

Determination of Antioxidant Activity by Oxygen Radical Absorbance Capacity (ORAC-FL), Cellular Antioxidant Activity (CAA), Electrochemical and Microbiological Analyses of Silver Nanoparticles Using the Aqueous Leaf Extract of *Solanum mammosum* L.

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Purpose: The importance of studying polyphenolic compounds as natural antioxidants has encouraged the search for new methods of analysis that are quick and simple. The synthesis of silver nanoparticles (AgNPs) using plant extracts has been presented as an alternative to determine the total polyphenolic content and its antioxidant activity.

Methods: In this study, aqueous leaf extract of *Solanum mammosum*, a species of plant endemic to South America, was used to produce AgNPs. The technique of oxygen radical absorption capacity using fluorescein (ORAC-FL) was used to measure antioxidant activity. The oxidation of the 2',7'-dichlorodihydrofluorescein diacetate (DCFH₂-DA) as fluorescent probe was used to measure cellular antioxidant activity (CAA). Electrochemical behavior was also examined using differential pulse voltammetry (DPV) and cyclic voltammetry (CV). Total polyphenolic content (TPH) was analyzed using the Folin-Ciocalteu method, and the major polyphenolic compound was analyzed by high performance liquid chromatography with diode array detector (HPLC/DAD). Finally, a microbial analysis was conducted using *Escherichia coli* and *Bacillus* sp.

Results: The average size of nanoparticles was 5.2 ± 2.3 nm measured by high-resolution transmission electron microscopy (HR-TEM). The antioxidant activity measured by ORAC-FL in the extract and nanoparticles were 3944 ± 112 and 637.5 ± 14.8 μ M ET/g of sample, respectively. Cellular antioxidant activity was 14.7 ± 0.2 for the aqueous extract and 12.5 ± 0.2 for the nanoparticles. The electrochemical index (EI) was 402 μ A/V for the extract and 324 μ A/V for the nanoparticles. Total polyphenolic content was 826.6 ± 20.9 and 139.7 ± 20.9 mg EGA/100 g of sample. Gallic acid was the main polyphenolic compound present in the leaf extract. Microbiological analysis revealed that although leaf extract was not toxic for *Escherichia coli* and *Bacillus* sp., minor toxic activity for AgNPs was detected for both strains.

Conclusion: It is concluded that the aqueous extract of the leaves of *S. mammosum* contains nontoxic antioxidant compounds capable of producing AgNPs. The methods using AgNPs can be used as a fast analytical tool to monitor the presence of water-soluble polyphenolic compounds from plant origin. Analysis and detection of new antioxidants from plant extracts may be potentially applicable in biomedicine.

Keywords: silver nanoparticles, *Solanum mammosum*, antioxidant activity, ORAC, voltammetry

Introduction

The therapeutic importance of antioxidants and the side effects reported from synthetic compounds has motivated the search for alternative sources.^{1–3} In this interesting analysis, plants and their biodiversity constitute an inexhaustible source of exploration.⁴ Silver nanoparticles (AgNPs) can be prepared with plant extracts through a chemical reduction process involving polyphenolic compounds,^{5,6} organic acids,⁷ and reducing sugars,⁸ among others.³ Plant extracts stabilize the size and shape of nanoparticles.^{9,10} Further, they provide them with additional properties depending on their nature.¹¹ In addition to their antimicrobial attributes,¹² AgNPs have optical properties that make them ideal tools for the detection of chemical reactions;¹³ they display the surface plasmon resonance (SPR) phenomenon, which consists of a collective oscillation of conduction electrons at the surface.^{14,15}

According to this principle, analyzing their formation is quite simple, so they can be used as an analytical method with species like polyphenolic compounds, especially considering how complex it is to study them in plant matrices.¹⁶ As antioxidants, polyphenolic compounds have the ability to remove free radicals¹⁷ and inhibit prooxidant enzymes.¹⁸ Oxidative stress is an imbalance between the generation and elimination of reactive species or free radicals, which both have a missing electron.¹⁹ These radicals can be caused by cellular metabolism, a clear example being reactive oxygen species (ROS) and reactive nitrogen species (RNS), among them the superoxide anion, hydroxyl radical, hydrogen peroxide, and nitric oxide.^{20,21} ROS and RNS cause severe cell damage associated with a wide list of diseases such as metabolic syndrome,²² cancer,²³ Alzheimer's disease,²⁴ and cardiovascular problems.²⁵ Antioxidant activity limits the generation of free radicals²⁶ and can be measured by chemical,²⁷ electrochemical,²⁸ or cellular methods.²⁹ Its effectiveness is evaluated with physiologically active antioxidants such as glutathione,³⁰ α -tocopherol,³¹ ergothioneine,³² and synthetics such as Trolox,³³ BHT,³⁴ and BHA.³⁵

Most studies that measure antioxidant activity using silver nanoparticles are based on methods such as DPPH (2,2-diphenyl-1-picryl-hydrazyl-hydrate), ABTS (2,2'-azino-bis [3-ethylbenzothiazoline-6-sulfonic acid]), and FRAP (ferric reducing ability of plasma), among others.^{36,37} The technique of oxygen radical absorption capacity using fluorescein (ORAC-FL)^{38,39} has been applied very infrequently to analyze the behavior of different nanoparticles, especially using plant extracts,^{28,40–43}

as have electrochemical methods such as differential pulse voltammetry (DPV) and cyclic voltammetry (CV).^{36,37} However, all these methods do not accurately reflect the antioxidant capacity at the cellular level, since physiological conditions such as pH, temperature, bioavailability and metabolism are not taken into account, so the use of in vitro models would allow a better approach for the determination of the antioxidant capacity.

Species of the genus *Solanum* are rich in polyphenolic compounds. *Solanum mammosum* L. (Sm) is a herbaceous species belonging to the Solanaceae family.^{38,39} In South America, it is commonly known as “teta de vaca” (“cow’s udder”) for the characteristic shape of its fruit.⁴⁰ The plant’s stem measures between 1 and 1.5 m in height, is fuzzy from the presence of glandular hairs, and has yellowish, smooth or curved thorns that are 10–15 mm wide at the base and approximately 22 mm long. Its leaves are oval-shaped with five to seven shallow lobes and irregular toothed margins; they are relatively thin and have an alternating arrangement and a truncated base. *S. mammosum* contains catechins, tannins, alkaloids, simple phenols, flavanones, flavonoids, cyanogenic heterosides, saponins, and triterpenes.^{39,40} It is known for its antioxidant,⁴¹ anticancer,^{42,43} antimalarial,⁴⁴ molluscicidal activity,^{45–48} and its toxic effects on pests^{49–51} and vectors.⁵² In vitro studies have also been conducted to structurally biotransform compounds with therapeutic properties from cell suspension cultures.^{53–58}

In the current study, AgNPs were used as sensors to determine the antioxidant activity of the aqueous extract of *S. mammosum* by ORAC-FL, cellular antioxidant activity by CAA, cyclic voltammetry (CV), and differential pulse voltammetry (DPV) as electrochemical analyses. The extract’s total polyphenolic content and phenolic profile were determined through high-performance liquid chromatography with diode array detector (HPLC/DAD). In addition, microbiological analysis was developed to test the toxicity of the plant extract used and the nanoparticles obtained.

Materials and Methods

Preparation of Aqueous Extract from *S. mammosum* Leaves

The protocol suggested by Pilaquinga et al⁵⁹ was used. Fresh *S. mammosum* leaves were collected in the Marianitas precinct (located at 0°05'46.3"N, 79°07'33.5"W; altitude: 135 m. a.s.l.). The collection was carried out with a Wildlife Management Patent N°06-2019-ICFAU-FLO DPAI/MAE from the Herbarium ECAA-PUCESI and authorization for

Scientific Research N° 006–2019-IC-FAU-FLO-DPAI/MAE from the Ministry of the Environment of Ecuador. A botanical voucher was previously deposited for reference at the Center for Research on Health in Latin America (CISEAL).⁵⁹ The samples were washed with distilled water to remove impurities and dried in the shade at room temperature (20°C) for one week. The sample was ground in a microfine grinder MF 10 (IKA, Königswinter, Germany). To prepare the aqueous extract, 1 g of sample was placed in 20 mL of distilled water and constantly stirred and heated on a heating plate (Glassco, Haryana, India) at room temperature for 1 h.

Microwave-Assisted Synthesis of AgNPs-Sm

To synthesize the AgNPs-Sm in colloid dispersion, the method suggested by Joseph and Mathew⁶⁰ was followed, with slight modifications. The parameters of the precursor concentration, amount of aqueous extract of *S. mammosum* leaves, pH, time, and power were set. For the optimized method, 20 mL of AgNO₃ 99.9% purity (Merck 99.9%, Darmstadt, Germany), 1 mmol L⁻¹ and 1 mL of the aqueous extract were used. The pH was adjusted to 8 with NaOH 1% (Merck 99.9%, Darmstadt, Germany), time 90 s, and power 800 W using a microwave MWI 28BL (Indurama, Cuenca, Ecuador). To obtain solid nanoparticles was necessary 2 mL of the leaf extract of *S. mammosum*, 20 mL of AgNO₃ 50 mM, pH 9, 40°C and stirring agitation during 30 minutes on a heating plate (Velp Scientifica, Usmate Velate, Italy). The solid residue was centrifuged for 10 minutes to 6000 rpm and decanted. Finally, the decanting was washed with distilled water and dried under infrared light at 30 cm of distance during 3 hours.

Characterization of AgNPs-Sm

To monitor the formation of the AgNPs-Sm, a Cary 60 double-beam UV-Visible spectrophotometer (Agilent Technologies, Santa Clara, CA, USA) was used. To determine the size of nanoparticles, a Tecnai G² F20 high-resolution transmission electron microscope (HR-TEM) (FEI Tecnai, Hillsboro, OR, USA) was used. The nanoparticles' hydrodynamic radius was measured with a dynamic light scattering meter (DLS) using an LB-550 nanoparticle size analyzer (HORIBA, Kyoto, Japan). X-ray diffraction (XRD) patterns were collected with a diffractometer (PANalytical EMPYREAN, Almelo, The Netherlands) with a Bragg-Brentano (Θ -2 Θ) configuration

set to 40 kV, 45 mA, and monochrome X-rays with a Cu K- α wavelength ($\lambda = 1.541 \text{ \AA}$); a Ni filter was used. The FTIR-ATR spectra were obtained with a spectrophotometer (Perkin Elmer, Spectrum BX FTIR, Waltham, MA, USA) with ATR coupling (resolution from 4 cm⁻¹ to 4000–400 cm⁻¹ and 10 scans per sample with intervals of 2 cm⁻¹).

Total Antioxidant Activity by Oxygen Radical Absorbance Capacity (ORAC-FL)

Antioxidant capacity was determined using the ORAC-fluorescence methodology (ORAC-FL).^{61–65} The analyses were performed using a 96-well white polystyrene plate reader (EnSpire multimode PerkinElmer, Waltham, MA, USA). The reaction was carried out in phosphate buffer of 75 mM (pH 7.4). The sample was homogenized and incubated at 37 °C for 5 min with 150 μ L of fluorescein solution (40 nM, final concentration). Then, 25 μ L of 2,2'-azobis(2-methylpropionamidine) dihydrochloride (Sigma-Aldrich 97%, St Louis, MO, USA) 18 mmol L⁻¹ was added; the fluorescence was registered with an excitation wavelength of 485/20 nm and an emission filter of 528/20 nm at a constant temperature of 37 °C in a Synergy HT multi-detection microplate reader (Bio-Tek Instruments, Winooski, VT, USA) every minute for 2 h. The results were expressed as μ mol L⁻¹ Trolox equivalent/g of sample. A Trolox calibration curve was used with a concentration range between 3 and 20 μ mol L⁻¹.

Cellular Antioxidant Activity (CAA)

Cellular antioxidant activity (CAA) was evaluated in VERO cell line culture (European Collection of Cell Cultures, ECACC 84113001) using 2',7'-dichlorodihydrofluorescein diacetate (DCFH₂-DA) as a fluorescent probe.⁶² The cells were plated in sterile, white, polystyrene, flat-bottom 96-well microplates (Nunc, Denmark) at a concentration of 50,000 cells per well and incubated for 24 h at 37 °C and 5% CO₂ in RPMI 1640 culture medium. The cells were washed with 150 mL of phosphate buffered saline (PBS) pH 7.4 and incubated for 1 h with 100 μ L of RPMI 1640 containing 20 μ M. The aqueous extract and AgNPs-Sm were added in a final concentration of 21 mg mL⁻¹. After 2 h of incubation, the medium was discarded, and the cells were gently washed twice with 100 μ L of PBS. Then, they were incubated with 2,2'-Azobis(2-methylpropionamidine) dihydrochloride (AAPH) (Sigma-Aldrich 97%, St Louis, MO, USA) at a final concentration of 600 μ M in PBS. Fluorescence was

measured immediately after addition in a spectrofluorometer (Bio-Tek Instruments, Winooski, VT, USA) in 96-well plates at 37 °C using an excitation of 485 nm and an emission of 538 nm. The evaluation was carried out every min for 1 hr, and the CAA values were calculated with Equation (1):

$$\%CAA = 100 - \left(\frac{\Delta F_{AH}}{\Delta F} * 100 \right) \quad (1)$$

where F is the fluorescence intensity in the presence of free radicals without the extract, and F_{AH} is the fluorescence intensity in the presence of free radicals, extract, and AgNPs-Sm. All measurements were done in the same period.

Antioxidant Activity by Differential Pulse Voltammetry (DPV) and Cyclic Voltammetry (CV)

For the analysis, the protocol suggested by Pilaquinga et al was followed.⁶⁶ A CH-Instruments potentiostat, model 700D (Tennison Hill Drive Austin, TX, USA) coupled to a conventional 15 mL three-electrode reaction cell was used. Glassy carbon (GC) was used as a working electrode, Ag/AgCl as a reference electrode, and a platinum wire (0.5 cm in diameter and 3 cm long) as a counter electrode. The glassy carbon electrode was polished with alumina paste (Al_2O_3 , 0.25 μm , Struers, Copenhagen, Denmark) to obtain a mirror-like surface (0.066 cm^2), followed by sonication for 5 min. Differential pulse voltammetry and cyclic voltammetry were performed in an electrolyte solution of sodium acetate 0.1 mol L^{-1} , with a range of -1.2 V to 1.3 V.

Total Polyphenolic Content (TPH)

Total polyphenolic content (TPH) was determined using the Folin-Ciocalteu spectrophotometric method.⁶⁷ Two hundred μL of Folin-Ciocalteu reagent (Merck 99.99%, Darmstadt, Germany) was added to 15 μL of extract/AgNPs; the mixture was homogenized and left to incubate for 5 min. Then, 40 μL of Na_2CO_3 20% (m/v) and 45 μL of distilled water were added; the reaction mixture was homogenized and incubated for 30 min at 37 °C. Afterward, absorbance at 760 nm was recorded with a Synergy HT microplate reader (Bio-Tek Instruments, Winooski, VT, USA).

Phenolic Profile Determination

The polyphenolic profile of the *S. mammosum* leaf extract was determined by high-performance liquid

chromatography (HPLC) Series 1260 Infinity II (Agilent Technologies, Santa Clara, CA, USA) coupled to a diode array detector (DAD) (Santa Clara, CA, USA). A Chromolith C18 endcapped column (150 $\times$ 4.60 mm, 5.0 μm) (Merck 99.99%, Darmstadt, Germany) was used for identification. The sample injection volume was 20 μL , and the column temperature was set to ambient conditions. The mobile phase composition corresponded to phase A-acetic acid 2% in water and phase B-acetonitrile, with a gradient elution of 92% A decreasing 5% every 5 min at a constant flow rate of 500 $\mu L min^{-1}$. OpenLab ChemStation analysis software, version A 01.03, was used for system control and data acquisition and processing.

Microbiological Analysis

Two isolates, *Escherichia coli* CECT 101 and *Bacillus* sp. CECT 40, both obtained from the Spanish Type Culture Collection (CECT, Valencia, Spain), were used. Minimum inhibitory concentration (MIC) analysis and agar dilution susceptibility tests were conducted as per the standard CLSI methods.⁶⁸ For the agar dilution tests, three distinct quantities (100, 10, and 1 micrograms) of each analyzed extract, nanoparticle, and compound were placed on different 6 mm Whatman® Antibiotic Assay Discs (Merck Life Science, Madrid, Spain). For the MIC analysis, a starting solution of 512 mg/L for each analyzed extract, nanoparticle, and compound was prepared.

Statistical Analysis

For the conditioning of microwave-assisted AgNPs-Sm synthesis, a 3K factorial design was utilized. Analyses of antioxidant activity with ORAC-FL, CV, DPV, and total polyphenolic content were performed in triplicate. To analyze the results, a Student's *t*-test with a 95% confidence interval was used. Data were expressed as mean $\pm$ standard deviation (SD). MIC experiments were performed in triplicates. Origin Pro 8.5 SR2 (Origin Lab Corporation, Washington, USA) was used for data analysis.

Results and Discussion

Characterization of AgNPs-Sm

The formation of AgNPs-Sm monitored by UV-Vis spectrophotometry is shown in Figure 1. The spectra absorption obtained with the optimized synthesis parameters such as potency, time, pH, precursor and *S. mammosum* extract volume are indicated from 1A to 1E respectively. According to the nanoparticle size estimate proposed by

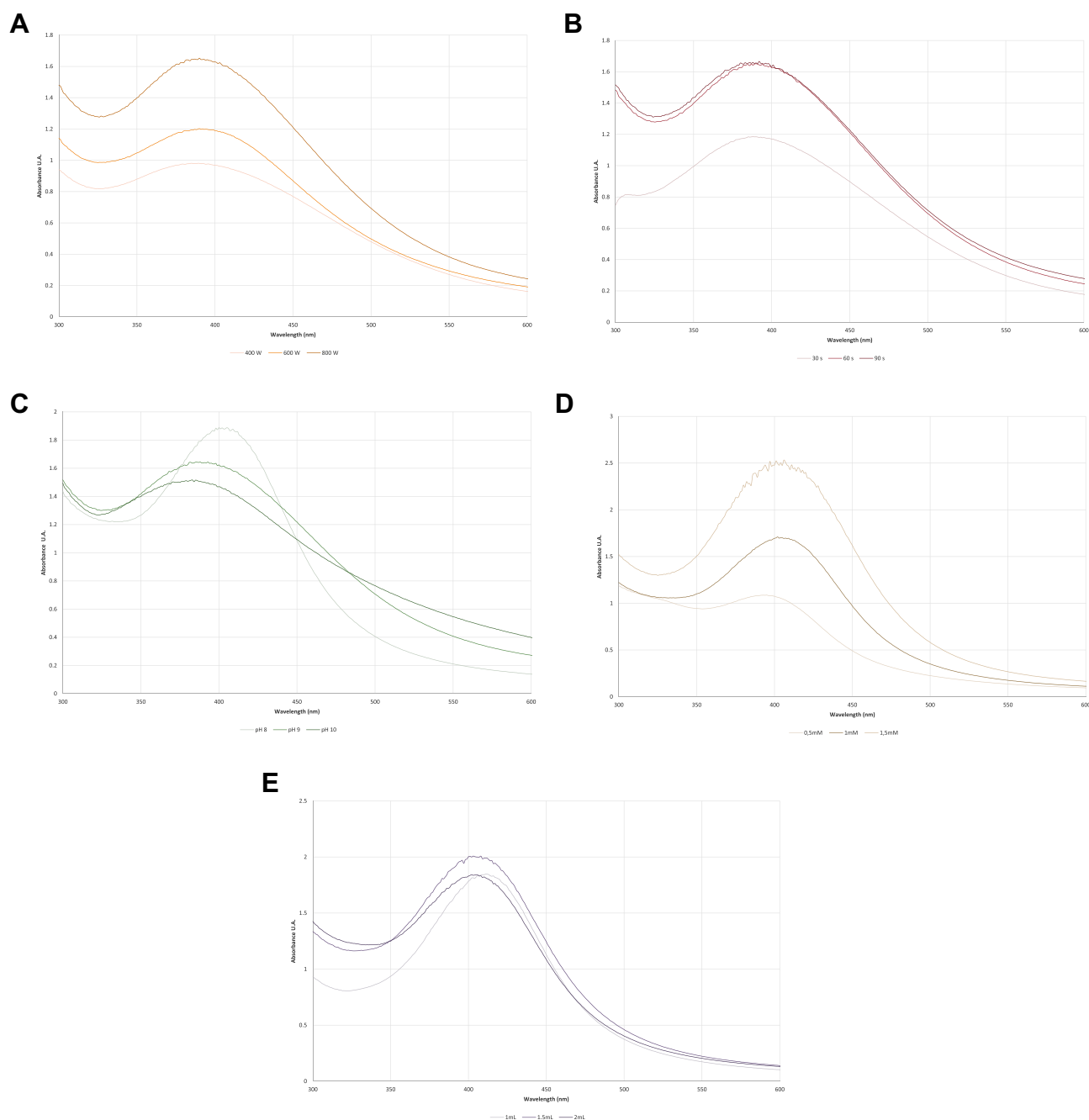


Figure 1 UV-Vis spectra of the AgNPs optimization. (A) potency, (B) time, (C) pH, (D) precursor and (E) *S. mammosum* extract volume.

Pradeep,⁶⁹ the surface plasmon suggests an approximate value of 30 to 50 nm with the organic coating. The actual size measured by HR-TEM was 5.2 ± 2.3 nm as shown in Figure 2A and the particle size histogram in Figure 2B. The morphology of the particles is quasi-spherical and polydispersed, as clearly observed. The d-spacing between the planes is calculated to be 0.25 nm, which closely corresponds to the spacing between the (111) Ag (0.236 nm) of the XRD pattern. Furthermore, some particles

displayed multiple-twinned crystalline planes, indicating that they consist of several differentially oriented crystals, see Figure 2C. The selected area electron diffraction (SAED) pattern of the as-prepared silver nanoparticles is shown in Figure 2D and confirms the nanoparticles' crystalline nature and that the observed structure is silver.^{3,66} The size measured by the DLS was $22.1 \text{ nm} \pm 2.3 \text{ nm}$, which indicates that the organic coating's size is approximately 16.9 nm,⁷⁰ which is consistent with the

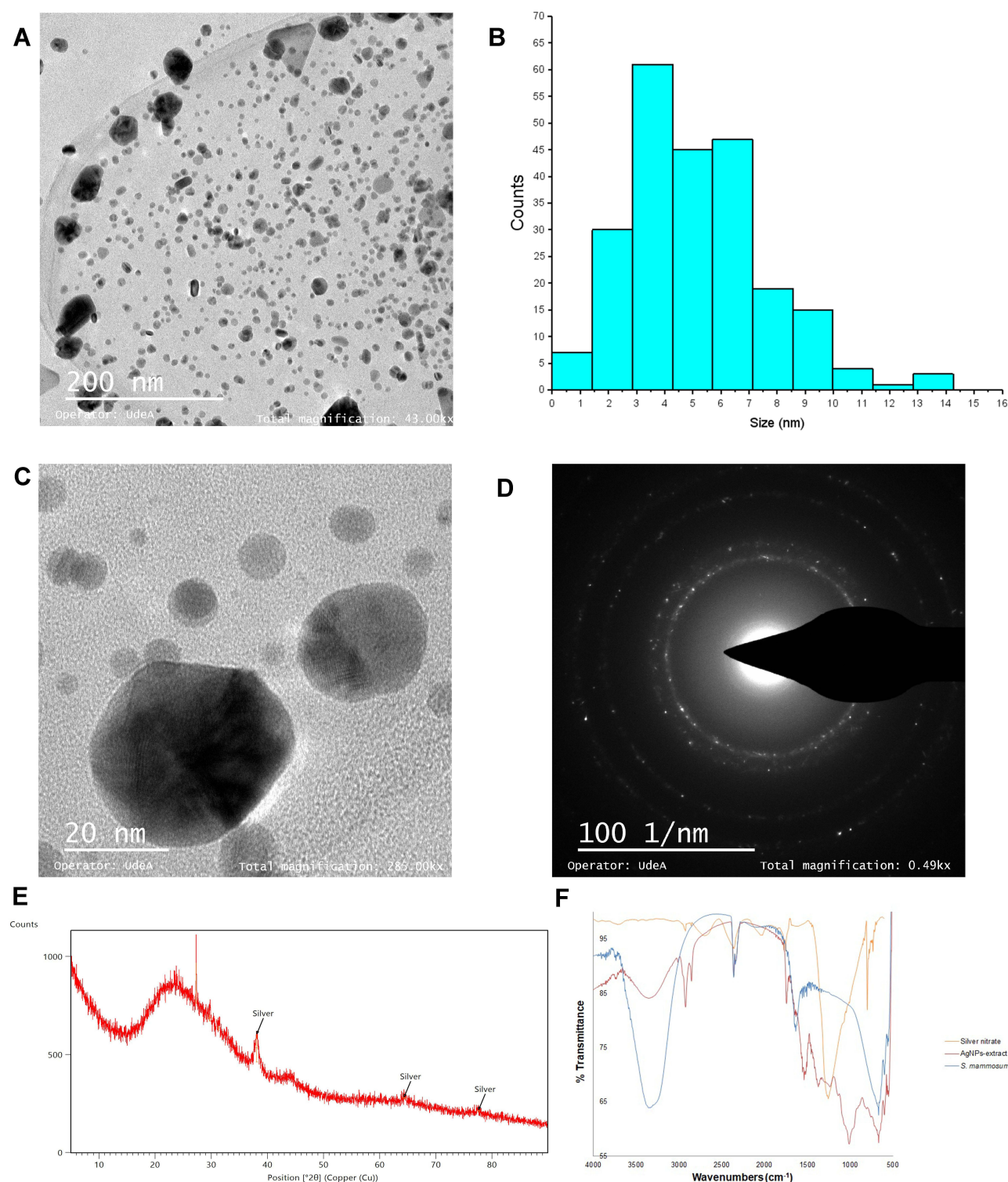


Figure 2 Characterization of AgNPs-Sm. **(A)** HR-TEM micrograph of AgNPs-Sm, **(B)** particle size histogram, **(C)** picture of a nanoparticle done with HR-TEM, **(D)** SAED of nanoparticle observed, **(E)** X-ray diffraction (XRD) pattern of AgNPs-Sm and **(F)** IR spectra of AgNO₃, AgNPs-Sm and *S. mammosum* extract.

nanoparticle size estimated by UV-Vis spectrophotometry. When comparing the size of the AgNPs-Sm obtained from the aqueous extract of *S. mammosum* fruit (15.3±4.8 nm)⁵⁹ to those from other *Solanum* species such as *S. melongena*

L. (24 nm)⁷¹ and *S. muricatum* (20–80 nm)⁷² that used microwave-assisted synthesis, the observed size is significantly smaller. The analysis of the diffraction pattern (XRD) in Figure 2E shows the observed peaks at angles

of 20 for the silver nanoparticles are as follows: 32.8 °, 38.0 °, 58.5 °, and 65.4 ° and correspond to the (111), (002), (122), and (220) reflection planes of a silver oxide cubic lattice referenced by ICDD 98-028-1041. The space group is Pn-3m and corresponds to space group number 224. The most intense peak corresponding to the predominant orientation of Ag nanocrystals is along plane (111), and the Debye-Scherrer equation predicts that the average size of the AgNPs-Sm is around 11 nm, which is consistent with the value obtained by HR-TEM. There is no evidence of other components, such as silver chloride, as possible competitors for the chemical reaction. This indicates again that the reaction is dominated by the synthesis of AgNPs nanocrystals. The FTIR measurement was conducted to identify the possible biomolecules of the *S. mammosum* extract responsible for the reduction and stabilization of AgNPs-Sm. The IR spectra comparison between AgNO₃, lyophilized AgNPs-Sm, and *S. mammosum* dry leaves is shown in Figure 2F. In the AgNPs-Sm IR spectrum, the band decreases to 3600 cm⁻¹, which corresponds to the O-H stretching of alcohols or phenols in relation to the extract, since they could act as reducing agents to synthesize AgNPs.^{47,72} At 1744 cm⁻¹, band of the quinone's (C=O) functional group could be observed in the spectrum of the plant extract, but it is absent in the AgNPs-Sm spectrum.⁷³

Antioxidant Activity by ORAC-FL

The ORAC-FL kinetic profile at different concentrations is shown in Figure 3A. The area under the fluorescence decay curve (AUC) was calculated by integrating the decrease in fluorescence. The Trolox calibration curve (3 μM a 20 μmol L⁻¹) is shown in Figure 3B. For the aqueous extract of *S. mammosum* leaves, antioxidant activity was determined to be 3944 ± 112 μM TE/g of sample. The antioxidant capacity through this methodology is directly related to the reaction's stoichiometry and the low reactivity of the probe used, not to the reactivity of the phenolic compounds present in the extract.^{74,75} The antioxidant activity of the AgNPs-Sm was 637.5 ± 14.8 μM TE/g of sample. This decrease is because the antioxidant compounds present in the extract were oxidized in the preparation of the AgNPs and, therefore, no longer had labile hydrogens to react with the peroxy radical. It should be noted that although the ORAC-FL methodology does not determine the reactivity of antioxidants, the formation of AgNPs could be an indirect measurement of the ability of this type of compound to react and is related to the content

of the reducing compounds determined through the Folin-Ciocalteu methodology. The decrease in antioxidant capacity is consistent with the results of Okiei et al⁷⁶ who measured ascorbic acid as it is related to the amount of vitamin C (antioxidant) in *Solanum aethiopicum*. Comparing the antioxidant activity of AgNPs using the aqueous extract of *Baccharis latifolia*⁸⁵ leaves by ORAC-FL, the activity decreased in a similar proportion from the extract to the nanoparticle, which corresponded to 87% versus 84% for AgNPs-Sm. Regarding the approximate values of antioxidant activity of AgNPs using crop plants, they fall within the range of 3000 to 8000 mg TE/mg mL using the same technique.⁷⁷

Cellular Antioxidant Activity (CAA)

The CAA kinetic profile of the aqueous extract and AgNPs-Sm is shown in Figure 3C. The 2',7'-dichlorodihydrofluorescein diacetate (DCFH₂-DA) probe was passively diffused through the cellular membrane and transformed into DCFH₂, a nonfluorescent compound that emits fluorescence when oxidized to 2,7-dichlorofluorescein (DCF) by intracellular ROS due to intracellular esterase. The percentage of CAA was determined to be 14.7 ± 0.2 for the aqueous extract and 12.5 ± 0.2 for the AgNPs-Sm. Both samples were at the same concentrations. The antioxidant activity was lower for AgNPs-Sm, which is consistent with that for ORAC-FL. Moncada-Basualto et al⁶² determined that cellular antioxidant activity has a relationship with lipophilicity (ie, major lipophilicity activity is associated with minor antioxidant activity), possibly due to a better interaction with the cell membrane, which could indicate the low percentages of antioxidant activity of the extracts and AgNPs-Sm, as well as the limited entry of the compounds into the cells, due to the polarity of the aqueous extracts. The main polyphenolic compounds in *Solanum* are quercetin, gallic acid, and rutin, among others.^{78,79} Wolfe et al⁸⁰ studied the cellular antioxidant activity of different polyphenolic compounds and determined that quercetin has high CAA compared to gallic acid and rutin. Our data obtained from the CAA for the aqueous extract and the AgNPs-Sm corroborate this finding.

Antioxidant Activity by Differential Pulse Voltammetry (DPV) and Cyclic Voltammetry (CV)

DPV and CV were used to evaluate the electrochemical behavior of the aqueous extract of *S. mammosum* leaves.

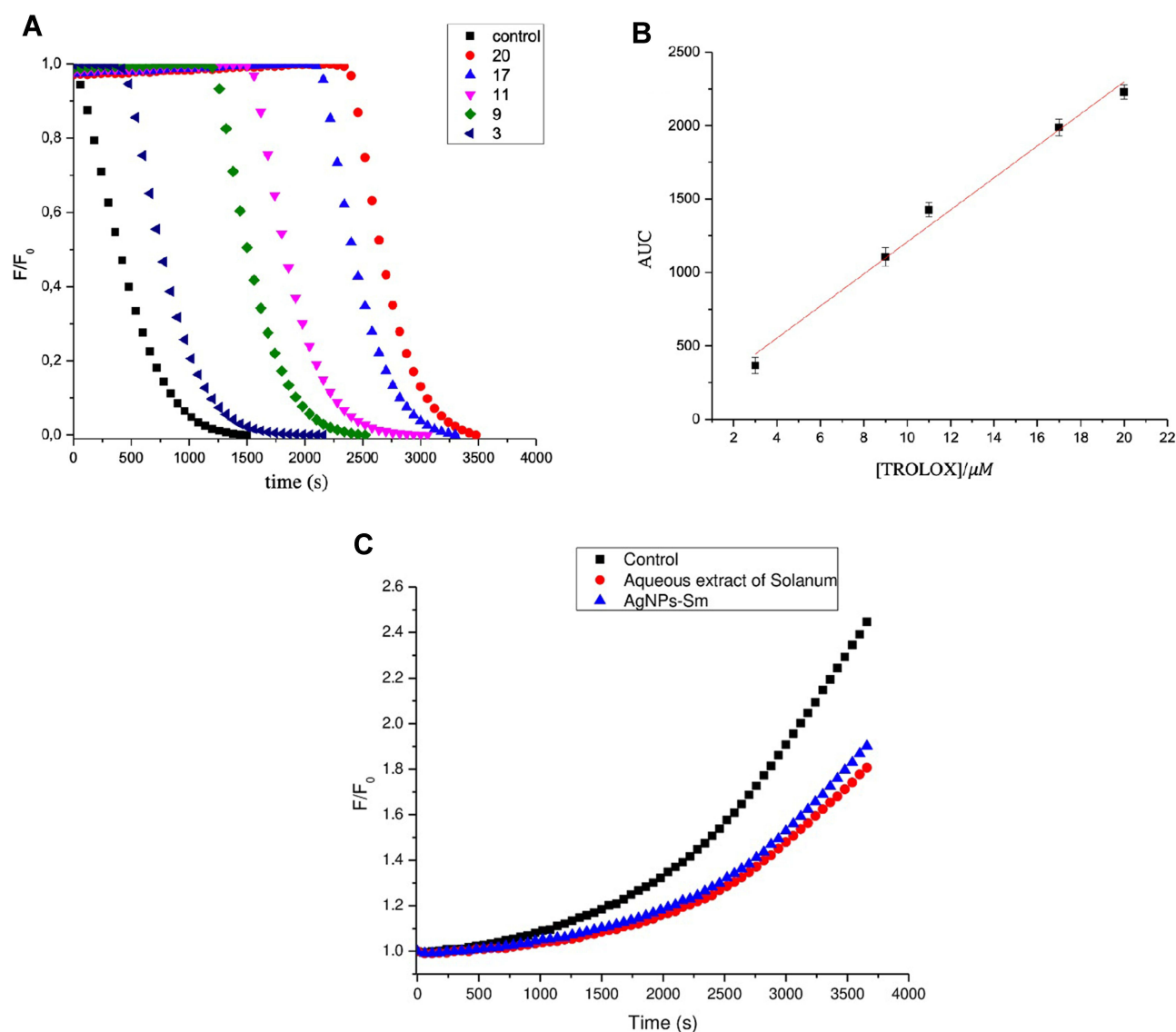


Figure 3 Oxygen radical absorption capacity using fluorescein (ORAC-FL) and cellular antioxidant activity (CAA) analysis. **(A)** Kinetic profile of ORAC-FL in different concentrations, **(B)** Trolox calibration curve and **(C)** kinetic profile of CAA of the aqueous extract and AgNPs-Sm.

In Figure 4, the DPV voltammograms, obtained at different initial scanning potentials using sodium acetate 0.1 mol L^{-1} , are compared (Figure 4A). For the *S. mammosum* leaf extract, when the voltammogram starts at $+0.5 \text{ V}$ (Figure 4B), two oxidation waves are observed, one at $+0.87 \text{ V}$ and the other at $+1.37 \text{ V}$; when the sweep starts at $+1.2 \text{ V}$, two additional waves appear (Figure 4C) that have a lower current intensity and lower potentials than the aforementioned waves. These results suggest the presence of four types of species with antioxidant capacity in the extract. The voltammogram of the AgNPs-Sm extract started at -0.5 V also shows the four voltamperometric waves described for the extract without

nanoparticles but with lower current values. Waves at lower oxidation potentials are associated with compounds with a greater capacity to donate electrons and vice versa, and those with higher currents indicate the concentration of antioxidants dissolved in the solution or the number of electrons exchanged per antioxidant. The electrochemical index (EI) was obtained by measuring these parameters using the voltammograms.⁸⁷ The calculated EI was $402 \mu\text{A/V}$ for the extract without nanoparticles and $324 \mu\text{A/V}$ for the AgNPs-Sm, which are values consistent with those obtained for these extracts through the ORAC-FL method; further, the extract with Ag nanoparticles showed lower oxidizing activity and lower content of oxidizing species.

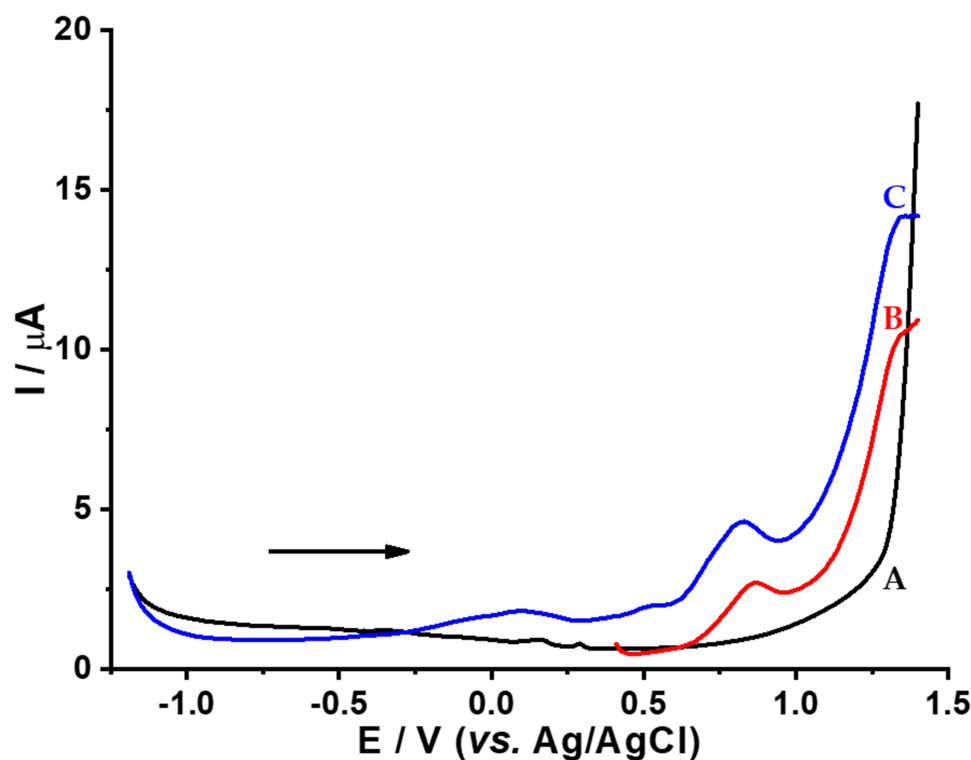


Figure 4 Differential pulse voltammetry analysis of *Solanum mammosum* extract at GC electrode in 0.10 mol L^{-1} . (A) Sodium acetate 0.1 mol L^{-1} , (B) initial potential scan $+0.5 \text{ V}$ and (C) initial potential scan -1.2 V . Scanning rate 50 mV s^{-1} vs Ag/AgCl at 25°C .

To calculate EI, the thermodynamic parameter E_{pa} expresses reducing power, while the kinetic and quantitative parameter I_{pa} expresses the speed of electron transfer and/or the amount of antioxidant content in the sample. When the EI calculation resulting from the extracts with and without nanoparticles was compared, the E_{pa} remained constant for both extracts, while the I_{pa} was lower for the extract with nanoparticles. According to these results and considering that “antioxidant activity” refers to the reaction speed between antioxidants and the oxidizable species and “antioxidant capacity” refers to the amount (in moles) of radicals recovered per one mole of antioxidant (that is, the stoichiometric factor), we can conclude that the *S. mammosum* extract has greater antioxidant activity than the extract that contains the nanoparticles; however, both possess antioxidant capacity and therefore the same type of antioxidant species. As the EI provides an effective measure of the antioxidant power (capacity and/or activity) of a given chemical species and is usually correlated with phenolic compounds, the corresponding analysis is presented in the following sections.

Figure 5 shows the CVs of the *S. mammosum* extract in relation to the blank sodium acetate 0.1 mol L^{-1} solution (Figure 5A) starting at potentials of $+0.0 \text{ V}$ (Figure 5C)

and sufficiently negative potentials, -1.5 V (Figure 5B), to promote electrochemical reduction of dissolved oxygen to radical species such as $\text{HO}_2^\bullet$ and $\text{HO}^\bullet$.^{81,82} The reducing direction of the voltammogram in Figure 5C shows a reduction wave to -0.7 V , which is also present in the voltammogram of the electrolyte medium with no extract, Figure 4A; this reduction wave is associated with the reduction of oxygen. In the oxidation direction, two waves are seen, the first between $+0.2 \text{ V}$ and 0.35 V and the second to 0.6 V , which, as in the voltammogram obtained by DVP (Figure 5B), are absent in the voltammogram started at $+0.0 \text{ V}$ (Figure 5C). This result can be interpreted in the following way: in the voltammogram started at $+1.5 \text{ V}$, the electrochemical reduction of the O_2 dissolved in the solution generates reactive radicals that react with the antioxidant species present in the extract, where the product of this reaction can generate other electroactive species that are responsible for the signal obtained in the oxidation sweep between the $+0.2 \text{ V}$ and 0.35 V potentials. On the other hand, in both the voltammogram initiated at $+0.0 \text{ V}$ and the one initiated at -1.5 V , the appearance of cathodic signals at the $+0.5 \text{ V}$ potential (Figures 5C), apparently coupled with the anodic processes, in the sweep in the oxidation direction, suggests

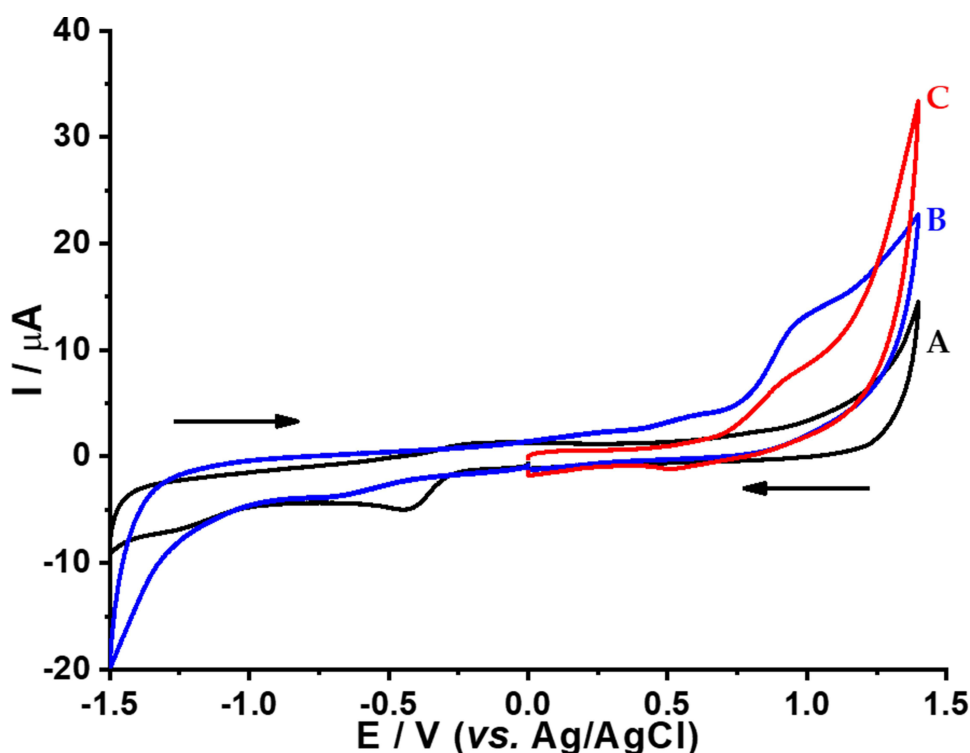


Figure 5 Cyclic voltammograms of *Solanum mammosum* extract in 0.10 mol L⁻¹ sodium acetate solution in 0.10 mol L⁻¹ at GC electrode. (A) blank sodium acetate 0.1 mol L⁻¹, (B) initial potential scan +0.5 V and (C) initial potential scan -1.2 V. Scanning rate 50 mV s⁻¹ vs Ag/AgCl at 25 °C.

that the redox processes involved have a certain degree of electrochemical reversibility. These results suggest that in the aqueous extract of *S. mammosum*, additional compounds are generated when electrochemical conditions are present that promote ROS generation, which again corroborates the presence of compounds that are highly reactive with ROS, that is, chemical components with high antioxidant capacity, in this extract.

Several studies have been conducted that use CV to detect antioxidant capacity in species belonging to the Solanaceae family. For example, Domenech-Carbó et al carried out an electrochemical characterization of the fruit of *S. lycopersicum* L. applying CV⁸³ and performed in situ monitoring of the reactivity of this species' phenolic compounds with ROS.⁸⁴

Total Polyphenolic Content (TPH)

The total polyphenolic content (TPH) of the aqueous extract of *S. mammosum* and AgNPs-Sm was determined, finding a respective content shown in Table 1. The difference in the value from de-extract to AgNPs-Sm is due to the decrease in the reducing capacity of the phenolic compounds by the formation of AgNPs, which is lower in their concentration by oxidation and in their antioxidant capacity by the electron

transfer mechanism. These results are consistent with those reported by Oszmianski et al⁸⁵ who characterized the phenolic compounds and antioxidant capacity of the fruits of *Solanum scabrum* and *Solanum burbankii*. When comparing the results obtained with AgNPs using different extracts, such as *S. intermedia* for example, the decrease in the total polyphenolic content of the extract to the nanoparticle is considerable, going from 2.398 ± 0.0028 to 0.507 ± 0.0031 (mg EGA/g of dried plant).

Polyphenolic Profile

The phenolic compounds' retention times, which have been previously described for the genus *Solanum*⁷⁸ (Table 2), were determined using the aforementioned elution program as part of a multicomponent analysis. The chromatogram corresponding to the *Solanum mammosum* extract showed two high-intensity peaks at 2.23 and 17.19 min. However, these peaks could not be attributed to any of the standards used. Only one of the observed peaks was attributable to gallic acid, as shown in Figure 6. The phenolic acid has also been identified in extracts of *Solanum lycopersicum* L., as well as rutin and quercetin.⁸⁶ The confirmation of the presence of only one of the phenolic compounds used as a standard may be due to the type of extraction performed;

Table 1 Total Polyphenolic Content in *Solanum Mammosum* and AgNPs-Sm

Sample	Total Polyphenolic Content (mg EAG/ 100 g Dry Mass)
<i>Solanum mammosum</i>	826.6 ± 20.9
AgNPs-Sm	139.7 ± 20.9

Table 2 HPLC Retention Times of Polyphenol Standards

Compound	Retention Time (min)
Gallic acid	2.90
Catechin	5.47
Quercetin	21.29
Resveratrol	26.03
Kaempferol	28.80

the extract was aqueous, and gallic acid is highly soluble in water, unlike polyphenolic compounds, whose water solubility is low. Chun Wang et al.⁷⁸ identified the presence of different polyphenols – gallic acid, protocatechuic acid, gallo catechin, caffeic acid, gallo catechin gallate, rutin,

quercetin, and naringenin in *Solanum nigrum* extracts using a different extraction method.

Microbiological Analysis

A nontoxic effect for the *Solanum mammosum* extracts, independent of the concentration and the analysis test used, was observed for both strains. An agar dilution test revealed a clear inhibitory effect of AgNO₃ (up to 1 microgram of AgNO₃ per disc) (Figure 7A) and a very weak growth inhibition of AgNPs-Sm (only with 100 micrograms of AgNPs-Sm) (Figure 7B) for *Bacillus* sp., likely due to the soluble Ag⁺ present in AgNPs-Sm, as previously described.⁸⁷ Unexpectedly, no inhibitory effect was observed with the agar dilution test (AgNO₃ and AgNPs-Sm) for *Escherichia coli*. Nevertheless, MIC analysis revealed a clear toxic effect for AgNO₃ and AgNPs-Sm, though for both strains, the inhibitory effect of AgNPs-Sm was weaker than that for AgNO₃. Thus, *Bacillus* sp. showed a strong inhibitory effect for AgNO₃ at 64 mg/L and a similar response for AgNPs-Sm at 128 mg/L (Figure 7C). For *Escherichia coli* (Figure 7D), the strongest inhibitory effect

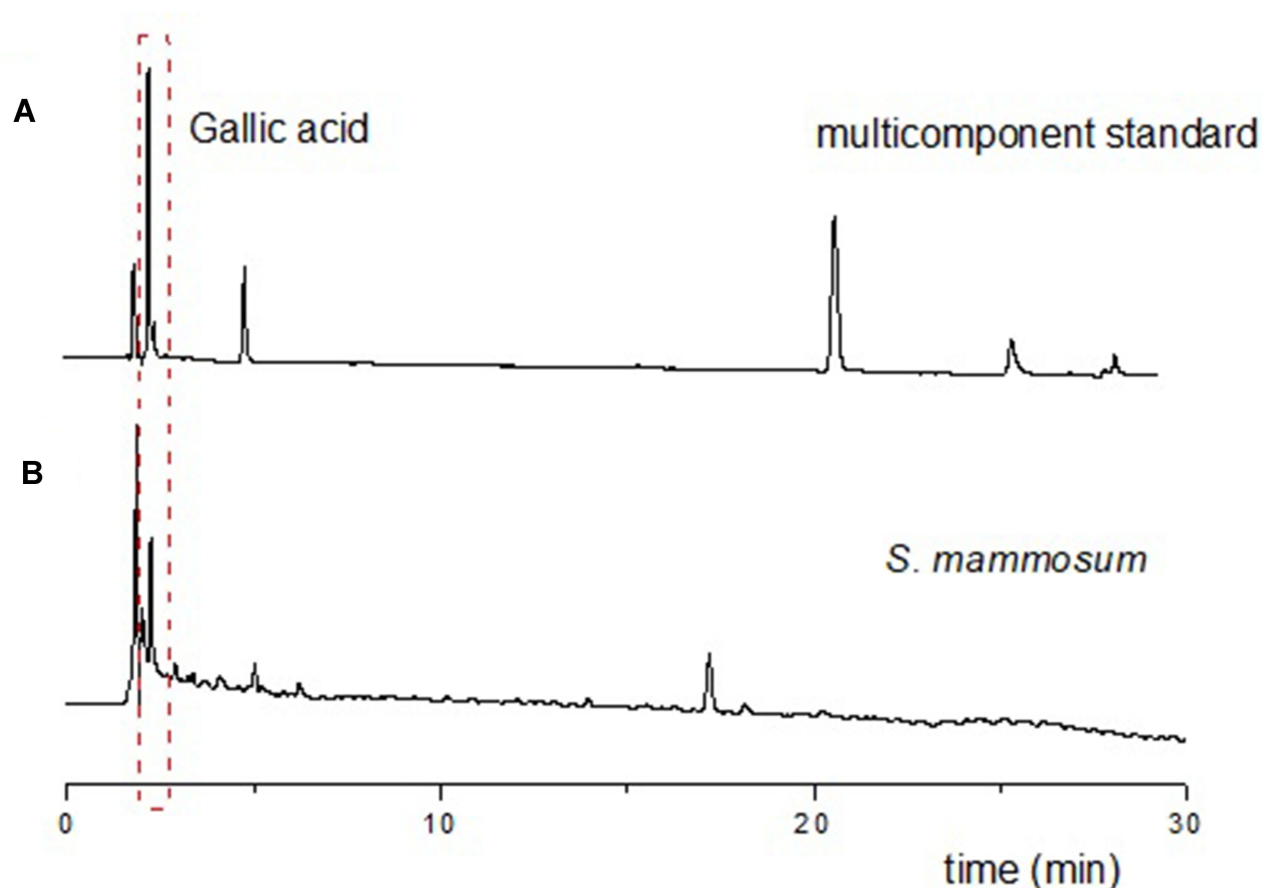


Figure 6 Antioxidant analysis. HPLC-MS chromatogram with (A) multicomponent standards and (B) *S. mammosum* extract.

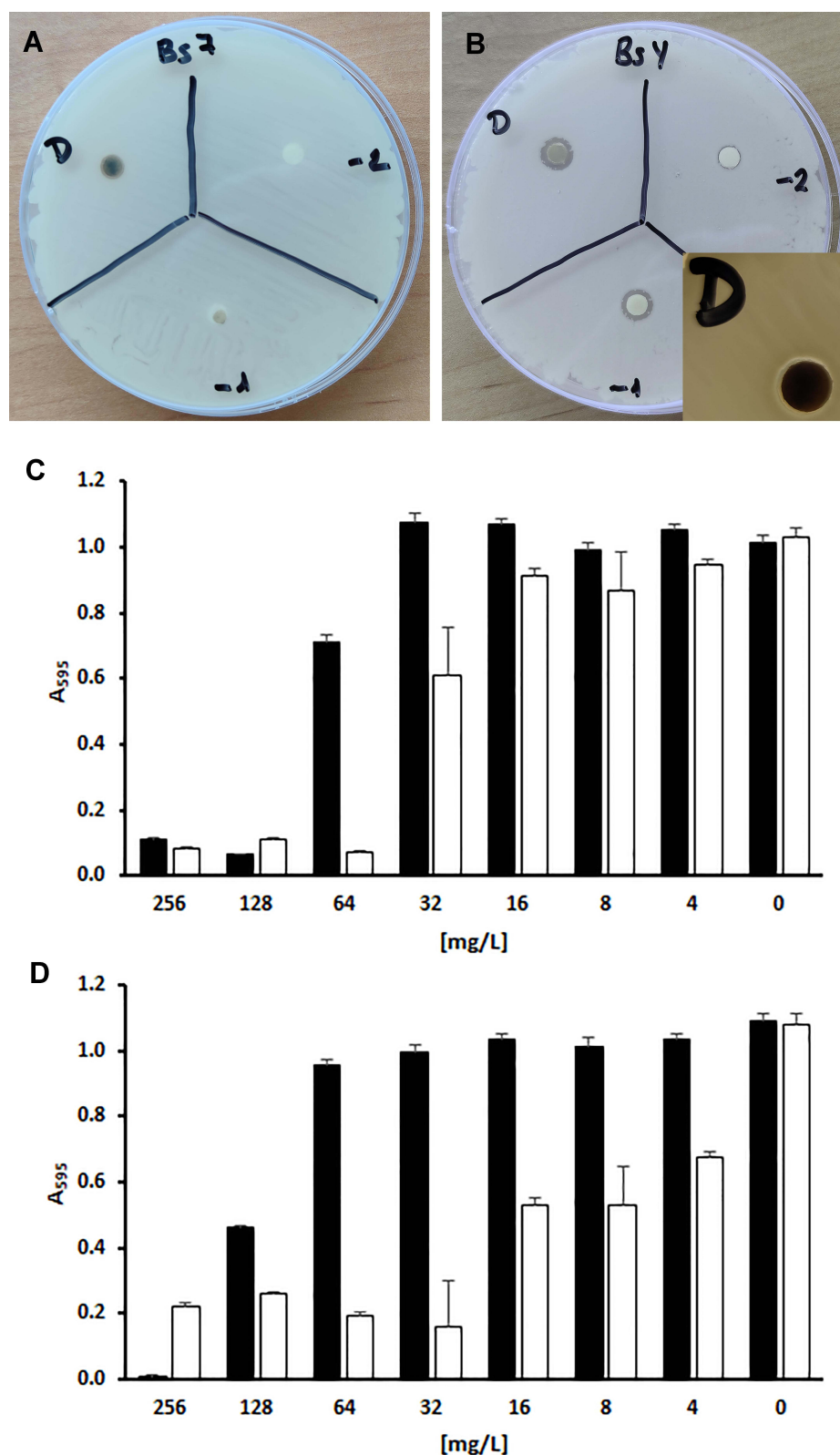


Figure 7 Microbial susceptibility tests. Panels (A and B) agar diffusion tests of *Bacillus* sp. (A) CECT 40 to AgNO₃ and (B) AgNPs-Sm; quantities load on the discs were 100 micrograms (discs labelled as (D), 10 micrograms (-1) and 1 microgram (-2). Panels (C and D) MICs for *Bacillus* sp. CECT 40 (C) and *Escherichia coli* CECT 101 (D) using AgNO₃ (white) and AgNPs-Sm (black); values are average and SD.

started at 32 mg/L for AgNO₃, although the measured optical density values suggest that a higher fraction of *Escherichia coli* cells were alive when compared to the *Bacillus* sp.-affected cultures (average of 0.2 and 0.1 absorbance units for *Bacillus* sp. and *Escherichia coli*, respectively) (Figure 7C and D). This difference might explain the lack of an inhibition zone for *Escherichia coli* in the agar dilution tests. Finally, although the inhibitory effect of AgNPs-Sm for *Escherichia coli* started at 128 mg/L, a concentration of 256 mg/L was needed to observe an inhibitory effect similar (higher) to that observed for AgNO₃ (Figure 7C). Thus, it was clear that the inhibitory effect of AgNPs-Sm was higher for *Bacillus* sp. than for *Escherichia coli*, as described in the previous literature,⁸⁸ and was weaker for both strains than that detected for AgNO₃.

Conclusions

The current study demonstrated the antioxidant activity of the aqueous extract of *S. mammosum* when obtaining AgNPs-Sm measuring 5.2 ± 2.3 nm. The values of the extract's antioxidant capacity analyzed by different techniques (ORAC-FL, CAA, DPV, CV, and TPH) decrease in relation to AgNPs, which could be an indirect measure of the ability of polyphenolic compounds such as gallic acid to react with the reducing compounds present. Likewise, it was determined that the extracts and nanoparticles generated antioxidant activity at the cellular level. Microbiological analysis revealed that the *Solanum mammosum* extract was nontoxic for *Bacillus* sp. and *Escherichia coli*. A weak growth inhibitory effect was observed for the AgNPs-Sm for both bacterial strains in the presence of Ag⁰ nanoparticles. The use of AgNPs to determine the antioxidant capacity of new molecules from plant extracts can be potentially applicable in biomedicine.

Abbreviations

AgNPs, silver nanoparticles; Sm, *Solanum mammosum* L.; SPR, surface plasmon resonance; ROS, reactive oxygen species; RNS, reactive nitrogen species; BHT, butylated hydroxytoluene; BHA, butylated hydroxyanisole; ORAC-FL, oxygen radical absorbance capacity using fluorescein; CAA, cellular antioxidant activity; CV, cyclic voltammetry; DPV, differential pulse voltammetry; HPLC, high-performance liquid chromatography; DAD, diode array detector; HR-TEM, high-resolution transmission electron microscope; DLS, dynamic light scattering meter; XRD, X-ray diffraction; FTIR-ATR, attenuated total reflection Fourier-transform infrared spectroscopy;

GC, glassy carbon electrode; SAED, selected area electron diffraction; ET, expressed as Trolox; EAG, expressed as gallic acid.

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Authors' Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure

The author reports no conflicts of interest in this work.

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