REVIEW

The Agonists of Peroxisome Proliferator-Activated Receptor-γ for Liver Fibrosis

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Abstract: Liver fibrosis is a common link in the transformation of acute and chronic liver diseases to cirrhosis. It is of great clinical significance to study the factors associated with the induction of liver fibrosis and elucidate the method of reversal. Peroxisome proliferator-activated receptors (PPARs) are a class of nuclear transcription factors that can be activated by peroxisome proliferators. PPARs play an important role in fibrosis of various organs, especially the liver, by regulating downstream targeted pathways, such as TGF-β, MAPKs, and NF-κB p65. In recent years, the development and screening of PPAR-γ ligands have become a focus of research. The PPAR-γ ligands include synthetic hypolipidemic and antidiabetic drugs. In addition, microRNAs, lncRNAs, circRNAs and nano new drugs have attracted research interest. In this paper, the research progress of PPAR-γ in the pathogenesis and treatment of liver fibrosis was discussed based on the relevant literature in recent years.
Keywords: liver fibrosis, PPAR-γ, HSCs, agonists

Introduction

The liver is the main metabolic organ in the body and has a strong regeneration ability. If the liver is stimulated by long-term injury and chronic inflammation, liver fibrosis will develop. In the liver fibrosis in rodent and human livers, the liver can reverse to normal function if the main chronic inflammation or injury trigger is eliminated. This conclusion has been confirmed in many studies.[1,2] These measures include lifestyle, interference, inhibition of hepatitis virus, and other etiological treatment. At the same time, liver fibrosis is accompanied by the activation and inhibition of a variety of pathways, such as the TGF-β/Smad3, NF-κB, PPARs, MAPK, NLRP3, hedgehog, Wnt/β-catenin, and Notch signaling.[1,3] Therefore, an in-depth study of the signal pathway of liver fibrosis can provide ideas for prevention and treatment.[4]

Peroxisome proliferator-activated receptors (PPARs) have a variety of biological effects and are closely related to adipocyte differentiation, glucose and lipid metabolism, carcinogenesis, atherosclerosis, and inflammatory response.[5,6] PPARs contains three subtypes (PPAR-α, PPAR-β/δ, and PPAR-γ) among them, which PPAR-γ has undergone the most in-depth research.[7–9] Studies have shown that PPAR-γ improves hepatic steatosis, reduces inflammation and fibrosis, and significantly increases the response rate of hepatitis virus, which plays an essential role in various acute liver injuries and liver cancer.[10–12] The pathologic process underlying of PPAR-γ is complex that involves the activation of HSCs, ECM deposition and other signaling pathways. In recent years, other research interests including

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intestinal flora and epigenetic modification play an important role in the reversal of liver fibrosis. Therefore, it is of great clinical significance to study drugs and mechanisms targeting PPAR-γ, which may be a target in the treatment of liver fibrosis.

Structure, Function, and Biological Activity of PPARs

PPARs belong to the hormone nuclear receptor superfamily and are a class of intranuclear receptors activated by peroxisome proliferators. In 1990, PPARs were successfully cloned in mouse liver by Issemann and colleagues.[13] To date, three subtypes of PPARs (PPAR-α, PPAR-β/δ, and PPAR-γ) have been found to be co-expressed in most tissues, but the tissue distribution is quite different. PPAR-γ is on 3p25 and has nine exons. Four kinds of mRNA isomers are produced by selective splicing (PPAR-γ1, PPAR-γ2, PPAR-γ3, and PPAR-γ4).[14] PPAR-γ is mainly distributed in adipose tissue, but is also found the colon, immune system, and retina. PPAR-γ plays a variety of biological functions, including regulating fat and glucose metabolism, inducing tumor cell differentiation and apoptosis, promoting ovulation and anti-atherosclerotic activity, improving heart failure and ventricular remodeling, and inhibiting inflammatory reactions.[15–18]

PPAR-γ-Related Ligands and Mechanism of Action

PPAR-γ plays an important role by binding and activating ligands, including natural and synthetic ligands. Natural ligands mainly include arachidonic acid and its metabolites, polyunsaturated fatty acid oxidation metabolites and prostaglandin metabolites. The synthetic ligands mainly include thiazolidinediones (troglitazone, rosiglitazone, and pioglitazone), nonsteroidal anti-inflammatory drugs, and leukotriene D4 receptor antagonists.[19] The N-terminal domain of PPAR-γ contains a protein kinase phosphorylation site that can be activated by mitogens. PPAR-γ, after being activated by ligands, forms a heterodimer with the retinoic acid receptor (RXR), which then binds with the peroxisome proliferator response element (PPRE) of the target gene promoter to activate target gene transcription of factors that have a variety of biological functions.[20,21] The current study also demonstrated that a variety of traditional Chinese medicine monomer extracts, which play an important role in PPARs.[22]

Mechanisms Underlying Hepatic Fibrosis and PPAR-γ

The pathologic process that characterizes liver fibrosis is complex, involving the activation and inhibition of many signaling pathways, from resting hepatic stellate cells (HSC) activation, extracellular matrix (ECM) deposition, and oxidative stress, to lipid metabolism and immune responses. In recent years, there are other research interests, including the role of intestinal flora, the establishment of anaerobic pro-inflammatory microenvironment in tissues, and epigenetic modification in the progress of fibrosis.[1,23] Intervention in any of these processes, many of which involve PPAR-γ, will play a positive role in the reversal of liver fibrosis.

Activation of HSCs

Hepatic fibrosis is the repair response of the liver to various chronic injury stimuli. The main feature of hepatic fibrosis is the continuous generation of ECM exceeding degradation, and the deposition of ECM in the liver develops into irreversible lesions. HSCs are the main source of the ECM and the central link in the occurrence and development of liver fibrosis.[24] Under normal conditions, HSCs account for 5%–15% of all the hepatic cells in the liver and are in a resting state. HSCs are responsible for the metabolism and storage of vitamin A and fat, and the synthesis and secretion of collagen, glycoproteins, MMPs, and TIMPs. When stimulated by a viral infection, excessive alcohol intake, toxic substances, or other detrimental factors, morphologic changes occur and HSCs will transform into myofibroblasts, which express the main components of α-SMA and the ECM, and promote the formation of liver fibrosis.[25] Recent studies have shown that PPAR-γ and GATA6 are necessary for inactivation of human HSCs and regression of liver fibrosis in mice.[26,27] PPAR-γ inhibits HSC activation, proliferation, and increases ECM production.[28,29] Wang and colleagues reported that PPAR-γ is also expressed in normal HSCs, while PPAR-γ expression gradually decreases with increased HSC activation stimulated by inflammation or injury, indicating that PPAR-γ inhibits HSC activation and maintains its quiescence.[30] Yavrom upregulated PPAR-γ expression by transfecting HSCs with PPAR-γ adenovirus vector, which confirmed that the translocation reduced the activation of HSCs; the decrease of type I collagen in veins was the most significant finding.[31] Yu and colleagues confirmed that PPAR-γ is expressed in primary resting

HSCs, but is absent in cultured activated HSCs. In contrast, the ectopic expression of PPAR-γ in activated HSCs reversed the phenotype of resting cells.[32] Therefore, PPAR-γ involvement in attenuating HSC activation is an important target whether or not liver fibrosis can be reversed. In addition, Zhang et al [reference #?] believed that platelet-derived growth factor (PDGF) receptor-γ inhibits angiogenesis signal transduction through trans-expression of PDGF-β receptor, resulting in decreased motility of HSCs and decreased expression of vascular endothelial growth factor, thus inhibiting angiogenesis induced by HSCs. Therefore, PPAR-γ can also be used as a molecular target for the prevention of hepatic fibrosis and vascular remodeling.[33]

Inflammation and Oxidative Stress

Oxidative stress and inflammation play an important role in the pathogenesis of the initiation and progression of many liver diseases.[34,35] ROS promotes the expression of IL-6 and IL-1β by stimulating intracellular signal cascades, such as NF-κB. In contrast, inflammatory cells, such as monocytes and lymphocytes, also produce additional ROS after activation.[36,37] PPAR-γ is a regulator of cell proliferation and immune responses, and can regulate the expression of related genes and participate in the inflammatory response. It has also been confirmed that PPAR-γ plays a strong role in inhibiting liver fibrosis by cross-talk with inflammatory-related factors, such as NF-κB and AP-1, inhibiting T cell activation and macrophage proliferation and secretion.[4,28,38,39]

Lipid Metabolism

The liver is an important site for fat metabolism, and plays an important role in fat digestion, absorption, synthesis, and transportation. Liver lipid peroxidation plays a key role in the pathological process of liver fibrosis. Free fatty acids induce high expression of CYP2E1 (the main component of the cytochrome P450 system) and enhance lipid peroxidation in fatty liver. These processes involve chemotactic neutrophils, induction of autoimmune responses, and direct activation hepatic macrophages and HSCs.[1,25] Fatty cirrhosis has a common histologic feature. That is, the cause of steatosis of liver cells is repeated attacks of the same cause. The first attack causes steatosis of liver cells, thus causing fatty liver. The second attack causes lipid peroxidation, leading to steatohepatitis and liver fibrosis. PPAR-γ is highly expressed in adipocytes and is a major regulator of adipogenesis.[14] PPAR-γ can activate the expression of phosphoenolpyruvate carboxy-kinase and glycerol kinase via transcription, which then participate in the production of triglycerides through different pathways. PPAR-γ regulates the glucose content under the action of lipoprotein lipase, then affects the final synthesis of triglycerides by affecting 3-phosphogly-cerate. Studies have confirmed that rosiglitazone can reduce liver lipid accumulation and peroxidation in patients with liver fibrosis, which facilitates the reversal of NASH.[26] The PPAR-α/γ agonist, saroglitazar, improves insulin resistance and steatohepatitis in a diet-induced NAFLD animal model.[40] Indeed, GDF10 blocks PPAR-γ activation in the liver and protects the liver from diet-induced liver injury.[41] These results suggest that PPAR-γ plays an effective role in anti-fibrosis by participating in lipid metabolism.

Immune Response

The regulatory function of immune cells plays an important role in the formation of liver fibrosis, including non-specific and specific immune cells. The non-specific immune cells mainly include macrophages, natural killer (NK) cells, dendritic cells, and other non-specific cells. Among the cell types, macrophages play an important role in the two-way regulation of liver fibrosis, while NK cells have a wide range of pathogen and activated cell clearance functions.[42,43] The specific immune cells are mainly T cells, which play an important role in the occurrence of anti-infective immunity, anti-tumor immunity, and autoimmune diseases. T cells cooperate with IL-17 to inhibit liver inflammation, but whether T cells can block the formation and mechanism underlying liver fibrosis remains to be studied.[44,45] Studies have confirmed that PPAR-γ participates in the innate immune response, promotes the differentiation and maturation of dendritic cells, and inhibits the uptake of antigen by dendritic cells. Concurrently, dendritic cells migrate to the draining lymph nodes to have an impact on antigen presentation. In addition, it can also have an impact on inflammatory factors and on the regulation of T cell balance and anti-T cell proliferation.[46,47]

PPAR-γ Targets Anti-Fibrosis Drugs

PPAR-γ signaling pathway has become a new hotspot in drug development; different types of agonists have different effects on liver.[48] In addition to the natural and synthetic ligands that have been confirmed, many effective components of Chinese herbal medicines can be used as

Natural Ligands

PPAR-γ agonists to affect liver fibrosis. At the same time, with the development of gene technology, microRNAs, lncRNAs, and nano new drugs have attracted research interest.[49]

Natural Ligands

Natural ligands of PPAR-γ include arachidonic acid and its metabolites, oxidative metabolites of polyunsaturated fatty acids, and prostaglandin metabolites. 15-Deoxy-Δ-12,14-prostaglandin J 2 (15d-PGJ2) is the most widely studied cyclopentenone prostaglandin, and is also the earliest identified endogenous ligand for PPAR-γ.[50] Studies have confirmed that 15d-PGJ2 inhibits TGF-β-induced expression of CTGF by inhibiting the phosphorylation of Smad2, which may have a favorable preventive effect on liver fibrosis induced by environmental toxicants.[51] Docosahexaenoic acid (DHA) and eicosatetraenoic acid (EPA) are trace bioactive substances and essential high unsaturated fatty acids in human, animals, and fish. He and his colleagues confirmed that long-term DHA and EPA supplementation inhibits the activation of HSCs via the TGF-β and ERK pathways in a PPAR-γ-dependent manner to alleviate liver fibrosis.[52–54] Leptin (LP) is a protein hormone secreted by adipose tissue. After entering the blood circulation, LP participates in the regulation of glucose, fat, and energy metabolism and inhibits the synthesis of adipocytes. In 2014, Zhou et al demonstrated that LP induced an increase in GATA2 and inhibited PPAR-γ expression, thus increasing the number of activated HSCs in vivo, which provides strong evidence that LP promotes liver fibrosis.[55] Wai et al subsequently established a GRX cell culture model with PPAR-γ gene overexpression, and confirmed that GATA-2 mediated LP expression and promoted the proliferation of HSCs through its binding site with the PPAR-γ promoter.[56] In addition, LP downregulates PPAR-γ gene expression in primary cultured rat HSCs by activating the PI-3K/Akt and ERK signaling pathways, which provides new insight into the mechanism underlying LP promotion of liver fibrosis.[57] Galanin and vitamin C, endogenous factors that negatively regulate the biological effects of LP bioenergy metabolism, inhibit the proliferation of HSCs and the expression of α-SMA by increasing PPAR-γ, which may be of potential significance for elucidating the detailed mechanisms underlying liver fibrosis in diseases with elevated circulating LP levels, such as non-alcoholic fatty hepatitis in obese patients.[54,58,59]

Synthetic Ligands

PPAR-γ synthetic ligands mainly include thiazolidine-diones (troglitazone, rosiglitazone, and pioglitazone), non-steroidal anti-inflammatory drugs, and leukotriene D4 receptor antagonists. In 2005, Marra demonstrated that troglitazone treatment reduces bile duct hyperplasia and fibrosis in a chronic cholestasis model.[60] At the same time, pioglitazone is thought to effectively reduce the levels of PDGF and TIMP-2 expression in a nonalcoholic fatty liver disease model, which helped to inhibit liver fibrosis.[39] Rosiglitazone, a highly effective PPAR-γ agonist, not only delays liver fibrosis induced by Schistosoma japonicum infection and bile duct ligation through downregulation of fibrosis-related genes, including TGF-β1, α-SMA, and CoI-α1, but also inhibits the activation of HSCs through miR-124-3p.[61–63] At the same time, when combined with erythromycin and all-trans retinoic acid in the treatment of liver fibrosis, rosiglitazone has a significant effect.[64,65] In addition, saroglitazar and ale stimulate PPAR-α/γ at the same time, which reduces liver fibrosis by inhibiting TGF-β1, PDGF, and neovascularization.[64,65] In addition to thiazolidinediones, new chemicals have been shown to activate PPAR-γ. GW570 has an anti-inflammatory effect on Kupffer cells in vivo during CCl4 injury, thus inhibiting astrocyte activation and inhibiting fibrosis after injury in a dose-dependent manner.[66] SC-236, a selective cyclooxygenase-2 inhibitor, alleviates liver fibrosis via non-parenchymal cell apoptosis and PPAR-γ activation.[67]

Traditional Chinese Medicine Extract

With advances in technology, it has been shown that some traditional Chinese medicine ingredients also lead to PPAR-γ ligand activation; curcumin is the most studied drug. In 2003 and 2004, Xu and Zheng found that curcumin significantly induces PPAR-γ gene expression in activated HSCs. Blocking the trans-activation activity with PPAR-γ antagonists significantly eliminates the inhibitory effect of curcumin on cell proliferation. It has been shown that curcumin has antioxidant potential and has no adverse effects on health, which makes curcumin a potential anti-fibrosis syndrome for the prevention and treatment of liver fibrosis.[68,69] Subsequently, many scholars have carried out a thorough exploration of the mechanism underlying curcumin function. Zhang believes that curcumin activates PPAR-γ to inhibit the angiogenesis of hematopoietic stem cells, thereby reducing sinusoidal angiogenesis in liver

fibrosis, while Qi has proved that PPAR-γ is a key factor in inhibiting HSC activation and DLK1 expression by blocking the Shh signal pathway.[70,71] Zhai proposed that co-activator-1α of PPAR-γ is key to the role of curcumin. Curcumin promotes the expression of SOD-1α via the AMPK pathway, which leads to an increase in SOD-2 transcription and activity.[72] These results may provide a new explanation for curcumin inhibition of HSC activation and hepatic fibrosis.

Other active Chinese medicines can play an anti-fibrosis role as a target of PPAR-γ. Polyphenol, rosmarinus acid (RA), and baicalin (BC) are important components of liver protection. The research shows that RA and BC significantly improve the histopathologic changes following CCl4 injury and BDL-induced liver fibrosis models. The underlying mechanism is to reduce the TGF-β1 and Wnt pathways and inhibit HSC activation.[73,74] Puerarin inhibits collagen deposition and plays an anti-fibrosis role by blocking the PI3K/Akt pathway.[75] Activation of PPAR-γ by tetra-methylpyrazine leads to the reverse expression of HIF-1α through a Smrt-dependent mechanism, which improves liver injury, angiogenesis, and vascular remodeling in rats with CCL4-induced liver fibrosis, and is thus of therapeutic significance for the peripheral cell biology and hepatic fibrosis of HSCs.[76] Resveratrol reduces the accumulation of lipid droplets in cells and PPAR-γ regulates the activation of HSCs and hepatic fibrosis by epigenetic regulation.[77] Hydroxysafflor yellow A inhibits carbon tetrachloride-induced liver fibrosis by regulating the PPAR-γ/P38 MAPK signaling pathway, while the antagonist, GW9662, significantly reduces the above effect.[78] Crocin treatment significantly reduces the serum liver enzyme level and improves liver CYP2E1 mRNA levels and the pathologic changes of liver tissue.[38] The alcohol extract of Rhus verniciflua fruit can prevent liver fibrosis induced by carbon tetrachloride in mice by inhibiting inflammatory factors [83]. Lycopene inhibits HSC activation, and regulates cell lipid storage and signal transduction.[79] In addition, picroside I, Achyranthes bidentata polysaccharide, rhubarb fruit extract, capsaicin, diosgenin, and umbelliferone participate in the regulation of PPAR-γ in liver fibrosis.[38,80–85]

MicroRNAs

Gene therapy is another major thrust of research, especially microRNAs, which have been used in the diagnosis and treatment of various diseases. Winkler and his colleagues [86] identified a miRNA network composed of 8 miRNA hubs and 54 target genes, and found that let-7, miR-30, miR-29c, miR-335, and miR-338 downregulate the key structure, signal, and remodeling components of the ECM. In addition, miR-130a and miR-130b were found to decrease PPAR-γ mRNA expression by targeting the expression of the 3 'untranslated region of PPAR-γ, a process that may be mediated by TGF-β1.[86] In activated artificial blood stem cells, the expression of PPAR-γ was negatively correlated with the expression of miR-34a and miR-34c, indicating that the miR-34 family may participate in the process of liver fibrosis by targeting PPAR-γ.[87] MiR-27b regulates the expression of PPAR mRNA in LX-2 cells and lays a foundation for further study of liver fibrosis.[88] MiR-29a increases PPAR-γ expression and negatively regulates HSC activation by inhibiting BRD4 and EZH2 functions, thus making it a promising target for drug therapy of liver fibrosis.[89] In human liver fibrosis, miR-942 inhibits PPAR-γ, then promotes the activation of HSCs induced by TGF-β1; an effect that is blocked by the inhibition of miR-942.[90] Therefore, if we can effectively regulate the related microRNAs, we can effectively improve liver fibrosis.

Other Ligands

In addition, there are some ligands which can play an effective role. For example, AT1 receptor blockers and the PPAR-γ activator, telmisartan, can reduce experimental liver fibrosis induced by Schistosoma mansoni infection.[91] Diosmin alleviates radiation-induced liver fibrosis by promoting PPAR-γ expression and inhibiting the typical Wnt signaling pathway activated by miR-17-5p.[92] Polymer nanoparticles co-administered with Hedgehog inhibitors and PPAR-γ agonists may be used to treat liver fibrosis by interfering with complex liver fibrosis cascade reactions.[93] The role of ligands in liver fibrosis is summarized in Table 1 and Figure 1.

Conclusions and Perspectives

PPAR-γ participates in the process of liver fibrosis by inhibiting HSCs activation and proliferation and increasing ECM production by regulating TGF-β and other pathways to achieve an anti-fibrosis effect.[94] According to the current research, PPAR-γ is closely related to liver fibrosis, so there is an incentive to study PPAR-γ ligands. In addition to the related ligands described in this review, many ligands, such as lncRNAs and circRNAs, are still under study. At present, it has only been preliminarily proved that they can act as endogenous competitive RNA through

Table 1 Summary of PPAR-γ Ligands in Liver Fibrosis

| Type	Agonists	Key Factor	Models	Dosage	Ref.
Natural	15d-PGJ2	TGF-β	Hep3B, HepG2, Huh-7	5–10 μM, 24 h	[51]
	DHA	TGF-β	HSC-LX2	0.05% v/v, 24 h	[53]
			ICR mice	20 mg/kg/d, 4w	
	Galanin	TGF-β	Primary HSCs, HSC-T6	1–10000 nM, 24 h	[54]
	Leptin	GATA-2	HSCs	100 ng/mL, 12 h	[55,56]
			C57BL mice	1 μg/g, 4 w	
	Vitamin C	TGF-β	SMP30 KO mice	1.5 g/L,16 w	[59]
Synthetic	Pioglitazone	PDGF	C57BL mice	3mg/kg/d, 4 w	[39]
	Thiazolidinedione	α-SMA	SD Rats	0.2% w/w, 4 w	[60]
	Rosiglitazone	TGF-β	Kunming mice	4 mg/kg/d, 6 w	[61,62]
		NF-κB	HepPPAR-γ KO mice	30 mg/kg/d, 2 w	
	Saroglitazar	TGF-β	SD Rats	3 mg/kg/d, 4 w	[64]
	Aleglitazar	TNF-α	SD Rats	0.3 mg/kg/d, 3 w	[65]
	GW570	TNF-α	SD Rats	10 mg/kg/d, 2 w	[66]
	SC-236	Apoptosis	SD Rats	6 mg/kg/d, 4 w	[67]
Traditional	Crocin	TNF-α	SD Rats	20–80 mg/kg/d, 4 w	[38]
	Curcumin	SOD2	Primary HSCs	15–20 μM, 24 h	[71,72]
			SD Rats	400 mg/kg/d, 4 w	
	Baicalin	TGF-β	SD Rats	25–100 mg/kg/d, 8 w	[73,74]
			Primary HSCs,	67.5–270 μM, 24 h	
	Puerarin	PI3K/Akt	SD Rats	200–800 mg/kg/d, 8 w	[75]
	Ligustrazine		HSC-T6	10–30 μM, 24 h	[76]
			SD Rats	50–200 mg/kg/d, 4 w	
	Resveratrol	IRT1	HSC-GRX	0.1 μM, 24 h	[77]
	Hydroxysafflor yellow A	P38 MAPK	SD Rats	10 mg/mL, 8 w	[78]
	Lycopene	Lipid metabolism	HSC-GRX	1–5 μmol/L, 24 h	[79]
	Picroside	TGF-β	C57BL mice	25–50 mg/kg/d, 8 w	[80]
	Ethanol extract	TNF-α	Kunming mice	400–800 mg/kg/d, 6 w	[82]
	Umbelliferone	TGF-β	SD Rats	25–100 mg/kg/d, 8 w	[83]
MiR-RNAs	miR-130	TGF-β	Primary HSCs, HSC-T6	0.1–100 pmol/L, 48 h	[86]
	miR-34	TGF-β	Primary HSCs, LX2	10–50 nmol/L, 48 h	[87]
	miR-27b	TGF-β	LX2	50–100 nmol/L, 48 h	[88]
	miR-29a	BRD4	Primary HSCs	25 nmol/L, 24 h	[89]
	miR-942	α-SMA	hHSCs, LX2	100 nmol/L, 24 h	[90]
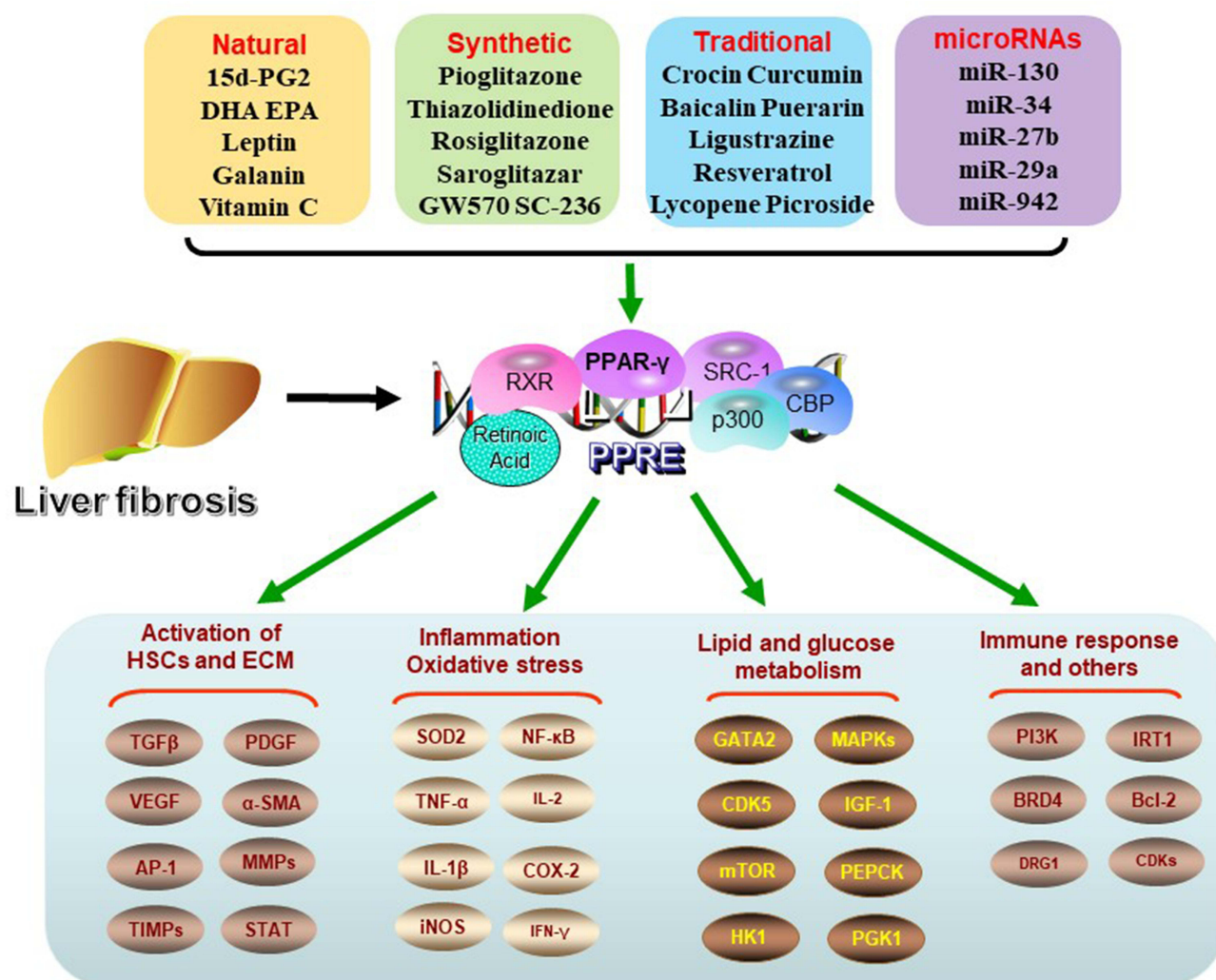
Figure 1 Regulation of PPAR-γ ligands and signal pathways. In liver fibrosis, PPAR-γ activation is inhibited. After activation by related ligands, PPAR-γ forms heterodimer with retinoic acid receptor (RXR), then plays a variety of biological roles by binding peroxisome proliferator response element (PPRE) to activate target gene transcription. Its biological processes involve the activation of HSCs, the deposition of ECM, oxidative and inflammatory reactions, glucose and lipid metabolism and immune response related pathways.

microRNAs, but the research on direct regulation is rare. We expect more effective ligands to be developed and applied in clinical treatment. The elucidation of the mutual regulation between PPAR-γ ligands and signal pathways, and the initiation of targeted gene therapy technology at the gene level will open a new way for the prevention and treatment of liver fibrosis and lay a foundation for the development of new anti-liver fibrosis drugs.

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Disclosure

The authors report no conflicts of interest in this work.

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