ORIGINAL RESEARCH

Clinical, Pathological and Genetic Characteristics of Pediatric Hepatocellular Carcinoma Associated with Hepatitis B Virus Infection

Pan Zhao D Yinying Lu¹ Chunya Wang² Limin Wang³ Jinfeng Li¹ Meina Li⁴

¹The Fifth Medical Center (formerly Beijing 302 Hospital), Chinese PLA General Hospital, Beijing, 100039, People's Republic of China; ²Beijing Anzhen Hospital, Capital Medical University, Beijing, 100029, People's Republic of China; ³Beijing Tsinghua Changgung Hospital, Tsinghua University, Beijing, 102218, People's Republic of China; ⁴Department of Health Service, Second Military Medical University, Shanghai, 200433, People's Republic of China **Introduction:** Hepatocellular carcinoma (HCC) remains the major challenge in the management of patients with hepatitis B virus (HBV) infection. To date, limited studies have been done on pediatric HBV-associated HCC specifically.

Methods: Pediatric patients younger than 16 years with HBV-associated HCC were included in the study. HBV integration detection was performed using a high-throughput viral integration detection (HIVID) method.

Results: Among the 13 included pediatric patients, boys predominated (10, 76.9%). The median age at diagnosis of HCC was 13 years and the youngest age was 6 years. Nine patients had initially seronegative hepatitis B e antigen (HBeAg) and 4 had seropositive HBeAg. All patients had cirrhosis and elevated alpha-fetoprotein. Splenomegaly was present in all patients. Intrahepatic HBsAg was not detected in any tumor tissues from 5 patients who underwent biopsy or excision, while it was positive in all matched non-tumor tissues. In the tumor and matched non-tumor tissues from 3 individuals, HBV integration was identified except in the neoplastic specimen from 1 patient. Integration into the reported genes associated with hepatocarcinogenesis was not found in the tumor tissues from the 3 patients. **Discussion:** Hypervigilance for HCC development is required in HBeAg-negative cirrhotic children. The findings based on the immunohistochemical and genetic results expand the knowledge of pediatric HCC development.

Keywords: hepatitis B virus, hepatocellular carcinoma, children

Introduction

Hepatitis B virus (HBV) infection remains a global public health problem and estimated 291 million people are chronically infected.^{1,2} Chronic HBV infection can increase the risk of developing cirrhosis and hepatocellular carcinoma (HCC).^{3–5} It has been reported that 0.01–0.03% of chronic HBV carriers develop HCC before adulthood.⁶ Despite passive-active immunoprophylaxis using hepatitis B vaccination with or without hepatitis B immunoglobulin (HBIg), up to 8–10% of newborns of HBV-infected mothers still acquire HBV infection.⁷ HBV-associated HCC is rare in childhood, but the prognosis is usually fatal. To date, the exact mechanisms that HBV causes malignant transformation remain uncertain, and inconsistent clinical characteristics or laboratory outcomes for pediatric patients with HBV-associated HCC often exist among different studies.^{8,9} These issues need to be further studied in more pediatric cases.

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Correspondence: Limin Wang; Pan Zhao Tel +86 10 56119097; +86 10 66933464 Email cynthia0929@sina.com; doczhaopan@126.com

The integration of HBV DNA into human genome was considered to result in oncogene activation, tumorsuppressor gene inactivation or other predisposition to chromosomal instability.¹⁰ Some studies have reported the phenomenon of HBV DNA integrated into HCC cells in adult patients.^{11,12} However, few studies were done on the cytogenetic background of HCC in the pediatric population specifically.¹³ In the present investigation, we collected the clinical, pathological and genetic data from pediatric cases with HBV-associated HCC, in order to better study the characteristics of the fatal disease in children.

Methods Study Population

A total of 13 pediatric patients with HBV-associated HCC were included in the study. All patients were diagnosed before 16 years of age. Among these patients, 6 developed HCC in our prospectively well-monitored cohort with chronic HBV infection, while the remaining 7 patients were retrospectively identified. Five patients had histologically confirmed HCC and 8 patients were diagnosed according to clinical, laboratory and imaging findings. Informed consent was obtained from each child's parents at admission. The study conformed to the ethical guidelines of the 1975 Declaration of Helsinki and was approved by the ethics committees of Beijing 302 Hospital. (currently the Fifth Medical Center of Chinese PLA General Hospital)

Laboratory Tests

Laboratory parameters, including biochemical and hematological indicators, viral profiles (HBV DNA (by polymerase chain reaction, Roche COBAS AmpliPrep, lower limit of detection: 20 IU/mL), hepatitis B surface antigen (HBsAg), anti-HBs, hepatitis B e antigen (HBeAg), anti-HBe, anti-HBc, HBsAg quantification (by Roche COBAS HBsAgII-Q, lower limit of detection: 0.05 IU/mL)), etc., were measured using standard laboratory procedures.

Imaging Examinations

Abdominal ultrasonography, computed tomography (CT) and magnetic resonance imaging (MRI) were carried out wherever necessary.

Pathological Evaluations

Formalin fixed and paraffin-embedded specimens were subjected to Haematoxylin&Eosin staining and immunohistochemical staining. Pathological evaluations were performed by an experienced hepatopathologist. Intrahepatic (including tumor and non-tumor liver tissues) HBsAg were detected by immunohistochemistry. Cytoplasmic staining with brown particles was considered as positive expression of intrahepatic HBsAg.

Viral Genome Sequencing

HBV DNA was extracted in serum from the patients. HBV pre-core and basal core promoter (pre-C/BCP) region and reverse-transcriptase (RT) region were sequenced according to the procedures reported in previous papers.^{14,15} Mutations at 11 sites, including nt1653, 1752, 1753, 1758, 1762, 1764, 1766, 1768, 1862, 1896 and 1899 were analyzed.

HBV Integration Detection

In order to detect the integration of HBV DNA into human genome, targeted HBV DNA fragment capture sequencing was performed. DNA was extracted from tumor tissues and matched non-tumor tissues using GeneReadTM DNA FFPE Kit (QIAGEN, Hilden, Germany). Sequence-capture probes according to eight types (A, B, C, D, E, F, G and H) genome sequences were produced by of HBV MyGenostics (Beijing, China). Detailed targeted HBV DNA fragment capture sequencing procedure was carried out according to a previous report.¹⁶ Then, highthroughput viral integration detection (HIVID) method was used for the detection of breakpoints.¹⁷ After removal of low-quality reads and duplication reads, sequenced datasets were compared against both HBV and human (hg19) reference sequences using Burrows-Wheeler Aligner (BWA). Integration sites were determined through Clipping Reveals Structure (CREST) analysis.

Statistical Analysis

Data analyses were performed using SAS 9.2 software (SAS Institute Inc, Cary, NC, USA). Continuous data were expressed as median (range). Categorical data were expressed as the number of subjects or percentages. Group comparisons were performed using Fisher's exact test. The survival curve was depicted based on Kaplan-Meier analysis. Tests were two-sided and a probability (*P*) value of less than 0.05 was considered statistically significant.

Results

Baseline Characteristics

Among the 13 pediatric patients, 10 were boys and 3 were girls. Male patients predominated in the study (76.9%).

The median age at diagnosis of HCC was 13 years and the youngest age was 6 years. Ten patients had HBsAgpositive mother, 1 had HBsAgpositive father and 2 declared no family history of HBV infection.

All patients had seropositive HBsAg at diagnosis of HCC. Nine patients (69.2%) had initially seronegative HBeAg and 4 had seropositive HBeAg. Seven patients were infected with genotype C HBV, 2 were infected with genotype B HBV and the other patients did not undergo the detection of HBV genotypes. Four patients had negative serum HBV DNA (<20 IU/mL). The median level of serum HBV DNA at diagnosis of HCC was 112 IU/mL (range, 0–3,860,000 IU/mL). Alpha-fetoprotein (AFP) was elevated in all patients (median, 1210 ng/mL; range, 72.73–1210 ng/mL (reference interval: <10 ng/mL)). Laboratory parameters at diagnosis are shown in Table 1.

All patients had evidence of cirrhosis, 5 of whom were pathologically diagnosed and 8 of whom were diagnosed according to clinical, laboratory and imaging findings. Splenomegaly was present in all patients. The 6 patients who were in the chronically HBV-infected prospective cohort developed cirrhosis before their diagnosis of HCC. Regarding symptoms, 5 individuals had abdominal pain, 4 were asymptomatic, 2 presented with fatigue and 2 presented with distended abdomen. Additionally, jaundice was present in 3 patients. Four patients were not known to be HBsAg-positive before their initial diagnosis of liver tumor.

Pathological and Genetic Findings

Tumor tissues and matched non-tumor tissues from 5 patients were collected, including 4 boys and 1 girl. Intrahepatic HBsAg was not detected by immunohistochemistry in any tumor tissues, while it was positive in all matched non-tumor tissues. Figure 1A and B shows HBsAg expression in the tumor tissues and matched nontumor tissues from 2 patients.

Four male patients underwent HBV genome sequencing. This detection was not successful in 2 patients because their serum HBV DNA levels were too low. Mutations in pre-C/BCP or RT regions were not found in 1 patient. The remaining treatment-naïve patient carried the lamivudine-resistant mutation of M204V in the tyrosine-methionine-aspartate-aspartate motif of the RT gene and no mutations in the pre-C/BCP region were found.

HBV capture sequencing was performed in the tumor tissues and matched non-tumor tissues from 3 male patients. HBV DNA integration was identified in the tumor tissues from 2 patients and non-tumor specimens from all the 3 patients. Distribution of chromosomal location of integration is shown in Table 2. The chromosomal locations of integration in the tumor tissues were not exactly same as those in the matched non-tumor tissues. There were respectively 0, 9, 19 integrants detected in the tumor tissues from the 3 individuals, and 2, 6, 10 integrants detected in the non-tumor tissues. In the tumor tissues, intronic HBV integration near CCNA2 (encoding cyclin A2), COP1 (encoding the RING finger protein constitutive photomorphogenic 1) or EPHA4 (encoding EPH receptor A4) was not found.

Treatments and Outcomes

The 6 prospectively followed patients received antiviral treatments before their diagnosis of HCC. None of them achieved HBsAg loss. All the 6 patients had a single nodule ranging from 2.1 to 4.6 cm. Five were alive in the follow-up after receiving radiofrequency ablation or transcatheter arterial embolization and 1 patient progressed rapidly to death without interventions. Among

Median	Range (Minimum-Maximum)	Reference Interval
112	0–3,860,000	<20
1014	97.84-8158	<0.05
39	24–105	<40
49	8.4–227	<40
39	26–44	35–55
16.4	5.5-113.5	<20.5
18	4–276	<10
1210	72.73–1210	<10
192	30-426	100-300
	112 1014 39 49 39 16.4 18 1210	112 0-3,860,000 1014 97.84-8158 39 24-105 49 8.4-227 39 26-44 16.4 5.5-113.5 18 4-276 1210 72.73-1210

Table I Laboratory Parameters of Pediatric Patients with HBV-Associated Hepatocellular Carcinoma at Diagnosis

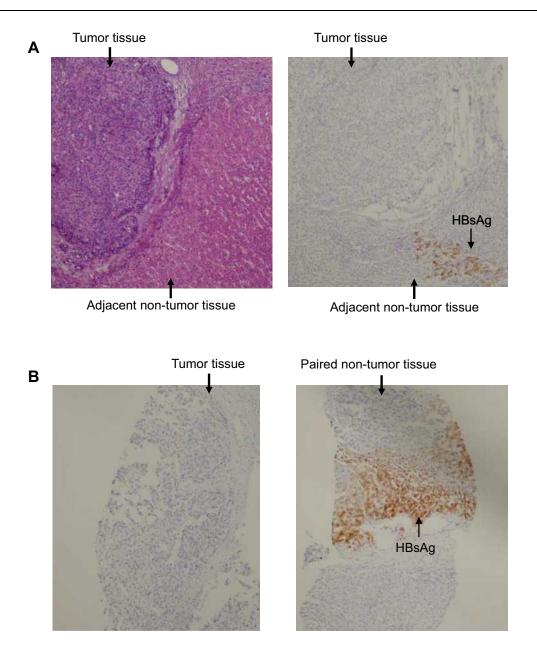


Figure I HBsAg expression in tumor tissues and matched non-tumor tissues from 2 pediatric patients with HBV-associated hepatocellular carcinoma. (A) Histological pictures of resected tumor tissues and adjacent non-tumor tissues stained by Hematoxylin-Eosin (× 200) and immunohistochemistry (× 200). (B) HBsAg in biopsy tumor tissues and paired non-tumor tissues (immunohistochemical staining, × 200). Brown particles represent HBsAg.

the 7 retrospectively identified patients, 4 had multiple nodules and progressed rapidly to death without interventions, 1 of whom had pulmonary metastases at presentation. The remaining 3 individuals all had a single nodule larger than 10 cm. Of them, 2 died without interventions and 1 died after excision. There was a statistical difference between prospectively followed patients and retrospectively identified patients regarding the mortality (P=0.021). Figure 2 shows the survival curve of all the included patients in the study.

Discussion

HCC is a leading type of primary cancer with increasing incidence worldwide and chronic HBV infection remains one of the major etiologic factors in HCC pathogenesis in endemic areas.¹⁸ In our study, the youngest patient was 6 years old, with a long history of cirrhosis. Of note, all patients in the study had cirrhotic background, which was discordant with the results from North America.⁹ Another study about pediatric HBV-associated HCC from Taiwan also reported cirrhosis at presentation in an absolute majority of included

	Chromosomal Location	Number of HBV Integrants
Tumor tissue		
Patient I	None detected	0
Patient 2	Chromosomes 2, 3, 4, 5, 7, 8, 17, 21	9 (two integration sites in Chromosome 7)
Patient 3	Chromosomes 1, 2, 5, 7, 8, 9, 10, 11, 14, 15, 20, 22	19 (three integration sites in Chromosome 7, two integration
		sites respectively in Chromosomes 1, 8, 10, 11 and 15)
Non-tumor tissue		
Patient I	Chromosomes 2, 3	2
Patient 2	Chromosomes 1, 2, 10, 17, 19, 21	6
Patient 3	Chromosomes 2, 3, 7, 10, 11, 14, 20, 21, 22	10 (two integration sites in Chromosome 7)

Table 2 Distribution of Chromosomal Location of Hepatitis B Virus (HBV) Integration

patients.¹⁹ It seemed that there was a difference regarding the onset background of pediatric HBV-associated HCC between North America and Asia. In addition, AFP was elevated in all patients in our study, whereas individuals in the American case series had normal AFP. These different clinical characteristics should be paid attention to by clinicians. On the other side, similar to the above two studies, male predominance and negative HBeAg predominance existed in our study. An investigation from a prospectively followed pediatric cohort with chronic HBV infection demonstrated that early e antigen seroconversion and/or cirrhosis may be risk factors for HCC.²⁰ According to these results, HCC had a predilection for HBeAg-negative children. The speculation needs further confirmation by accumulating evidence. Though studies showed that some mutations in the preC/BCP region after HBeAg-seroconversion were associated with liver fibrosis, we could not raise a concern about the association between mutations in the preC/BCP region and HCC development based on limited available experimental data in the study.²¹

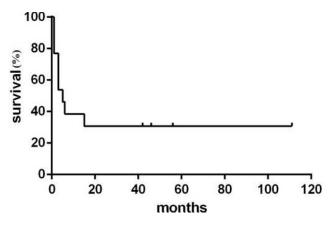


Figure 2 Survival curve of the patients in the study based on Kaplan-Meier analysis.

HBsAg is an important biomarker of HBV infection. Absence or reduced expression of HBsAg in neoplastic tissues can be considered as reflection of the interruption of HBV life cycle in HCC cells. In adults, the significant association between serum HBsAg level and HCC was elucidated by several studies, while negative results were also obtained by other studies.²²⁻²⁵ Whether HBsAg can promote direct carcinogenesis remains unclear, though some molecular mechanisms of HCC development mediated by HBsAg or mutant HBsAg have been explored.^{26–28} In the present study, intrahepatic HBsAg were detected out in none of collected tumor tissues. Three previous papers also reported the significantly reduced expression of HBsAg in HBV-associated HCC tissues compared with matched non-tumor tissues, and one paper attributed it to HBV integration in tumor cells.^{29–31} However. no viral integrants were found in the tumor specimen from a patient in our study. According to the results from one of these papers that HBV receptors were deficient in tumor but over-expressed in peritumor,³⁰ HBV integrants in tumor tissues should be probably fewer than those in non-tumor tissues. In fact, our results are not fully consistent with the speculation. Additionally, integration into the reported genes associated with hepatocarcinogenesis, such as those encoding cyclin A2, was not found in the tumor tissues from the 3 patients. Therefore, the link of intrahepatic HBsAg, HBV integration and HCC development in children deserves further concerns.

Early recognition is vital for pediatric patients with HCC. In the study, the retrospectively identified cases had significantly worse prognosis than the well-monitored cases. Moreover, 4 individuals had no idea about their HBsAg status until their initial consultation in hepatology clinic. Therefore, surveillance is necessary for children with chronic HBV infection and hypervigilance is required for the HBeAg-negative cirrhotic cases specifically. Regarding the treatment, no exact criteria to guide intervention in the population have been issued up to now. A previous investigation showed that complete tumor resection improved the survival in pediatric HCC.³² However, for children with unresectable HCC, no further suggestions were made. Appropriate treatment approaches in pediatric HCC should be studied.

The study is limited regarding the sample size. Nevertheless, we first specifically explored HBV integration in pediatric HBV-related HCC using a high-throughput method and found some interesting results. These findings have important implications and should warrant further investigations.

In conclusion, hypervigilance for HCC development is required in HBeAg-negative cirrhotic children. The association of intrahepatic HBsAg, HBV integration and HCC development deserves further studies.

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Disclosure

The authors report no conflicts of interest in this work.

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