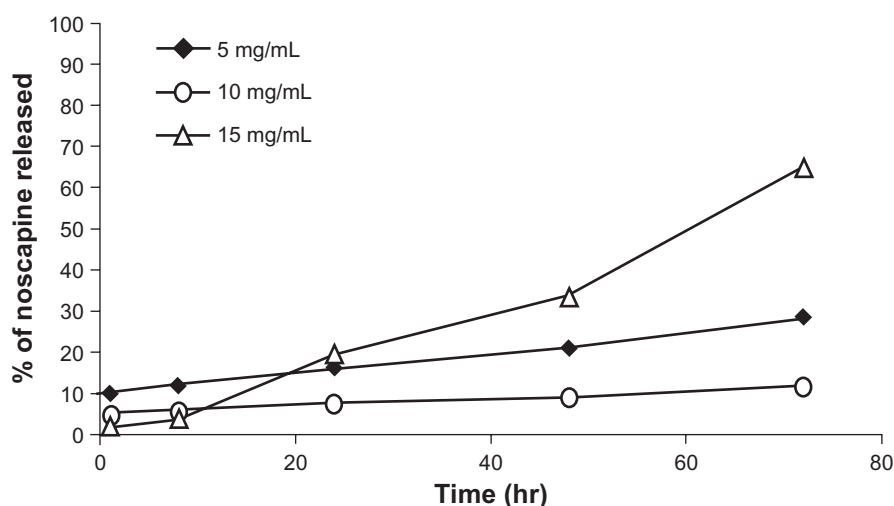
## Conclusion

HSA nanoparticles can be successfully used to encapsulate noscapine and be prepared by the coacervation method. Preparation pH appeared to have an influence on size and particle yield; however it did not induce any effect in the drug-loading capacity. In addition, the *in vitro* drug delivery studies indicated controlled slow release profiles. In conclusion noscapine-loaded HSA nanoparticles have the

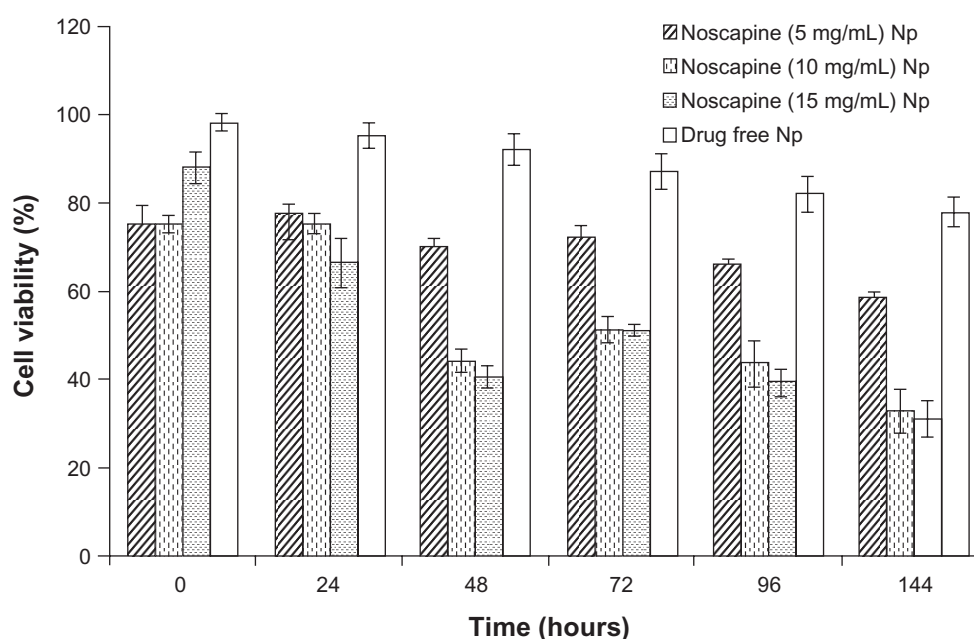
potential to be used to deliver a maximum amount of noscapine to target sites at a rate and concentration that permits optimal therapeutic efficacy, while reducing any undesirable side effects to a minimum. Further experiments using surface modification of noscapine nanoparticles are being conducted for tumor cell-specific targeting applications, and for further evaluation of cell cytotoxicity and efficacy in cancer cells.

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**Figure 6** Drug release profile of noscapine-loaded human serum albumin (HSA) nanoparticles (noscapine concentration 5–15 mg/mL) over predetermined time intervals. Noscapine-loaded HSA nanoparticles prepared with 10 mM NaCl solution and 100 mg of HSA protein at pH 8.



**Figure 7** Breast cancer efficiency of human serum albumin (HSA) nanoparticles-loaded with noscapine (5–15 mg/mL concentration) and controls (drug-free nanoparticles). Nanoparticle concentrations of 50 µg/mL were seeded in SK-BR-3 breast cancer cell line incubated at 24, 48, 72, 96, and 144 h.

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