

# 3D Multicellular Spheroid for the Study of Human Hematopoietic Stem Cells: Synergistic Effect Between Oxygen Levels, Mesenchymal Stromal Cells and Endothelial Cells

Emilia Barreto-Duran<sup>1</sup>Claudia Camila Mejia-Cruz<sup>1</sup>Luis Fernando Jaramillo-Garcia<sup>2</sup>Efrain Leal-Garcia<sup>3</sup>Alfonso Barreto-Prieto<sup>1</sup>Viviana Marcela Rodriguez-Pardo<sup>1</sup>

<sup>1</sup>Grupo de Inmunobiología y Biología Celular, Facultad de Ciencias, Pontificia Universidad Javeriana, Bogotá D.C., Colombia (South America);

<sup>2</sup>Departamento de Patología, Facultad de Medicina, Pontificia Universidad Javeriana, Hospital Universitario San Ignacio, Bogotá D.C., Colombia (South America); <sup>3</sup>Departamento de Ortopedia y Traumatología, Facultad de Medicina, Pontificia Universidad Javeriana, Hospital Universitario San Ignacio, Bogotá D.C., Colombia (South America)

**Introduction:** The human bone marrow microenvironment is composed of biological, chemical and physical factors that act in a synergistic way to modulate hematopoietic stem cell biology, such as mesenchymal stromal cells (MSCs), endothelial cells (ECs) and low oxygen levels; however, it is difficult to mimic this human microenvironment in vitro.

**Methods:** In this work, we developed 3D multicellular spheroid (3D-MS) for the study of human hematopoietic stem cells (HSCs) with some components of perivascular niche. HSCs were isolated from umbilical cord blood, MSCs were isolated from human bone marrow and a microvasculature EC line (CC-2811, Lonza®) was used. For the formation of a 3D structure, a magnetic levitation culture system was used. Cultures were maintained in 21%, 3% and 1% O<sub>2</sub> for 15 days. Culture volume, sphericity index and cell viability were determined. Also, human HSC proliferation, phenotype and production of reactive oxygen species were evaluated.

**Results:** After 15 days, 3D-MS exhibited viability greater than 80%. Histology results showed structures without necrotic centers, and higher cellular proliferation with 3% O<sub>2</sub>. An increase in the expression of the CD34 antigen and other hematopoietic antigens were observed to 1% O<sub>2</sub> with MSCs plus ECs and low ROS levels.

**Conclusion:** These findings suggest that 3D-MS formed by MSCs, ECs and HSCs exposed to low concentrations of oxygen (1–3% O<sub>2</sub>) modulate human HSC behavior and mimics some features of the perivascular niche, which could reduce the use of animal models and deepen the relationship between the microenvironment of HSC and human hematological diseases development.

**Keywords:** hematopoietic stem cells, niche, bone marrow, microenvironment, 3D culture

## Introduction

Hematopoiesis is a very active and efficient process that generates approximately  $4-5 \times 10^{11}$  hematopoietic cells per day.<sup>1</sup> This complex system is regulated by the interaction between hematopoietic stem cells (HSCs) and other cell populations that reside in the bone marrow (BM), soluble factors and physicochemical conditions such as oxygen perfusion.<sup>2</sup> HSCs are distributed in endosteal and perivascular niches<sup>3-5</sup>; however, recent evidence suggests that HSCs have a preferential location in the perivascular niche,<sup>6-8</sup> which are enriched with different populations of mesenchymal stromal cells (MSCs) and endothelial cells (ECs).<sup>9-12</sup>

Correspondence: Viviana Marcela Rodriguez-Pardo  
Email vivianar@javeriana.edu.co

Several studies have shown the close relationship between HSCs, MSCs and ECs and the influence of these relationships on hematopoiesis.<sup>13</sup> MSCs are the major source of SCF and SDF-1 (CXCL12)<sup>14</sup> in the bone marrow, which once deleted, greatly affect the quantity and engrafting capacity of HSCs.<sup>11,14,15</sup> It has also been shown that ECs regulate self-renewal, proliferation and phenotype modulation of HSCs through direct contact or soluble factors.<sup>16–20</sup>

Contrary to the findings about the murine HSC niche, information about the microenvironment of human HSCs is limited. It has been shown that 86% of human primitive CD34+ HSCs are located in the perivascular region close to CD271+/CD146+ MSCs,<sup>21,22</sup> which secrete factors that promote the maintenance of human HSCs and increase their ability to repopulate in vivo.<sup>23–25</sup> Furthermore, HSCs are particularly sensitive to low oxygen levels that induce intracellular signals that promote the maintenance of their self-renewal, stemness, phenotype modulation, glycolytic metabolism and low production of reactive oxygen species (ROS).<sup>7,26,27</sup> Recent evidence shows that the most primitive HSCs are located in the perivascular niches in close relationship with MSCs, ECs and low levels of oxygen.<sup>27,28</sup>

To establish an in vitro system that mimics the microenvironment of human HSCs, we previously established a 3D culture system using magnetic levitation for the study of the human HSC microenvironment.<sup>29</sup> In this work, we evaluated the synergistic effect between MSCs, endothelial cells and different levels of oxygen levels on the viability, proliferation, phenotype modulation and production of reactive oxygen species of human HSCs using 3D multicellular spheroid (3D-MS).

## Methods

### Generation of 3D Multicellular Spheroids

HSCs (CD34+) were isolated from umbilical cord blood (UCB) (Table 1), and MSCs were isolated from human bone marrow (hBM) (Table 2), after accepting the informed consent for the voluntary donation of UCB and hBM was conducted in accordance with the Declaration of Helsinki and approval of the ethics committee of Hospital Universitario San Ignacio, Act No 18 (2014/154). HSC and MSC isolation, expansion and characterization have been previously demonstrated by our group.<sup>29–34</sup> HMVEC-dBAd (CC-2811 Lonza®) cell line was expanded in endothelial cell growth medium

**Table 1** Umbilical Cord Blood (UCB) Donors for HSC Isolation

No.	Gestation Time (Weeks)	Volume UCB (mL)	CD34+ Cells (Total Counts)	CD34+ (Purity %)
1	38	34	$6.00 \times 10^5$	82
2	38	76	$3.20 \times 10^6$	97
3	39	16	$1.40 \times 10^6$	71
4	39	25	$6.00 \times 10^5$	60
5	37	31	$4.00 \times 10^5$	86
6	38	40	$1.00 \times 10^6$	96
7	38	36	$1.80 \times 10^6$	90
8	40	44	$2.00 \times 10^6$	97
9	40	52	$8.00 \times 10^5$	73
10	39	60	$1.30 \times 10^6$	71
11	38	61	$1.90 \times 10^6$	87
12	39	31	$1.10 \times 10^6$	94
13	38	64	$1.66 \times 10^6$	97
14	37	42	$4.00 \times 10^6$	93
15	38	17	$5.00 \times 10^6$	95
16	39	45	$1.10 \times 10^6$	98
17	40	55	$2.00 \times 10^6$	79
18	38	47	$9.00 \times 10^5$	85
19	39	75	$1.00 \times 10^6$	90
20	39	24	$8.00 \times 10^5$	89

**Table 2** Bone Marrow Donors for MSC Isolation and Expansion

No.	Age Donors (Years)	Gender (Female / Male)	Volume BM (mL)	MNC (Total Counts)
1	56	F	27	$1.16 \times 10^8$
2	59	F	40	$4.81 \times 10^7$
3	53	F	20	$4.58 \times 10^7$
4	75	F	30	$2.18 \times 10^7$
5	35	F	41	$3.10 \times 10^7$
6	60	F	20	$9.40 \times 10^6$

(EGMTM-2 BulletKit™, Lonza®). HpDLF (fibroblast, CC-7049 Lonza®) cell line was expanded in modified Eagle's medium (Merck®), and this was used as a negative control. The 3D structures were generated with a magnetic levitation system containing MSCs, HSCs and ECs (ratio 1:2:2).<sup>29</sup> 3D structure control groups were generated: fibroblasts, HSCs and ECs (Fb-HSC-ECs), and MSCs-HSCs. 3D-MS were cultured at three O<sub>2</sub> concentrations (1%, 3% and 21%) for 15 days. For hypoxic conditions, 3D-MS were cultured in a hypoxia chamber (chamber C-Chamber Incubator Subchamber, BioSpherix ©) using a mixture of gases (O<sub>2</sub>, N<sub>2</sub> and CO<sub>2</sub>).

## Morphological Evaluation

Volume and sphericity index (SI) were performed according to Kelm et al,<sup>35</sup> using ImageJ program (Fiji, version 1.0) to calculate the area and perimeter from the photographs obtained from each of the 3D structures on day 5 ( $n = 88$ ), 10 ( $n = 72$ ) and 15 ( $n = 48$ ).

## Histological and Immunohistochemical (IHQ) Analysis

Pathology Department at the Hospital Universitario San Ignacio (HUSI) (Bogotá, Colombia) supported the development of histochemical tests. 3D-MS were fixed with formaldehyde, dehydrated with ethanol, cleared with xylenes (Thermo Scientific™ Excelsior™ AS Tissue Processor), embedded in paraffin (Thermo Scientific™ HistoStar™ Embedding Workstation) and sectioned into 3  $\mu\text{m}$  thick sections with a microtome (Leica® RM2125 RTS). Serial slides of 3D structures sections were then processed with hematoxylin and eosin (H&E), anti-Ki67 (Flex Ki-67 antigen, MxH/MIB-1, Dako®) and anti-vimentin (Flex vimentinMx/V9, Dako®).

## Cell Viability and Hematopoietic Antigens Evaluation

LIVE/DEAD® Cell Imaging Kit (R37601 Thermo Scientific®) was used following the manufacturer's instructions. This viability staining was performed on the complete and living 3D structure. For hematopoietic antigens, 3D-MS serial slides were incubated at room temperature with primary antibodies (Anti-CD34 APC clone 581 Thermo Scientific®, anti-Ki67 clone 4A1 Thermo Scientific®, anti-CD133 clone AC133 Miltenyi Biotec®, anti-CD33 clone WM53 Thermo Scientific® and anti-CD7 clone MEM-186, Thermo Scientific®). Alexa Fluor 514 goat anti-mouse IgG (Thermo Scientific®) was used as a secondary antibody for CD133, CD33 or CD7 detection. Anti- $\alpha$ -tubulin (Alexa Fluor 488 clone B512, Invitrogen®) was used for cytoskeletal labeling, and DAPI (4',6-diamidino-2-phenylindole, ref D1306, Invitrogen®) was used for nuclei identification. Slide-mounted 3D structure sections were sealed with 10  $\mu\text{L}$  of ProLong Gold antifade reagent (Invitrogen®) and covered with coverslips.

## Reactive Oxygen Species (ROS) Evaluation

3D structure sections were treated with CellROX™ Deep Red Reagent (Thermo Scientific®) at room temperature.

The cells were fixed with 3.7% formaldehyde, permeabilized with 0.1% Triton X-100, and stained with anti- $\alpha$ -tubulin and DAPI following the protocol mentioned above.

## Image Acquisition and Analysis Strategy for Confocal Microscopy

An Olympus FV1000 confocal microscope with 488-, 543-, 635- and 405-nm lasers was used to collect images of sphere and sphere sections. Viability imaging and ROS imaging was performed using a UPLFLN 20X objective with a numerical aperture (NA) of 0.50 every 5  $\mu\text{m}$  to generate reconstructions in XYZ planes. Between 25 and 30 images were obtained to assess viability (complete spheres), and 15 to 20 images were obtained to assess ROS (sphere sections). For proliferation and hematopoietic phenotype modulation, Z stack images of sphere sections were collected with UPLSAPO 60X and UPLFLN 20X objectives using an NA of 1.35 in the XY focal plane. 3D reconstruction and data analysis were performed with FlowView (Olympus®) and Fiji 1.0, respectively. For the evaluation of viability, the percentage of green area (living cells) and red area (dead cells) was calculated in separate channels using the “threshold” and “analyze particles” tools to select the estimated area for each color. Hematopoietic antigens and ROS were calculated using the following equation: antigen expression area % =  $\frac{\text{antigen expression area}(\mu\text{m}^2)}{\alpha\text{-tubulin expression area}(\mu\text{m}^2)} \times 100\%$

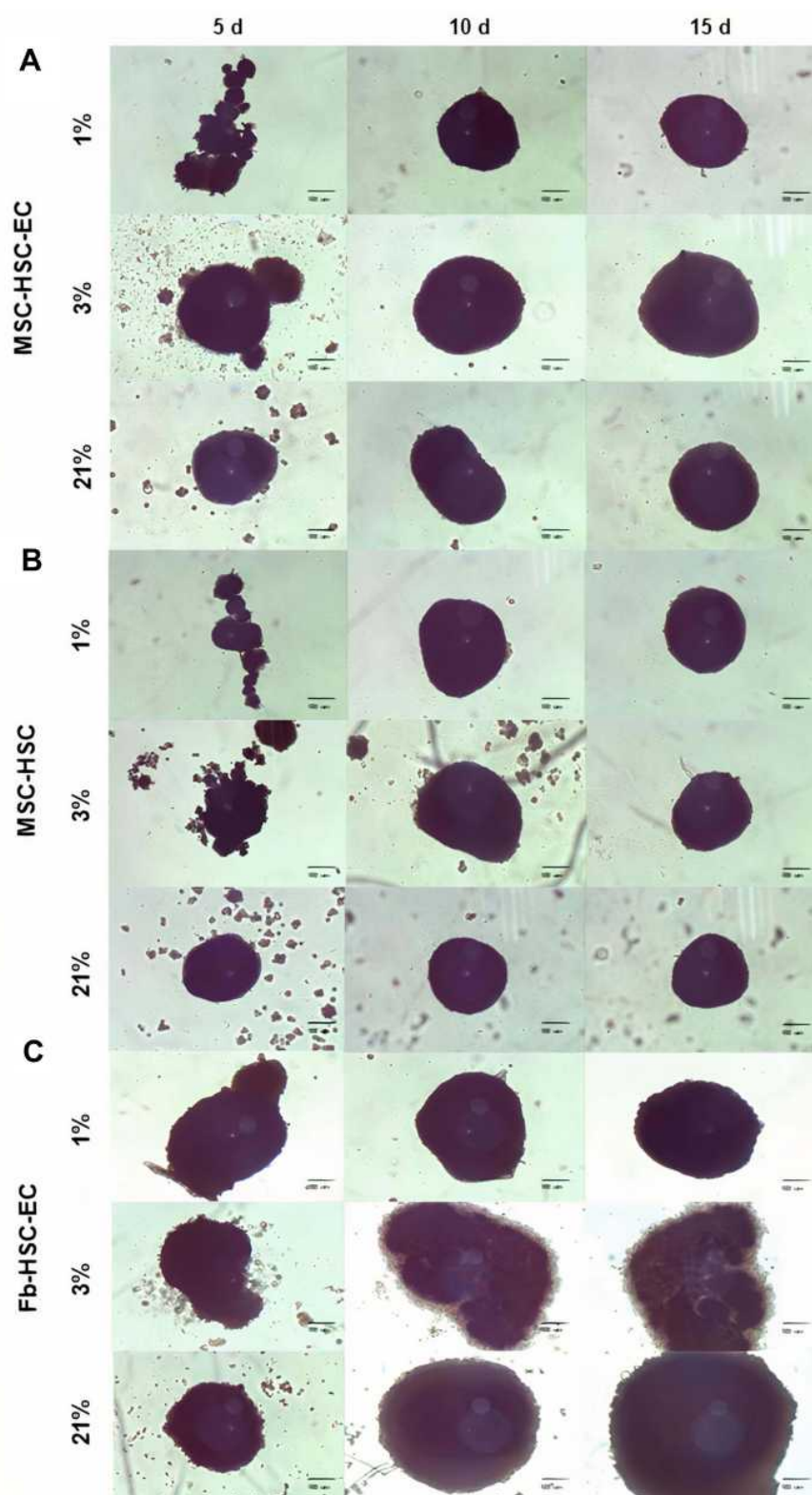
## Statistical Analysis

Shapiro–Wilk tests were applied to determine the normality of the data. The comparisons between two samples were made using Wilcoxon tests. For the comparison of more than three groups of data, Kruskal–Wallis (KW) tests, Friedman test with Dunn's test was used. All the data were analyzed and plotted with GraphPad Prism 6™ software. \*  $p = 0.05$ , \*\*  $p = 0.01$ , and \*\*\*  $p = 0.001$ .

## Results

### Structure and Viability of 3D Cultures

After 15 days in culture, homogeneous 3D structures were obtained under the three oxygen conditions; mesenchymal cells control formed by fibroblasts, hematopoietic cells and endothelial cells reached a larger size (Fb-HSC-EC) (Figure 1). The volumes of MSC-HSC-EC were between 0.1 and 0.5  $\text{mm}^3$  in all culture conditions and the sphericity index (SI) was between 0.6 and 0.8 during 15 days. Fb-HSC-EC 3D



**Figure 1** Structure of 3D cultures. Structures at days 5, 10 and 15 in three O<sub>2</sub> conditions (1%, 3% and 21% O<sub>2</sub> levels). **(A)** Experimental condition: MSC-HSC-EC, **(B)** Endothelial cells control: MSC-HSC, **(C)** Mesenchymal cells control: Fb-HSC-EC. (Olympus® CKX31, 10×) (scale bar: 100 μm).



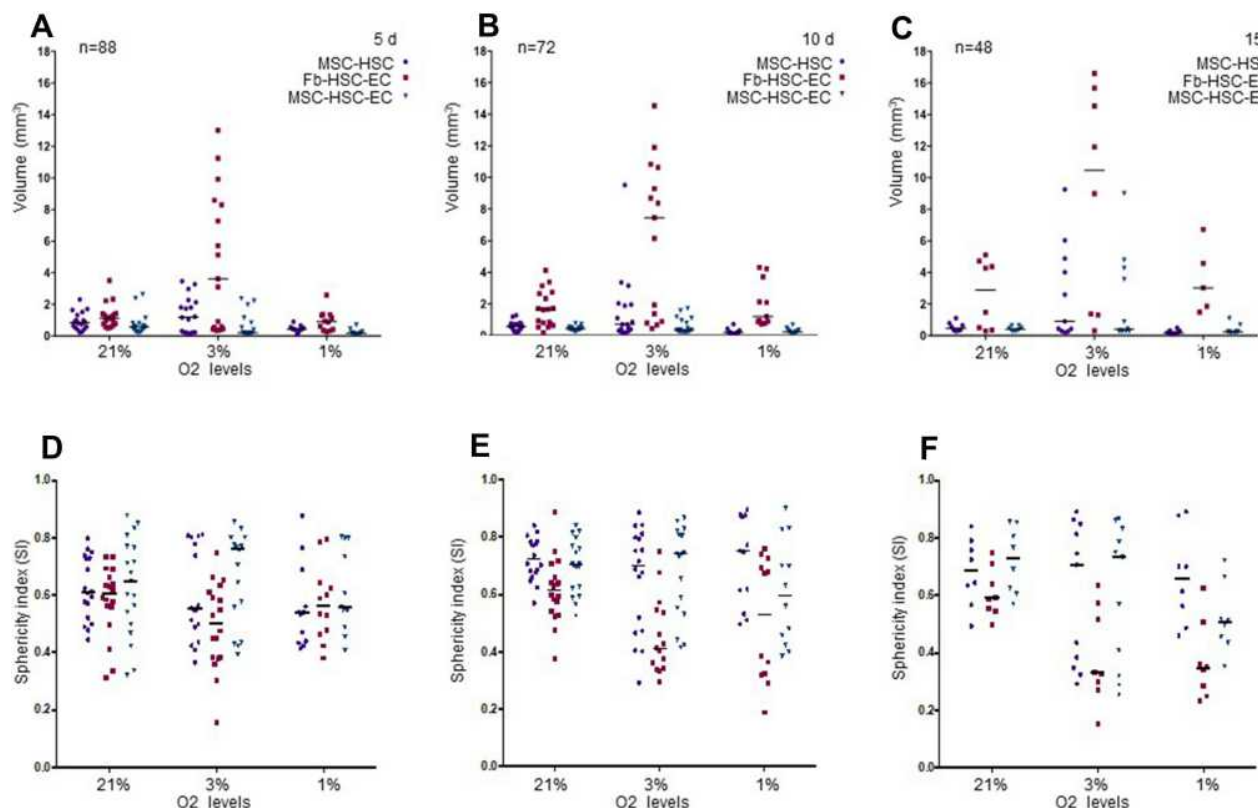
structures had an approximate volume between 1.7 and 7.5 mm<sup>3</sup> at day 10 of culture and between 3.0 and 10.5 mm<sup>3</sup> after 15 days, with a sphericity between 0.4 and 0.8. Some variations in size and sphericity were observed between oxygen culture concentrations, although significant differences were observed in the volume of the 3D structure between the MSC-HSC-EC condition versus controls at day 5 (Figure 2). All evaluated 3D structures had a viability >80%, and no significant differences were found in cell viability between experimental organoid prototype and controls ( $p > 0.9999$ ) (Figure 3).

## Histology and Immunohistochemistry

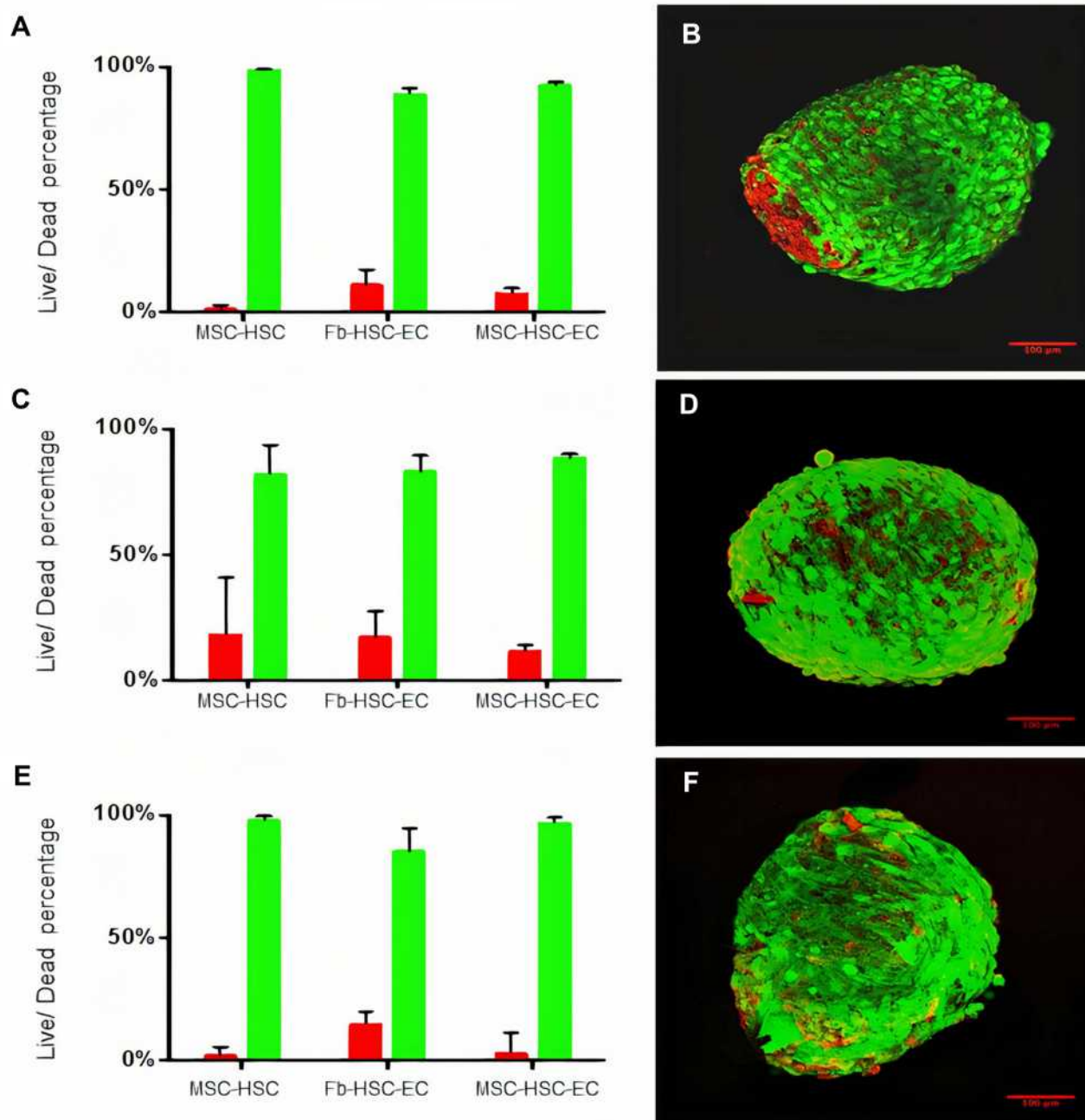
H&E staining demonstrated that all evaluated 3D-MS showed a cellular organization similar to that found in tissue (Figure 4A). Brown areas were accumulations of metallic nanoparticles in the extracellular matrix of the structure. There was no evidence of a necrotic center in 3D structures (Figure 4A). Vimentin expression was strongly positive compared to controls (Figure 4B–D).

## Evaluation of Proliferation and Phenotype Modulation of Human HSC in 3D Structures

A lower expression of Ki67 was observed in cultures with 1% O<sub>2</sub> but Ki67 expression by immunohistochemistry was not conclusive because the nanoparticles did not allow the protein to be visualized in the nucleus (Figure 5A–D). Global expression of Ki67 by confocal microscopy allowed to demonstrate significant differences between the oxygen conditions, especially in MSC-HSC-EC structure exposed to the lowest levels (1–3%) (Figure 5E and F). After 15 days, MSC-HSC-EC structure showed a higher expression of CD34 antigen with 1% O<sub>2</sub> in comparison with 21% O<sub>2</sub> (Figure 6A and B). For the CD133 antigen there was a higher expression at 3% and 1% O<sub>2</sub> than there was at 21%, and it was generally shown that there was higher expression in experimental MSC-HSC-EC than there was in the controls (Figure 6C and D). Regarding the expression of CD33, there was a statistically significant difference in the MSC-HSC-EC structures maintained at 21% and 1% O<sub>2</sub> (Figure 6E and



**Figure 2** Volume and sphericity index (SI) of 3D cultures. (A–C) Volume after 5, 10 and 15 days. Friedman tests were performed to compare O<sub>2</sub> concentrations: 5, 10 and 15 days. Friedman tests were performed to compare MSC-HSC-EC structures and controls: 5 days ( $p=0.0278$ ), 10 days and 15 days ( $p>0.05$ ). (D–F) Sphericity Index (SI) after 5, 10 and 15 days. Friedman tests were performed to compare O<sub>2</sub> concentrations and MSC-HSC-EC structures vs controls after 5, 10 and 5 days ( $p>0.05$ ).



**Figure 3** Viability of 3D cultures. (A and B) Cell viability in 1% O<sub>2</sub>. (C and D) Cell viability in 3% O<sub>2</sub>. (E and F) Cell viability 21% O<sub>2</sub>. (B, D, F) LIVE/DEAD® viability kit. Red: dead cells, Green: live cells. Olympus confocal microscope FV1000, 20× (scale bar: 100μm).

F). For the CD7 antigen, there tended to be greater expression at 3% and 1% O<sub>2</sub>, with statistically significant differences in the MSC-HSC-EC structure between 21% and 1% O<sub>2</sub> (Figure 6G and H).

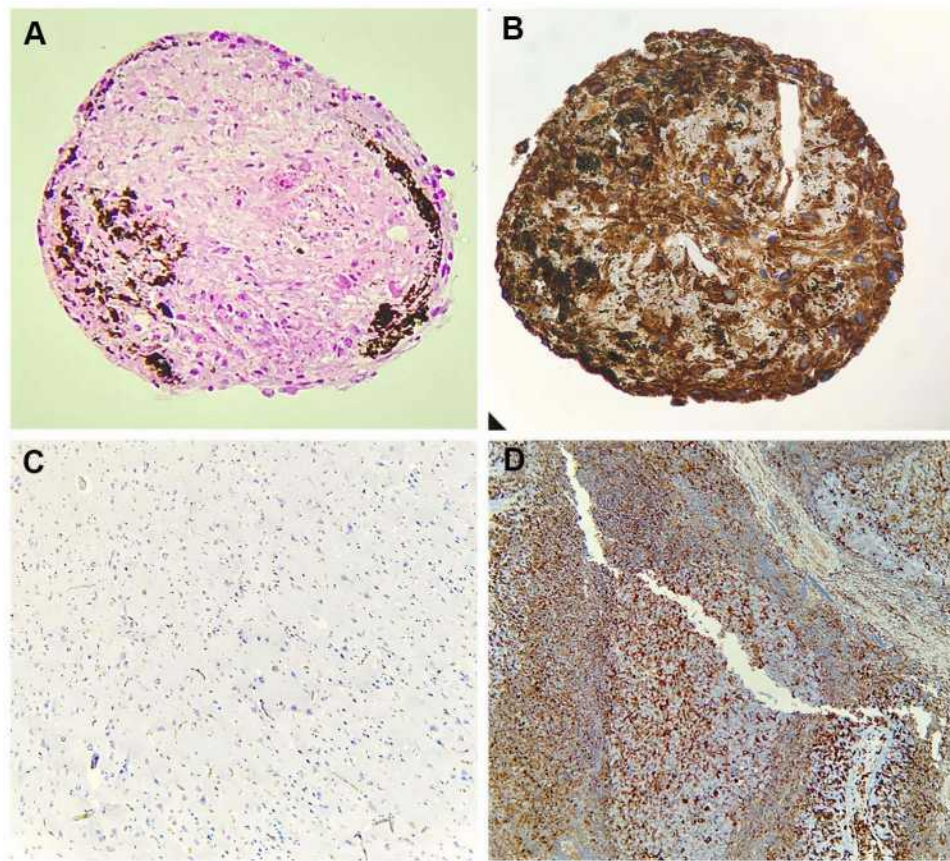
## Reactive Oxygen Species

No statistically significant differences were found in ROS production between experimental and control 3D-MS.

Nevertheless, although no statistically significant differences were found in ROS production with different oxygen conditions ( $p = 0.1944$ , Friedman test), a lower production was observed in 3D-MS with 1% O<sub>2</sub> (Figure 7).

## Discussion and Conclusions

The niche of human HSCs in bone marrow is influenced by different cell populations; however, the perivascular niche



**Figure 4** Histological characterization of 3D structures. (A) Hematoxylin and eosin stain (20 $\times$ , Olympus<sup>®</sup> CX21), (B) Vimentin expression of MSC-HSC-EC structures after 15d with 3% O<sub>2</sub> (20 $\times$ , Olympus<sup>®</sup> CX21), (C) Negative control for vimentin (brain tissue) (10 $\times$ , Olympus<sup>®</sup> CX21), (D) Positive control for vimentin (lymph node tissue) (10 $\times$ , Olympus<sup>®</sup> CX21).

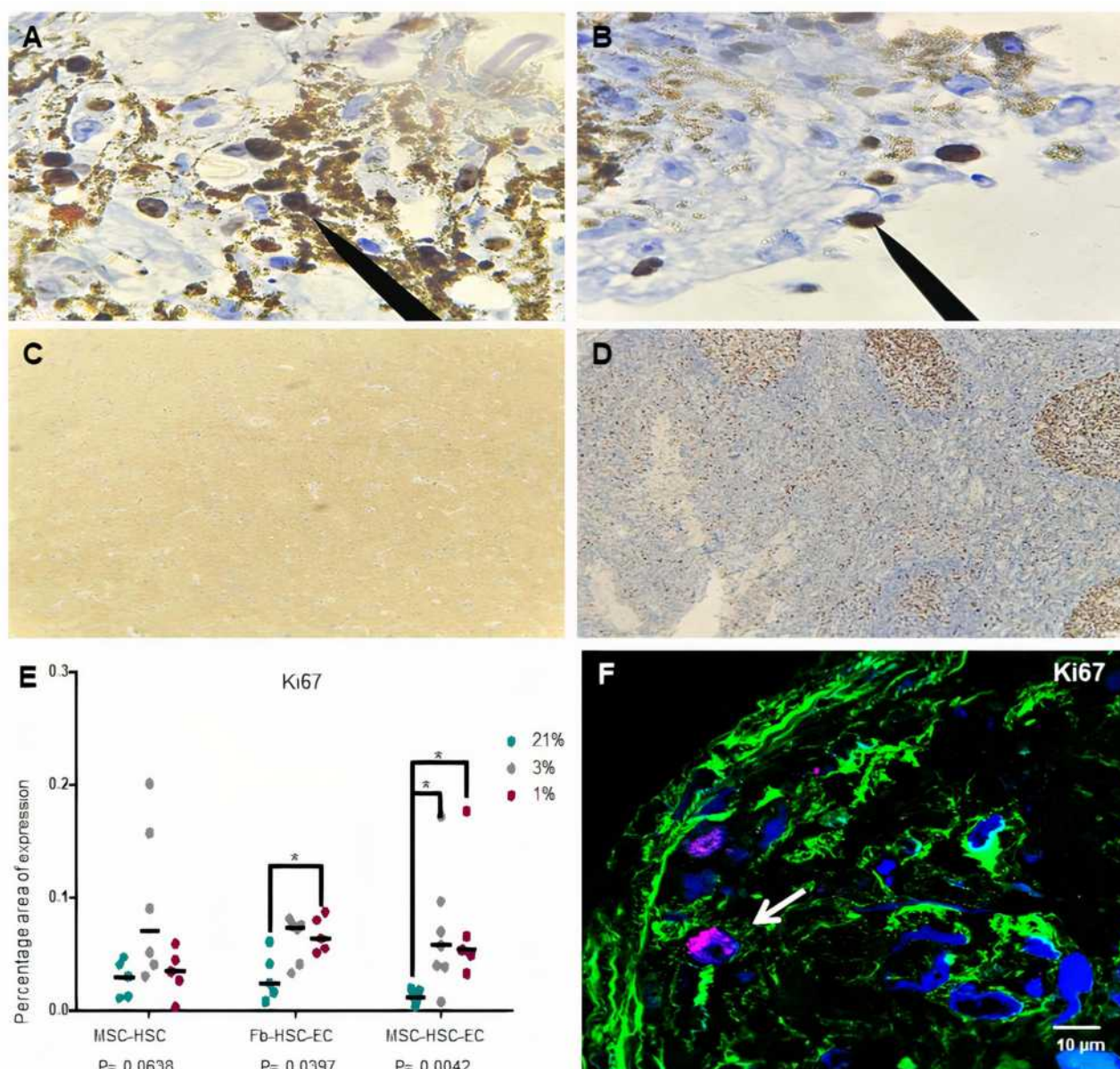
(MSC and ECs) is increasingly important given the different phenomena that occur there and that are related to functional hematopoiesis.<sup>36,37</sup> In this work, we used the synergistic relationship between MSCs and ECs with different concentrations of O<sub>2</sub> on human HSCs in 3D-MS,<sup>38</sup> enabling the *in vitro* study of human HSC microenvironment.

In relation to the structure of the 3D-MS obtained, it has been previously shown that under hypoxic conditions (2–3% O<sub>2</sub>), various cell populations could increase the expression of adhesion molecules.<sup>39,40</sup> The MSC-HSC-EC structures showed a similar sphericity index (0.6–0.8) after 5, 10 and 15 days of culture at all oxygen concentrations; however, in the controls, the sphericity index fluctuated according to the culture time and O<sub>2</sub> concentration (Figure 2). These results could possibly be related to a synergistic effect between oxygen levels and the interaction between MSCs, HSCs and ECs on the expression of adhesion molecules that is not observed in the condition with Fb or without ECs; however, addressing this possibility requires the determination of adhesion molecules that act in the 3D structure.

Interestingly, and contrary to other models of 3D cultures, we do not observe a necrotic center inside the structures. Necrotic centers in these types of structures appear in spheres with volume between  $0.112 \pm 0.013 \text{ mm}^3$ <sup>341,42</sup>; however, the average volume of the 3D-MS generated in this work was between 0.1 and 0.5 mm<sup>3</sup> (MSC-HSC-CE). We propose that the absence of necrotic centers in the obtained spheres is possibly related to the secretion of soluble factors of the three cell populations and is promoted by their cell–cell interaction, which maintains the internal viability of the 3D structure and high expression of vimentin in all experimental conditions (Figures 3 and 4).

In this work, we demonstrate the effect of physiological oxygen levels of the human bone marrow (1% and 4%),<sup>27</sup> which some call hypoxia, on the proliferation and modulation of the human HSC phenotype. Statistically significant differences were observed in Ki67 expression levels in the different experimental conditions and controls (confocal microscopy) (Figure 5E and F) with greater expression of Ki67 at 1% and 3% O<sub>2</sub> in comparison with 21%.





**Figure 5** Ki67 expression in 3D structures. (A and B) Ki67 expression of MSC-HSC-EC structures after 15 days and 1% O<sub>2</sub> by immunohistochemistry (arrows) (100×, Olympus® CX21), (C) Ki67 negative control by immunohistochemistry (brain tissue) (10×, Olympus® CX21), (D) Ki67 positive control by immunohistochemistry (lymph node tissue) (10×, Olympus® CX21), (E) Ki67 expression by confocal microscopy (percentage area of expression), (F) Paraffin section (3 μm thick) of MSC-HSC-EC structure analyzed by confocal microscopy (Magenta: Ki67, green: α-tubulin, and blue: DAPI-nuclei) (40×, Olympus confocal microscope FV1000).

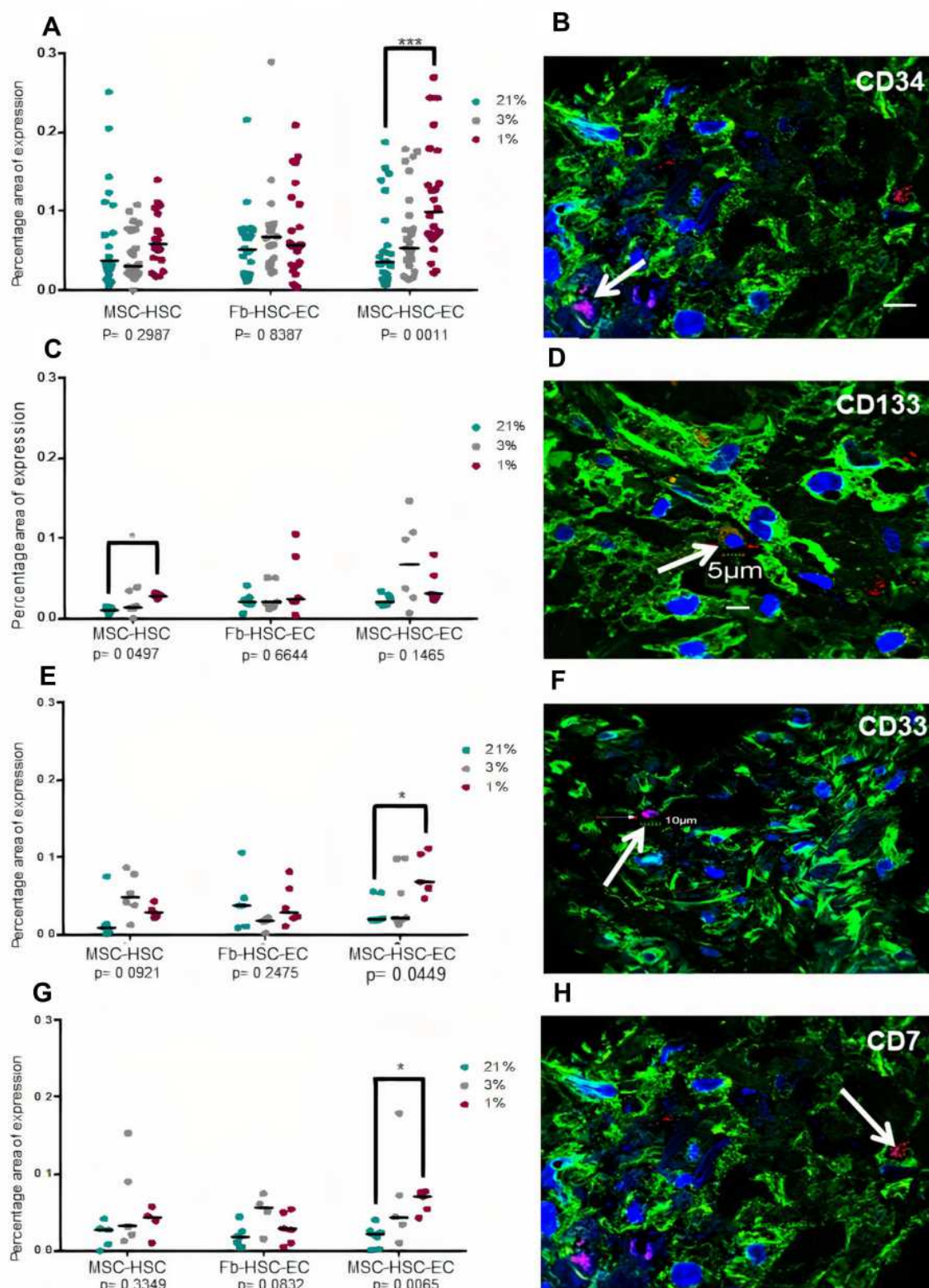
Interestingly, we show that oxygen levels between 1% and 3% increase the expression of CD34, CD133, CD33 and CD7 over the levels observed in conventional culture conditions (21% oxygen), which suggests synergy with MSCs and ECs, given that this increase was greater in the MSC-HSC-EC condition (experimental 3D-MS) (Figure 6).

Previous studies have shown that the increase in the expression of CD34 and CD133 in human HSCs is associated with a greater capacity of repopulation of these cells in models of immunodeficient mice.<sup>43,44</sup> Similarly,

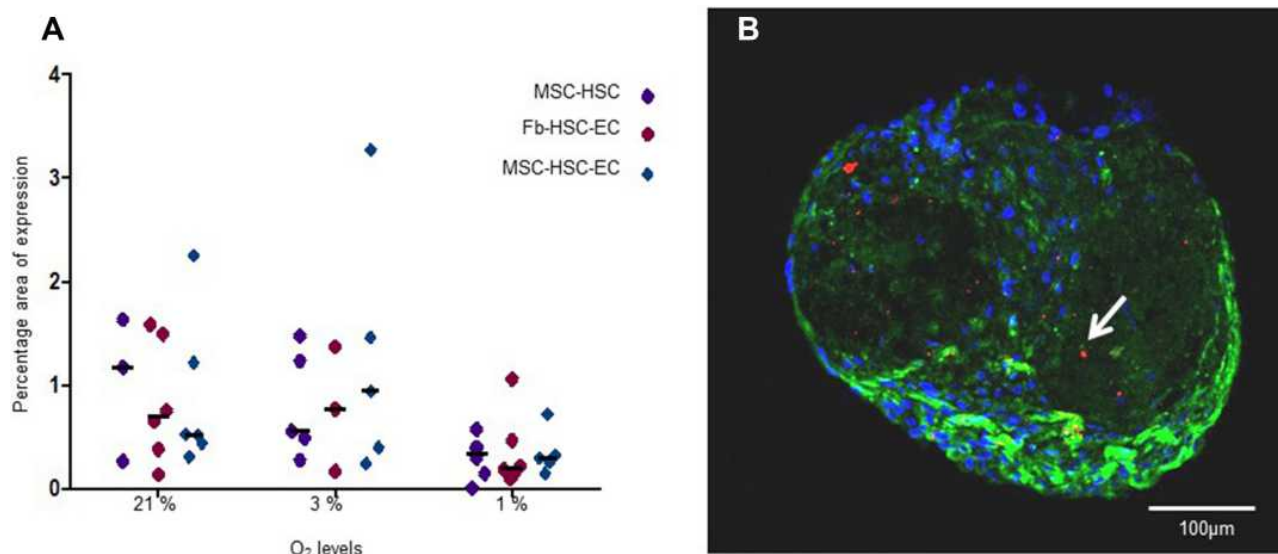
although the expression of CD33 and CD7 is associated with myeloid and lymphoid progenitors, respectively, it has been shown that the expression of CD33 in human HSCs from umbilical cord blood is also associated with long-term culture capacity (LTC-IC).<sup>45,46</sup>

We propose in this work that the 3D-MS formed by MSCs, ECs and HSCs exposed to low concentrations of oxygen (1–3% O<sub>2</sub>) modulate human HSC expression of hematopoietic antigens associated with the engraftment and stemness capacity in an environment with low ROS





**Figure 6** Hematopoietic antigens expression in 3D structures. (A and B) CD34 antigen, (C and D) CD133 antigen, (E and F) CD33 antigen, (G and H) CD7. (B, D, F, H) Paraffin section (3 μm thick) of MSC-HSC-EC structure analyzed by confocal microscopy after 15 days. Magenta or red: hematopoietic antigen (arrows), green: α-tubulin, and blue: DAPI-nuclei (40X, Olympus confocal microscope FV1000).



**Figure 7** ROS levels in 3D structures. **(A)** ROS expression area in structures cultivated in 1%, 3% and 21% O<sub>2</sub>. Differences between MSC-HSC-EC and controls,  $P > 0.9999$ , as determined by Friedman tests; differences between O<sub>2</sub> concentrations,  $P = 0.1944$ , as determined by Friedman tests. **(B)** Paraffin section (3 μm thick) of MSC-HSC-EC culture stained with CellROX<sup>®</sup>. Green: α-Tubulin, blue: DAPI (nuclei), and red: ROS (20×, Olympus confocal microscope FV1000).

levels (Figure 7), and although ROS levels were detected globally and statistically significant differences were not determined, a lower production was observed in 3D-MS with 1% O<sub>2</sub>, it has been described that this “hypoxic” environment favors the maintenance of the long-term primitive human HSC pool and mimics some features of the perivascular niche of human HSCs.

The development of human 3D-MS that mimic the microenvironment of HSCs has great potential because they may allow the testing of new drugs for the treatment of diseases such as leukemia, reduce the use of animal models, and deepen our understanding of the relationship between the microenvironment of the HSC and the evolution of different human hematological diseases.

## Abbreviations

3D-MS, 3D multicellular spheroid, HSC, hematopoietic stem cells, MSCs, mesenchymal stromal cells; ECs, endothelial cells.

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## Disclosure

The authors declare no conflicts of interest.

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