

# miR-622 suppresses tumor formation by directly targeting VEGFA in papillary thyroid carcinoma

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**Background:** MicroRNAs (miRNAs) were reportedly to play crucial roles in papillary thyroid carcinoma (PTC) tumorigenesis and development. Therefore, the discovery of miRNAs may provide a new and powerful tool for diagnosis and treatment of PTC.

**Purpose:** The aim of this study was to investigate the biological function and underlying mechanism of miR-622 in PTC.

**Materials and methods:** The expression levels of miR-622 in PTC patient tissues and cell lines were determined by quantitative real-time PCR (qRT-PCR). The biological function including cell proliferation, colony formation, migration and invasion, as well as underlying mechanism of miR-622 in PTC, were also evaluated by a series of in vitro and in vivo experiments.

**Results:** miR-622 expression level was significantly downregulated in PTC tissues and cell lines. Decreased miR-622 expression was associated with advanced clinical stage and lymph node metastasis ( $P < 0.01$ ). The overexpression of miR-622 in TPC-1 cells inhibited cell proliferation, migration and invasion in vitro, as well as suppress tumor growth in vivo. Moreover, we also demonstrated that miR-622 specifically targeted the 3'-UTR regions of vascular endothelial growth factor A (VEGFA) and inhibited its expression both mRNA level and protein levels. Overexpression of VEGFA reversed miR-622-mediated inhibition effect on cell proliferation, migration and invasion in thyroid cancer cells. More importantly, VEGFA expression was significantly increased and inversely correlated with the levels of miR-622 in PTC tissues.

**Conclusion:** These results show that miR-622 acts as a tumor suppressor in thyroid cancer, at least in papillary targeting VEGFA, and suggest that miR-622 may serve as a potential target for treatment of thyroid cancer patients.

**Keywords:** thyroid cancer, miR-622, VEGFA, proliferation, invasion

## Introduction

Thyroid cancer (TC) is the most common endocrine disease and the fifth most frequent cancer in women. Papillary thyroid carcinoma (PTC) is the most prevalent type of tumor among thyroid malignancies, accounting for ~80% of all TC cases.<sup>1</sup> The incidence of PTC has increased in many countries in the past few years due to, among others, an increase in environmental pollution, which causes suppression of immune system, and unhealthy diet.<sup>2</sup> Despite significant improvement in overall survival of patients after treatment with surgery, chemotherapy, and/or radiotherapy, metastatic thyroid carcinoma has a poor prognosis.<sup>3</sup> Therefore, studies regarding the mechanism of action of the underlying causes of metastasis of PTC is highly warranted in order to find a more effective therapy for PTC.

MicroRNAs (miRNAs) are a group of small (18–25 nucleotides in length), highly conserved noncoding RNA molecules that regulate gene expression by binding to partially complementary sequences of target mRNAs.<sup>4</sup> Increasing evidence have revealed

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that miRNAs regulate a wide range of physiological activities in the cell, such as cell growth, apoptosis, lipid metabolism, tumorigenesis, and metastasis.<sup>5,6</sup> Alterations in miRNA have been shown to play crucial roles in tumor invasion, migration, and metastasis of TC.<sup>7-9</sup>

The expression of miR-622 is downregulated in colorectal cancer,<sup>10</sup> gastric cancer,<sup>11</sup> esophageal squamous cell carcinoma,<sup>12</sup> glioma,<sup>13</sup> and hepatocellular carcinoma,<sup>14</sup> which suggests that miR-622 acts as a tumor suppressor in these types of cancers. However, the role of miR-622 in TC is largely unknown. Therefore, in this study, we aimed to investigate the clinical significance of miR-622 in patients with PTC and to study the role and mechanism of action of miR-622 in TC progression.

## Materials and methods

### Patients and samples

Forty-two patients with PTC who had undergone thyroidectomy at the China–Japan Union Hospital of Jilin University between January 2015 and January 2016 participated in this study. Matched TC samples and adjacent nontumor tissues (>3 cm distant from the tumor margin) were obtained during the operation and immediately stored at  $-80^{\circ}\text{C}$  until use. The pathological stage and grade were evaluated by an experienced pathologist. Thirty-two out of 42 patients were diagnosed with TNM stages I and II tumors and 10 patients with TNM stages III and IV tumors. Among the lymph node metastasis was observed in case of 11 patients, whereas no such metastasis was observed in case of 31 patients. Written informed consent was obtained from all patients in this study. This study was approved by the ethical committee of China–Japan Union Hospital of Jilin University.

### Cell culture

Three human TC cell lines (8505C, TPC-1, and SW1736), thyroid gland epithelial cell line (Nthy-ori3-1), and human embryonic kidney (HEK) 293T cells were purchased from American Type Culture Collection (ATCC) and were cultured in DMEM (Thermo Fisher Scientific, Waltham, MA, USA) supplemented with 10% FBS (Thermo Fisher Scientific) at  $37^{\circ}\text{C}$  in 5%  $\text{CO}_2$ .

### Construction of plasmid and establishment of stable cell lines

The sequence of VEGFA (lacking 3' untranslated region [UTR]) was amplified and cloned into pcDNA3.1 (Thermo Fisher Scientific) to generate the pcDNA3–VEGFA recombinant plasmid. The recombinant plasmid (10  $\mu\text{g}$ ) was transfected into TPC-1 cells using Lipofectamine 2000 transfection

reagent (Thermo Fisher Scientific), according to the manufacturer's instructions. Lentivirus carrying miR-622 or negative control mimic (miR-NC) was packaged into HEK 293T cells using the lentiviral packaging kit (GeneChem, Shanghai, People's Republic of China), according to the manufacturer's manual. Stable cell lines were established by infection of TPC-1 with 0.5  $\mu\text{L}$  lentivirus ( $1 \times 10^8$  TU/mL), followed by puromycin (1  $\mu\text{g}/\text{mL}$ ) selection.

### Quantitative real-time PCR

Total RNA from tissues and cell lines was extracted using the TRIzol reagent (Thermo Fisher Scientific) and was reverse transcribed using TaqMan MicroRNA Reverse Transcription Kit (Thermo Fisher Scientific). The expression of miR-622 was quantified using TaqMan microRNA assay (Ambion) with the ABI 7900 Sequence Detection System (Thermo Fisher Scientific). To detect VEGFA mRNA, the total RNA was reverse transcribed to cDNA using the HiScript QRT SuperMix (Vazyme, Nanjing, People's Republic of China) and then real-time PCR was performed using SYBR Green I Fluorescent Assay kit (DR041A; Takara) and quantified using ABI 7900 Sequence Detection System. The expression of miR-622 or VEGFA was calculated relative to the expression of U6 snRNA or GAPDH, respectively, using the comparative threshold method ( $2^{-\Delta\Delta\text{CT}}$ ).

### Cell proliferation

Cell proliferation was determined using an MTT cell proliferation kit (Roche Applied Science, Indianapolis, IN, USA), by following the manufacturer's instructions. Briefly, the transfected cells were seeded in 96-well plates at a density of  $2.0 \times 10^4$  cells/well. At indicated time points (24, 48, and 72 h post-transfection), the cells were incubated with 20  $\mu\text{L}$  MTT reagent for 4 h, followed by the addition of 200  $\mu\text{L}$  dimethyl sulfoxide (DMSO) to each test well. The absorbance was measured at 490 nm using a Thermo Scientific Multiskan spectrophotometer (Thermo Fisher Scientific).

### Cell cycle analysis

Cells were harvested (48 h post-transfection) in flow cytometry tubes and centrifuged at  $1,000 \times g$  for 5 min to pellet the cells. The pellets were washed with PBS and fixed with 70% cold ethanol at  $4^{\circ}\text{C}$  overnight. After washing again with PBS, the cells were incubated with RNAaseA (0.1 mg/mL) for 30 min, followed by incubation with propidium iodide (50  $\mu\text{g}/\text{mL}$ ) for 30 min at room temperature. Cell cycle analysis was performed using a flow cytometer (FACSsort; Becton Dickinson, Franklin Lakes, NJ, USA). The percentage of cells at G0/G1, S, and G2/M phases was analyzed using the FlowJo software 3.1 (TreeStar, Inc., Ashland, OR, USA).

## Cell migration and invasion assays

A wound healing assay was performed to assess cell migration. Briefly,  $2 \times 10^5$  transfected cells were seeded in 60 mm dishes and cultured in DMEM for 24 h. A linear wound was created by scraping the confluent cell monolayer. Cells were washed with PBS and cultured in serum-free DMEM for additional 24 h. Wound closure was measured by photographing five randomly selected fields at the time of wounding (time 0) and 24 h after wounding.

Cell invasion was determined using BD BioCoat™ Matrigel invasion chambers (Becton Dickinson) in accordance with the manufacturer's instructions. Briefly, the transfected cells were seeded onto the upper side of the invasion chamber with Matrigel (1 mg/mL), and the medium containing 10% FBS was added to the lower chamber as a chemoattractant. After 48 h, the invasive cells that migrated to the lower side of the chamber were fixed using 20% methanol and stained with 0.1% crystal violet. The invasive cells were imaged under a Nikon phase-contrast microscope and counted under five randomly selected fields of view at 200× magnification.

## Luciferase reporter assay

The human VEGFA 3'-UTR oligonucleotides containing the wild-type (WT) or mutant (Mut) miR-622 binding sites were synthesized by Ribobio Co. (Guangzhou, People's Republic of China) and then inserted into the psiCHECK-1 vector (Promega, Madison, WI, USA). For luciferase reporter assay, TPC-1 cells were seeded in 96-well culture plates and cultured for 24 h and then transfected with the WT or Mut VEGFA 3'-UTR construct along with miR-622 mimic or miR-NC using Lipofectamine 2000 (Thermo Fisher Scientific), according to the manufacturer's recommendation. Firefly and renilla luciferase activities were determined at 48 h post-transfection using the Dual-Luciferase Reporter Assay kit (Promega).

## Western blot

Total proteins were extracted from cultured cells using RIPA buffer (Beyotime, Shanghai, People's Republic of China) and the protein concentration was determined using a BCA protein assay kit (Pierce, Rockford, IL, USA). Equal amounts of protein (30 µg) were separated by SDS-PAGE and transferred onto a nitrocellulose membrane (Amersham Biosciences, Piscataway, NJ, USA). After blocking with 5% nonfat milk, the blots were probed with mouse antibodies against VEGFA and GAPDH (1:1000; Santa Cruz Biotechnology Inc., Dallas, TX, USA), followed by incubation with an HRP-conjugated

goat-anti-mouse secondary antibody (1:5000; Santa Cruz Biotechnology Inc.) for 2 h at room temperature. The protein bands were detected with enhanced chemiluminescence reagents (Pierce) and were quantified using the Scion Image software and normalized to GAPDH levels.

## Animal studies

All experiments were approved by the Animal Care and Utilization Committee of Jilin University (Changchun, People's Republic of China) and the protocols complied with the guidelines for the welfare and use of animals in cancer research (ad hoc committee of the National Cancer Research Institute, UK).<sup>15</sup>

TPC-1 cells ( $2 \times 10^5$ ) stably expressing miR-622 or miR-NC were subcutaneously injected into the BALB/c-nu mice. Tumor growth was determined by measuring the tumor length (L) and width (W) weekly and calculating its volume (V) by using the formula:  $V = (L \times W^2)/2$ . Mice were sacrificed 35 days after inoculation and tumors were excised, weighed, and stored at  $-80^\circ\text{C}$  for further analysis.

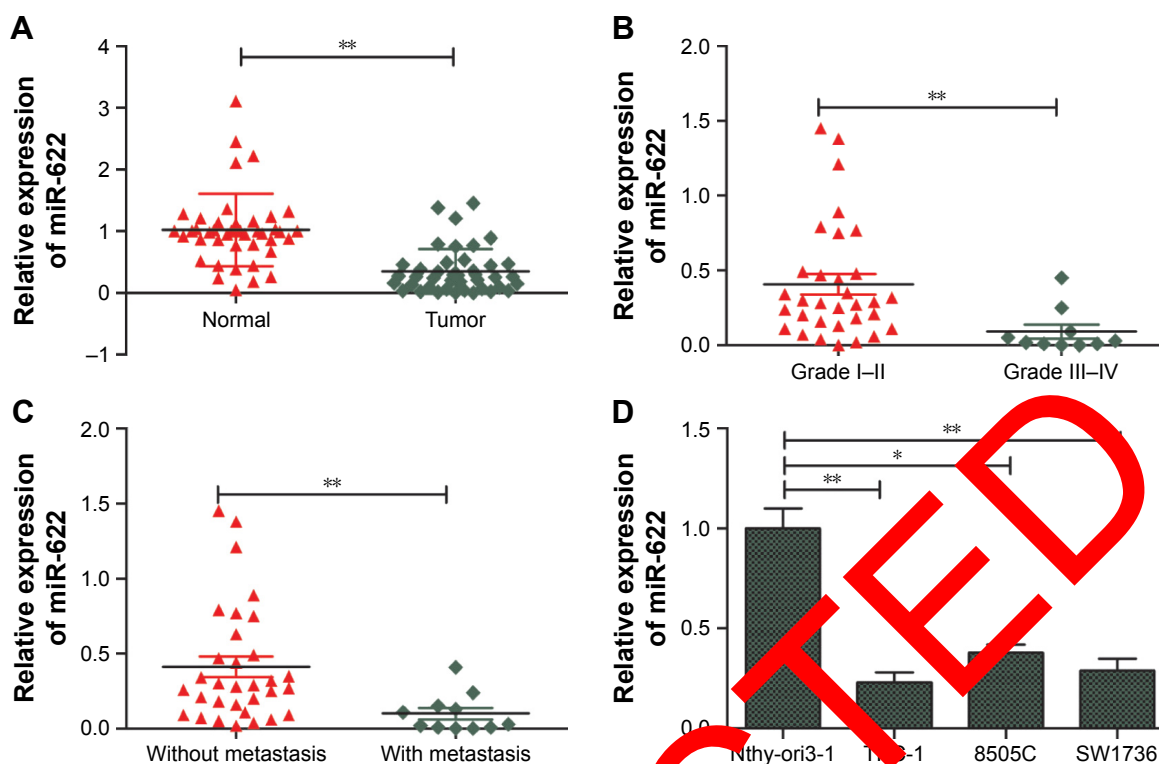
## Statistical analysis

All data are expressed as the mean  $\pm$  standard deviation (SD) from at least three independent experiments. Statistical analysis was performed using the SPSS 19.0 package of the Social Sciences 19.0 Windows (IBM Corporation, Armonk, NY, USA) was used to perform statistical analyses. Differences between the two groups were compared using Student's *t*-test, and Tukey's one-way ANOVA was used when more than two groups were compared. The relationship between the expression of miR-622 and VEGFA was assessed by the Pearson's correlation analysis. *P*-values  $< 0.05$  were considered statistically significant.

## Results

### Expression of miR-622 is decreased in TC tissues and cell lines

The expression of miR-622 in 42 paired PTC tissues and corresponding adjacent normal tissues was determined by qRT-PCR. According to our results, the expression of miR-622 in TC tissues was found to be decreased when compared with adjacent normal tissues ( $p < 0.01$ , Figure 1A). In addition, the expression of miR-622 was found to be downregulated in advanced TNM stages (Figure 1B). Meanwhile, TC tissues with lymph node metastasis showed lower levels of miR-622 than that of tissues without lymph node metastasis (Figure 1C). Moreover, the expression of miR-622 was found to be decreased in all three TC cell lines, namely, 8505C, TPC-1, and SW1736 when compared with normal



**Figure 1** The expression levels of miR-622 were downregulated in papillary thyroid carcinoma (PTC) and cell lines. **Notes:** (A) The relative expression levels of miR-622 in PTC tissues and corresponding adjacent normal tissues. (B) The relative expression levels of miR-622 in PTC tissues with different TNM stage. (C) The relative expression levels of miR-622 in PTC tissues with or without lymph node metastasis. (D) The relative expression levels of miR-622 in three thyroid cancer cell lines 8505C, TPC-1, and SW1736 and thyroid gland epithelial cell line Nthy-ori3-1 were detected by qRT-PCR. \* $p < 0.05$ , \*\* $p < 0.01$ .

thyroid gland epithelial cell line, Nthy-ori3-1 ( $p < 0.05$ ; Figure 1D).

## Overexpression of miR-622 inhibits thyroid cell proliferation

To investigate the biological role of miR-622 in TC, we generated TPC-1 cells overexpressing control miRNA or miR-622 by transfection with lentiviruses carrying miR-NC or miR-622, respectively (Figure 2A). MTT assay demonstrated that the overexpression of miR-622 significantly inhibited TPC-1 cell proliferation ( $p < 0.05$ ; Figure 2B). Moreover, it significantly improved the percentage of cells arrested at the G0/G1 stage and decreased the percentage of cells at the S stage of the cell cycle in TPC-1 cells (Figure 2C).

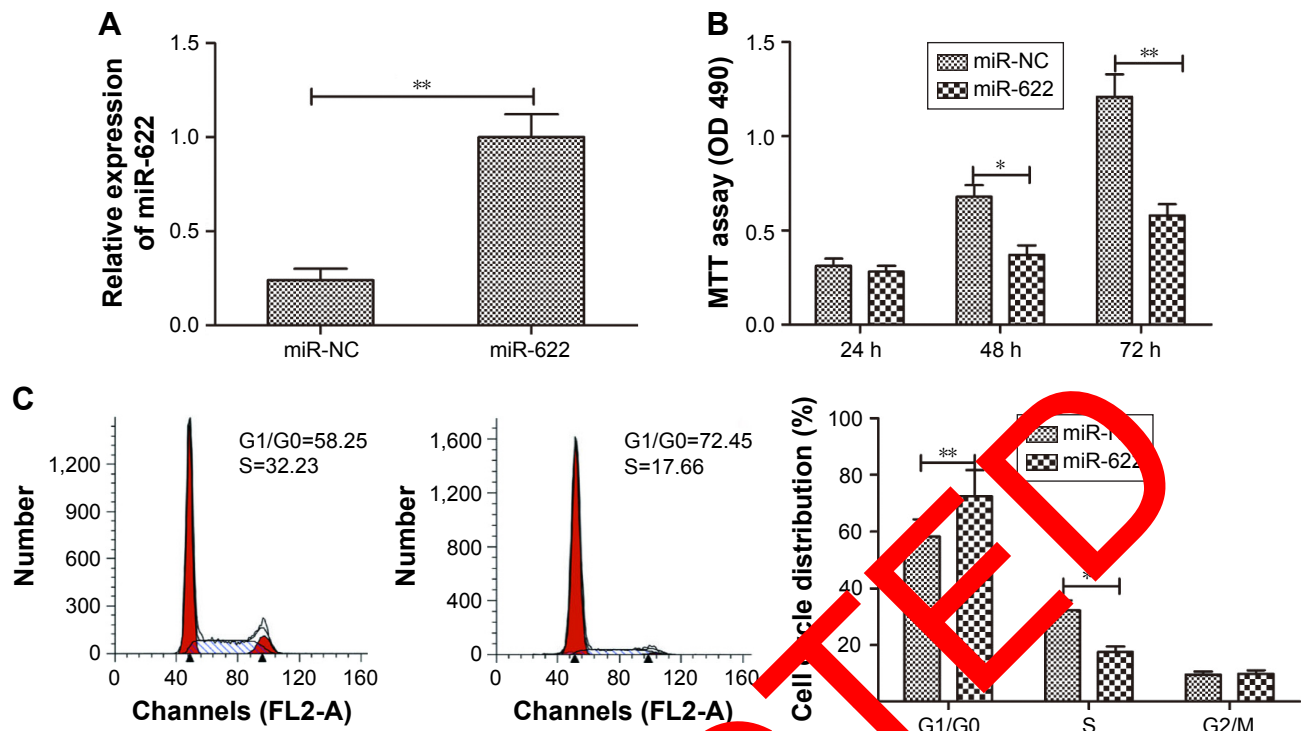
## miR-622 suppresses TC cell migration and invasion

Next, we investigated the role of miR-622 in TC migration and invasion through wound healing and transwell invasion assays, respectively. We found that the overexpression of miR-622 significantly decreased migration and invasion compared with the miR-NC group (both  $p < 0.01$ ; Figure 3A and B).

## VEGFA was directly targeted by miR-622 in TC cells

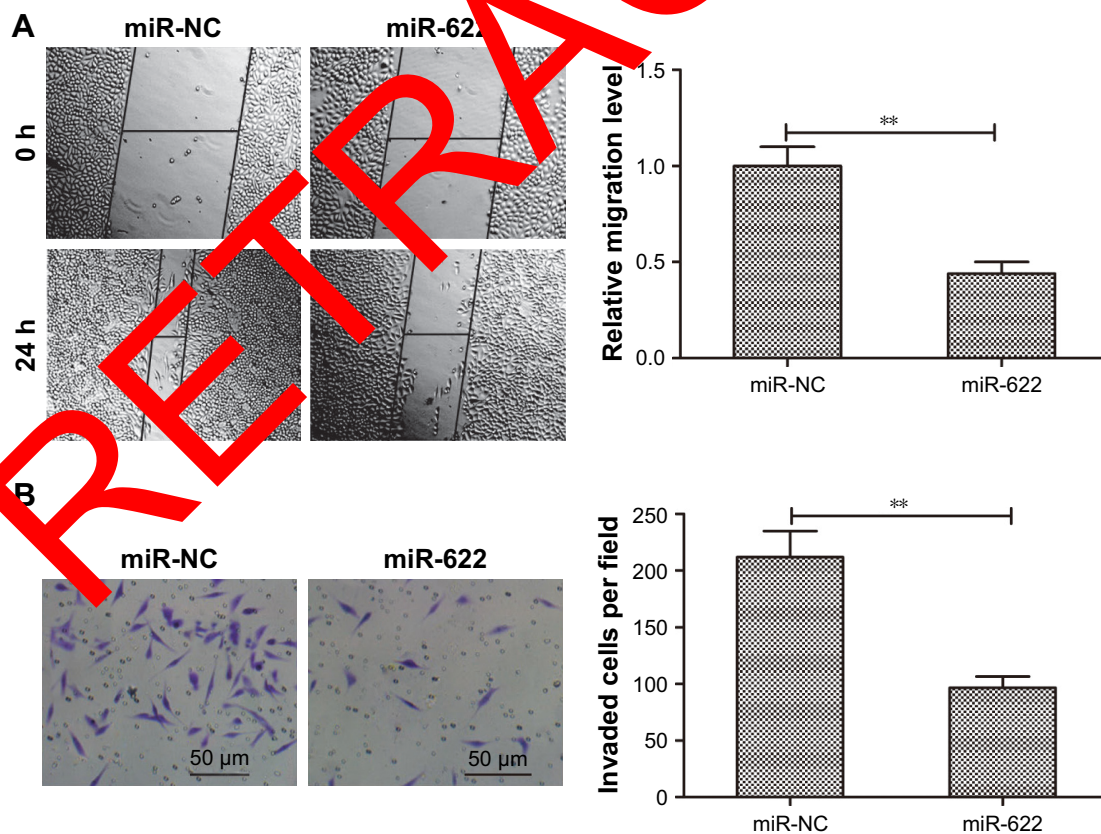
By TargetScan and miRanda software database screening, we found that VEGFA was a predicted target of miR-622e and that there was a binding site of miR-622 in the 3'-UTR of VEGFA (Figure 4A). To confirm that miR-622 binds to the 3'-UTR of VEGFA, luciferase reporter assays were performed in miR-NC or miR-622 overexpressing TPC-1 cells transfected with WT or Mut VEGFA-3'-UTR. As predicted, miR-622 bound to VEGFA 3'-UTR, resulting in markedly decreased luciferase activity (Figure 4B). Moreover, the mRNA and protein levels of VEGFA in TPC-1 cells were downregulated upon miR-622 infection (Figure 4C and D). VEGFA mRNA expression in 42 pairs of PTC tissues and the corresponding adjacent nontumor tissues were also detected by qRT-PCR. As shown in Figure 4E, VEGFA mRNA expression was found to be significantly increased in the PTC tissues compared to that of adjacent nontumor tissues. Pearson's correlation analysis revealed that the expression of miR-622 was inversely correlated with the VEGFA mRNA levels in PTC tissues ( $n = 42$ , Figure 4F). These results implied that VEGFA was a direct target of miR-622 in TC.





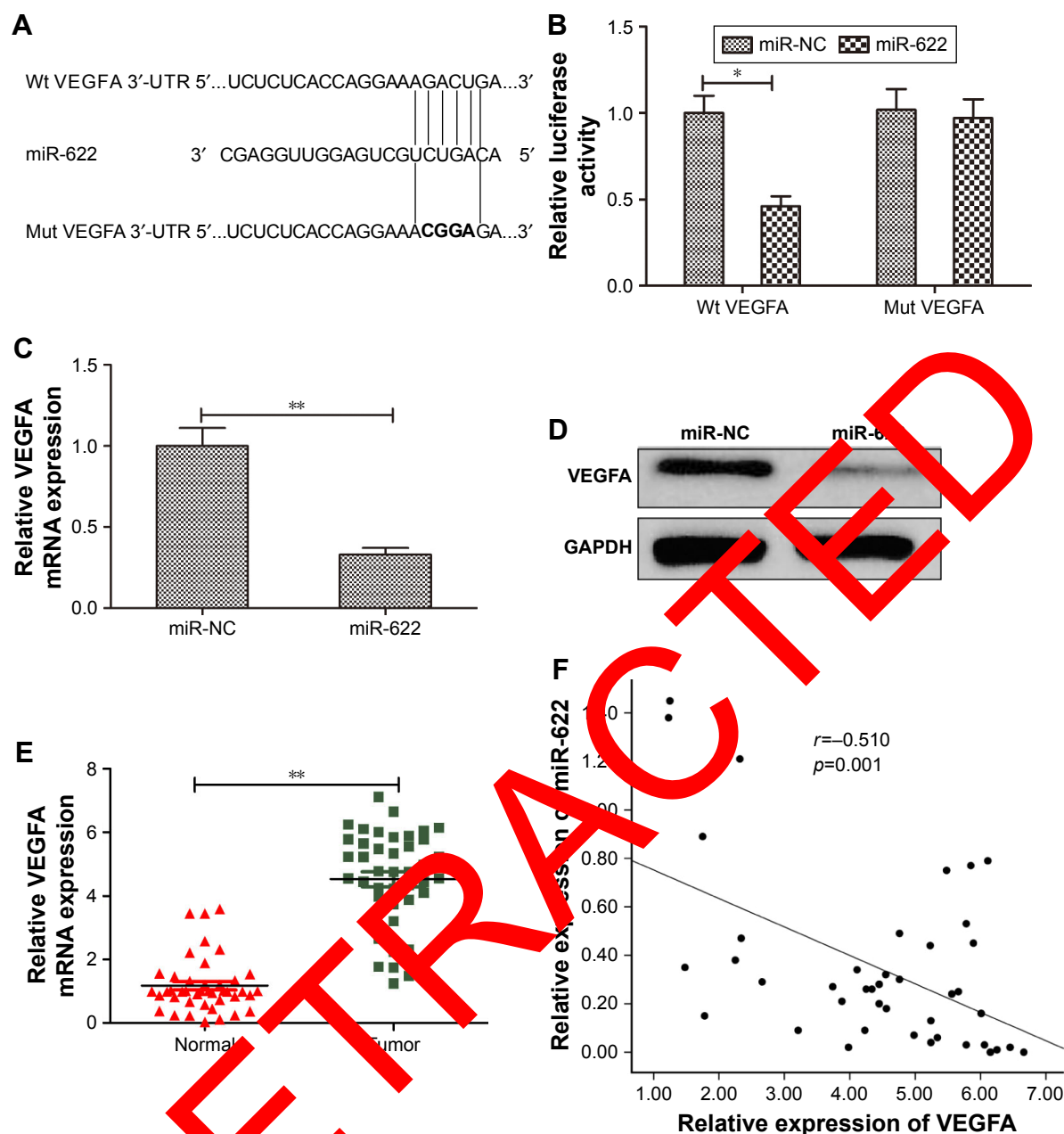
**Figure 2** miR-622 overexpression inhibits thyroid cancer cell proliferation.

**Notes:** (A) The relative expression levels of miR-622 in TPC-I cells infected miR-622 and miR-NC. (B, C) Cell proliferation and cycle arrest were determined in TPC-I cells infected with miR-622 or miR-NC. \* $p < 0.05$ , \*\* $p < 0.01$ .



**Figure 3** miR-622 overexpression inhibits thyroid cancer cell migration and invasion.

**Notes:** (A) Cell migration was determined in TPC-I cells infected with miR-622 or miR-NC by wound healing assay. (B) Cell invasion was determined in TPC-I cells transfected miR-622 mimic or miR-NC mimic by transwell invasion assay. \*\* $p < 0.01$ .



**Figure 4** miR-622 targets VEGFA 3'-UTR in TPC-1 cells.

**Notes:** (A) Sequence alignment of putative wild-type (Wt) and mutant (Mut) miR-622-binding sites in the 3'-UTR of VEGFA. (B) The luciferase activities in TPC-1 cells infected with miR-622 or miR-NC and luciferase reporters containing Wt or Mut VEGFA-3'-UTR, respectively. (C, D) The VEGFA expression at mRNA and protein levels was determined in TPC-1 cells transfected with miR-622 or miR-NC. (E) The relative expression levels of VEGFA mRNA in PTC tissues and adjacent normal tissues were detected by qRT-PCR. (F) The Pearson's Correlation analysis for the relationship between miR-622 levels and VEGFA mRNA levels in PTC tissues. \* $p < 0.05$ , \*\* $p < 0.01$ .

**Abbreviations:** PTC, papillary thyroid carcinoma; VEGFA, vascular endothelial growth factor A; UTR, untranslated region; qRT-PCR, quantitative real-time PCR.

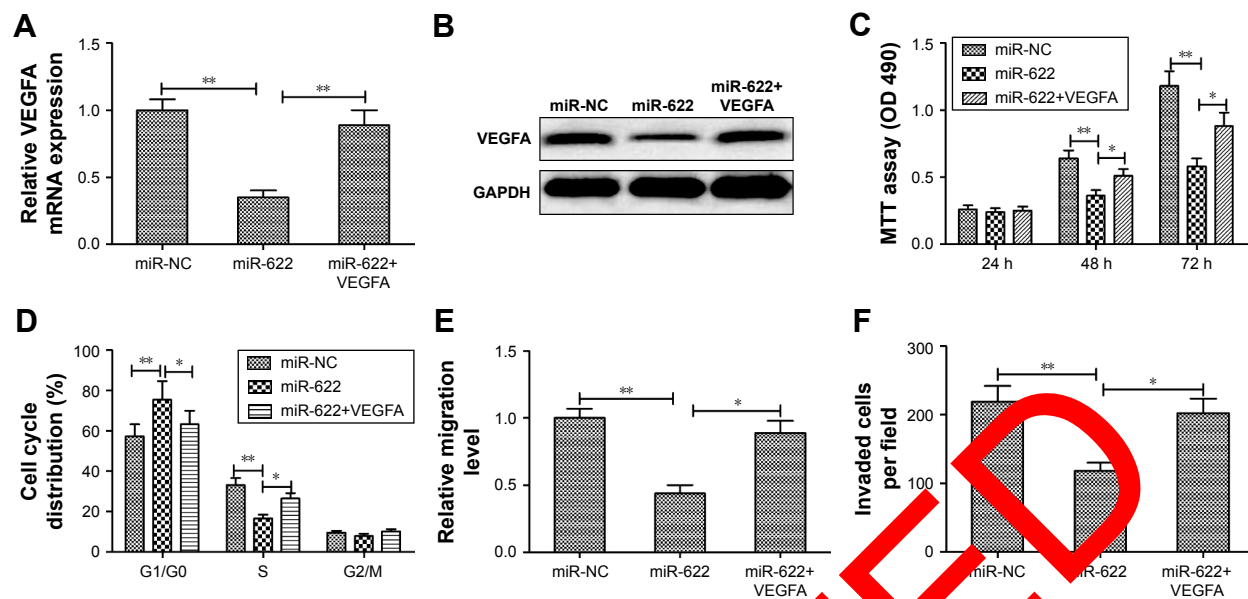
## Overexpression of VEGFA reverses the tumor-suppressive effect of miR-622 in TC

To investigate the role of miR-622 in TC cells, TPC-1 cells with high expression of miR-622 were transfected with the VEGFA overexpression vector (pcDNA3.1-VEGFA). We observed that in TPC-1 cells transfected with the VEGFA overexpression plasmid, VEGFA expression that was found to be decreased by the overexpression of miR-622 was restored

(Figure 5A and B). Furthermore, overexpression of VEGFA reversed the effect of miR-622 overexpression on proliferation, cell cycle arrest, migration, and invasion of TPC-1 cells (Figure 5C–F). These data showed that miR-622 inhibits TC cell growth and metastasis by decreasing VEGFA expression.

## miR-622 inhibits tumor growth in vivo

To assess whether miR-622 suppresses tumor growth in vivo, we created tumor xenograft mouse models by subcutaneously



**Figure 5** Overexpression of VEGFA reverses the tumor-suppressive effect of miR-622 in thyroid cancer.

**Notes:** (A, B) The VEGFA expression on mRNA and protein levels was determined in TPC-1 cells transfected with VEGFA plasmid without 3'-UTR in combination with either miR-622 or miR-NC. (C–F) Cell proliferation, cycle arrest, migration, and invasion were determined in TPC-1 cells transfected with VEGFA plasmid without 3'-UTR in combination with either miR-622 or miR-NC. \* $p < 0.05$ , \*\* $p < 0.01$ .

**Abbreviations:** VEGFA, vascular endothelial growth factor A; UTR, untranslated region.

injecting TPC-1 cells stably expressing miR-622 or miR-NC.

The results show that tumor growth decreased in the TPC-1/miR-622 group when compared with TPC-1/miR-NC group

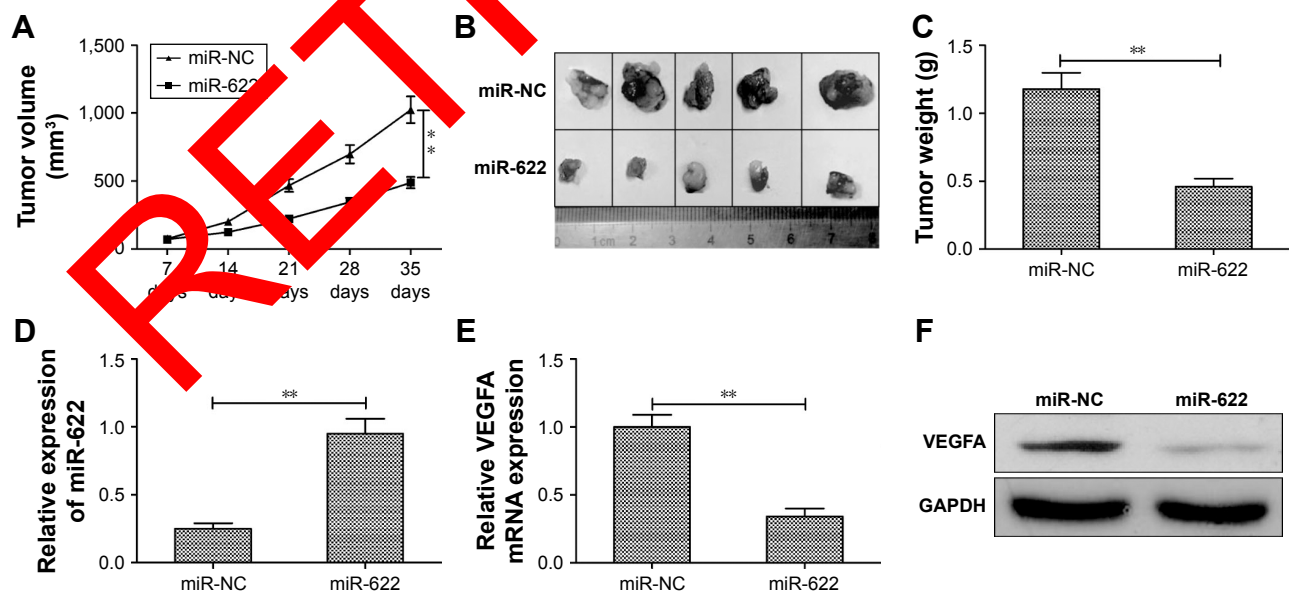
(Figure 6A–C). We also detected the expression of miR-622 and VEGFA in tumor tissues from nude mice and found that the expression of miR-622 was upregulated (Figure 6D), whereas the levels of VEGFA mRNA and VEGFA protein were found to be decreased in the TPC-1/miR-622 group

(Figure 6E and F), compared with the TPC-1/miR-NC group.

These results suggest that miR-622 suppresses tumor growth in vivo by regulating VEGFA.

## Discussion

Emerging evidence has revealed that miRNAs play an important role in the initiation and development of TC by regulating target genes that are involved in cellular



**Figure 6** miR-622 inhibits tumor growth in vivo.

**Notes:** (A) The growth curves of xenograft tumors were established after the subcutaneous injection of indicated cells. (B) Representative images of tumor tissues. (C) The tumor weights of tumor tissues. (D) The relative expression levels of miR-622 in tumor tissues from nude mice by qRT-PCR. (E, F) The VEGFA expression at mRNA and protein levels was determined in tumor tissues from nude mice detected by qRT-PCR and Western blot. \* $p < 0.05$ , \*\* $p < 0.01$ .

**Abbreviation:** VEGFA, vascular endothelial growth factor A; qRT-PCR, quantitative real-time PCR.

proliferation, apoptosis, migration, and metastasis, and cell cycle arrest,<sup>7-9</sup> suggesting that investigating the biological role of miRNAs in TC might contribute toward finding novel diagnostic markers and therapeutic agents for TC. In this study, we performed a series of in vitro and in vivo experiments to investigate the role of miR-622 in PTC. To the best of our knowledge, this is the first study reporting that the expression of miR-622 was downregulated in PTC tissues and cell lines, and that the decrease was associated with the TNM stage and lymph node metastasis. We also showed that miR-622 inhibited TC growth in vitro and in vivo by repressing VEGFA. These findings suggest that miR-622 can be a new therapeutic target for TC.

Accumulating evidence shows that the expression of miR-622 decreased due to its tumor suppressive role in several types of cancer.<sup>9-14</sup> For example, Song et al showed that the overexpression of miR-622 could significantly reduce cell proliferation rate of esophageal squamous cell carcinoma, enhance cell apoptosis, and impair cell invasion and migration by targeting E2F1.<sup>12</sup> Wang et al suggested that upregulated miRNA-622 expression inhibited cell proliferation, migration, and invasion of glioblastoma by repressing K-Ras.<sup>16</sup> Cheng et al indicated that increased expression of miR-622 inhibited lung cancer cell migration and invasion in vitro and in vivo by repressing hypoxia-inducible factor-1 $\alpha$ .<sup>17</sup> Liu et al indicated that miR-622 acts as a tumor suppressor in hepatocellular carcinoma by targeting CXCR4.<sup>13</sup> However, the role and mechanism of action of miR-622 in PTC remained unclear. In this study, we found that the expression of miR-622 was downregulated in PTC tissues and cell lines, and its expression was associated with the TNM stage and lymph node metastasis. Moreover, we also showed that the overexpression of miR-622 significantly inhibited cell proliferation, migration, and invasion in vitro as well as suppressed tumor growth in vivo. These results implied that miR-622 acts as a tumor suppressor in PTC.

There is mounting evidence demonstrating that miRNAs exert their biological roles in cancer by regulating target genes.<sup>18</sup> Therefore, investigating the relationship between the miRNA and its target is crucial in understanding the mechanism underlying the action of miRNA in cancer.<sup>19</sup> Through bioinformatic prediction (TargetScan and miRanda), we identified that VEGFA 3'-UTR possessed a miR-622 response element. Subsequently, through the luciferase reporter assay, we found that miR-622 could bind to the VEGFA 3'-UTR and could inhibit the activity of the VEGFA 3'-UTR. Moreover, the overexpression of miR-622 significantly decreased

VEGFA mRNA and protein levels in TPC-1 cells. An inverse relationship between VEGFA and the expression of miR-622 was observed in PTC tissues. These data suggest that miR-622 targets VEGFA in PTC.

VEGFA has been reported to play crucial roles in regulating vascular development during embryogenesis and in the formation of new blood vessels from preexisting vascular networks.<sup>20,21</sup> Accumulating evidence indicated that upregulation of VEGFA expression was associated with poor prognosis and death of the patient from metastasis in various cancers.<sup>22,23</sup> In addition, VEGFA has been found to contribute to tumor growth and metastasis by inducing angiogenesis via the VEGFR-1 and VEGFR-2.<sup>21,24</sup> In TC, VEGFA expression was increased in the TC tissues, and overexpression of VEGFA was correlated with the pathological parameters and metastatic status of the thyroid carcinomas.<sup>25,26</sup> In this study, we identified VEGFA as a target of miR-622 in TC. Overexpression of VEGFA partially abrogated the suppression effect on proliferation, cell cycle arrest, migration, and invasion of TPC-1 cells induced by miR-622. Moreover, miR-622 suppressed tumor growth in vivo by suppressing VEGFA. These data indicate that miR-622 acts as a tumor suppressor in TC by targeting VEGFA.

The primary limitation of our study is that we analyzed few samples and cell types. Therefore, more elaborate studies are necessary to further explore the potential therapeutic and prognostic role of miR-622 in TC progression. In conclusion, our results demonstrated that miR-622 acts as a tumor suppressor in PTC by inhibiting cell proliferation, migration, and invasion via suppressing VEGFA, suggesting that miR-622 can be a potential therapeutic target for PTC treatment.

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## Disclosure

The authors report no conflicts of interest in this work.

## References

1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin*. 2015;65(2):87–108.
2. Liu S, Semenciw R, Ugnat AM, Mao Y. Increasing thyroid cancer incidence in Canada, 1970–1996: time trends and age-period-cohort effects. *Br J Cancer*. 2001;85(9):1335–1339.



3. Pemayun TG. Current diagnosis and management of thyroid nodules. *Acta Med Indones*. 2016;48(3):247–257.
4. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell*. 2004;116(2):281–297.
5. Oliveto S, Mancino M, Manfrini N, Biffo S. Role of microRNAs in translation regulation and cancer. *World J Biol Chem*. 2017;8(1):45–56.
6. Malan-Müller S, Hemmings SM, Seedat S. Big effects of small RNAs: a review of microRNAs in anxiety. *Mol Neurobiol*. 2013;47(2):726–739.
7. de la Chapelle A, Jazdzewski K. MicroRNAs in thyroid cancer. *J Clin Endocrinol Metab*. 2011;96(11):3326–3336.
8. Leonardi GC, Candido S, Carbone M, et al. microRNAs and thyroid cancer: biological and clinical significance (Review). *Int J Mol Med*. 2012;30(5):991–999.
9. Wei WJ, Shen CT, Song HJ, Qiu ZL, Luo QY. MicroRNAs as a potential tool in the differential diagnosis of thyroid cancer: a systematic review and meta-analysis. *Clin Endocrinol (Oxf)*. 2016;84(1):127–133.
10. Wang Y, Sun J, Wei X, et al. Decrease of miR-622 expression suppresses migration and invasion by targeting regulation of DYRK2 in colorectal cancer cells. *Onco Targets Ther*. 2017;10:1091–1100.
11. Guo XB, Jing CQ, Li LP, et al. Down-regulation of miR-622 in gastric cancer promotes cellular invasion and tumor metastasis by targeting ING1 gene. *World J Gastroenterol*. 2011;17(14):1895–1902.
12. Song C, Lu P, Shi W, et al. MiR-622 functions as a tumor suppressor and directly targets E2F1 in human esophageal squamous cell carcinoma. *Biomed Pharmacother*. 2016;83:843–849.
13. Liu H, Liu Y, Liu W, Zhang W, Xu J. EZH2-mediated loss of miR-622 determines CXCR4 activation in hepatocellular carcinoma. *Nat Commun*. 2015;6:8494.
14. Zhang R, Luo H, Wang S, et al. MiR-622 suppresses proliferation, invasion and migration by directly targeting activating transcription factor 2 in glioma cells. *J Neurooncol*. 2015;121(1):63–72.
15. Workman P, Aboagye EO, Balkwill F, et al; Committee of the British Cancer Research Institute. Guidelines for the welfare and use of animals in cancer research. *Br J Cancer*. 2010;102(11):1555–1577.
16. Wang X, Xin Z, Xu Y, Ma J. Upregulated miRNA-622 inhibited cell proliferation, motility, and invasion via repressing Kirsten rat sarcoma in glioblastoma. *Tumour Biol*. 2016;37(5):5963–5970.
17. Cheng CW, Chen PM, Hsieh YH, et al. Foxo3a-mediated overexpression of microRNA-622 suppresses tumor metastasis by repressing hypoxia-inducible factor-1 $\alpha$  in ERK-responsive lung cancer. *Oncotarget*. 2015;6(42):44222–44238.
18. Moles R. MicroRNAs-based therapy: a novel and promising strategy for cancer treatment. *Microna*. 2017;6(2):102–109.
19. Tomasetti M, Amati M, Santarelli L, Neuzil J. MicroRNA in metabolic re-programming and their role in tumorigenesis. *Int J Mol Sci*. 2016;17(5):E754.
20. Olsson AK, Dimberg A, Kreuger J, Claesson-Welsh L. VEGF receptor signalling – in control of vascular function. *Nat Rev Mol Cell Biol*. 2006;7(5):359–371.
21. Leung DW, Cachianes G, Kuang WL, Goeddel DJ, Ferrara N. Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science*. 1989;246(4935):1306–1309.
22. Hicklin DJ, Ellis LM. Role of the vascular endothelial growth factor pathway in tumor growth and angiogenesis. *J Clin Oncol*. 2005;23(5):1011–1027.
23. Hanahan D, Folkman J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. *Cell*. 1996;86(3):353–364.
24. Kim M, Jang SK, Miller P, et al. VEGFR links self-renewal and metastasis by inducing Sox2 to repress miR-452, driving Slug. *Oncogene*. 2017;36(36):5199–5211.
25. Hsueh C, Lin JD, Lin IC, et al. Vascular endothelial growth factors and angiopoietins in presentations and prognosis of papillary thyroid carcinoma. *J Surg Oncol*. 2011;103(5):395–399.
26. Erdem H, Gundogdu C, Sipal S. Correlation of E-cadherin, VEGF, COX-2 expression to prognostic parameters in papillary thyroid carcinoma. *Exp Mol Pathol*. 2011;90(3):312–317.

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