Corneal Donation: Current Guidelines and Future Direction

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Purpose: This review aims to outline current practices and guidelines of corneal donation and eye banking, describes the implications of COVID-19 and emerging diseases on the corneal donor pool, and discusses future trends to improve and increase the efficiency of the processes involved in corneal donation and eye banking.

Summary: Corneal screening, preservation, corneal storage, and prevention of systemic disease transmission from donor to recipient have been crucial in shaping the policies of the FDA and eye banks across the world. Eye banks globally have developed varying guidelines and criteria for evaluating the viability of donor corneas. Variables such as the age of the donor, medical history, and potential disease transmission are important screening parameters. While known infectious diseases may be transmissible through the cornea, emerging infectious diseases that are not well studied may be more transmissible than other infections.

In particular, coronavirus has impacted corneal transplantation as SARS-CoV-2 expression has been detected in corneal tissue and conjunctiva. In recent years, partial-thickness corneal transplantations have been introduced. Lamellar grafts and other corneal layers are now utilized for transplantation of the specific areas that are damaged.

Keywords: donor cornea, eye bank, COVID-19, cornea transplant, keratoplasty, donor recipient

Introduction

Corneal blindness is one of the leading global causes of blindness, with nearly 12.7 million people requiring a transplant. However, the ratio of corneas available to those required is a mere 1 in 70.1 Corneal transplantation has enabled us to replace damaged parts of the cornea, allowing for restoration of sight. For successful corneal transplantation, several factors must be considered, including screening parameters, preservation technique storage, transport, and the evaluation of corneal tissue using serological testing and imaging. Initial clinical screening of the patient’s medical history is crucial in determining past or recent potential for infectious disease transmission via corneal transplant. The Eye Bank Association of America (EBAA) plays a vital role in creating the standards for the evaluation of donor tissue, distribution, and quality assurance. Eye banks all over the world play a similar role in evaluating corneal tissue for transplantation.

Previously, surgeons typically transplanted full-thickness corneas in order to restore vision. Corneal transplantation methods are evolving to replace only specific layers of the cornea. It is important to note that COVID-19 has affected corneal transplantation due to the presence of SARS-CoV-2 in conjunctival swabs, which has been confirmed by several different studies.2–4 However, evidence of direct
transmission is yet to be determined. Due to the various potential risks, several eye banks, including the EBAA, have developed specific guidelines to prevent disease transmission via the ocular surface.

**Evolution of Corneal Preservation**

For several years, corneal transplantation was a time-sensitive process that required obtaining tissue from deceased donors, immersing the tissue in saline, and immediately transferring the tissue to the recipient within hours of harvest. However, the 1930s marked a change whereby collected corneal tissue was stored in moist chambers in ice. Such methods of preservation for more extended periods are even more prevalent today. The US Food and Drug Administration (FDA) guidelines allow for corneal preservation in solution for a maximum time per period of 14 days. In 2017, Rosenwasser et al conducted the Cornea Preservation Time Study (CPTS), a national clinical trial through which they showed that corneal donor tissue can be stored for up to eleven days to ensure proper transplantation of cornea and restoration of vision. However, despite such data, surgeons in the United States have largely refrained from using corneas preserved for longer than 7 days. Several scientists have suggested that such practices are rooted in opinion instead of scientific evidence.

Current corneal transplantation techniques utilize various approaches for the preservation and storage of donor cornea. In order for proper corneal transplantation to occur, the corneal endothelium must be viable. Preservation of endothelium is crucial in transplantation because the proliferation of endothelial cells is limited in situ as they are arrested in the G1 phase of the cell cycle.

There are three primary approaches to corneal preservation: organ culture, hypothermia, and cryopreservation. Organ culture is a method of preservation whereby corneas are incubated in tissue culture medium which can contain additional supplements such as antibiotics or antimycotics. Hypothermia is another technique whereby donor eyes are stored at low temperatures as it significantly decreases the cell’s demand to use metabolic energy. Cryopreservation requires storage at subfreezing temperatures, typically below −80°C. Cryopreservation, the preservation of tissues by cooling at low temperatures, provides longer storage times compared to organ culture and hypothermia. However, cryopreservation has been shown to be harmful as freezing corneas leads to endothelial damage via intracellular freezing or solution effect injury. Several studies have shown that cryopreservation is the most effective method of storing corneal stromal lenticules. Cryopreservation is crucial for deep anterior lamellar keratoplasty (DALK), a partial-thickness transplantation of corneal stroma. Mohamed-Noriega et al examined the collagen structure of stromal lenticules retrieved from refractive lenticule extraction (ReLEx) following a one-month period of cryopreservation. They concluded that the architecture of the collagen and viability of keratocytes was retained after cryopreservation. In contrast, hypothermic storage at 2–8 °C is widely used and accepted as the more effective means of preservation. However, standard methodology for lenticule harvest and preservation is yet to be established as methods are continuously evolving.

In the US, eye banks primarily utilize Optisol-GS as the storage solution, which consists of dextran, chondroitin sulphate, and vitamins. The McCarey–Kaufman medium (M-K medium) consisting of culture medium 199 and dextran 40 is also utilized in many countries but has a limited storage time of 2–4 days. In 2005, Wagoner et al determined the mean Optisol-GS storage time to be 237.1 ± 41.6 hours (7–14 days). The Optisol-GS solution alone lacks antifungal supplements. Several cases of corneal transplantation have reported candida keratomycosis. Antifungal additives to Optisol-GS, such as amphotericin, have been proven to limit the growth of fungal colonies, particularly those of the *Candida* species, one of the most frequent causes of post-corneal transplant fungal infections. For stromal lenticules, cryopreservation in glycerol solution has proved to be cheaper and more effective. Glycerol has antimicrobial characteristics due to its role as a dehydrating agent and is thereby more potent in long-term preservation. Other commonly utilized solutions for cryostorage include Dulbecco’s Modified Eagle’s Medium (DMEM), serum-free medium (SFM), and Dексsol. Greenbaum et al show that corneal epithelium can be stored effectively in both Optisol-GS and Dексsol with no significant differences in viability and preservation of epithelium among both solutions. However, it is important to recognize that a thorough comparison of preservation methods for corneal epithelium is yet to be reported.

Organ culture is a third preservation approach most commonly used in European eye banks where eyes are rinsed with saline, placed in povidone–iodine solution, and later suspended in Eagle’s minimum essential medium.
(MEM). Through this medium, corneas can be preserved for a maximum period of 4 weeks. In comparison to hypothermia, organ culture allows for the identification of bacteria and fungi, which could cause infection following transplantation. Organ culture medium can be utilized for storing Bowman Layer (BL) grafts. Located between the epithelial basement membrane and the anterior corneal stroma, BL is primarily comprised of collagen fibers, providing structural support. BL transplantation, a minimally invasive method where BL is inserted into a stromal pocket, is used to treat keratoconus. Similarly, Descemet’s membrane, the basement membrane of the corneal endothelium, is often isolated for use in Descemet Membrane Endothelial Keratoplasty (DMEK), a partial-thickness transplant used to treat damaged corneal endothelium. DMEK grafts can also be stored in organ culture medium or growth factor-containing enhanced medium (En-OC). Romano et al compared various forms of preservation in DMEK procedures by testing endothelial viability among endothelium placed in organ culture versus hypothermic conditions. They concluded that endothelium stored in organ culture was viable for 30 days compared to 14 days in hypothermic storage.

**Corneal Evaluation and Eye Bank Guidelines**

Prior to transplantation, eye banks perform a thorough screening of the eye donors for eligibility. The EBAA provides detailed guidelines and medical standards for corneal transplantation (Table 1). The EBAA requires that each donor is identified by name. Their medical standards call for a consistent method of examination and documentation of donor eligibility within each eye bank. The guidelines in Table 2 summarize the necessary information and procedures required for effective donor eligibility determination, but is not limited to those listed in the table. Generally, eye banks in the US perform three primary tests, including serological testing, review of donor’s medical history, and physical inspection of corneas using imaging technique. Serological testing is performed using donor blood collected during tissue procurement. It is important to note that corneal transplantation does not require that donors and recipients have the same blood type because the cornea is an immune-privileged environment and does not contain blood vessels. The risk of transplant failure due to ABO incompatibility is not increased. The donor is screened for hepatitis B, hepatitis C, syphilis, and HIV, among other infectious diseases. After preliminary screenings, the cornea is examined utilizing various imaging techniques to assess the general quality of corneal tissue.

### Table 1 Eye Bank Association of America Donor Screening Guidelines

<table>
<thead>
<tr>
<th>Requirement</th>
<th>Requirements</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>EBAA Testing</strong></td>
<td>The following tests must be completed and must return a negative or non-reactive test result. A</td>
</tr>
<tr>
<td></td>
<td>(1). Anti-HIV-1, Anti-HIV-2</td>
</tr>
<tr>
<td></td>
<td>(2). Hepatitis B Surface Antigen (HBsAg)</td>
</tr>
<tr>
<td></td>
<td>(3). Anti-HCV</td>
</tr>
<tr>
<td><strong>FDA Testing</strong></td>
<td>The FDA recommends the following tests:</td>
</tr>
<tr>
<td></td>
<td>(1). HIV, Type 1: FDA licensed screening test for anti-HIV-1 or combination test for anti-HIV-1 and anti-HIV-2, FDA licensed Nucleic Acid (NAT) test for HIV-1 or combination NAT</td>
</tr>
<tr>
<td></td>
<td>(2). HIV, Type 2: anti-HIV-2 test or combination test</td>
</tr>
<tr>
<td></td>
<td>(3). HBV: FDA licensed test for HBsAg, total antibody to Hepatitis B core antigen (anti-HBc), and NAT screening for HBV</td>
</tr>
<tr>
<td></td>
<td>(4). HCV: Screening Test for HBsAg, total antibody to Hepatitis B core antigen (anti-HBc), and NAT donor test</td>
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<tr>
<td></td>
<td>(5). Treponema pallidum: screening test for syphilis</td>
</tr>
</tbody>
</table>

### Eye Banks and Eye Bank Associations

Eye banks play a crucial role in the evaluation and distribution of ocular tissue for transplantation. There are different types of eye banks in the United States, such as non-profit, for-profit corporations, consortium, and university-based eye banks. For instance, Lions Eye Bank consists of more than 60 eye banks worldwide, all operated by a non-profit foundation. In contrast, CorneaGen serves as a for-profit business entity with the goal of eliminating corneal blindness. The Eye Bank Association of America (EBAA) represents 83 eye banks across the United States as well as 14 international eye banks. Within the United States, eye banks must be credentialed by the EBAA as well as the FDA. The credentialing process occurs every 3 years, whereby an
Table 2: Eye Bank Association of America Donor and Donor Eye Eligibility Guidelines

<table>
<thead>
<tr>
<th>Infectious Disease Testing</th>
<th>See Table 1 for the Following Conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Physical Assessment of Donor</td>
<td>The donor must be examined for signs of:</td>
</tr>
<tr>
<td></td>
<td>(1). HIV</td>
</tr>
<tr>
<td></td>
<td>(2). Infectious Hepatitis</td>
</tr>
<tr>
<td></td>
<td>(3). Intravenous Drug Use</td>
</tr>
</tbody>
</table>

| Tissue Evaluation | Corneal tissue must be evaluated through an array of methods including slit lamp examination, endothelial cell density, and pachymetry measurement. |

<table>
<thead>
<tr>
<th>Donor History Evaluation</th>
<th>Donor’s name and medical history are required. Donor information from at least one of the following is included:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(1). Medical examiner’s physical assessment of death report</td>
</tr>
<tr>
<td></td>
<td>(2). Medical examiner’s investigative report</td>
</tr>
<tr>
<td></td>
<td>(3). Police Investigation Report along with (1) or (2)</td>
</tr>
<tr>
<td></td>
<td>(4). Donor Risk Assessment Interview</td>
</tr>
<tr>
<td></td>
<td>(5). Medical record or hospital chart</td>
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<tr>
<td></td>
<td>(6). Treating physician interview</td>
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<tr>
<td></td>
<td>(7). Review of donor information from any of these categories by the medical director</td>
</tr>
</tbody>
</table>

**Contraindications:**

1. Rabies
2. Hepatitis B
3. Hepatitis C
4. Creutzfeldt–Jacob disease
5. Retinoblastoma
6. Bacterial/fungal keratitis
7. Bacterial/fungal endophthalmitis
8. HIV
9. Herpes Simplex Virus
10. Prion Disease
11. Human T-Cell Lymphotropic Virus
12. Ocular Adenocarcinoma
13. Malignant Tumors of Anterior Segment
14. Reye Syndrome
15. Subacute sclerosing panencephalitis
16. Progressive multifocal leukoencephalopathy
17. Leukemias
18. Active disseminated lymphomas
19. Active infectious endocarditis
20. Active septicemia
21. Dementia
22. Down Syndrome
23. Congenital Rubella
24. Recipient of non-synthetic dura mater graft
25. Prior refractive surgery
26. Neurological diseases of unknown cause

**Important Considerations:**

1. Donors are deemed ineligible if they have received a tattoo within the past 12 months using unsterile instruments.
2. Transgender populations are largely omitted in EBAA guidelines.

**Note:** Data from Eye Bank Association of America (EBAA).
eye banker and corneal surgeon perform site inspections to assess compliance to the medical standards of the EBAA, standards which have been endorsed by the American Academy of Ophthalmology. The process consists of two stages, including pre-inspection as well as on-site inspection. The eye bank completes a pre-inspection questionnaire (PIQ) reporting on staff qualifications as well as testing processes and maintenance. During the on-site inspection, operating procedures, donor records, equipment, and administrative documentation are reviewed in detail. The team concludes by completing a site inspection questionnaire (SIQ) to document their observations, and further outlines any necessary corrective actions and preventive actions (CAPA). EBAA Accreditation Board later convenes to grant an eye bank with one of three forms of accreditation: 3 years, 1 year, or denial of accreditation.

It is also important to note the coroner’s role in eye banking. The coroner is crucial in authorizing the pre-autopsy release of corneal tissue; thus, the coroner’s office and eye bank should maintain good communication. For instance, the Rocky Mountain Lions Eye Bank recommends that coroners sign a protocol with each eye bank to ensure that specific standards are met. This is significant as time impacts the preservation of corneal tissues.

**Corneal Tissue Imaging Analysis for Different Types of Transplants**

After screening for eligibility, slit-lamp biomicroscopy and ancillary imaging are crucial tools in the complete evaluation of the cornea. Specifically, slit-lamp biomicroscopy is utilized to detect endothelial disease and further examine the epithelium and stroma. The presence of corneal edema, scarring, stress lines, arcus, pterygia, neovascularization, striae, central guttata, polymegathism, pleomorphism, and infiltrates may all indicate issues of viability. The EBAA also asks that eye banks define and mark a “clear zone” on the cornea, measuring the diameter of the cornea that is absent of conditions such as scarring, stress lines, and other pathologies. Specular microscopy is another imaging technique essential for examining endothelial cell morphology and viability. Pachymetry is a third technique wherein a probe is used to measure the thickness of the cornea. Corneal thickness is important as it may indicate corneal swelling as seen in conditions such as Fuchs’ dystrophy. Transplantation methods involving specific corneal layers require varying criteria for evaluation (Table 3).

While evaluation criteria for whole donor corneas remain the same across these different procedures, many additional criteria are unique to the corneal layer transplanted. During Bowman’s Layer Transplantation (BLT), the graft is harvested via separation of the BL from the anterior stroma layer taken from either corneoscleral rims or entire donor globes. Sharma et al have shown that both methods result in very similar success rates, reporting rates of 69.4% and 72.2% for globes and corneoscleral rims, respectively. The primary evaluation criteria for BL grafts include tearing of the BL and thickness of graft. A thick BL graft may contain additional stromal tissue, leading to poor quality of vision following transplantation. Tearing of the BL leads to damage of tissue, thus rendering it non-viable. Damage to Bowman’s Layer can also indicate damage to the stroma as the BL is formed early in development and cannot be repaired later.

DMEK transplantations present another set of criteria due to the intricacy of the procedure. Donor tissue selection is thus crucial for the DMEK process. Specifically, the DMEK procedure recommends that tissues be harvested from donors of age 55 and older because tissue from those who are younger tends to create compressed scrolls, thus posing intraoperative challenges as the scroll must later unfold in the procedure. However, this topic is a source of debate as other scholars, specifically Schaub et al, found that younger donor age did not, in fact, alter the outcome of DMEK within the first postoperative years. Schaub et al reviewed 1084 consecutive DMEKs and found that corneal donors with minimum age of 17 years were viable donors. The endothelial cell density can be examined through specular microscopy and successful transplantation often requires cell densities greater than 2500 cells/mm².

While slit-lamp imaging provides a broad inspection of the tissue, specular microscopy is essential for identifying corneas with marginal functional reserve and evaluating endothelial cell density. Corneal endothelium contains a large number of hexagonal cells of specific size, and thus significant variation in cell size (polymegathism) further indicates non-viable tissue. Various eye banks require a minimum endothelial cell density for corneal endothelium transplantation. In a recent study published in 2018, Batista et al have shown that high-resolution two-photon imaging is another technique capable of examining...
Table 3 Criteria for Evaluation of Corneal Tissue for Transplantation Procedures

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Contraindications</th>
</tr>
</thead>
<tbody>
<tr>
<td>Penetrating Keratoplasty (PK)</td>
<td>Infiltrates, pterygia, neovascularization, foreign bodies, stromal scars in graft area, detachment of Descemet's membrane, endothelial dystrophy, low endothelial cell density, Down syndrome, ectatic dystrophy (keratoconus, keratoglobus), previous refractive surgery (laser and incisional)</td>
</tr>
<tr>
<td>Anterior Lamellar Keratoplasty (ALK/DALK)</td>
<td>Infiltrates, pterygia, neovascularization, foreign bodies, stromal scars in graft area, Down syndrome, ectatic dystrophy, previous refractive surgery</td>
</tr>
<tr>
<td>Descemet's stripping endothelial keratoplasty (DSEK/DSAEK)</td>
<td>Infiltrates, scars impacting posterior stroma in graft area, detachment of Descemet's membrane, low endothelial cell density, low rim size and corneoscleral disc size (necessary for mounting on anterior chamber)</td>
</tr>
<tr>
<td>Descemet's membrane endothelial keratoplasty (DMEK)</td>
<td>Infiltrates, foreign bodies, tears in Descemet's membrane in graft area, low endothelial cell density</td>
</tr>
<tr>
<td>Keratolimbal Allograft (KLA)</td>
<td>Infiltrates, small scleral rim, conjunctiva not intact over the rim, melanoma, cancer of solid organ</td>
</tr>
<tr>
<td>Keratoprosthesis (K-Pro)</td>
<td>Infiltrates, pterygia, neovascularization, foreign bodies, significant thinning of the cornea, prior refractive surgery, Down syndrome, ectatic dystrophy</td>
</tr>
</tbody>
</table>

Note: Data from Eye Bank Association of America (EBAA).

The impact of femtosecond lasers in corneal transplantation procedures should be considered. Femtosecond lasers can be utilized for several anterior refractive procedures such as penetrating keratoplasty, laser in situ keratomileusis (LASIK), DALK, and DSEK. The laser allows for bladeless incisions with very high precision. Furthermore, femtosecond lasers allow a single donor tissue to be utilized for several recipients undergoing various forms of corneal transplantation. The femtosecond laser is reported to provide better wound stability along with decreased recovery times.

Current Criteria for Corneal Donation in the United States

Eye banks in the US typically require that donors be younger than 65 years of age. However, several debates surrounding the topic have emerged as scholars disagree on whether donor age is a proper indicator of the quality of donor cornea. For instance, Mannis et al determined that the ten-year success rate of corneal transplantation, specifically penetrating keratoplasty, was determined to be 75% for transplants from donors aged 34–71 years old. It is reported that nearly 75% of donors in the US fall within this age range and one-third of the donors fall within the age range of 61–70. The study utilized 1090 corneas from donors aged 12–75. Furthermore, they found that the success rate was higher for donors aged 12–33 years (96% for 80 donors) in comparison to donors aged 72–75 (62% for 120 donors). However, the study concluded that age is not a crucial factor in the success of transplantation for penetrating keratoplasty due to the similar success rate for donors aged 34–71. Thus, data indicate that cornea donors older than 65 years can result in successful transplantation. As surgeons typically refrain from utilizing corneal tissue from older donors to maximize success rates, it is expected that the pool of younger donors will decrease significantly in comparison to donors 65 years and older.

On May 20, 1994, the FDA has also instituted guidelines, which has led to a decrease in the pool of donor corneas. The FDA established a policy whereby men who have had sex with men (MSM) in the past 5 years...
are prohibited from serving as corneal donors. While introduced several decades ago, the policy remains in place and is enforced by the FDA. In the 2018 calendar year alone, the policy led to a removal of approximately 1600 potential donors from the donor pool. During the introduction of the policy, HIV tests were deemed unreliable for up to 6 months following exposure.47 However, Puente et al discuss that modern virology techniques can effectively detect HIV after a few days of exposure. Thus, they call for reformulation of such policies as they prevent thousands of donor corneas from being utilized.47

The FDA and EBAA have not established guidelines surrounding the ability of transgender individuals to donate corneal tissue. FDA policies typically call for gender self-identification.48 However, for instance, regarding the MSM policy, no clear guidelines exist for transgender individuals who may not identify as a specific gender.49 Consequently, this expresses the need to revise policies and include transgender populations.49

Risks for Disease Transmission

The potential for transmission of infection through corneal transplantation has been documented by various studies. Current documented cases of transmission include hepatitis B, Creutzfeldt-Jakob disease (CJD), and rabies.50 CJD is a fatal neurodegenerative disorder caused by an abnormal form of prion.51 Contraindications to transplantation include herpes simplex virus, hepatitis B, syphilis, tuberculosis, smallpox, and malaria, among many others.52 Keratoplasty was the first procedure that proved transmission of virus through corneal transplantation.53 Remeijer et al were the first to show that HSV-1 (herpes simplex virus) can be transmitted through penetrating keratoplasty.54 Several years later, EBAA medical advisory board reported 31 cases of postoperative fungal keratitis (n=14) and endophthalmitis (n=17) out of 221,664 corneal transplants.55 Currently, the SARS-CoV-2 strain presents a significant threat to corneal transplants.

The EBAA has also determined that many viruses can be transmitted during corneal transplantation, including the West Nile, Ebola, Zika, and Vaccinia viruses.56 However, according to the EBAA, there is a lack of sufficient evidence to confirm the transmission of such viruses through corneal transplants.22

Individuals with tattoos have been a topic of contention regarding the ability to serve as a corneal donor. Tattoos may significantly increase the likelihood of the donor transmitting viruses such as hepatitis B and C, HIV, and molluscum contagiosum.57 The EBAA guidelines recommend that individuals who have received a tattoo or any body piercings in the past 12 months without the use of sterile procedures or sterile instruments (ie, use of shared instruments unsterilized between use) be considered ineligible for corneal donation.22

SARS-CoV-2 and Emerging Eye Bank Guidelines

COVID-19 manifestations include conjunctivitis.58 Studies have detected the presence of COVID-19 in conjunctival swabs.2–4,59–61 A case report indicated the presence of SARS-CoV-2 RNA in tears for up to 27 days after initial onset.61 However, the evidence does not prove whether SARS-CoV-2 can be directly transmitted through an ocular surface.62

SARS-CoV-2 infects the cell through the ACE2 receptor upon priming of cellular protease using transmembrane serine protease 2 (TMPRSS2).62 A recent study published in 2021 investigated the expression of SARS-CoV-2 entry genes in corneal and conjunctival epithelium. Collin et al utilized single RNA Seq and ATAC-Seq datasets to discover co-expression of ACE2 and TMPRSS2.62 They also found that inflammatory signals upregulated the expression of ACE2+TMPRSS2+ cells, showing that the ocular surface epithelium is a site of SARS-CoV-2 entry.62 Sawant et al examined expression of SARS-CoV-2 in corneal tissue using RT-PCR.63 The study used tissues from 33 surgical-intended donors removed from the donor pool due to EBAA or positive COVID test results.63 They detected a positivity rate of 13% (17 of 132 ocular tissues), indicating a notable prevalence of SARS-CoV-2 in ocular tissue and emphasizing the importance of donor screening guidelines.63

Studies have also indicated that the virus infects the upper respiratory tract, and consequently coughing or sneezing may lead to contamination of the ocular surface.58 While such data points to the presence of virus in the corneal epithelium and ocular surface, there is no direct evidence proving transmission of the virus through corneal transplantation. Regardless, due to uncertainty regarding potential dangers of transmission, the EBAA and the Global Alliance of Eye Bank Associations (GAeba) have advised that donors who are PCR positive for SARS-CoV-2 be removed from the donor pool.55 The EBAA thereby excludes various groups of individuals...
Several studies have also examined the potential to reduce the expression of SARS-CoV-2 in corneal tissue. Sawant et al investigated the potential of povidone–iodine in the inactivation of SARS-CoV-2. They isolated the right eyes as the control group and the left eyes as the variable, utilizing a double-soak procedure with povidone–iodine. The team found that all left eyes produced negative results, whereby one of the right eyes produced a positive result. However, due to a small sample size of only 10 patients, the data could not determine a proper correlation. Thus, this study has many implications for future studies as povidone–iodine could prove to be a potential inactivator of COVID-19 if tested with a larger sample size. While povidone–iodine may be promising in reducing SARS-CoV-2 expression, other barriers pose significant challenges. Due to screening conditions and regulations within eye banks such as the EBAA, the donor eligibility review process is often delayed, leading to longer preservation times.

**Table 4 Eye Bank Association of America (EBAA) Guidelines for Evaluation of Viable Donors**

<table>
<thead>
<tr>
<th>Criteria</th>
<th>Recommendations for Corneal Viability of Donors with Potential COVID-19 Infection</th>
</tr>
</thead>
<tbody>
<tr>
<td>PCR Test</td>
<td>RT-PCR test must be performed either 24 hours after death or within 28 days prior to death. Donors should be excluded if the test is inconclusive.</td>
</tr>
<tr>
<td>Signs</td>
<td>If donor contracted any of the following signs within 28 days prior to death, donor cornea is not viable. The signs include: (1) acute respiratory distress syndrome (ARDS), (2) pneumonia, (3) detection of “ground glass opacities” via pulmonary computed tomography (PCT).</td>
</tr>
<tr>
<td>Symptoms</td>
<td>If donor had various symptoms consistent with criteria below within 28 days prior to death, donor cornea is not viable. One from the below: (1) Fever or chills, (2) difficulty breathing or shortness of breath, (3) Coughing, (4) Loss of taste or smell OR two from the below: (1) Headache, (2) Sore throat, (3) Congestion, (4) Nausea, (5) Fatigue, (6) Muscle Aches.</td>
</tr>
<tr>
<td>Contact</td>
<td>Donor being within six feet of a COVID-19 case or in direct contact with secretions from COVID-case.</td>
</tr>
<tr>
<td>Vaccination</td>
<td>At the time of death, donor would be considered as fully vaccinated under two conditions: (1) 2 weeks have passed since donor’s second dose in a 2-dose series such as the Pfizer or Moderna vaccines, (2) 2 weeks have passed after a single-dose vaccine such as Johnson &amp; Johnson’s Janssen vaccine.</td>
</tr>
</tbody>
</table>

Note: Data from Eye Bank Association of America (EBAA) (URL: https://restore.sight.org/covid-19-updates/). From the donor pool (Table 4). Several eye bank associations in various countries, such as the European Eye Bank Association (EEBA), have developed their own guidelines. The EEBA primarily requires a PCR test. If the PCR test is positive 14 days prior to death, the donor is ineligible. In addition, a nasopharyngeal swab postmortem is typically performed. However, individual risk assessment is necessary to determine the eligibility of the donor. It is important to note that postmortem swabs are yet to be validated. From the donor pool (Table 4). Several eye bank associations in various countries, such as the European Eye Bank Association (EEBA), have developed their own guidelines. The EEBA primarily requires a PCR test. If the PCR test is positive 14 days prior to death, the donor is ineligible. In addition, a nasopharyngeal swab postmortem is typically performed. However, individual risk assessment is necessary to determine the eligibility of the donor. It is important to note that postmortem swabs are yet to be validated. From the donor pool (Table 4). Several eye bank associations in various countries, such as the European Eye Bank Association (EEBA), have developed their own guidelines. The EEBA primarily requires a PCR test. If the PCR test is positive 14 days prior to death, the donor is ineligible. In addition, a nasopharyngeal swab postmortem is typically performed. However, individual risk assessment is necessary to determine the eligibility of the donor. It is important to note that postmortem swabs are yet to be validated.

**Future Guideline Changes**

Currently, the primary purpose of eye banks is storage, evaluation, and distribution of corneal tissues for transplantation. However, there has been a recent emerging focus on the storage of stromal lenticules. Corneal lenticules may be utilized for new indications such as presbyopia, corneal allogenic intrastromal ring segment implantation (CAIRS), hyperopic SMILE, myopic SMILE, tissue augmentation, and mechanical reinforcement for ectatic disorders.

In the aftermath of COVID-19, several changes in eye banking may be necessary. Ang et al reported that collaboration between eye bank technicians and surgeons must be prioritized. Furthermore, proper training methods are necessary for surgeons and technicians alike. Patient education is also crucial as information regarding risks of transmission must be conveyed clearly and effectively. Ang et al expressed the need to routinely evaluate donor viability criteria as the supply chain must continue.

Most importantly, the donor pool can be expanded through significant improvement of corneal evaluation methods and techniques. Furthermore, increasing the storage time of both full-thickness and corneal layers is an important next step in advancing corneal transplantation efficiency. Corneal tissue culture is cited as a promising approach to optimizing the function of corneal tissue such as the endothelium. Thus, eye banks may integrate corneal tissue culture into their infrastructure to optimize their operation. The pandemic has further given rise to the need for a new infrastructure upon which corneal transplantation can continue to restore vision.
Data Sharing Statement
Not applicable.

Code Availability
Not Applicable

Author Contributions
All named authors meet the International Committee of Medical Journal Editors criteria for authorship for this manuscript, take responsibility for the integrity of the work, and have given final approval to the version to be published. All authors contributed to drafting or revising the article, analyzing the literature, gave final approval of the version to be published, agreed to the selected journal and agree to be accountable for all aspects of the work.

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