Aerobic training increases skin perfusion by a nitric oxide mechanism in type 2 diabetes

Sheri R Colberg1
Laura C Hill2
Henri K Parson3
Kathleen S Thomas1
Aaron I Vinik3
1Old Dominion University, Norfolk, Virginia, USA; 2State University of New York at Cortland, New York, USA; 3Eastern Virginia Medical School, Norfolk, Virginia, USA

Abstract: It is well known that a number of locally released vasodilatory and vasoconstrictive compounds can affect skin perfusion. This study investigated the effects of aerobic training on the contribution of nitric oxide (NO), prostaglandins (PG), and endothelial-derived hyperpolarizing factor (EDHF) in stimulated dorsal foot skin perfusion in individuals with type 2 diabetes (T2DM). Ten previously sedentary, older individuals with T2DM (57.0 ± 3.1 years) and nine sedentary controls (53.5 ± 3.2 years) were tested before and after undertaking six months of moderate aerobic training three times weekly in a supervised setting. All subjects underwent measurement of baseline (32°C) and heat-stimulated (40°C and 44°C) dorsal foot skin perfusion starting one hour after ingestion of a single, oral 325 mg dose of aspirin, a known inhibitor of PG synthesis. Before aspirin ingestion, a subcutaneous microdialysis probe was inserted into each foot dorsum to administer either saline (PG pathway only blocked by aspirin in the left foot) or L-NAME (N(G)-nitro-l-arginine methyl ester; thereby inhibiting both PG and NO pathways in the right foot). Normative data collected previously on subjects undergoing saline administration via microdialysis without aspirin ingestion served as a control group. Significantly lower responsiveness of maximal perfusion was found with the EDHF pathway alone unblocked compared with NO and EDHF unblocked after training. Maximal suppression attributable directly to NO, PG, and EDHF was not significantly different when examined by subject group and training status. However, contributions of NO, PG, and EDHF to maximal perfusion were significantly increased, decreased, and unchanged by aerobic training, respectively, with diabetic and control subjects combined due to nonsignificant differences between groups. Improvements in maximally stimulated dorsal foot skin perfusion resulting from six months of aerobic training appear to have primarily an NO basis, with lesser contributions from PG following training, regardless of diabetes status.

Keywords: skin perfusion, aerobic training, type 2 diabetes, nitric oxide, prostaglandins, endothelial-derived hyperpolarizing factor, microdialysis

Introduction

It is well known that a number of locally released vasodilatory and vasoconstrictive compounds can affect skin perfusion. Among the vasodilators are nitric oxide (NO), prostacyclins, prostaglandins (PG), bradykinin, and endothelial-derived hyperpolarizing factor (EDHF), which appear to vary in importance depending on their cutaneal location. Delivery of nutrients, removal of metabolic waste, temperature regulation, and healing are all affected by maintenance of skin perfusion, but when type 2 diabetes mellitus (T2DM) has an underlying presence, alterations in foot skin perfusion may result in a greater risk of ulceration and poor cutaneous healing.
Previously, it has been postulated that participation in chronic aerobic training (ie, regular exercise for longer than six months) improves stimulated dorsal foot skin perfusion through local release of vasodilatory compounds including NO, PG, and EDHF. A prospective study reported a significantly lower skin perfusion during local heating to 44°C in the dorsal feet of sedentary individuals with T2DM compared with active people without diabetes. This underlying defect was present despite normal increases in interstitial NO in the foot dorsum under stimulated conditions, suggesting that NO is either ineffective or not significantly involved in this acute response. A prior bout of acute maximal exercise also appears to improve responsiveness to local heating of the dorsal foot in active individuals with T2DM, but not in their sedentary counterparts.

These previous studies have suggested that participation in chronic aerobic training may regulate stimulated (heated) dorsal foot skin perfusion through alterations in either PG or EDHF pathways, given that interstitial cutaneous NO levels (measured via a subcutaneous probe) are not apparently significantly depressed by the presence of T2DM. Prior research also suggests that chronic exercise participation (lasting a year or more) may change the relative importance of PG and EDHF in maximal dorsal foot skin perfusion during local heating more so than NO. Therefore, the purpose of this study was to investigate the chronic effect of relatively short-term (six months), moderate aerobic training on the relative importance of these skin vasodilators in previously sedentary, older individuals with and without T2DM.

### Methods

**Subjects**

Ten sedentary T2DM and nine control subjects who were free of known cardiovascular disease, severe peripheral neuropathy, unstable proliferative retinopathy, end-stage renal disease, and uncontrolled hypertension participated in this study. In addition, no subjects were using insulin, angiotensin II receptor blockers, or angiotensin-converting inhibitors, due to their potential enhancing effects on nerve blood flow. Their sedentary status pretraining was verified using a modified version of the Harvard Alumni Physical Activity Questionnaire. By self-report, “sedentary” subjects had failed to engage in aerobic activities lasting 30 or more minutes at least three times per week for at least six months prior to the study.

After all subjects signed the informed consent approved by the Institutional Review Boards at both Eastern Virginia Medical School and Old Dominion University, they then underwent a history and physical examination, body fat estimation (via skin folds), exercise testing, and fasting blood work. Their resting and exercise characteristics are shown in the Table 1. Peripheral and autonomic neuropathy was assessed as previously described.

### Aerobic exercise testing and training

After undergoing cardiopulmonary exercise testing on a stationary cycle with measurement of peak oxygen consumption, subjects participated in moderate aerobic training for six months, consisting of three weekly sessions of 45 minutes of training at 50%–65% of heart rate reserve (ie, a set percentage of the difference between resting and maximal heart rate

### Table 1 Subject characteristics

<table>
<thead>
<tr>
<th>Subject number</th>
<th>CON Pre</th>
<th>CON Post</th>
<th>DM Pre</th>
<th>DM Post</th>
</tr>
</thead>
<tbody>
<tr>
<td>(Male/Female)</td>
<td>9 (5/4)</td>
<td>10 (3/7)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>57.0 ± 3.1</td>
<td>53.5 ± 3.2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Height (cm)</td>
<td>172.2 ± 4.1</td>
<td>172.0 ± 2.8</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>89.9 ± 13.6</td>
<td>88.3 ± 6.8</td>
<td>99.1 ± 8.0</td>
<td>98.3 ± 8.0</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>32.4 ± 4.3</td>
<td>33.6 ± 2.2</td>
<td>37.6 ± 2.2</td>
<td>33.1 ± 2.1</td>
</tr>
<tr>
<td>Heart rate (bpm)</td>
<td>71.2 ± 2.8</td>
<td>64.2 ± 3.2</td>
<td>82.3 ± 1.8</td>
<td>77.6 ± 1.8</td>
</tr>
<tr>
<td>Blood glucose (mM)</td>
<td>4.6 ± 0.1</td>
<td>4.9 ± 0.2</td>
<td>7.1 ± 0.1⁷</td>
<td>6.7 ± 0.7⁷</td>
</tr>
<tr>
<td>Serum insulin (μIU/ml)</td>
<td>6.4 ± 1.1</td>
<td>6.4 ± 1.5</td>
<td>15.6 ± 4.5⁶</td>
<td>10.9 ± 0.8³</td>
</tr>
<tr>
<td>HbA₁c (%)</td>
<td>5.9 ± 0.2</td>
<td>5.8 ± 0.1</td>
<td>7.3 ± 0.3⁰</td>
<td>7.4 ± 0.3¹</td>
</tr>
<tr>
<td>VO₂ peak (mL/kg/min)</td>
<td>19.0 ± 2.2</td>
<td>19.3 ± 2.1</td>
<td>17.1 ± 1.3</td>
<td>19.5 ± 1.5</td>
</tr>
<tr>
<td>E/i ratio</td>
<td>1.16 ± 0.02</td>
<td>1.18 ± 0.03</td>
<td>1.15 ± 0.03</td>
<td>1.19 ± 0.04</td>
</tr>
<tr>
<td>Total Neuropathy Score</td>
<td>4.1 ± 1.0</td>
<td>3.5 ± 0.6</td>
<td>4.1 ± 0.7</td>
<td>3.4 ± 0.7</td>
</tr>
</tbody>
</table>

**Notes:** *P < 0.05 versus controls pretraining; **P < 0.05 versus controls post-training.

**Abbreviations:** CON, control subjects; DM, diabetic subjects; Pre, pre-training; Post, post-training; bpm, beats per minute; HbA₁c, glycated hemoglobin; VO₂, peak, peak oxygen consumption; E/i ratio, expiration-to-inspiration ratio.

---

Colberg et al

Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy 2010:3

Dovepress
that is added to the resting value). All training sessions were supervised by exercise personnel and heart rate monitored to ensure that training sessions were completed in the prescribed target range. Subjects were allowed to choose among different modes of training, including treadmill walking, stationary cycling, elliptical strider, and seated rowing exercise. Subjects were required to undertake a chosen activity for a minimum of 10 minutes before another could be substituted during any given training session.

**Skin perfusion measurements**

Skin perfusion was measured using laser Doppler assessment with probes positioned on each mid-metatarsal foot dorsum both before and after completion of training. The latter was conducted 48–72 hours after completion of the last exercise session so that only chronic training effects were being measured rather than the residual effects of acute exercise. Baseline flow was assessed under the probe during sequential heating to 32°C for 50 minutes, 40°C for 10 minutes, and 44°C for 40 minutes to induce neurogenic vasodilation, starting about one hour after subcutaneous microdialysis probe insertion on each foot dorsum to administer L-NAME (N(G)-nitro-l-arginine methyl ester, right foot) or saline (left foot), as done previously.8

Briefly, a small dose of L-NAME a known NO synthase inhibitor, dissolved in 2 mL of normal saline, was administered over 100 minutes starting one hour after subjects ingested a single 325 mg oral dose of generic aspirin to inhibit all but the EDHF pathway in the right foot and the PG pathway alone in the left foot. To prepare the infusate, 4 mL of sterile 9% saline were added to a single vial containing 220 mg of L-NAME, and after mixing, 2 mL were withdrawn using the microsyringe. Infused at a rate of 2 μL per minute over 100 minutes, about 200 μL of the L-NAME solution was administered locally to a small area of skin under the laser Doppler probe by the end of the testing period.

Normative data collected previously on subjects undergoing saline administration via microdialysis without aspirin ingestion served as a control for this study.8 The normative value used to determine percent suppression was 150 arbitrary perfusion units during maximal stimulation caused by localized heating to 44°C.

**Statistical analyses**

Separate repeated one-way analysis of variance (ANOVA) measures were employed to compare skin perfusion during each heating condition (ie, baseline at 32°C, heating to 40°C, heating to 44°C) between groups (T2DM versus controls) both pre- and post-training. Pooled data with subject groups combined were analyzed with results from maximal heating to 44°C only. Resting subject characteristics (T2DM versus controls) were analyzed using unpaired t-tests. Significance for all statistical analyses was set at \( P < 0.05 \).

**Results**

T2DM subjects had significantly higher fasting serum glucose, insulin, and glycated hemoglobin levels than controls both pre-and post-training \( (P < 0.05) \). Body fatness was similar between groups pre-training, as were resting heart rate, peak oxygen consumption, and neuropathy scores, and none of these measures were significantly altered by training.

No significant differences in skin perfusion under baseline (32°C) conditions or at 40°C were observed (data not shown). With blockage of PG or PG and NO pathways during maximally stimulated conditions (local heating to 44°C), significantly greater suppression of dorsal foot skin perfusion was observed with the non-PG, non-NO (EDHF) pathway alone unblocked compared with non-PG pathways (NO and EDHF) after training compared with the sedentary state in both controls and T2DM subjects (Figure 1).

However, as shown in Figure 2, no significant differences were found when suppression of maximally stimulated dorsal skin perfusion was examined as a percentage of suppression attributable directly to each vasodilator individually, both pre- and post-training. Pooled data with subject groups combined were analyzed with results from maximal heating to 44°C only. Resting subject characteristics (T2DM versus controls) were analyzed using unpaired t-tests. Significance for all statistical analyses was set at \( P < 0.05 \).

**Figure 1** Suppression of maximally stimulated (44°C) dorsal skin perfusion by subject group pre- and post-training. “Non-PG” conditions include oral ingestion of aspirin and systemic suppression of PG synthesis by aspirin (left foot), while “non-PG/non-NO” conditions have inhibition of PG by aspirin and add local blockade of NO via microdialysis infusion of L-NAME, a NO synthase inhibitor (right foot). These results include oral aspirin ingestion in all subjects and have not yet been compared with normative data collected without aspirin. Subject groups are differentiated by both diabetes and training status.

**Notes:** Black bars, control subjects pre-training; gray bars, diabetic subjects pre-training; spotted bars, control subjects post-training; striped bars, diabetic subjects post-training. \( *P < 0.05 \) versus same group in non-PG; \( †P < 0.01 \) versus same group in non-PG.

**Abbreviations:** PG, prostaglandins; NO, nitric oxide; L-NAME, (N(G)-nitro-l-arginine methyl ester); CON, controls; DM, diabetes mellitus.
determined using normative data collected previously in similar subject groups without oral aspirin ingestion. For these analyses, subject groups were differentiated by both diabetes (T2DM versus controls) and pre- and post-training status.

For subsequent analyses, the T2DM patients and controls were grouped due to nonsignificant differences between subject groups based on diabetes status. The relative contributions of NO, PG, and EDHF to maximal skin perfusion after six months of moderate aerobic training were determined using further analyses that again included the normative data without oral aspirin ingestion to block PG. In these analyses, the contributions of NO, PG, and EDHF were significantly increased, decreased, and unchanged by training, respectively, as shown in Figure 3. Using blockage of PG (with aspirin) or PG and NO (with aspirin and L-NAME), significantly less responsiveness of maximal perfusion was evident with the latter EDHF pathway only unblocked (36.5% ± 5.7%) post-training compared with both NO and EDHF unblocked after training (66.9% ± 3.5%; P < 0.05).

**Discussion**

This study examined the importance and relative contributions of different vasodilatory compounds to dorsal foot skin perfusion during baseline and local heat-stimulated conditions in individuals with and without T2DM before and after six months of moderate, supervised aerobic training. We hypothesized that such training may alter the relative importance of PG, NO, and EDHF similarly to that previously observed in chronic exercisers, ie, increased PG, reduced EDHF, and unchanged NO contributions to stimulated perfusion responses.8

Contrary to our expectations, six months of training undertaken by sedentary individuals in the current study resulted instead in a significantly increased contribution to stimulated perfusion apparently coming from an NO mechanism, with reduced PG and unchanged EDHF. Consistent with these findings, Heidarianpour et al11 reported that 10 weeks of aerobic exercise training improves NO-dependent dilatation, possibly via the phosphoinositide 3-kinase pathway. However, when previously examining the effects of similar, moderate aerobic training done three times weekly for 10 weeks in participants with T2DM, we found no significant changes in measured levels of cutaneous interstitial NO in the dorsal foot during local heating despite differences in stimulated perfusion.12

Gooding et al3 investigated the vasodilatory mechanisms involved in maximal skin hyperemia in response to localized heating to greater 40°C or above, which is similar to the technique employed in the current study. In nondiabetic subjects, NO, but not PG, histamines, or an axon reflex, appeared to contribute most to the increase in blood flow in response to heating. Although NO levels are not necessarily altered by diabetes,3 defects in stimulated perfusion may result from a diminished local effectiveness of NO, particularly given that endothelium-dependent dilation in skin can be enhanced by eight weeks of moderate training and reversed by detraining.3,6,12 The current results are suggestive of a shift to a greater reliance on NO (via enhanced
release and/or effectiveness) following relatively short-term participation in moderate aerobic exercise training and a lesser relative contribution of PG, although more chronic participation (lasting a year or more) may later increase the importance of PG. The current study design did not allow for a direct measurement of NO to confirm that it remained unchanged, but all of our prior research supports this conclusion in similarly-aged diabetic and non-diabetic populations studied.12

Perfusion responses to such localized heating are largely controlled by small C-fiber nociceptors,13 and in the skin of diabetic subjects, significant C-fiber impairment and attenuated NO-mediated skin vasodilation have been reported.14 The defect in skin vasodilation is detectable before the development of diabetes and is partially correctable with insulin sensitizers, may be associated with C-fiber dysfunction (ie, the dermal neurovascular unit), coexists with variables of the insulin resistance syndrome, and is partially correctable with insulin sensitizers. This defect may involve an imbalance among NO, substance P, calcitonin gene-related peptide, angiotensin II, endothelin, and other vasodilatory or vasoconstrictive compounds.4,10 The participants in the current study were all studied during maximal stimulated skin perfusion in the dorsal foot. The results of the present study examining dorsal foot skin perfusion concur with these observations related to the effects of localized heating on forearm skin responses. With regard to the interactions of NO and PG on skin vasodilation, in healthy humans, chronic low-dose aspirin therapy attenuates the reflex cutaneous vasodilation during whole-body heating that is mediated through both the cyclooxygenase-dependent pathway of PG formation and NO synthase-dependent mechanisms.17 While vasodilation of skin in response to local heating is highly NO-dependent, it may also reflect the interaction of NO and PG.15,18 Cutaneous acetylcholine-mediated endothelium-dependent vasodilation has been reported to be highly NO-dependent and also strongly related to the interactions of NO with prostaglandins.18 Thus, an interaction between these two vasodilatory compounds during localized heating of dorsal foot skin to 44°C is also a possibility, albeit an untested one, in this study.

Admittedly, the introduction of a microdialysis catheter into the cutaneal surface of the dorsal foot undeniably resulted in an immediate PG release and influx of interstitial fluids. Microdialysis probe insertion into the ventral forearm is known to cause a significant increase in skin blood flow, erythema, flare, and skin edema,19 and a 90–120 minute period of equilibration is required in human skin to allow the vascular reactions of the skin to stabilize.20 While it is unlikely that the skin perfusion completely normalized after the probe’s insertion in the current study, PG dissipation was likely to have been relatively rapid during the initial hour of stabilization. Skin perfusion returned largely back to normal values before baseline testing was started after an hour, and another hour elapsed before maximal heating to 44°C was initiated, thereby allowing 120 minutes after the probe insertion before the maximal responses were tested.

As for the EDHF pathway, it may play a larger role only when NO or PG is compromised,2 which NO did not appear to be in this study, and EDHF may serve as more of a back-up mechanism, particularly with aging.21 Both NO and PG are known to play important roles in skeletal muscle blood flow regulation during moderate intensity exercise, and EDHF does not compensate for the impaired formation of NO and prostaglandins in muscle.22 The relevance of these findings to cutaneal responses has not been established. In the presence of a diabetic state, toe skin perfusion has been shown to increase less in response to PG analogs when the ankle brachial index is lower,23 which is also a possible factor affecting the current findings, but one that was not able to be investigated in the current study design.

A recent study of the effects of ruboxistaurin on skin microvascular blood flow evaluated the relationship between endothelial and neural control of this perfusion in diabetic patients with diabetic peripheral neuropathy.24 They found a correlation between C-fiber-mediated and endothelium-dependent cutaneous perfusion, suggesting that improvements in endothelial function may affect the microcirculation not only locally, but also via the neurovascular arcade. As for the potential effects of neuropathic changes in the present study, skin perfusion was similar between control and diabetic groups, and measures of both central and peripheral
neuropathy were similar between subject groups, making neuropathic changes unlikely to have affected the results.

In conclusion, six months of moderate aerobic exercise training appears not to alter baseline or maximal dorsal foot skin perfusion responses via EDHF, but does affect release of or sensitivity to NO and PG, regardless of diabetes status. Thus, it is likely that improvements in dorsal foot skin perfusion resulting from six months of moderate training involve a non-EDHF pathway, most probably NO for short-term (including six months of training) and PG for longer-term (ie, in chronic exercise) adaptations. Moreover, these findings suggest that regular exercise participation may have a bigger impact on maximal skin perfusion than diabetes alone, and may counter the negative effect that T2DM can exert on perfusion, at least in dorsal foot skin.

Acknowledgments
This work was fully supported by a clinical research grant from the American Diabetes Association. We would also like to acknowledge Amanda L Boyd and Kathleen R Sharp for their assistance in collecting these data.

Disclosure
The authors report no conflict of interest in this work.

References