


Ginkgo biloba Extract 50 (GBE50) Ameliorates Insulin Resistance, Hepatic Steatosis and Liver Injury in High Fat Diet-Fed Mice

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Background: *Ginkgo biloba* extract 50 (GBE50) has a variety of pharmacological functions such as anti-inflammatory, antioxidant and maintenance of glucose and lipid metabolism homeostasis. However, the therapeutic effects and mechanisms of GBE50 on non-alcoholic fatty liver disease (NAFLD) remain unknown. Therefore, in this study, we evaluated the therapeutic effects of GBE50 in NAFLD by using a high-fat diet (HFD) mice model.

Methods: C57BL/6J mice were fed a HFD diet for 15 weeks and were given respectively 25, 50, and 100 mg/kg GBE50 daily by gavage from 3 to 15 weeks. After the administration, blood samples and liver tissues were collected for biochemical detection, histological measurement, immunohistochemistry and Western blot, respectively.

Results: We found that GBE50 treatment could ameliorate insulin resistance (IR), glucose intolerance, lipid accumulation, hepatic steatosis and liver injury in HFD-fed mice. Further mechanism exploration discovered that the hepatoprotective effects of GBE50 on NAFLD may be related to the strengthening of IRS-1 signal activation and the weakening of NF- κ B, Akt and endoplasmic reticulum stress signals activation.

Conclusion: GBE50 is a potentially powerful therapeutic agent for the treatment of NAFLD.

Keywords: *Ginkgo biloba* extract 50, non-alcoholic fatty liver disease, insulin resistance, hepatic steatosis, inflammatory response, liver injury

Introduction

Non-alcoholic fatty liver disease (NAFLD) refers to a chronic liver disease that is caused by metabolic disorders and characterized by diffuse bullae steatosis and lipid accumulation in liver cells, excluding alcohol consumption, hepatitis virus, drugs and other specific factors that damage the liver.^{1,2} NAFLD is often closely related to metabolic diseases such as obesity, insulin resistance (IR), hypertension and dyslipidemia, so NAFLD is a chronic hepatic disease with metabolic abnormalities.³ Even, NAFLD is a potential factor for primary hepatocellular carcinoma.⁴ Moreover, due to the improvement of people's living standard and changes in lifestyle and diet structure in recent years, diseases caused by abnormal lipid metabolism are gradually increasing, and the prevalence of metabolic syndrome-related diseases, such as obesity and diabetes, was increasing day by day, resulting in a rapid rise in the prevalence of NAFLD.⁵ In particular, the prevalence of NAFLD in people with abdominal obesity is as high as 75–100%.⁶ Globally, NAFLD affects about 25% of the adult population.⁷ In addition, the prevalence of NAFLD also showed an increasing trend among young

children.⁸ Therefore, NAFLD has become a worldwide public health problem. Currently, the drugs used in the clinical treatment of NAFLD, such as membrane insulin sensitizers, antioxidants, biological agents, weight-loss drugs, anti-inflammatory and liver-protecting drugs, are beneficial to the relief of symptoms, but cannot completely reverse NAFLD.⁹ Therefore, the current prevention and treatment methods for NAFLD are not very satisfactory.

The pathogenesis of NAFLD is very complex, which is mainly believed to be the disorder of hepatic lipid metabolism caused by IR and further triggered by a series of cytotoxic events such as hepatocyte inflammation, oxidative stress and liver injury.¹⁰ For example, the accumulation of long-chain fatty acids in hepatocytes and their oxidation in the endoplasmic reticulum (ER) lead to an increase in reactive oxygen species (ROS) production, which leads to oxidative stress and endoplasmic reticulum stress (ERS), and ultimately to lipid inflammation, cell damage, and apoptosis in hepatocytes.¹⁰ Therefore, hepatic lipid metabolism disorder is a key pathological change of NAFLD, and IR and ERS are the major cytotoxic events leading to NAFLD.^{10,11} Based on the major pathological changes in lipid metabolism, inhibition of IR and ERS have become the key targets for NAFLD treatment.

Ginkgo biloba extract 50 (GBE50) is the new *Ginkgo biloba* extracts, containing 44% Ginkgo flavonoid glycosides (including quercetin and isorhamnin derivatives) and 6% lactones (including Ginkgolide A, B, C and J, and ginkgolide). GBE50 has a wide variety of pharmacological effects including anti-oxidant, anti-inflammatory, vasodilating, maintaining lipid metabolism homeostasis, enhancing insulin sensitivity, etc.^{12,13} Based on the pathological characteristics of NAFLD and the pharmacological effects of GBE50, we hypothesized that GBE50 may have an anti-NAFLD effect, while the effects and mechanisms of GBE50 on NAFLD remain unknown. Therefore, the present study was mainly explored whether GBE50 had a hepatoprotective effect on NAFLD and its mechanism.

Materials and Methods

Experimental Animals and Experimental Design

Six-week-old male C57BL/6J mice with SPF grade were purchased from Charles River Laboratory Animal Technology Co., Ltd. (Zhejiang, China), and kept in a humidity and temperature-controlled SPF grade animal room (12h light/dark cycle, the room temperature was

maintained at 23°C–25°C, humidity was maintained at 55%–65%). The mice were allowed free access to food and water before being given a special diet. After 5 days of adaptive feeding, mice were divided into normal control group (NC), NC+100 mg/kg GBE50 group, NAFLD model group, high fat diet (HFD)+25 mg/kg GBE50 group, HFD+50 mg/kg GBE50 group, and HFD+100 mg/kg GBE50 group, with 11 mice in each group. GBE50 was purchased from SPH Xingling Sci. and Tech. Pharmaceutical Co., Ltd (Shanghai, China). NAFLD mouse model was established by HFD induction. Mice in the NC group and the NC+100 mg/kg GBE50 group were fed a normal chow diet for 15 weeks. Mice in the NAFLD model group, the HFD+25 mg/kg GBE50 group, the HFD+50 mg/kg GBE50 group, and the HFD+100 mg/kg GBE50 group were fed a HFD diet (60% normal chow diet, 35% fat and 5% cholesterol) for 15 weeks. Mice in the NC+100 mg/kg GBE50 group, the HFD+25 mg/kg GBE50 group, the HFD+50 mg/kg GBE50 group, and the HFD+100 mg/kg GBE50 group were given gavage of 100 mg/kg, 25 mg/kg, 50 mg/kg, and 100 mg/kg GBE50 every day from 3 to 15 weeks, respectively; the NC group and the NAFLD model group were given gavage of normal saline of the same volume at the same feeding period.

Blood Biochemical Detection

After the end of the drug regimen, the mice were deprived of water and food for 12 h, and the mice were anesthetized with sodium pentobarbital (60 mg/kg) by intraperitoneal injection (i.p.). Then, blood was collected from the orbit by using vacuum anticoagulant tubes. The blood samples were placed in vacuum anticoagulant tubes for 10 min and centrifuged at 1500 g/min at room temperature for 20 min to collect serum. The levels of serum alanine aminotransferase (ALT), aspartate aminotransferase (AST), triglyceride (TG), total cholesterol (TC), free fatty acids (FFA), high-density lipoprotein cholesterol (HDL-C) and low-density lipoprotein cholesterol (LDL-C) were measured by biochemical assay according to the corresponding kit's instructions (cat No. C009-3-1, C010-3-1, A110-1-1, A111-1-1, A042-2-1, A112-1-1, A113-1-1; Nanjing Jiancheng Bioengineering Institute, Nanjing, China) by using an automatic biochemical analyzer (AU480, Beckman Coulter, Miami, FL, USA), respectively. The levels of fasting blood glucose (FBG) and fasting insulin (FINS) were quantified by kits assay according to the corresponding kit's instructions (cat No. YC0713, YS02720B; Shanghai YaJi Biotechnology Co., Ltd.,

Shanghai, China). The degree of IR was evaluated by using a homeostasis model assessment of IR index (HOMA-IR). The HOMA-IR was calculated as $\text{FBG (mmol/L)} \times \text{FINS (}\mu\text{U/mL)/22.5}$.

Glucose Tolerance Test (GTT) and Insulin Tolerance Test (ITT) Assays

For GTT and ITT assays, five mice (12 h overnight-fasted mice) of each group at the end of the drug regimen were intraperitoneally injected with glucose (1.5 g/kg, Sigma-Aldrich, St. Louis, MO, USA) or insulin (1.0 IU/kg, Sigma-Aldrich), then, the blood glucose levels of caudal vein were detected by using a blood glucose monitor (AlphaTRAK, Abbott Molecular, Abbott Park, IL, USA) at 0 (baseline), 30, 60, 90 and 120 min after the injection.

Liver Tissue Collection

At the end of the experimental period, all the mice were anesthetized with sodium pentobarbital by i.p. Then, the mice were sacrificed and the livers were harvested. After weighing each liver, it was divided into two parts. One part of liver tissue (left lobe) was used for subsequent biochemical assays and Western blot analysis, while the other part (right lobe) was fixed with 4% paraformaldehyde for 24 h and then used for histological measurement and immunohistochemistry.

Liver Tissue Biochemical Detection

The mouse IL-6, IL-1 β and TNF- α ELISA kits (cat No. H007, H002, H052-1; Nanjing Jiancheng Bioengineering Institute) were used to detect the levels of IL-6, IL-1 β and TNF- α in liver tissues. After liver tissues were homogenized by RIPA reagent (cat No. P0013C; Beyotime, Shanghai, China), the liver homogenates were collected for ELISA assay according to the corresponding kit's instructions, respectively. Briefly, liver homogenates were added into the plates that had been coated with the corresponding antibodies, and incubated at room temperature for 2 h. Then, the solution was removed and incubated with Biotin-labeled goat anti-rabbit IgG (H+L) antibody at 37°C for 45 min. Followed, rinsed the well with PBS for 3 times, and incubated with HRP-conjugated Streptavidin antibody at 37°C for 30 min. Finally, TMB solution was added and incubated at room temperature for 20 min. The OD values of the wells were measured at wavelength of 490 nm by using a microplate reader (Bio-Tek, Winooski, VT, USA). Protein concentrations of liver homogenates

were quantified by BCA assay (cat No. P0012; Beyotime). According to the protein concentrations of liver homogenates, the contents of hepatic IL-6, IL-1 β and TNF- α in unit mg protein were calculated.

For detection of TC and TG in liver tissues, the liver tissues were homogenized in a 10-fold volume (v/w) of anhydrous ethanol for 5 min, and the liver homogenates were collected for TC and TG detection by using the corresponding kit (cat No. A110-1-1, A111-1-1; Nanjing Jiancheng Bioengineering Institute) according to the manufacturer's instructions, respectively. For detection of FFA in liver tissues, the liver tissues were homogenized in a 10-fold volume (v/w) of cold PBS for 5 min, and the liver homogenates were collected for FFA detection using an FFA assay kit (cat No. A042-2-1; Nanjing Jiancheng Bioengineering Institute) according to the manufacturer's instruction. The protein concentrations of liver homogenates were measured by BCA assay (Beyotime). According to the protein concentrations of liver homogenates, the contents of hepatic TC, TG and FFA in unit mg protein were calculated.

Hematoxylin–Eosin (HE), Masson Trichrome and Sirius Red Staining

Paraffin-embedded liver tissues were sliced into 5 μm thickness sections. After deparaffinization and hydration, liver sections were stained with HE, Masson Trichrome, and Picrosirius Red according to the corresponding staining kits' instructions (cat No. G1120, G1340, S8060; Solarbio), respectively. The histological features of liver were observed and the staining images were captured by a BX53MRF-S microscope (Olympus). Six random fields of each section were selected, and hepatic steatosis, inflammation and fibrosis of each field were quantified by using the Image J software (version 1.8.0, NIH, USA).

Oil Red O Staining

Paraffin-embedded liver tissue were sliced into 10 μm thickness sections. After deparaffinization and hydration, liver sections were rinsed with 60% isopropyl alcohol for 20–30 s, followed by staining with Oil red O solution (cat No. G1261; Solarbio) at room temperature for 10 min. The excess dye liquor was slightly washed with 60% isopropyl alcohol, and then washed with distilled water to remove the isopropyl alcohol. The nucleus was stained with hematoxylin and differentiated with 1% hydrochloric acid alcohol. Finally, rinsed with water for 10 min and dried at room temperature. The lipid accumulation in liver tissues

was observed and the Oil red O staining images were captured by a BX53MRF-S microscope (Olympus). Six random fields of each section were selected, and the Oil red O-positive staining (lipid droplet) area of each field was counted by using the Image J software (version 1.8.0).

Immunohistochemical Staining

The procedure of immunohistochemical staining was described as previously.¹⁴ Briefly, paraffin-embedded liver sections (5 μ m thickness) were deparaffinized and rehydrated, then disposed with 3% H₂O₂ for 10 min followed by blocking with 5% donkey serum for 30 min at room temperature, finally, incubated with primary antibodies against F4/80 (macrophage marker; cat No. ab16911; 1:200, Abcam, Cambridge, MA, USA), CD45 (leukocyte marker; cat No. #70257; 1:250, Cell Signaling Technology, Danvers, MA, USA), and Ly6G (neutrophils marker; cat No. #87048; 1:150, Cell Signaling Technology) overnight at 4 °C. After incubation with the Biotin-labeled goat anti-rabbit IgG (H+L) and HRP-conjugated Streptavidin antibodies (cat No. PK-4001; ZSGB-BIO Inc, Beijing, China), the immunostaining was visualized by using DAB solution (cat No. SW1040; Solarbio), and the sections were counterstained with hematoxylin solution (cat No. H8070; Solarbio). The inflammatory cell infiltration in liver tissues was observed and the immunostaining images were captured by a BX53MRF-S microscope (Olympus). Six random fields of each section were selected, and the immunostaining-positive area of each field was quantified by using the ImageJ software (version 1.8.0).

Western Blot Analysis

RIPA reagent (cat No. P0013C; Beyotime) was employed to isolate the total proteins from the liver tissues of each group. Nuclear and cytoplasmic proteins were isolated by using a Nuclear and Cytoplasmic Protein Extraction Kit (cat No. P0027; Beyotime). The concentrations of the isolated proteins were measured by using a BCA Protein Quantitative Kit (cat No. P0012; Beyotime) according to the kit's instruction. Then, the isolated proteins were used to analyzed by Western blot. The protocol of Western blot was described as previously.¹⁵ The primary antibodies including eukaryotic translation initiation factor-2 α (eIF2 α) (cat No. sc-133132), activating transcription factor 4 (ATF-4) (cat No. sc-390063), fatty acid synthase (FAS) (cat No. sc-8009), sterol regulatory element-binding protein-1c (SREBP-1c) (cat No. sc-365513), acetyl-Coenzyme A carboxylase- α (ACC α) (cat No. sc-271965),

C/EBP homologous protein (CHOP) (cat No. sc-7351) and glucose regulator protein 78 (GRP78) (cat No. sc-13539) (1:1000; all from Santa Cruz, CA, USA), p-eIF2 α (cat No. #9721), insulin receptor substrates 1 (IRS-1) (cat No. #2382), p-IRS-1^{Tyr608/612} (cat No. #3203), p-IRS-1^{Ser307} (cat No. #2381), p-Akt^{Thr308} (cat No. #13038), p-Akt^{Ser473} (cat No. #4060), AKT (cat No. #9272), nuclear factor kappa-B (NF- κ B) p65 (cat No. #59674), inhibitor of NF- κ B (I κ B α) (cat No. #4812), and p-I κ B α (cat No. #2859) (1:2000; all from Cell Signaling Technology), were used in this experiment. β -Actin (cat No. AF0003; 1:8000; Beyotime) was used as an internal reference, and the relative expression levels of the target proteins were quantified by using the ImageJ software (version 1.8.0).

Statistical Analysis

The data were presented as mean \pm standard deviation (SD). Statistical analysis was performed using the SPSS 20.0 software. Comparisons among multiple groups were performed by one-way ANOVA with post hoc Fisher's Least Significant Difference test. *P* value less than 0.05 was considered to be statistically significant.

Results

GBE50 Attenuated IR in HFD-Fed Mice

Glucose metabolism disorder and IR are the typical symptoms of NAFLD.¹⁶ In this study, FBG and FINS were significantly increased in the NAFLD model group ($P < 0.01$), but were significantly decreased after treatment with GBE50 ($P < 0.05$ or 0.01) (Figure 1A and B). Furthermore, systemic IR was evaluated by the HOMA-IR index, our data (Figure 1C) showed that HFD induced a higher HOMA-IR value ($P < 0.01$), suggesting a significant increased level of IR, which was significantly reduced by GBE50 treatment ($P < 0.05$ or 0.01). The data of GTT and ITT (Figure 1D and E) revealed that there were no differences between NC group and NC+100 mg/kg GBE50 group; however, GBE50 administration cleared blood glucose efficiently after glucose or insulin injection. Taken together, these data suggested that GBE50 administration could increase glucose clearance and insulin sensitivity.

GBE50 Ameliorated HFD-Induced Hepatic Lipid Metabolism Disorder

Hepatic lipid metabolism disorder is a key pathological change of NAFLD.^{10,11,17} In this study, HFD not only

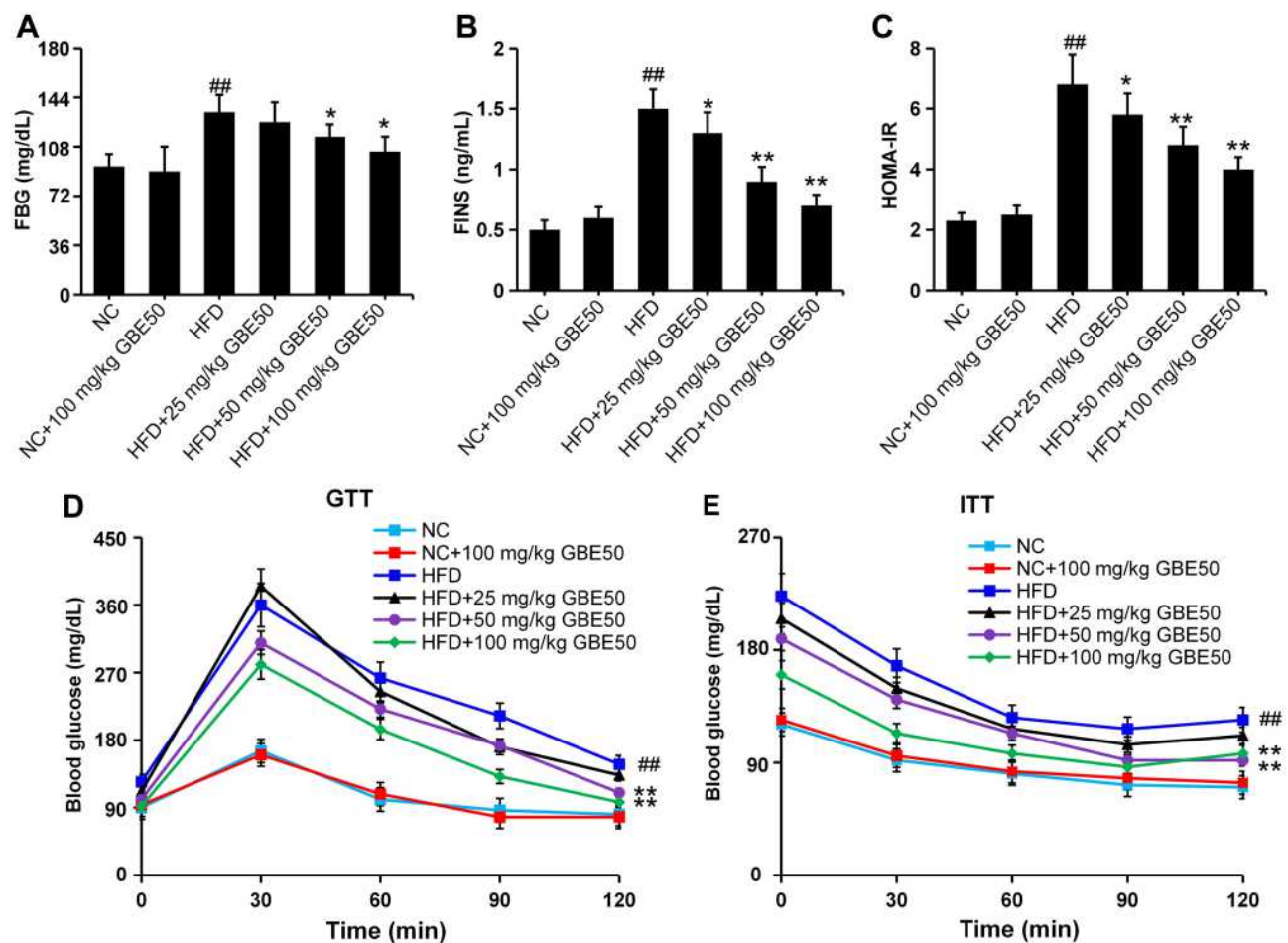


Figure 1 GBE50 attenuated IR in HFD-fed mice. (A–C) The levels of fasting blood glucose (FBG) (A), fasting insulin (FINS) (B) and the homeostasis model assessment of IR (HOMA-IR) values (C); $n=11$ mice/group. (D and E) The results of glucose tolerance test (GTT) (D) and insulin tolerance test (ITT) (E); $n=5$ mice/group. All data are presented as mean \pm SD, * $P<0.05$, ** $P<0.01$ vs HFD alone group; $^{##}P<0.01$ vs NC group. **Abbreviations:** NC, normal control group; HFD, high-fat diet.

increased hepatosomatic index (liver weight/body weight) (Figure 2A) but also promoted the deposition of lipid droplets (Figure 2B and C), which was significantly ameliorated by GBE50 treatment ($P<0.05$ or 0.01). Growing research findings showed that the abnormal elevation of TC and TG contributes to the development of NAFLD.^{18,19} Our results showed that serum TG and TC levels in the NAFLD model group were significantly increased ($P<0.01$), while GBE50 treatment significantly inhibited the HFD-induced elevated TG and TC levels ($P<0.05$ or 0.01) (Figure 2D and E). This phenomenon was also present in liver tissues (Figure 2F and G). LDL-C and HDL-C are also known to be key indicators affecting the progression of NAFLD, so we examined the effect of GBE50 on LDL-C and HDL-C levels in mice fed a HFD diet. As shown in Figure 2H

and I, compared with NC group, serum HDL-C in NAFLD model group was significantly decreased and LDL-C was significantly increased ($P<0.01$); compared with NAFLD model group, serum HDL-C was significantly increased and LDL-C was significantly decreased after GBE50 treatment ($P<0.05$ or 0.01). In addition, GBE50 markedly inhibited HFD-induced the increase level of FFA in serum and liver tissues (Figure 2J and K). We also examined the fat synthesis-related proteins FAS, SREBP-1c and ACC α in this study, as shown in Supplemental Figure 1, GBE50 could markedly inhibited HFD-induced the increased expression levels of FAS, SREBP-1c and ACC α ($P<0.05$ or 0.01). Overall, these data confirmed that GBE50 administration could ameliorate HFD-induced hepatic lipid metabolism disorder.

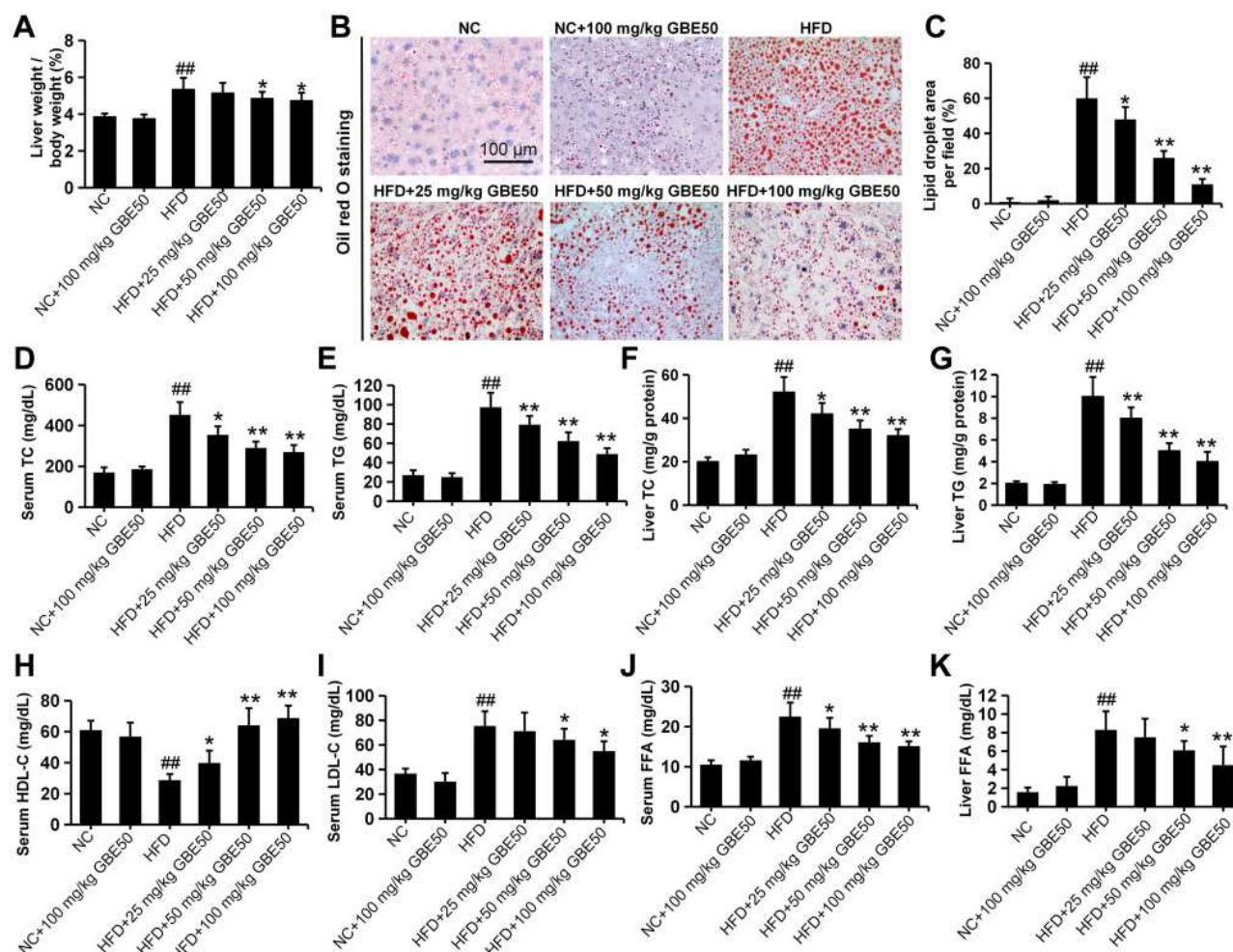


Figure 2 GBE50 ameliorated HFD-induced hepatic lipid metabolism disorder. (A) The results of the ratio of liver weight to body weight. (B and C) Representative Oil red O staining images (B) and the quantification of lipid droplet (C). (D–K) The levels of serum total cholesterol (TC) (D), serum triglyceride (TG) (E), liver TC (F), liver TG (G), serum high-density lipoprotein cholesterol (HDL-C) (H), serum low-density lipoprotein cholesterol (LDL-C) (I), serum free fatty acids (FFA) (J), and liver FFA (K). All data are presented as mean±SD, n=11 mice/group, * $P<0.05$, ** $P<0.01$ vs HFD alone group; ### $P<0.01$ vs NC group.

Abbreviations: NC, normal control group; HFD, high-fat diet.

GBE50 Ameliorated HFD-Induced Liver Injury

We directly observed the liver histopathological changes by HE, Masson Trichrome and Picrosirius Red staining, and the results (Figure 3A–D) showed that, liver tissues in the NAFLD model group presented a large number of vacuolated hepatocyte changes (HE staining) and fibrosis (Masson Trichrome and Picrosirius Red staining), which was significantly attenuated by GBE50 treatment ($P<0.05$ or 0.01). We also evaluated the NAS scores and the degree of fibrosis in various groups. The data (Supplemental Table 1) showed that, compared with the NAFLD model group, the NAS scores and the degree of fibrosis were lower in the GBE50 treatment group, especially in the high-dose group, 72.7% of the mice without fibrosis (8/11). ALT and AST are the two

important indicators of liver injury.²⁰ To investigate the effect of GBE50 on hepatic injury during NAFLD, we then examined the two indicators in serum. As shown in Figure 3E and F, GBE50 could significantly inhibit the increased activity of ALT and AST induced by HFD ($P<0.05$ or 0.01), suggesting that GBE50 could ameliorate HFD-induced liver injury.

GBE50 Attenuated HFD-Induced Inflammation in Liver

Chronic inflammation is generally associated with NAFLD and can further promote liver injury.²¹ Therefore, we further examined the inflammatory response in liver tissues in HFD-fed mice. As shown in Figure 4A–D, GBE50 could alleviate HFD-induced infiltration of macrophages and neutrophils ($P<0.05$ or 0.01). This effect of GBE50

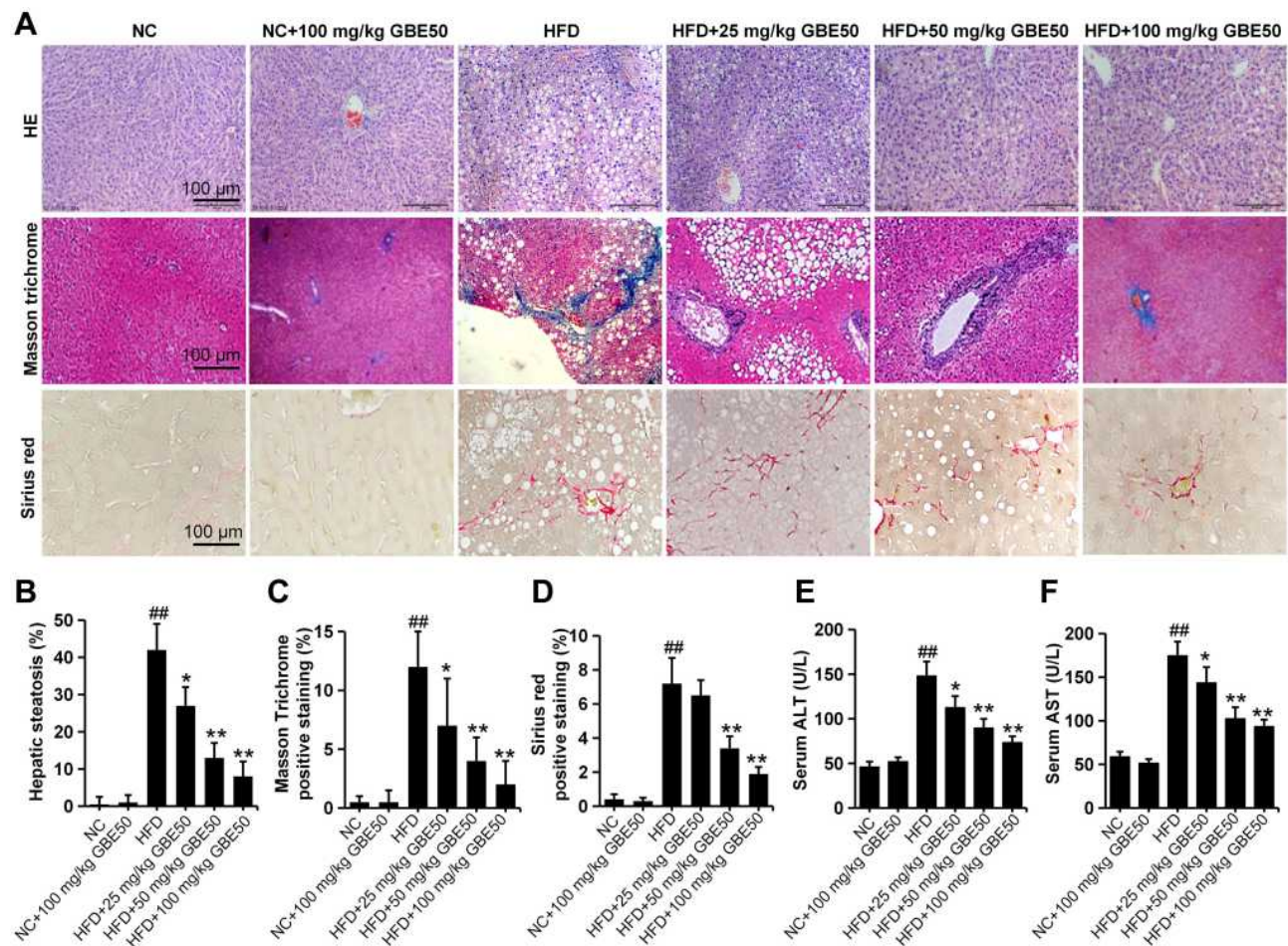


Figure 3 GBE50 ameliorated HFD-induced liver injury. (A–D) Representative liver sections images of hematoxylin-eosin (HE) staining, Masson trichrome staining and Sirius red staining (A), the quantitative data of hepatic steatosis from the HE staining (B), Masson trichrome positive staining area (C), and Sirius red staining positive staining area (D). (E and F) The levels of serum alanine aminotransferase (ALT) (E) and aspartate aminotransferase (AST) (F). All data are presented as mean±SD, n=11 mice/group. **P*<0.05, ***P*<0.01 vs HFD alone group; ###*P*<0.01 vs NC group.

Abbreviations: NC, normal control group; HFD, high-fat diet.

was further confirmed by ELISA, and the expression levels of pro-inflammatory cytokines (IL-6, IL-1 β and TNF- α) in liver tissues of HFD-fed mice were significantly decreased after GBE50 treatment (*P*<0.01) (Figure 4E–G). As we know that NF- κ B signaling pathway is one of the most important signaling pathways in the regulation of inflammatory response. Our data (Supplemental Figure 2) show that GBE50 could significantly promote the nuclear translocation of NF- κ B p65 and decreased the phosphorylation level of I κ B α in liver tissues of HFD-fed mice (*P*<0.05 or 0.01).

GBE50 Regulated the Hepatic Insulin Signaling and ERS in HFD-Fed Mice

IR is also a key pathological mechanism leading to NAFLD.^{10,11,16} To assess the hepatoprotective of GBE50

on NAFLD, the activation of IRS-1 pathway was determined. As seen in Figure 5A–D, GBE50 administration activated insulin signaling in HFD-fed mice, as evidenced by mild increase in the expression levels of p-IRS-1^{Tyr608} and reduced the expression levels of p-IRS-1^{Ser307} in the liver tissues; moreover, an acute injection of insulin dramatically activated insulin signaling, which was further enhanced after GBE50 administration. Akt signaling pathway is one of the most important downstream signals of IRS-1^{22,23}, therefore, we further tested the activity of Akt. As shown in Figure 5E–G, GBE50 could significantly decrease the phosphorylation level of Akt at Thr308/Ser473 in liver tissues of HFD-fed mice (*P*<0.05 or 0.01).

ERS response is one of the key steps leading to liver injury in NAFLD.^{10,11,17} We then examined the ERS-related proteins p-eIF2 α , ATF-4, CHOP and GRP78. The

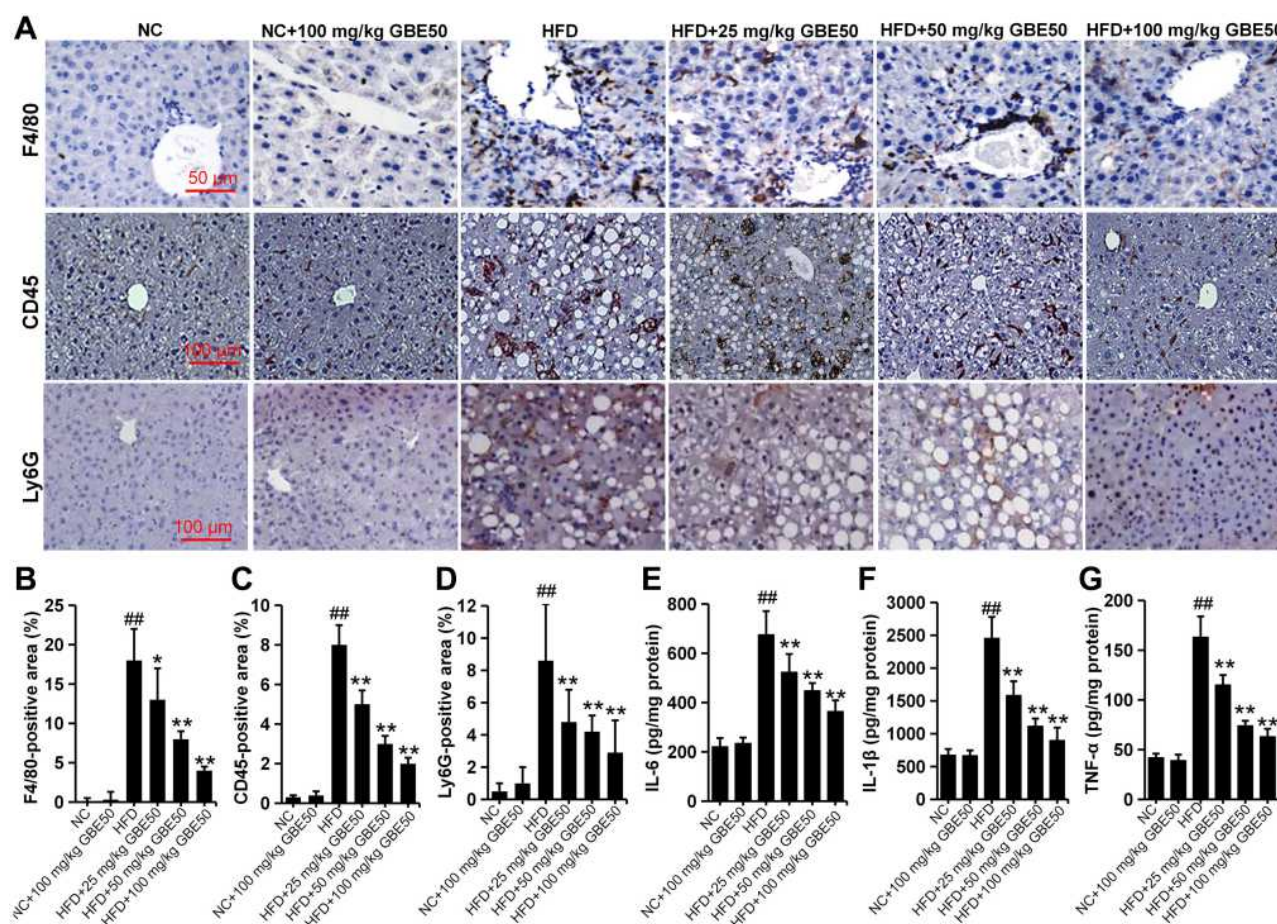


Figure 4 GBE50 attenuated HFD-induced inflammatory cell infiltration and the expression levels of pro-inflammatory cytokines in liver. (A–D) Representative immunohistochemical staining images of the expressions of F4/80, CD45, and Ly6G (A), and the quantification of F4/80 (B), CD45 (C), and Ly6G (D) positive staining areas in liver tissues. (E–G) The levels of IL-6 (E), IL-1β (F) and TNF-α (G) in liver tissues. All data are presented as mean±SD, n=11 mice/group, *P<0.05, **P<0.01 vs HFD alone group; ##P<0.01 vs NC group.

Abbreviations: NC, normal control group; HFD, high-fat diet.

results showed HFD induced higher expression levels of p-eIF2α, ATF-4, CHOP and GRP78, suggesting a significant increase level of ERS, which was significantly reduced by GBE50 treatment ($P<0.01$) (Figure 6A–G).

Discussion

NAFLD has become a serious public health problem, which affects about 25% of the adult population worldwide.⁷ Unfortunately, there is no specific drug currently available for the treatment of NAFLD. Natural Chinese herbal medicines have been proven to have a wide range of pharmacological effects that are safe, effective and low toxicity.²⁴ Screening small-molecule compounds or monomers with anti-inflammatory, anti-oxidation, maintenance of lipid homeostasis and enhancement of insulin sensitivity from natural Chinese medicinal

materials may be a potential strategy to search for anti-NAFLD drugs. Our present study demonstrated that, GBE50, the new *Ginkgo biloba* extracts, had the pharmacological effects of inhibiting IR, ameliorating hepatic steatosis and relieving liver injury in HFD-fed mice. And these data here suggested that GBE50 was a potential anti-NAFLD drug.

Hepatic steatosis and lipid accumulation are the key pathological changes in NAFLD, and also the key factors that trigger subsequent liver function decline and liver injury.^{10,11,17} Metabolic disorders induced by HFD can increase the abnormal levels of TC, TG and LDL-C, and decrease the level of HDL-C. Especially, the abnormal deposition of TC, TG and LDL-C in the liver can promote hepatic steatosis, and the continuous accumulation of TC, TG and LDL-C and the continuous decrease of HDL-C also can promote the sensitivity of the liver to

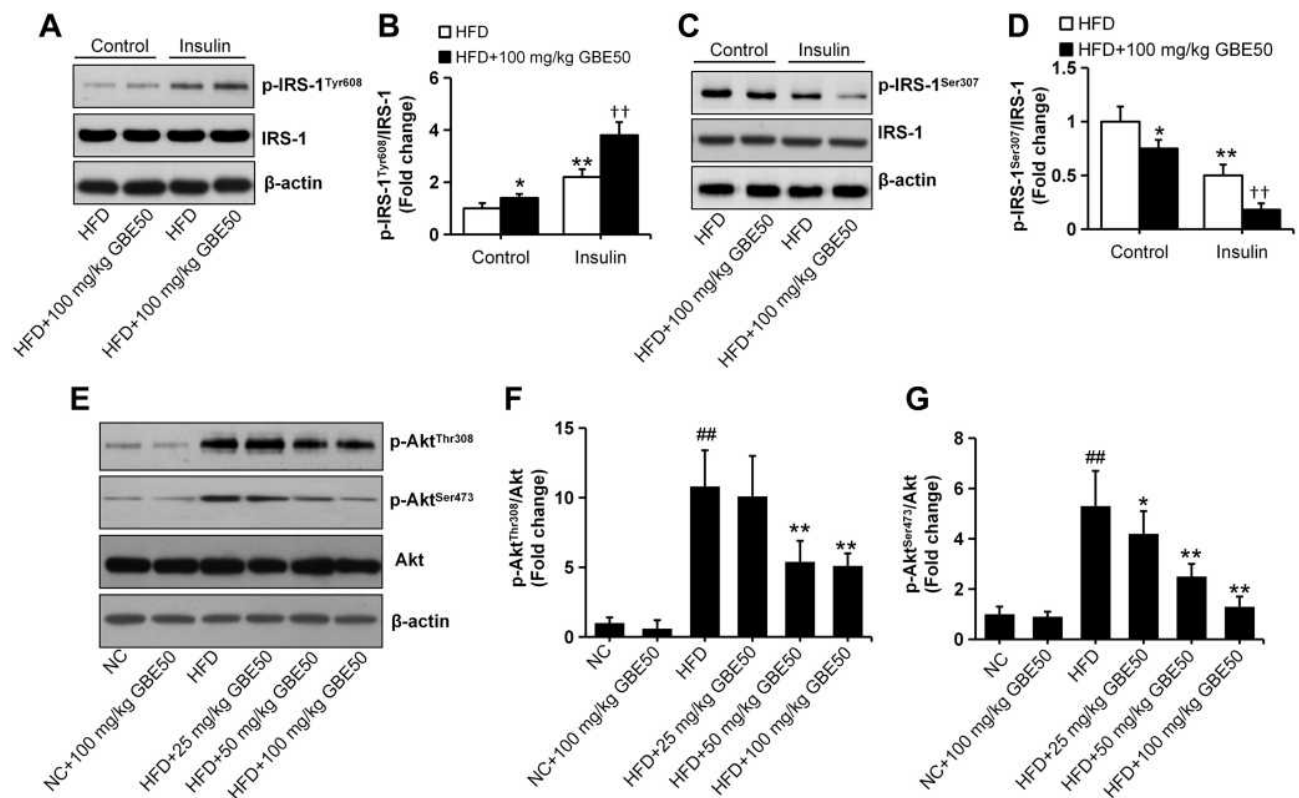


Figure 5 GBE50 promoted IRS-1 signal activation and inhibited Akt signal activation in HFD-fed mice. (A–D) The phosphorylation levels of IRS-1 (Tyr608) (A and B) and IRS-1 (Ser307) (C and D) under baseline (control) and in response to an intraperitoneal injection of insulin (1.0 IU/kg for 15 min) in HFD alone group and HFD+100 mg/kg GBE50 group; n=5 mice/group. (E–G) The expression levels of p-Akt (Thr308) (E and F) and p-Akt (Ser473) (E and G) proteins in liver tissues in various groups were detected by Western blot; n=6 mice/group. All data are presented as mean±SD, *P<0.05, **P<0.01 vs HFD alone group; ††P<0.01 vs HFD+insulin group; ###P<0.01 vs NC group.

Abbreviations: NC, normal control group; HFD, high-fat diet.

inflammation and the progression of NAFLD.^{18,19} This study showed that GBE50 reduced TC, TG and LDL-C levels, increased HDL-C levels, and also alleviated hepatic steatosis and lipid accumulation in HFD-fed mice. HFD can lead to an increased level of circulation FFA, while the excessively high levels of circulation FFA will increase the synthesis of TG and TC in liver, and cause IR, which will lead to the disorder of lipid metabolism, and finally lead to liver lipid deposition.^{25,26} In addition, abnormal amounts of FFA accumulate in liver cells, which can cause lipid peroxidation and oxidative stress, resulting in degeneration and necrosis of liver cells and inflammatory response.^{25,26} In the current study, we found that GBE50 decreased the FFA level in serum and liver tissues in HFD-fed mice. To investigate the mechanism of GBE50 inhibits lipid accumulation, the expression levels of FAS, SREBP-1c and ACC α proteins were evaluated in this study. Our results found that the expression levels of FAS, SREBP-1c and ACC α proteins were obviously decreased in HFD-fed mice after GBE50 treatment, suggesting that GBE50

decreased de novo lipid synthesis may reduce the expression levels of FAS, SREBP-1c and ACC α .

Liver injury and liver dysfunction are the important driving factors for the progression of liver diseases, including NAFLD.^{10,11,17,27,28} ALT and AST are important indicators for the evaluation of liver injury and liver function in the biochemical examination of NAFLD, and long-term HFD can cause liver injury and liver function decline and increase the level of ALT and AST.²⁰ The results of this study showed that GBE50 could significantly reduce ALT and AST levels in serum of HFD-fed mice, suggesting that GBE50 could reduce liver injury induced by HFD. In addition, liver steatosis induced by HFD is closely related to chronic liver inflammation, which increases the sensitivity of liver cells to pro-inflammatory cytokines mediated injury, and persistent chronic inflammation further exacerbates the progression of NAFLD.^{10,11,16,17,21} Therefore, this study further observed whether GBE50 had a protective effect on the inflammatory response of liver in NAFLD, and the

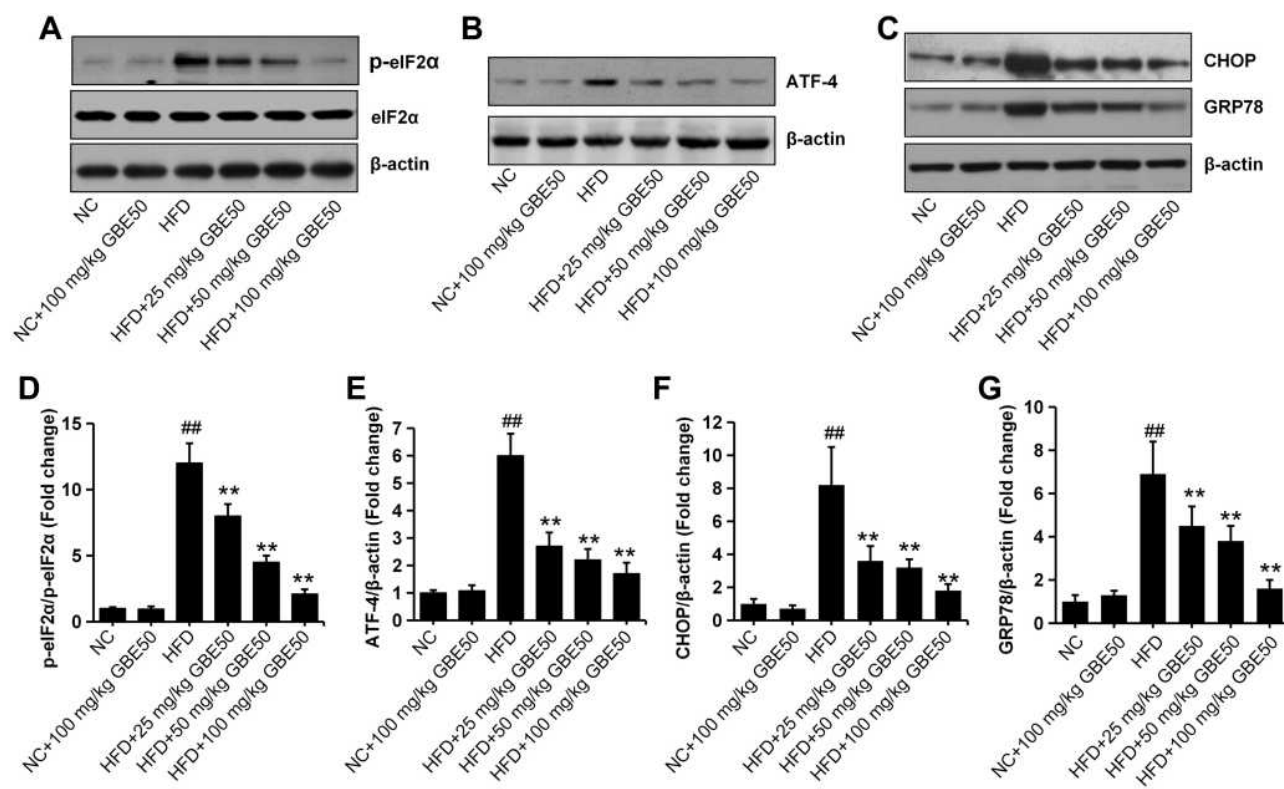


Figure 6 GBE50 attenuated endoplasmic reticulum stress in HFD-fed mice. (A–C) Representative Western blot bands of the expressions of p-eIF2α, eIF2α, ATF-4, CHOP and GRP78 in liver tissues in various of groups. (D–G) The quantification of the expression levels of p-eIF2 (D), ATF-4 (E), CHOP (F) and GRP78 (G). All data are presented as mean±SD, n=6 mice/group, ***P*<0.01 vs HFD alone group; ^{##}*P*<0.01 vs NC group.

Abbreviations: NC, normal control group; HFD, high-fat diet.

results showed that GBE50 alleviated the pathological changes of inflammatory cell infiltration and reduced the levels of pro-inflammatory cytokines IL-6, IL-1β and TNF-α in liver tissues in HFD-fed mice, indicating that GBE50 could effectively inhibit liver inflammation caused by HFD. To explore the mechanism by which GBE50 inhibits HFD-induced liver injury, we analyzed the activity of NF-κB, a classical signal that affected the inflammatory response. HFD has been reported to activate NF-κB signal and further aggravated the progression of NAFLD through NF-κB signal mediated inflammatory response and fibrosis.^{29,30} Inhibition of NF-κB signaling activity can reduce the production of pro-inflammatory factors, liver injury and block the progression of NAFLD.³¹ In this study, we found that GBE50 attenuated NF-κB signaling activity in liver tissues of HFD-fed mice, suggesting that GBE50 may control liver inflammatory response and liver injury of NAFLD through negative regulation of NF-κB signaling activity.

IR, as one of the initiating factors of NAFLD, is defined as a reduced sensitivity to insulin and associated

with defective insulin signal transduction.^{10,11,16} Insulin plays an important role in regulating glucose and lipid homeostasis in the liver.¹⁶ IRS-1 is the main effector of insulin receptors and is responsible for insulin signal transduction.^{10,11,16} When insulin binds to insulin receptor, phosphorylation of IRS-1 at Tyr608 can activate insulin pathway, in contrast, phosphorylation of IRS-1 at Ser307 is considered to block the insulin signaling, leading to IR.^{16,32} In the current study, the data showed that GBE50 enhanced phosphorylation of IRS-1 at Tyr608 and further inhibited phosphorylation at Ser307 induced by acute injection of insulin, suggesting that GBE50 could attenuate IR by promoting insulin signaling activation in HFD-fed mice.

ERS response is one of the major toxic events leading to liver injury in NAFLD.^{10,11,17} Hypoxia, lipid overloading, inflammation and other pathological factors can disrupt ER homeostasis, resulting in the accumulation of unfolded and misfolded proteins, which can lead to ERS in hepatocytes.^{33,34} In response to this stress, unfolded protein response (UPR) is activated. The UPR involves 3 transmembrane ER pressure sensors: inositol-requiring

enzyme 1 (IRE1), (PKR)-like ER kinase (PERK) and ATF-6.³⁵ Among them, PERK plays an important role in the corresponding ERS response. When ERS is activated, PERK is separated from GRP78, and then, the cytoplasmic eIF2 α is phosphorylated.³⁶ The phosphorylated eIF2 α can inhibit protein translation or activate ATF-4 to regulate target gene transcriptional level in response to ER folding load.³⁷ It has been found that activation of eIF2/ATF-4 signals could promote adipogenic differentiation and intracellular lipid synthesis of breast epithelial cells by up-regulating the expression of FAS.³⁸ In animal experiment, Oyadomari et al³⁹ observed that ERS agonist-induced mice presented fatty liver, while eIF2 dephosphorylation significantly ameliorated the fatty liver. Furthermore, phosphorylation of eIF2 α could up-regulate the expression of ATF-4, and then activate the expression of SREBP-1c, ACC2, FAS and other lipid-synthesis-related genes.⁴⁰ ATF-4 knockout was resistant to HFD-induced obesity, hypertriglyceridemia, and NAFLD in mice.⁴¹ In addition, high levels of phosphorylated eIF2 α in the liver can exacerbate IR and enhance NF- κ B activity in NAFLD, which can further aggravate hepatic steatosis and liver injury.^{33,42,43} The above evidences indicated the role of eIF2/ATF-4 in hepatic lipid synthesis and metabolism, IR and liver injury in NAFLD. The results of this study also showed that p-eIF2, ATF-4, CHOP and GRP78 proteins were highly expressed in liver tissues of HFD-fed mice, while they were all decreased after GBE50 treatment, suggesting that GBE50 could ameliorate HFD-induced NAFLD by weakening ERS.

In conclusion, HFD-induced NAFLD mice were characterized by lipid accumulation and hepatic steatosis, accompanied by liver injury, inflammation, glucose intolerance and IR, which were significantly ameliorated after GBE50 treatment. The therapeutic effects of GBE50 on NAFLD may be related to the strengthening of IRS-1 signal activation and the weakening of NF- κ B and ERS signal activation. These findings suggested that GBE50 may be a potentially powerful therapeutic agent for NAFLD.

Compliance with Ethical Standards

All animal experiments were approved and performed according to the guidelines of the institutional animal care and use committee of the Hunan Provincial People's Hospital, with the protocol number: HNPH/IAEC/LS/2018-102.

Consent for Publication

This study is approved by all authors for publication.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure

The authors have declared that no competing interests exist.

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