

Preparation, Synergism, and Biocompatibility of in situ Liquid Crystals Loaded with Sinomenine and 5-Fluorouracil for Treatment of Liver Cancer

Jiaojiao Cao^{1,*}
Jie Huang^{1,*}
Shuangying Gui^{1,2}
Xiaoqin Chu^{1,2}

¹Department of Pharmaceutics, Anhui University of Chinese Medicine, Hefei, 230012, People's Republic of China;

²Institute of Pharmaceutics, Anhui Academy of Chinese Medicine, Hefei, 230012, People's Republic of China

*These authors contributed equally to this work

Purpose: Transarterial chemoembolization is the preferred treatment for patients with middle and advanced-stage hepatocellular carcinoma (HCC); however, most hepatic artery embolization agents have various disadvantages. The purpose of this study was to evaluate phytantriol-based liquid crystal injections for potential use in treatment of HCC.

Methods: Using sinomenine (SN) and 5-fluorouracil (5-FU) as model drugs, three precursor in situ liquid crystal injections based on phytantriol (P1, P2, and P3) were prepared, and their in vitro biocompatibility, anticancer activity, and drug release investigated, to evaluate their feasibility for use in treatment of HCC. The properties of the precursor injections and subsequent cubic liquid crystal gels were observed by visual and polarizing microscopy, in an in vitro gelation experiment. Biocompatibility was evaluated by in vitro hemolysis, histocompatibility, and cytotoxicity assays.

Results: Precursor injections were colorless liquids that formed transparent cubic liquid crystal gels on addition of excess water. The three precursor injections all caused slight hemolysis, without agglutination, and were mildly cytotoxic. Histocompatibility experiments showed that P1 had good histocompatibility, while P2 and P3 resulted in strong inflammatory responses, which subsequently resolved spontaneously. In vitro anti-cancer testing showed that SN and 5-FU inhibited HepG2 cells in a time- and concentration-dependent manner and had synergistic effects. Further, in vitro release assays indicated that all three preparations had sustained release effects, with cumulative release of >80% within 48 h.

Conclusion: These results indicate that SN and 5-FU have synergistic inhibitory effects on HepG2 cells, which has not previously been reported. Moreover, we describe a biocompatible precursor injection, useful as a drug carrier for the treatment of liver cancer, which can achieve targeting, sustained release, synergistic chemotherapy, and embolization. These data indicate that precursor injections containing SN and 5-FU have great potential for use in therapy for liver cancer.

Keywords: carcinoma, hepatocellular, drug carriers, delayed-action preparations, chemoembolization, therapeutic, SN, 5-FU

Introduction

Liver cancer,^{1,2} including hepatocellular carcinoma (HCC), cholangiocarcinoma, and other hepatic malignancies, is the sixth most common malignant tumor. Among primary liver cancers, HCC is one of the most common, according to cancer statistics updated in 2019, the estimated number of new HCC case ranks at 14th, while the estimated number of deaths from HCC ranked 5th among cancer-related

Correspondence: Xiaoqin Chu
Master Instructor
Tel +86 13856929921
Fax +86 551-68129028
Email chuxq420@163.com

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deaths. Currently, hepatic resection is the main treatment for patients with early-stage HCC, while transarterial chemoembolization (TACE) is preferred for patients with middle- and advanced-stage tumors,³ however, most traditional hepatic artery embolization agents have disadvantages, including low embolic selectivity, incomplete embolization, and insufficient embolization time, among others.^{4,5} Phytantriol (PYT)^{6,7} is an amphiphilic molecule with both hydrophilic hydroxyl groups and hydrophobic carbon chains. When PYT is dispersed in a small amount of water, a very low viscosity magnetic isotropic solution is formed, referred to as a precursor solution, which has the advantages of good mobility and easy injection; however, when PYT is dispersed in sufficient water and other organic solvents, it can form a high viscosity, rigid liquid crystal gel, which is non-toxic and biocompatible. Hence, the PYT precursor solution has the potential for use in TACE, and has been the subject of several reports.^{8–11}

As the most commonly used uracil anti-metabolite drug, 5-Fluorouracil (5-FU)^{12–14} has a broad spectrum of anti-tumor effects. Further, 5-FU is the most widely used drug for treatment of HCC, and has clear therapeutic effects, making it the first choice for treatment of advanced HCC; however, minimal bioavailability, low selectivity, obvious side effects, and drug resistance limit its clinical application. Thus, there is an urgent need to explore alternative therapeutic approaches to improve patient outcomes. Sinomenine (SN)¹⁵ is the main active alkaloid extracted from *Sinomenium acutum*; it has anti-inflammatory, analgesic, immunosuppressive, antihypertensive, and antiarrhythmic effects, and is primarily used for treatment of rheumatoid arthritis, chronic gastritis, morphine addiction, and other diseases. Recently, SN has been demonstrated to have strong and extensive anti-tumor effects,^{16,17} and when used in combination with 5-FU, achieves superior efficacy. Further, some studies have demonstrated that SN has potential as a 5-FU-based chemotherapy sensitizer,^{18–20} however, the effects of SN combined with 5-FU on liver cancer cells have not previously been reported.

Our previous studies have shown that liquid crystal gels based on PYT can be used in the articular cavity,²¹ skin,²² and periodontitis;²³ however, no systematic evaluation of the biocompatibility of PYT has been conducted. The primary objective of the present study was to construct a liquid embolic agent, based on PYT, using SN and 5-FU as model drugs, and investigate its biocompatibility, anticancer activity, and drug release, to assess its feasibility for use in TACE as a new therapeutic modality for patients with liver cancer.

Materials and Methods

Materials

Ethanol was provided by Sinopharm Chemical Reagent Co., Ltd. (Hefei, China). PYT (GC > 95%) was purchased from Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan). SN was supplied by Chengdu Must Biotechnology Co., Ltd. (Chengdu, China). 5-FU was produced by Shanghai Ika Biotechnology Co., Ltd. (Shanghai, China). Sterilized water for injection was purchased from Shanghai Xinyi Jinzhu Pharmaceutical Co., Ltd. (Shanghai, China). Methanol (chromatographic grade) was purchased from Sigma-Aldrich (Shanghai, China). Urethane (purity > 99.6%) was purchased from Shanghai TargetMol Biotechnology Co., Ltd. (Boston, USA). The HepG2 cell line was purchased from JENNIE-O Biological Technology Co., Ltd. (Guangzhou, China).

All animals were purchased from the Experimental Animal Center of Anhui Medical University (Hefei, China), and all procedures were approved by the Committee on Ethics of Animal Experiments of Anhui University of Chinese Medicine, and conducted in compliance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Preparation of Precursor Formulations

According to the literature^{8–11} and previous research,²¹ three solutions with ratios of PYT:ethanol:water (w/w) of 64:16:20 (P₁), 48:32:20 (P₂), and 16:64:20 (P₃), were selected for use in development of a TACE carrier. Briefly, PYT and ethanol were mixed in a water bath at 37°C ± 0.5°C, and the designated amounts of water simultaneously preheated. Melted PYT and ethanol were vortexed, to generate a uniform oily phase, which was then vortex-mixed with the prescribed amount of water. Mixtures were then centrifuged at 3000 rpm for 10 min, to obtain the precursor liquids. Drug-loaded preparations (drug-loaded P₁, drug-loaded P₂, and drug-loaded P₃) were formulated by first dissolving the drugs in water, and then performing the operations described above. All samples were equilibrated at room temperature for at least one week and filtered through a sterile filter (0.22 µm), prior to use in experiments.

Hemolysis Test

Preparation of Red Blood Cell Suspensions

In vitro hemolysis assays were conducted using red blood cells (RBCs). The hemolytic activity of samples was

analyzed by quantification of the released hemoglobin by spectrophotometric measurement. Briefly, one New Zealand rabbit (male, 2.2 kg) was used in this study. We collected 18 mL blood by heart puncture, and placed it into a pre-prepared centrifuge tube, containing 2 mL 3.8% sodium citrate, to prepare fresh anticoagulated blood. Blood was slowly stirred using a glass rod to remove fibrin, then diluted in physiological saline (PS) at a 1:1 volume ratio, and centrifuged at 3000 rpm for 10 min. The supernatant was removed, the remaining sample diluted 1:10 in PS, and then purified and washed by centrifugation (3000 rpm, 10 min, 5 times) to collect the RBCs. Finally, precipitated RBCs were diluted with PS to prepare a 2% RBC suspension.²⁴

Experimental Design

Before the experiment, fluid extracted from the precursor solution²⁵ was prepared by adding approximately 0.5 g of drug-free precursor solution in 5 mL PS, incubating at 37°C for 72 h, then filtering through a sterile filter (0.22 µm). To control for the effects of other components in the precursor solution, PYT, 64% ethanol, 32% ethanol, and 16% ethanol were treated in the same manner as described above. Groups used for the experimental design of the hemolysis experiment are shown in Table 1. Briefly, 1.5 mL of extracts were added to tubes labeled A–G, with 1.5 mL PS in test tube H, as a negative control, and 1.5 mL distilled water in test tube I, as the positive control group. Then, 2% RBCs suspension was added into each tube, the mixtures gently vortexed, and incubated for 4 h at 37°C. All samples were analyzed in triplicate.

Table 1 Hemolysis Experiment Group Design

Extraction (mL)	Test Tube Number								
	A	B	C	D	E	F	G	H	I
P1	1.5	0	0	0	0	0	0	0	0
P2	0	1.5	0	0	0	0	0	0	0
P3	0	0	1.5	0	0	0	0	0	0
PYT	0	0	0	1.5	0	0	0	0	0
16% Ethanol	0	0	0	0	1.5	0	0	0	0
32% Ethanol	0	0	0	0	0	1.5	0	0	0
64% Ethanol	0	0	0	0	0	0	1.5	0	0
PS	0	0	0	0	0	0	0	1.5	0
Distilled Water	0	0	0	0	0	0	0	0	1.5
2% RBCs Suspension	1.5	1.5	1.5	1.5	1.5	1.5	1.5	1.5	1.5

Determination of Hemolysis Rate

First, the presence or absence of hemolysis in each group of tubes was observed by visual inspection of RBC fragmentation and sedimentation. If RBCs sank, tubes were gently shaken to determine whether they could be resuspended and assess whether agglutination had occurred. Next, tubes were centrifuged at 1000 rpm for 10 min, 150 µL aliquots of supernatant from each sample transferred to a 96-well plate, and absorbance recorded using a microplate reader (Biotek, ELX800, USA). The maximum absorption wavelength (λ_{\max}) was determined by UV/Vis spectroscopy as follows: supernatants from the positive control group were placed in quartz cuvettes with a 1 cm path in a total volume of 2.0 mL, then UV/Vis spectra (200–800 nm) were recorded to determine the λ_{\max} value. To express the in vitro hemolytic properties of our samples, the percentage of hemolysis was calculated as follows:²⁶

$$\text{Hemolysis (\%)} = (S_{\text{absorbance}} - H_{\text{absorbance}}) / (I_{\text{absorbance}} - H_{\text{absorbance}}) \times 100$$

where $S_{\text{absorbance}}$, $H_{\text{absorbance}}$, and $I_{\text{absorbance}}$ refer to the absorbance values of the sample, negative control group, and positive control group, which were the mean values of measured absorbance from test tubes A–G, H, and I, respectively.

Observation of Red Blood Cell Morphology

The sediment collected in the tubes by centrifugation (as described above) was resuspended in PS, a drop placed on a glass coverslip, and another coverslip placed over it. The coverslips were then gently pulled apart horizontally and allowed to air dry. This technique provides uniform smear thickness and staining. Smears were then flooded with Wright's stain for 2 min, followed by flooding with water for 4 min, drying, mounting on a glass slide, and examination by light microscopy.

In vivo Injectability and Histocompatibility

Subcutaneous injection in Sprague Dawley rats is routinely used to characterize foreign body reactions to polymeric biomaterials. The size of the animals allows for clinically relevant injection volumes (500–1000 µL) to be tested;²⁷ hence, the foreign body response to the precursor formulations created using varying concentrations of PYT/ethanol/H₂O was assessed, using a subcutaneous pouch model in Sprague-Dawley rats (male, 200 ± 20 g). The precursor formulations were sterilized before injection as liquid embolic agents. Rats were anesthetized by

intraperitoneal injection of pentobarbital sodium solution (30 mg/kg). There were four experimental groups: P1, P2, and P3, which were injected with 500 μ L of the indicated solutions, and PS, which was injected with 500 μ L PS, as a negative control.

The subcutaneous tissue surrounding the hydrogel was evaluated by histochemical staining. In brief, rats were sacrificed at the indicated time points and 0.5 g subcutaneous tissue samples from close to the hydrogel cut into fragments and fixed in paraformaldehyde solution at 4°C for further analysis. Tissue samples were subjected to hematoxylin-eosin (HE) staining to determine the degree of the inflammatory response. All images were acquired using a general optical microscope.

In vitro Cytotoxicity

The in vitro cytotoxicity of samples was determined by MTT assay. The principle of this assay is based on the fact that succinate dehydrogenase in the mitochondria of living cells can reduce exogenous MTT to water-insoluble blue-purple crystalline formazan, which is deposited in cells, while dead cells cannot perform this function. Dimethyl sulfoxide (DMSO) can dissolve the formazan in the cells, and its light absorption value is then measured using an enzyme-linked immunosorbent assay at a wavelength of 540 nm, with the results indirectly reflecting the number of living cells.

Cell Culture

The human hepatoma cell line, HepG2, was used for in vitro cytotoxicity experiments. The cell line was grown in Dulbecco's Modified Eagle medium (DMEM), supplemented with 10% fetal bovine serum, 100 μ g/mL penicillin, and 100 μ g/mL streptomycin. Cells were incubated in a saturated humidity incubator (37°C, 5% CO₂), washed with phosphate-buffered saline (PBS), and digested using 0.25% trypsin to form a single cell suspension. Cells in logarithmic growth phase were tested.

Preparation of Sample Extracts

With reference to the relevant standards (GB/T 16886.5—2017 and ISO 10993-5:2009), 1 g of the blank preparation, P1, P2, P3, and PYT were injected into 5 mL of DMEM, and leached at 37°C for 48 h. Then, samples were filtered through 0.22 μ m sterile membranes, and the filtrates used as stock solutions. Next, the stock solutions were diluted in different proportions (1:0; 1:1; 1:2; 1:4;

1:8; 1:16; 1:32; 1:64; 1:128, v/v) of DMEM culture medium, as test solutions.

In vitro Cytotoxicity Assay

Briefly, 100 μ L of DMEM was added to the peripheral wells of a 96-well tissue culture microtiter plate, and 100 μ L of HepG2 cell suspension (1×10^4 cells/mL) added to the remaining wells. After incubating for 24 h, the medium was removed and 100 μ L of the different concentrations of stock solutions diluted with fresh medium added to each well, with 100% DMEM as a negative control ($n = 3$). After incubation for 72 h, 20 μ L MTT solution (5 mg/mL) was added to each well and the cells incubated at 37°C for an additional 4 h. Subsequently, the MTT solution was removed, replaced with 100 μ L DMSO, and the mixture shaken at a low speed for 10 min on a shaker to fully dissolve the crystalline formazan. The optical density (OD) at 540 nm of each well was measured using a microplate reader. To express the in vitro hemolytic properties of our samples, cell viability was calculated using the following formula:

$$\text{Cell viability (\%)} = \text{OD}_{570\text{S}} / \text{OD}_{570\text{N}} \times 100$$

where OD_{570S} represents the average optical density of each tested sample group, and the OD_{570N} represents the average optical density of the negative control group. According to the ISO standard (GB/T 16886.5—2017/ISO 10993-5:2009), cell viability was then classified according to percentage of viable cells, to reflect the degree of toxicity of the material as follows: 0, 100%; 1, 80–100%; 2, 50–80%; 3, 30–50%; 4, 0–30%.

In vitro Anticancer Activity

To test the in vitro anticancer effects of SN and/or 5-FU solutions, according to the literature^{12–20} and preliminary experimental results, the stock solution concentration of 5-FU was 640 μ g/mL, while that of SN was 1280 μ g/mL, and the stock combination solution containing was 640 μ g/mL 5-FU + 1280 μ g/mL +SN. Four groups were included in the experiment as follows: negative control group (only DMEM culture medium added), 5-FU monotherapy group, SN monotherapy group, and the combination group. All solutions were formulated and diluted with DMEM medium at ratios of 1:0, 1:1, 1:2, 1:4, 1:8, 1:16, 1:32, 1:64, and 1:128 (v/v), then filtered through a 0.22 μ m microporous membrane before use in experiments. Cell proliferation was determined in vitro by MTT assay, as described above, except that cell viability was detected at 24, 48, and 72 h. Based on

the results of these experiments, drug loading of SN in precursor injections was set to 0.5%, that of 5-FU to 0.25%, and the combined drug loading to 0.5% + 0.25% (SN + 5-FU). Drug-loaded precursor solutions were extracted as described above and diluted with the culture solution to obtain a gradient of sample concentrations.

To analyze the drug interaction between SN and 5-FU, the co-administration efficacy analysis software, CalcuSyn 2.1, was used to analyze the inhibition rates (IRs) according to Chou and Talalay,^{28,29} and combination index (CI) values, a dose-effect curve, and a median-effect plot were obtained. The median-effect plot is a graph of $x = \log(D)$ vs $y = \log(fa/fu)$, based on the median effect equation (a general equation for the dose-effect relationship):

$$fa/fu = (D/D_m)^m$$

where fa is the fraction affected by the dose, fu is the fraction unaffected ($fu = 1 - fa$), D is the drug concentration or dose, D_m is the median-effect dose or concentration (usually expressed as ED_{50} or IC_{50}), m is an exponent signifying the sigmoidicity (shape) of the dose-effect curve; $m = 1$, >1 , and <1 indicate hyperbolic, sigmoidal, and negative sigmoidal shapes, respectively. CI is a quantitative measure of the degree of drug interaction, in terms of additive effect ($CI = 1$), synergism ($CI < 1$), or antagonism ($CI > 1$), for a given endpoint of the effect measurement.

In vitro Drug Release

The dialysis membrane method was used to study in vitro drug release. In summary, approximately 0.5 g of the precursor injection containing SN and/or 5-FU was injected into a dialysis bag (MWCO 1000), which was suspended in a centrifuge tube containing an appropriate amount of PS, and subjected to a release test in a shaker at constant temperature ($37^\circ\text{C} \pm 0.5^\circ\text{C}$, 100 rpm). Simultaneously, equal amounts of free SN and 5-FU solution were treated in the same manner. All of the release medium was removed at predetermined intervals and replenished with an equal amount of fresh isothermal release medium. All samples were filtered through a $0.45 \mu\text{m}$ microporous membrane and assayed using a validated HPLC method²² to calculate the cumulative release percentage. Then, the function of the cumulative release percentage and time was drawn to predict the release mechanism.

Statistical Analysis

All data are expressed as mean \pm SD, and statistical analyses were performed using SPSS 22.0. $P < 0.05$ or $P < 0.01$ were considered to indicate significant differences.

Results

Precursor Formulations

As shown in Figure 1, the precursor injections prepared in this study were all colorless, uniform solutions, which transformed into transparent gels after adding excess water at 37°C , followed by centrifugation at 8000 rpm for 10 min to remove excess water. The precursor injection and gel were observed under a polarized light microscope, and all showed dark fields, indicating that the precursor injections were isotropic solutions, and that the gels were in cubic phase.²¹

In vitro Hemolysis Test

Intravascular hemolysis increases the amount of plasma-free hemoglobin, which can cause toxic reactions or affect the kidneys and other organs. Further, the release of phospholipids can promote thrombosis. Moreover, when hemolysis causes a clinically significant decrease in the number of RBCs, it can lead to anemia and impair oxygen-carrying capacity, with secondary effects on the brain and other organs or tissues.³⁰ Since precursor injections are used for arterial embolization and direct contact with blood, we performed a red blood cell hemolysis experiment using the precursor injection, to assess its hemocompatibility.

As shown in Figure 2A, the negative control solution (tube I) was red, and there was no RBC residue at the bottom of the tube; however, the upper solutions of the other groups were colorless and clear, accompanied by sinking RBCs, that could be resuspended by shaking the tube, indicating good hemocompatibility and no RBCs agglutination. RBCs collected in the base of the tube were stained using Wright's dye. RBC fragments, including red cytoplasm and blue nuclei, were observed under the microscope in the negative control group, while the morphology of RBCs in the other groups was normal (Figure 2B). The hemolysis rate of the precursor formulations at 538 or 575 nm (λ_{max} at 538 or 575 nm) was lower than 2% (Figure 2C), while that in the PYT group was much lower (0.10% at 538 nm and 1.15% at 575 nm), and the hemolysis rate of the different concentrations of ethanol was slightly higher ($<10\%$) (Figure 2D).

In vivo Injectability and Histocompatibility

Protuberances were formed immediately after injection of 0.5 mL of the precursor solutions into the dorsal skin of rats; however, only P1 generated a circular bulge, while P2 and P3 were dispersed into flakes (Figure 3A). After 15 min, rat skin samples with liquid crystalline gel attached were excised to

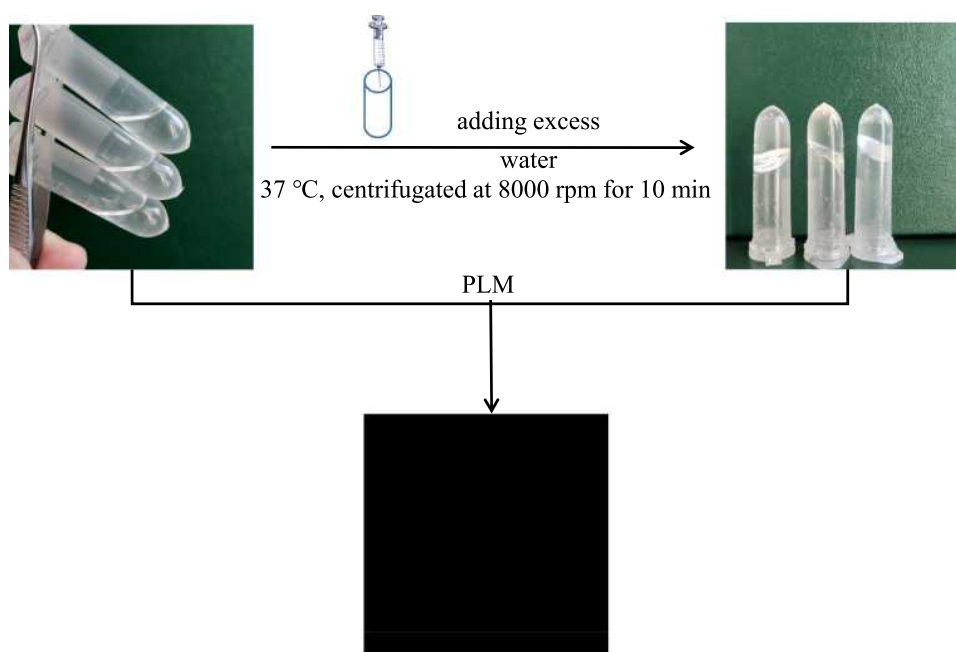


Figure 1 The appearance of the precursor injection before and after the formation of the in situ liquid crystal gel.

observe the formation of the gel. P1 formed a spherical liquid crystalline gel, while P2 and P3 formed plate-like gels, and P3 spread across a wider area (Figure 3B). Overall, each precursor injection formed a liquid crystal gel, with gelling properties, in vivo. The saline injection also formed a bulge; however, it quickly dispersed (Figure 3A).

Changes of the injection site for each precursor injection were visually observed during subsequent experiments, and pathological changes in tissue samples isolated from the gel injection site analyzed by HE staining (Table 2 and Figure 3C). The results of the histocompatibility experiment revealed erythema and a few inflammatory cells in the P1 group on the first day; however, samples in this group soon returned to normal and no other pathological reactions were observed at later stages, indicating superior histocompatibility. In contrast, the P2 and P3 groups showed strong stimulation from the 1st to 20th days, with skin reactions manifesting as edema, necrosis, and ulcers, and HE staining revealing substantial inflammatory cell infiltration; however, this recovered spontaneously after 20 days, with the P3 group taking longer to recover than the P2 group.

In vitro Cytotoxicity

In vitro Cytotoxicity of Blank Preparations

To evaluate the cytotoxicity of the precursor injections rapidly and economically, in vitro tests of the precursor

injection solutions were conducted using an MTT assay. The results are shown in Figure 4. After 72 h of incubation, the viability of cells treated with the three types of blank preparation was >80% (grade 1), and increased with rising dilution ratio, suggesting good cell compatibility of the blank preparation, with only mild cytotoxicity. Furthermore, the cytotoxicity of the PYT material was also measured and the same conclusion reached, supporting the safety of the matrix agents (cell viability > 80%; grade 1).

In vitro Anticancer Activity

The viability of HepG2 cells after treatment with SN and/or 5-FU solutions at different concentrations and times is shown in Figure 5A. As expected, SN and/or 5-FU inhibited cell viability, which decreased with increasing concentrations of SN, 5-Fu, or the two combined. Incubation time also significantly influenced cell viability, with prolonged incubation time associated with decreased viability. Further, cell viability was more strongly reduced by the combination of SN and 5-FU than either alone, indicating that their inhibitory effects on HepG2 cells may be synergistic. Cell viability rates of extracts treated with different drug-loaded solutions, for various incubation times, at several dilution ratios following application of precursor solutions are presented in Figure 5 (Figure 5B–D). Extracts from each group of drug-loaded preparations

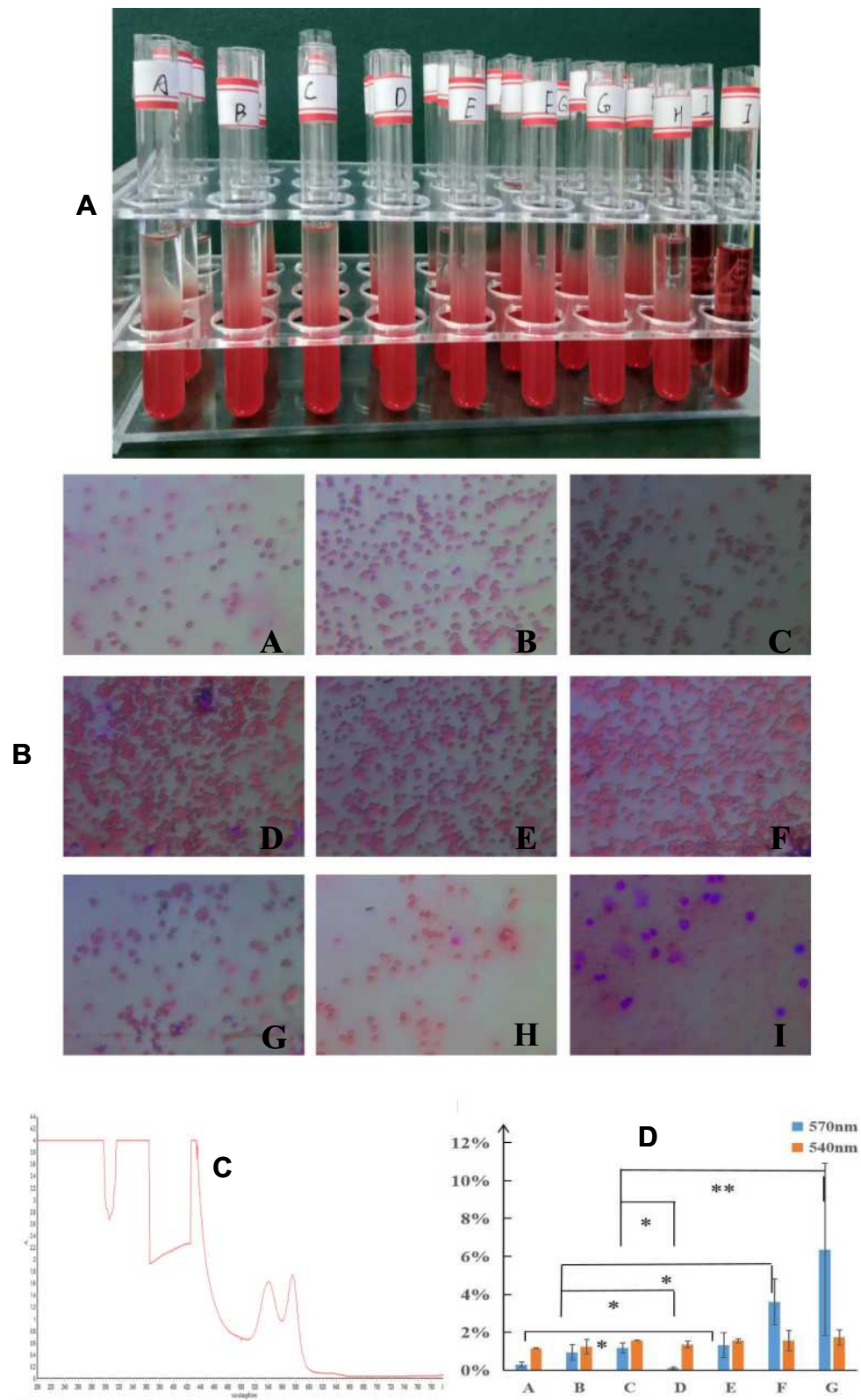


Figure 2 Hemolysis testing: (A) appearance of each group of samples in RCBs solution; (B) RCBs morphology in each group; (C) determination of the λ_{max} ; (D) hemolysis rate (means \pm SD, $n = 3$; * $p < 0.05$, ** $p < 0.01$).

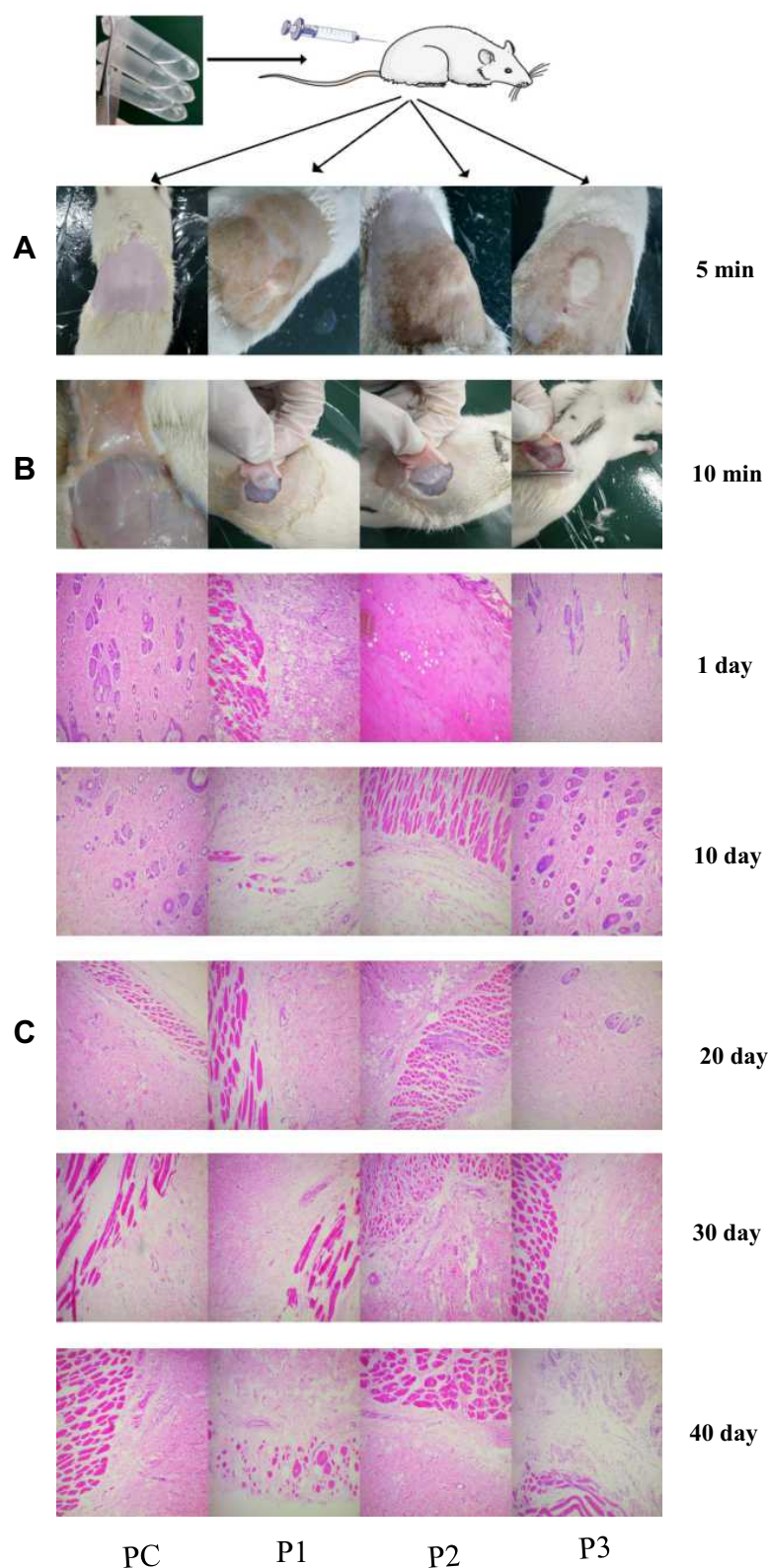


Figure 3 The results of histocompatibility test: **(A)** appearance after subcutaneous injection for 5 min; **(B)** liquid crystal gel state after subcutaneous injection for 15 min; **(C)** HE staining results of tissue surrounding the liquid crystal gels after subcutaneous injection of the precursor injection into SD rats at the indicated number of days (scale bar: 1:100).

Table 2 The Results of Histocompatibility Experiment

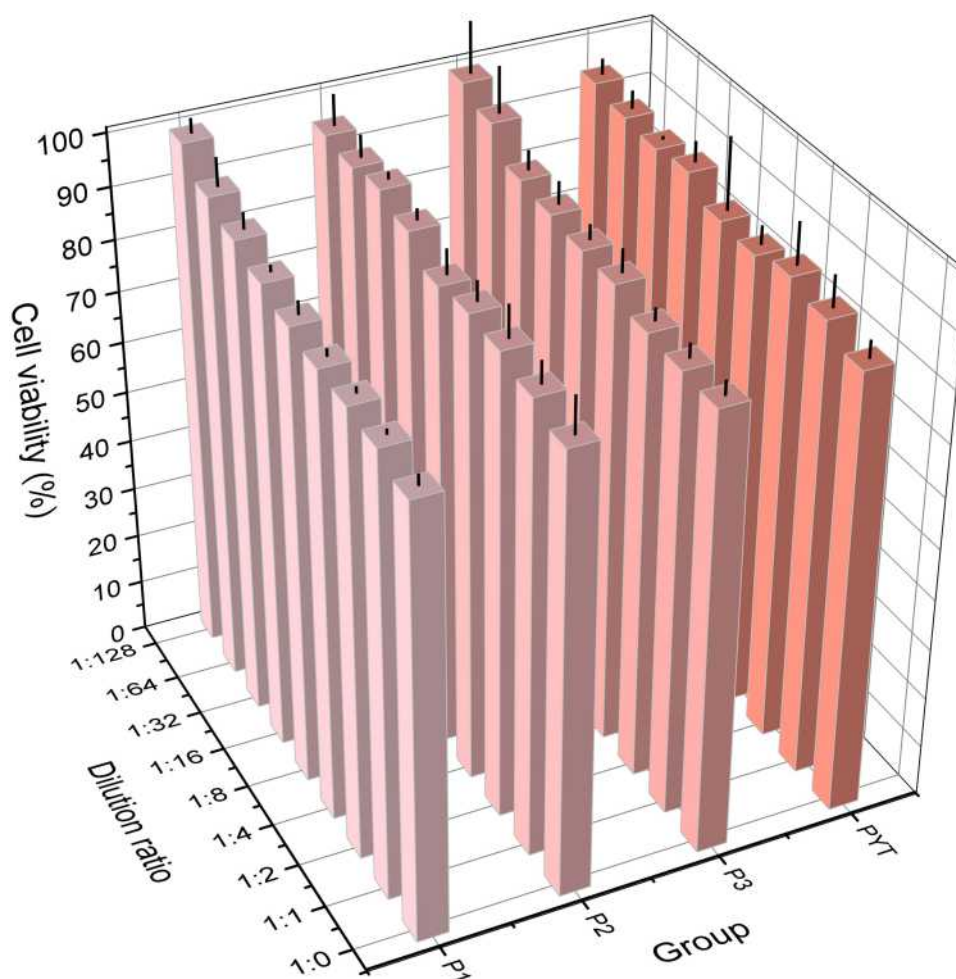
Group Time	P1		P2		P3	
	Skin*	HE [#]	Skin	HE	Skin	HE
1 day	++	++	+++	++	+++	++
10 day	+	+	++++	++++	++++	++++
20 day	+	+	++++	++++	++++	++++
30 day	+	+	++	++	+++	+++
40 day	+	+	+	+	++	++

Notes: *: + Basically no lesions; ++ erythema, eschar; +++ necrosis, oedema; ++++ corrosion, ulceration. [#]: +basically no inflammatory cells; ++A few lymphocytes; +++lymphocyte infiltration, visible neutrophils; ++++ neutrophil infiltration, visible phagocytic cells.

had clear inhibitory effects on the growth of liver cancer cells. Further, prolonged incubation time and decreased dilution ratio, led to additional reductions in cell viability rates in each group. Furthermore, in vitro cell viability in response to the combined drug-loaded preparation was lower relative to formulations containing each drug

alone. This effect was similar to the in vitro anticancer effects of SN and 5-FU solutions; that is, it was concentration-dependent, time-dependent, and synergistic.

Next, we analyzed the interaction between SN and 5-FU using CalcuSyn 2.1 software, generating median-effect plots, dose-effect curves, and algebraic estimates for each drug solution at different times (Figures 6A, 24 h; Figures 6B, 48 h; Figures 6C, 72 h). The inhibitory effect of each drug solution on HepG2 cells was typically concentration- ($r > 0.9$) and time-dependent. Moreover, the CI values for all incubation times were <1 (Table 3). Quantification of CI values at ED₅₀ (24 h, 0.1239; 48 h, 0.3534; 72 h, 0.0370), ED₇₅ (24 h, 0.0564; 48 h, 0.2924; 72 h, 0.4764), and ED₉₀ (24 h, 0.0262; 48 h, 0.2794; 72 h, 0.9283) for each drug combination are presented in Table 3. Further, combination of the two drugs led to a significant reduction in Dm value ($p < 0.05/0.01$). Overall, these findings indicate that the inhibitory effects of SN and 5-FU on HepG2 cells were synergistic.

**Figure 4** Cytotoxicity of the blank precursor injections and PYT (means \pm SD, $n = 3$).

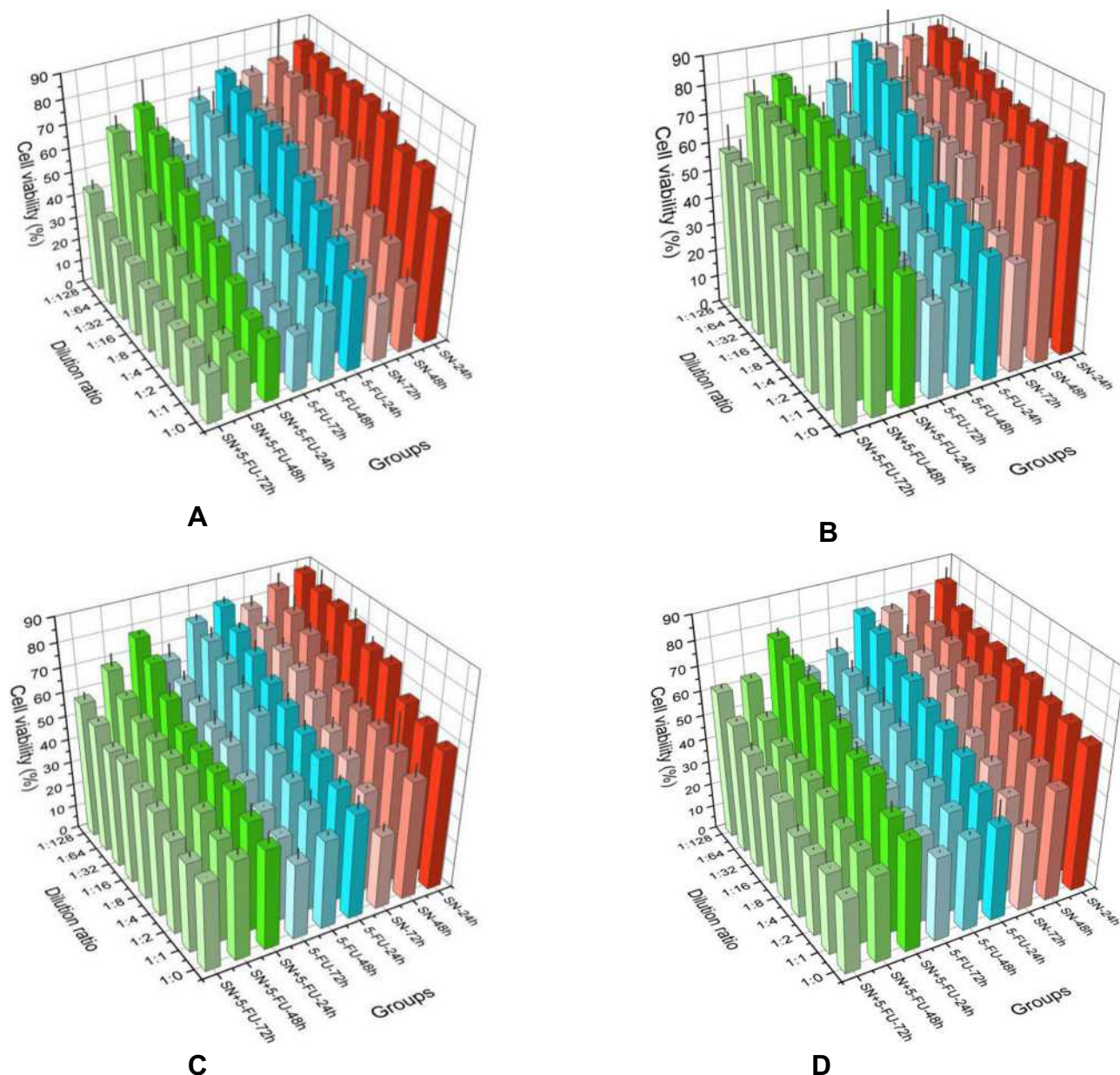


Figure 5 The results of in vitro anticancer activity at different time and dilution ratios: (A) SN and/or 5-FU solution; (B) drug-loaded P1; (C) drug-loaded P2; (D) drug-loaded P3 (means \pm SD, $n = 3$; at the same incubation time of the same prescription: SN or 5-FU VS SN+5-FU, * $p < 0.05$, ** $p < 0.01$).

In vitro Drug Release

As described above, extracts of drug-loaded preparations suppressed the growth of liver cancer cells, relative to extracts from the blank control preparation. This could be a consequence of release of the drugs from the preparation into the liver cancer cells; therefore, we studied the in vitro release of the drug-loaded preparations. Each drug-loaded preparation (both double- and single-loaded) contained 0.5% SN and/or 0.25% 5-FU. The in vitro release profile of the two drugs within 48 h was as follows: release of both drugs reached

approximately 90% within 48 h, and there was a significant burst phenomenon at 0.5 h (where release of the two drugs was $>30\%$) (Figure 7A, SN; Figures 7B, 5-FU). The amounts of the drugs released were close to 100% within 4 h, indicating that the drug-loaded preparations exhibited significant sustained release ($p < 0.05$).

In addition, to visually express the release behaviors and patterns of the preparations, release data from the first 12 h were fitted to three commonly used mathematical models: Higuchi, first-order, and zero-order equations. Linear

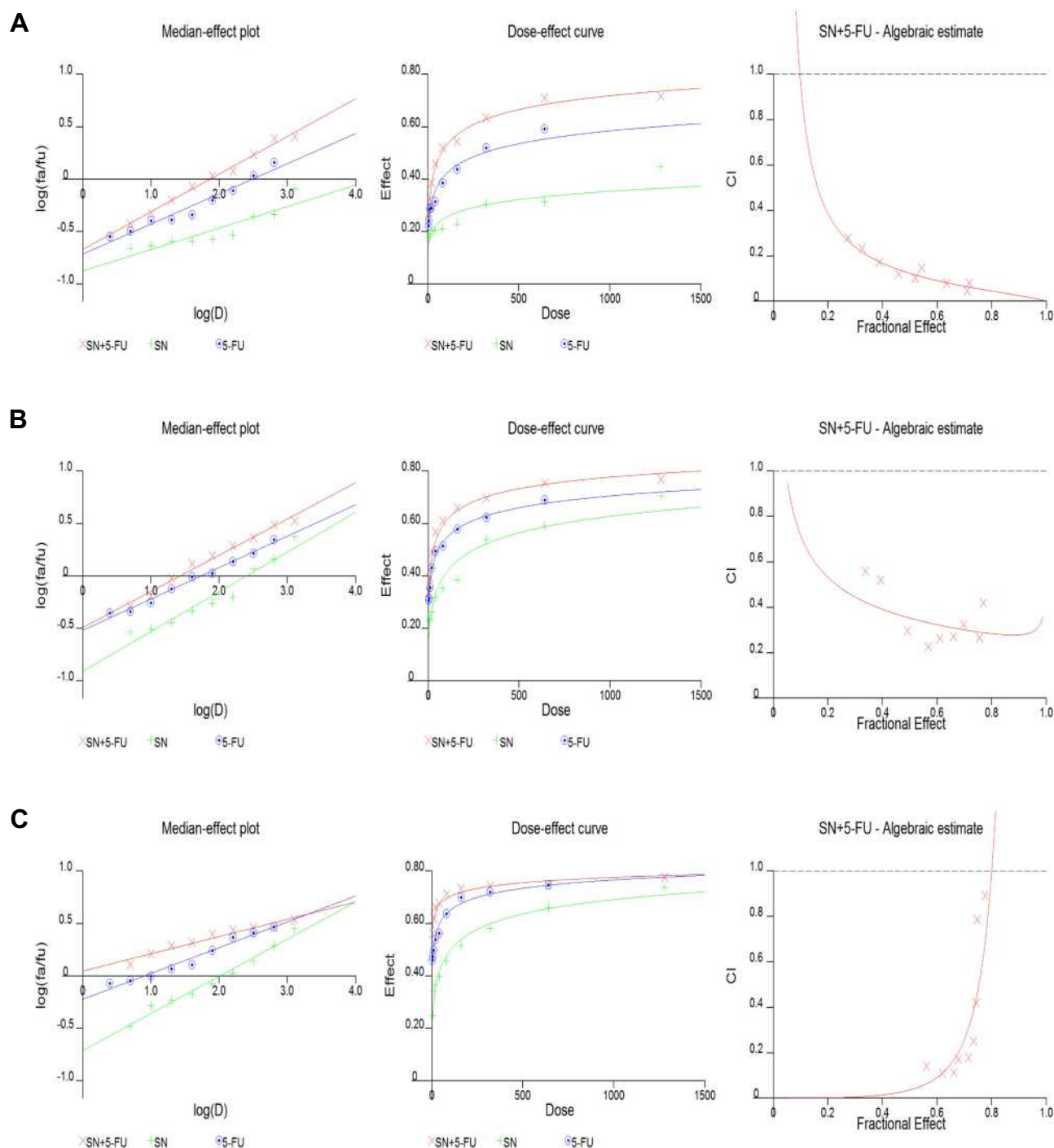


Figure 6 Median-effect plot, dose-effect curve and algebraic estimate generated by CalcuSyn 2.1 software for Hep G2 cells treated by various different dilution ratios of SN and/or 5-FU solution at (A) 24h, (B) 48h and (C) 72h.

correlation coefficients are presented in Table 4. The r values for the SN and 5-FU release curves indicated that release of these drugs conforms to first-order kinetics. Although the r value for the Higuchi equation was the largest for an individual group, it did not differ substantially from that for the first-order equation. Overall, the release of the drug

was consistent with the first-order equation, indicating that drug release was dominated by swelling.³¹

Discussion

The precursor injections prepared in this experiment were clear, transparent solutions, that could form cubic liquid

Table 3 The Results of Hep G2 Cells Treated by Various Different Dilution Ratios of SN and/OR 5-FU Solution by CalcuSyn 2.1 Software (Including CI, r, Dm, M; Compared with the Combined Group of the Same Culture Time, * $p < 0.05$, ** $p < 0.01$)

Drug	Time (h)	CI Values at			r	Dm	m
		ED ₅₀	ED ₇₅	ED ₉₀			
SN	24	N/A	N/A	N/A	0.9046	18908.0000**	0.2041
	48	N/A	N/A	N/A	0.9697	250.3044**	0.3781
	72	N/A	N/A	N/A	0.9990	101.8780**	0.3538
5-FU	24	N/A	N/A	N/A	0.9779	303.4755**	0.2871
	48	N/A	N/A	N/A	0.9927	53.7864*	0.2990
	72	N/A	N/A	N/A	0.9836	7.8814**	0.2456
SN+5-FU	24	0.1239	0.0564	0.0262	0.9946	72.8535	0.3585
	48	0.3534	0.2924	0.2794	0.9907	26.5875	0.3448
	72	0.0370	0.4764	0.9283	0.9725	0.5050	0.1640

crystals on removal of excess water. Previous rheological experiments³² have shown that precursor injections exhibit Newtonian fluid properties at high shear rates, where they exhibit lower viscosity; however, when the cubic liquid

crystals formed, solid and elastic properties were dominant, indicating that all three liquid embolic agents had good injectability and adhesion, as well as being able to resist blood flow clearance.

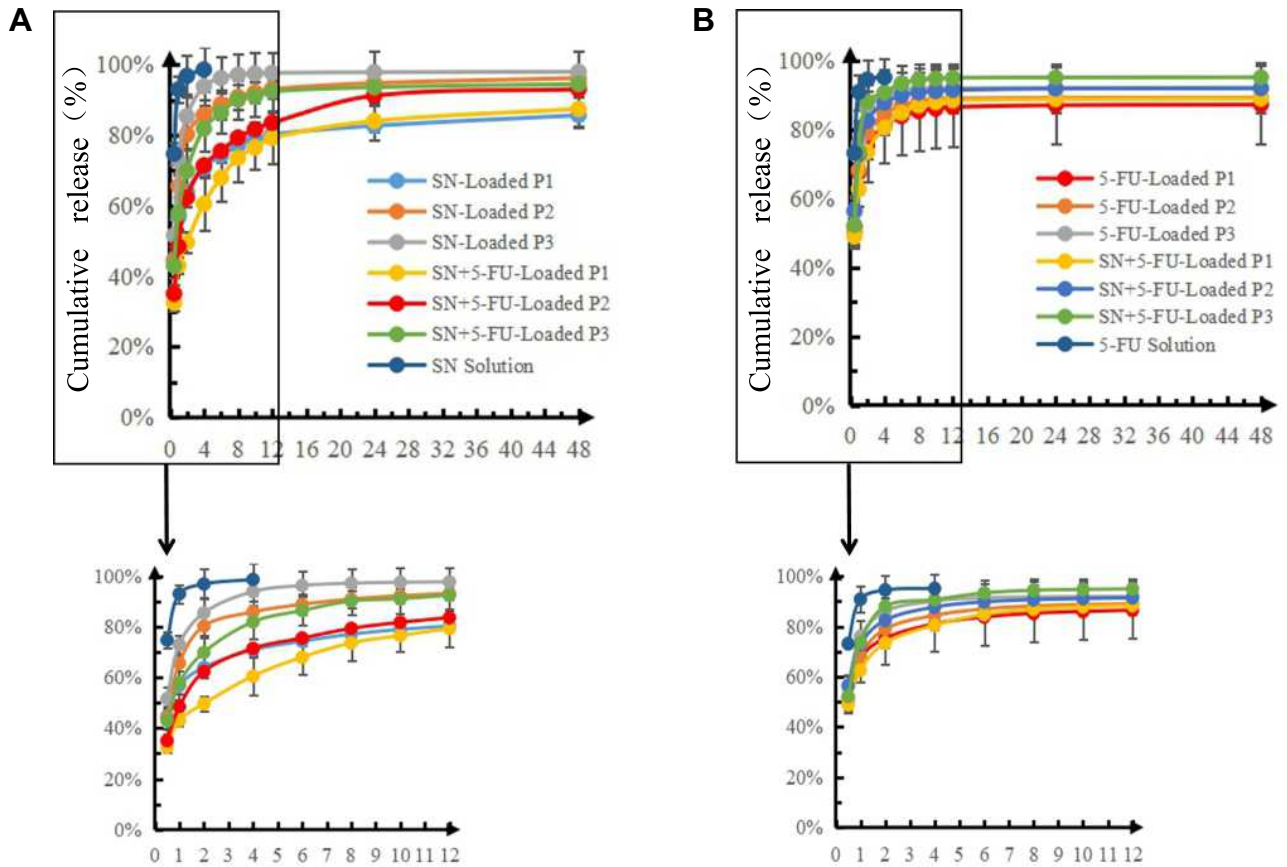


Figure 7 The results of in vitro drug release of SN (A) and 5-FU (B).

Table 4 r Values of Release Curves Fitted by Different Equations

Drug	Group	r		
		Higuchi Equation	First Order Equation	Zero Order Equation
SN	SN-Loaded P1	0.9507	0.9438	0.8887
	SN-Loaded P2	0.8772	0.9228	0.7925
	SN-Loaded P3	0.8626	0.9243	0.7704
	SN+5-FU-Loaded P1	0.9886	0.9856	0.9498
	SN+5-FU-Loaded P2	0.9491	0.9528	0.8843
	SN+5-FU-Loaded P3	0.9432	0.9618	0.8736
5-FU	5-FU-Loaded P1	0.8926	0.8917	0.8106
	5-FU-Loaded P2	0.8667	0.8728	0.7776
	5-FU-Loaded P3	0.8078	0.8105	0.7088
	SN+5-FU-Loaded P1	0.9241	0.9276	0.8485
	SN+5-FU-Loaded P2	0.8627	0.8714	0.7723
	SN+5-FU-Loaded P3	0.8279	0.8736	0.7332

To ensure the accuracy of the hemolysis test, we determined the λ_{\max} of oxyhemoglobin by full-wavelength scanning. The results showed that the hemolysis rates of the preparations and PYT were well below the safe value (5%) specified in the relevant standard (GB/T 16886.4—2003/ISO 10993-4:2002), while different concentrations of ethanol resulted in slight hemolysis (<10%), indicating that the hemolysis in response to the preparations may be attributable to the ethanol in the solution. Reports indicate that liquid crystal nanoparticles based on PYT exhibit some cytotoxicity and hemolysis, likely because the meso-phase structure of the nanostructures is present in biofilms, and when the liquid crystal nanoparticles undergo image transformation, the high concentration of cationic lipids can interact with cell membranes and destroy their integrity, leading to cell lysis and death.^{33,34}

In vivo gelation properties and histocompatibility were investigated by subcutaneous injection of precursor solutions into the backs of rats. The results showed that all the precursors could form liquid crystalline gels; however, only P1 formed a spherical shape, while P2 and P3 dispersed into flakes, possibly related to the gelation time of each solution; if the gelation time is faster, gels can be formed in situ before the injection disperses, whereas with slow gelation times, injections exhibit more dispersibility. This suggests that the time of injection should be considered when performing arterial embolization in vivo. If the bolus time is too fast, the injection may flow to other areas before the gel is formed, resulting in ectopic embolization. Histopathological results showed that P1 had good histocompatibility, while P2 and P3 elicited inflammatory

responses; however, the latter subsequently recovered spontaneously. High ethanol concentrations generally cause severe inflammation and subsequent necrosis.^{35,36} Comparison of the P1, P2, and P3 solutions indicates that the inflammation associated with P2 and P3 may be caused by excessive ethanol in their solutions. Therefore, P1 is recommended for subsequent in vivo studies.

Unlike traditional methods, the preparations were first diluted with medium before use for in vitro cytotoxicity tests (including anticancer experiments), according to the proposal of Zhai et al.³³ Injection of precursor solutions is accompanied by release of a large amount of drug during the formation of liquid crystalline gel,^{8,9} as confirmed by our in vitro drug release tests; therefore, if the liquid crystal gel forms before application in an anti-cancer experiment, the amount of drug released may be reduced, potentially influencing the accuracy of the findings. Based on our preliminary results from experiments conducted using direct injections, the water in the medium was absorbed to form a liquid crystal gel, resulting in a decrease in medium content, which may starve the cells and affect their proliferation. To investigate the true state of drug-loaded preparations, we did not induce the precursor injection to form liquid crystal gel before the experiment, rather we directly extracted the injection and then carried out the experiment.

Our results show that SN and 5-FU have clear synergistic effects across a broad concentration range, which were time- and concentration-dependent. Various studies have shown that expression levels of Bax and Bcl-2 are up- and down-regulated, respectively, during the apoptosis

induced by SN and 5-FU,^{18–20} indicating that the mechanisms underlying this effect may be associated with increased apoptosis via the mitochondrial pathway and downregulation of thymidylate synthase mRNA expression. It has been suggested that, in the course of 5-FU chemotherapy for patients with liver cancer, addition of SN can greatly reduce the dose of 5-FU required, produce a synergistic effect, and reduce the side effects of 5-FU. Indeed, our data show that use of SN and 5-FU in our precursor injection for TACE led to good synergistic effects. Further, the blank formulations showed very little cytotoxicity, indicating that the effects of the drug-loaded formulation are likely mainly attributable to drug release. Hence, this combination has potential for treatment of liver cancer.

Since both SN and 5-FU are poorly soluble in water, high drug loading would lead to drug precipitation, particularly when they are used in combination; therefore, we used a drug loading formulation containing 0.5% SN and 0.25% 5-FU. The results of the in vitro release test showed that drug release exhibited a burst phenomenon and that higher levels of ethanol in the solution led to increases in the amount and rate of drug release. During the process of water absorption by the precursor solution to form a liquid crystal gel, ethanol in the preparation is exchanged with water, releasing large amounts of the drug into the medium, resulting in the observed burst behavior. In addition, higher proportions of ethanol in solutions are associated with lower viscosity and higher fluidity, which increases the contact area between the preparation and the release medium, leading to elevated drug release.^{8,33} These data suggest that, when this preparation is used in vivo, it can quickly reach the blood concentration required in the human body and produce effective treatment, while simultaneously reducing the side effects of 5-FU.

Conclusion

The results obtained in this investigation demonstrate that all three of the precursor injections we selected exhibit good biocompatibility and good gelation properties, both in vitro and in vivo. Our data also prove that SN and 5-FU synergistically inhibit the growth of HepG2 cells. Further, we evaluated the in vitro anticancer activity and release of SN- and/or 5-FU-loaded precursor injections and our results show that all three formulations can significantly inhibit the growth of HepG2 cells, while the combined drug-loaded group had synergistic effects. The in vitro release profile was generally consistent with first-order-

release kinetics, and demonstrated a significant sustained release effect. The results of this study emphasize the biocompatibility of these three precursor injections for use in TACE and the interaction of SN and 5-FU in inhibition of HepG2 cell growth, providing a theoretical reference for clinical treatment of liver cancer; however, further research is required, including study of the feasibility of embolization and in vivo pharmacokinetic and pharmacodynamic investigations. We will conduct extensive experimental research in the future, with the aim of developing our liquid embolic agent for use in TACE and improving therapeutic efficacy.

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Disclosure

The authors report no conflicts of interest in this work.

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