

Reversal of Chemotherapy Resistance to Cisplatin in NSCLC by miRNA-195-5p via Targeting the FGF2 Gene

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Objective: To explore the mechanism of miR-195-5p in the pathogenesis non-small cell lung cancer (NSCLC) and cisplatin resistance.

Methods: The function of miR-195-5p in NSCLC and cisplatin resistance were determined by MTT, scratch assay, transwell assay, and nude mice xenograft experiments. miR-195-5p target gene was identified by dual-luciferase reporter assays and real-time PCR analysis.

Results: miR-195-5p content was lower in A549/DDP than that in A549 cells, with reduced chemotherapy sensitivity and increased cell invasion and migration ability. The loss-of-function and gain-of-function assays illustrated that miR-195-5p might have increased the chemosensitivity to cisplatin in the A549/DDP cells and decreased cell migration and invasion. FGF2 is a negatively correlated action target of miR-195-5p. miR-195-5p might affect EMT by inhibiting FGF2. Overexpression of FGF2 resulted in enhanced cisplatin resistance in the cells, while miR-195-5p might have reversed this resistance.

Conclusion: Overall, miR-195-5p might target FGF2 to reduce cisplatin resistance in A549/DDP cells and enhance chemosensitivity.

Keywords: miR-195-5p, NSCLC, cisplatin, chemosensitivity, FGF2

In 2018, there were 774,000 lung cancer (LC) cases and 691,000 deaths due to LC in China.¹ Due to the lack of effective non-invasive screening and early treatment in most areas, the prognosis of LC is poor. Another primary reason for the poor prognosis of LC is chemotherapy resistance. Due to the distant metastasis and recurrence of lesions after local resection, surgical resection alone cannot effectively improve patients' survival. A combination of surgery and adjuvant and neoadjuvant chemotherapy is an effective means to improve the therapeutic effect. However, most patients will gradually develop drug and chemotherapy resistance in the later stage of treatment, resulting in the treatment's ultimate failure.²

MicroRNAs (miRNAs) are endogenous non-protein-coding RNAs with a length of 21–25 nt in eukaryotes that regulate more than 60% of protein-coding genes, influence physiological responses to development, growth, aging, and disease, and play an important role in tumor resistance to chemotherapy by regulating apoptosis, the cell cycle, drug transport, and drug resistance gene expression.³

Specifically, miR-195-5p shows low expression in many malignant tumors, eg, LC, prostate cancer, liver cancer, gastric cancer, ovarian cancer, and cervical cancer, and is closely correlated with tumorigenesis, progression, metastasis, and prognosis.

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It also has an impact on the sensitivity of chemotherapeutic agents.⁴ However, the role of miR-195 in chemotherapy resistance in LC has not been heavily studied.

Fibroblast growth factor 2 (FGF2) is one of the 23 members of the FGF family. In addition to participating in many pathophysiological processes, including angiogenesis and tissue repair, FGF2 also regulates immune cell activity, influences the function of the immune system, and is very important to inhibit tumor resistance to chemotherapy.⁵ We focused on the effect of miR-195 in the chemotherapy resistance of non-small cell LC (NSCLC) and its relationship with the target gene FGF2, aiming to provide new ideas for NSCLC treatment.

Methods

Cell Culture

The human NSCLC cell line A549 and cisplatin (DDP)-resistant cell line A549/DDP were purchased from Tianjin Saier Biotechnology Co., China, and cultured in RPMI-1640 solution. To maintain the cells' normal phenotypic characteristics, the cell culture medium contained 10% fetal bovine serum (FBS), streptomycin (100 mol/mL), and penicillin (100 u/mL), and the culture was placed in a constant-temperature incubator at 37°C and 5% CO₂. Cells were routinely cultured for one week in the medium without DDP.

Plasmid Construction and Cell Transfection

miRNA mimic oligonucleotide duplexes and small interfering RNA (siRNA) were chemically synthesised by GenePharma (Shanghai, China). Cells were seeded in six-well plates and cultured until 50% to 70% confluency. Then cells were transfected by Lipofectamine 2000 (Invitrogen, Carlsbad, New Mexico, USA). The transfection mixture was replaced with a medium containing 10% foetal bovine serum (FBS) after 6 h to 8 h.

The commercially synthesized miR-195-5p-mimics and miR-195-5p-ASO were purchased from GenePharma (Shanghai, China). The pSilencer/shR-FGF2 (shR-FGF2) plasmids expressing siRNA targeting FGF2 were constructed by annealing double-strand hairpin cDNA and inserting it into the pSilencer 2.1-U6 neo vector (Ambion, Austin, TX, USA). The relevant control vector is pSilencer 2.1-U6 neo, which was named "pSilencer-NC" in short in paper. The full-length sequences of human FGF2 cDNA (NM_002006) were obtained by

RT-PCR and cloned into pcDNA3 at HindIII and KpnI sites. The resulting plasmid was termed pcDNA3/FGF2. All primers for PCR amplification are provided in Table 1. Transient transfection was performed in antibiotic-free Opti-MEM medium (Invitrogen) with the Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA) following the manufacturer's protocol. The transfection mixture was replaced with a medium containing 10% foetal bovine serum (FBS) after 6 h. After 48 h after transfection, cells were collected for Western blot (WB) analysis and reverse transcription polymerase chain reaction.

RNA Isolation and RT-qPCR

The total RNAs was extracted from the A549 and A549/DDP cells using the miR-Vana miRNA Isolation Kit (Ambion, Austin, TX, USA). Complementary DNA (cDNA) was obtained by reverse transcription of total cellular RNA using M-MLV reverse transcriptase (Promega, Madison, WI, USA). After the reverse transcription, the expression of miR-195-5p and FGF2 were detected by RT-qPCR using an iQ5 Real-Time PCR Amplifier with U6 as the internal reference. All the primers (Table 1) were purchased from AuGCT, Inc. (Beijing, China).

Western Blotting Analysis

After washing the A549 and A549/DDP cells with pre-cooled phosphate-buffered saline (PBS) three times, each group of cells was lysed on ice with RIPA buffer (Saier Biotechnology Co. Tianjin, China) for 30 min. The cell protein sample was obtained by transferring the cell lysate to a 1.5mL centrifuge tube. Then, the protein sample went SDS denaturing 10% polyacrylamide gel electrophoresis (SDS-PAGE) until the target proteins were effectively separated. The proteins were electrotransferred to a PVDF membrane, and blocked for 2 h with 5% skim milk at room temperature. After the proteins were incubated with primary antibodies at 4°C overnight, the blots were washed four times with PBS, incubated with horseradish peroxidase (HRP)-labelled secondary antibody at room temperature for 2 h and visualised through enhanced chemiluminescence (Saier Biotechnology Co. Tianjin, China). The grayscale values of the protein bands were analyzed using the Image J, and the expression of each target protein was calculated with GAPDH or β -actin used as the internal reference. The following primary antibodies

Table I The Primers and Oligonucleotides Used in This Work

Name	Sequence (5'-3')
has-miR-195-5p-mimics	UAGCAGCACAGAAAUUUGGC
has-miR-195-ASO	GCCAAUAAUUUCUGUGCUGCUA
has-miR-195-RT	GTCGTATCCAGTGCAGGGTCCGAGGTGCTGCTGGATACGACGCCAATAT
has-miR-195-Fwd	TGCGGTAGCAGCACAGAAATATTGGC
U6 RT	GTCGTATCCAGTGCAGGGTCCGAGGTATTGCTGCTGGATACGACAAAATATGGAAC
U6 Fwd	TGCGGGTGCTCGCTTCGGCAGC
Reverse	CCAGTGCAGGGTCCGAGGT
PmiRGLO-FGF2-3'UTR-top	AAACTAGCGGCCGCTAGTTAATTTAAATATTTTCTGCTAGTTAACTATGT
PmiRGLO-FGF2-3'UTR-bot	CTAGACATAGTTAACTAGCAGCAAAATATTTTAAATTAAGTAGCGGCCGCTAGTTT
PmiRGLO-FGF2-3'UTR-mut-top	AAACTAGCGGCCGCTAGTTAATTTAAATATTTTACACGAAGTTAACTATGT
PmiRGLO-FGF2-3'UTR-mut-bot	CTAGACATAGTTAACTTCGTGTAATTTTAAATTAAGTAGCGGCCGCTAGTTT
shR-FGF2-top	GATCCTATAGCTCAGTTTGGATAATTCTCGAGAATTATCCAACTGAGCTATATTTTGA
shR-FGF2-bot	AGCTTCAAAAATATAGCTCAGTTTGGATAATTCTCGAGAATTATCCAACTGAGCTATAG
pcDNA3-FGF2 -S	TATAAGCTTCT GGTGGGTGTG GGGGGTGGAGAT
pcDNA3-FGF2-AS	CGGGGTACCGCTCTTAGCAGACATTGGAAG
FGF2-qPCR-Fwd	AAGAGCGACCCTCACATCAAG
FGF2-qPCR-Rev	CGTTTCAGTGCCACATACCAAC
β -actin-qPCR-Fwd	CGTGACATTAAGGAGAAGCTG
β -actin-qPCR-Rev	CTAGAAGCATTTGCGGTGGAC

were used in the Western blot assay: (1) rabbit anti-GAPDH (Tianjin Saier Biotech; 1:1000), (2) rabbit anti-E-cadherin (Tianjin Saier Biotech; 1:1000), (3) rabbit anti-vimentin (Tianjin Saier Biotech; 1:1000), (4) rabbit anti-FGF2 (Tianjin Saier Biotech; 1:1000).

Luciferase Assay

Bioinformatic analysis software TargetScanHuman 7.2, target gene prediction software, and miRanda were applied to perform the bioinformatic functional analysis of miR-195-5p and predict target genes (<https://www.targetscan.org>). Luciferase reporter assays were co-transfected in 48-well plates with 0.5 μ g of mimics-NC or miR-195-5p mimics and 0.5 μ g of reporter vector FGF2-3'UTR or FGF2-3'UTR mut. The activities of the firefly luciferase and the Renilla luciferase were detected using the Dual-Glo system (Promega, Madison, WI) 48 h post-transfection.

The Cell Function Tests

Methyl Thiazole Tetrazole (MTT) Assay

The cells, following transfection, were seeded at a density of 3,000 cells per well, and cell activity was detected by MTT at 48 h after transfection (incubated with 10 μ L MTT solution at a concentration of 10 mg/mL at 37°C for 4 h). The original culture solution was then removed, and added 100 μ L DMSO to each well. After shaking for 10 min, on a shaker to fully dissolve the methylzan the blue-purple crystalline material. The absorbance values of each well were measured at 570 nm via an uQuant Universal Microplate Spectrophotometer (Bio-Tek Instruments, Winooski, USA).

The Scratch Healing Assay

Transfected cells were scraped and at least three artificial wounds were photographed using 10 high-power fields at 0 h, 24 h and 48 h after scratch.

The Invasion Assay

For the invasion assay, 6×10^4 A549 cells or A549/DDP cells were seeded on upper chamber of the inserts coated by 40 μ L Matrigel (DB Biosciences) diluted to 2 μ g/ μ L. Meanwhile, the bottom of the inserts were incubated in medium containing 20% FBS. After cultured for 48 h, the number of cells migration or invasion to the bottom of chamber was counted per field.

Mouse Model of Tumor Xenograft

After transfection, 1×10^7 A549 and A549/DDP cells were injected into the endermic of 6-week-female BALB/c nude mice (4 per group). Tumor size was measured every four days after two weeks of injection. The tumor volume was calculated as follows: $\text{length} \times \text{width}^2 \times 1/2$. All mice were sacrificed 5 weeks after implantation, the tumors were isolated from the mice and stored at -80°C . Which is approved by Animal Use Committee of Tianjin Medical University Cancer Institute and Hospital (Approved No.: NSFC-AE2020103) and complied with the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Statistical Analysis

All of the experiments were repeated three times. The SPSS 21.0 and GraphPad Prism7 software were adopted for the statistical analysis. The measurement data conformed to the normal distribution were expressed as $\bar{X} \pm s$, and *t*-test was used to compare means between two groups. For the comparison of means among multiple groups, the homogeneity of variance was tested first, and then one-way ANOVA was conducted; $P < 0.05$ was considered statistically significant.

Results

Biological Efficacy in A549/DDP and A549 Cells

To explore the differences between A549/DDP and A549 cells. The results showed that the IC_{50} of DDP on the A549 cells was 0.8 $\mu\text{g/mL}$, while the IC_{50} of DDP on the A549/DDP cells was 2.2 $\mu\text{g/mL}$. These results showed that the inhibitory effect of DDP on the activity of A549 cells was stronger than that of A549/DDP cells and A549/DDP cells were less sensitive to DDP than A549 cells. (Figure 1A). The scratch assay revealed that the migration rate of the A549 cells after 24 h was 20%, significantly lower than the migration rate of 57% in the A549/DDP

cells. This suggested that the A549 cells had a lower cell migration ability compared to the A549/DDP cells (Figure 1B). Moreover, the invasion ability of the A549 cells was weaker than that of the A549/DDP cells (Figure 1C).

The Effects of miR-195-5p on the Cell Function

The expression of miR-195-5p in the A549/DDP cells was lower than that in the A549 cells (Figure 2A). MTT assay showed increased sensitivity and decreased the viability of A549/DDP cells after transfection with miR-195-5p mimics via the action of cisplatin. Meanwhile, the A549 cells transfected with the miR-195-5p-ASO had a reduced sensitivity, increased cells viability, and increased cisplatin resistance (Figure 2B and C). The migration and invasion ability of the A549/DDP cells decreased after transfection with miR-195-5p mimics, while the migration and invasion ability of the A549 cells transfected with the miR-195-5p-ASO increased significantly (Figure 2D and E).

We then examined whether the molecular changes typical to affected EMT occurred in A549 and A549/DDP cells after transfected miR-195-5p or miR-195-5p-ASO. miR-195-5p increased the expression levels of E-cadherin, but decreased the level of vimentin, while the opposite results were observed in A549 cells after transfected miR-195-5p-ASO (Figure 2F). These results suggested that miR-195-5p inhibits the epithelial-like/mesenchymal-like phenotype.

Prediction and Verification of Target Genes of miR-195-5p

The bioinformatics functional analysis showed that the mature sequence of miR-195-5p is „UAGCAGCACAG AAAUAUUGGC,,, which is highly conserved among species. Target prediction using bioinformatics methods suggested the candidate targets for miR-195-5p, we chose FGF2 as putative target for further study after considering available functional knowledge. The luciferase activities were decreased when FGF2 3'UTR and miR-195-5p were co-transfected, whereas co-transfection with miR-195-5p-ASO caused a increase compared with the control (Figure 3A). In contrast, mutation of the miR-195-5p binding site in the FGF2 3' UTR abolished the effect of both miR-195-5p over-expression and miR-195-5p-ASO on the fluorescence intensity of the

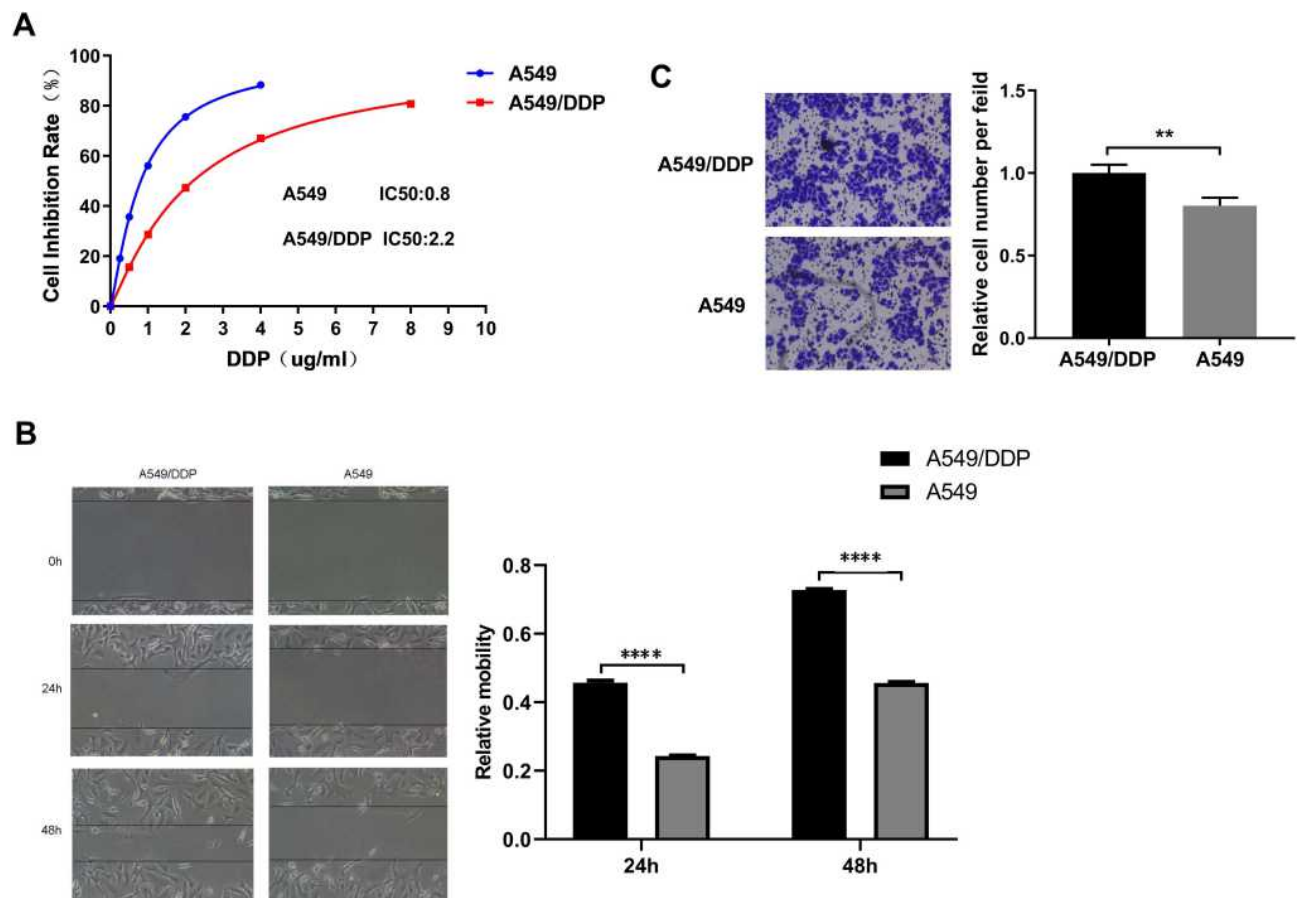


Figure 1 The difference in biological efficacy A549 and A549/DDP cells. **(A)** The MTT assay was used to detect the difference in cisplatin chemosensitivity of the two cell lines. The cisplatin sensitivity of A549/DDP by the MTT assay, IC₅₀ = 2.2 μg/mL. The cisplatin sensitivity of A549 by the MTT assay, IC₅₀ = 0.8 μg/mL **(B)** Wound healing assays were performed to reveal the relative mobility. The line graphs show the mean ± SD of three scratches from one representative experiment. **(C)** 24 h after transfection, A549 and A549/DDP cells (in 200 μL serum-free medium) were seeded into inserts and incubated for 60 h, chambers were disassembled, and the membranes were stained with 2% crystal violet solution for 5 min and placed on a glass slide. Cells at the lower surface of the membrane were counted under a light microscope at five random fields. All data represent mean ± SD of three independent experiments. **p < 0.01. The invasion ability of the A549 cells was weaker than that of the A549/DDP cells. All data represent mean ± SD of three independent experiments. ****p < 0.0001.

luciferase activities (Figure 3B). These results suggest that miR-195-5p may directly target FGF2. Next we further determined the effect of miR-195-5p on endogenous FGF2 expression. RT-qPCR showed that FGF2 mRNA levels were decreased in A549 and A549/DDP cells transfected with miR-195-5p (Figure 3C and D). Furthermore, Western blotting assays showed that miR-195-5p decreased FGF2 expression (Figure 3E and F). These results indicate that miR-195-5p suppresses FGF2 expression at both the mRNA and protein levels in both cells. Furthermore, protein and mRNA expression results of FGF2 in the A549 and A549/DDP cells detected by WB and RT-qPCR demonstrated a significant difference in the expressions between the A549 and A549/DDP cells (Figure 3G).

Effect of MiR-195-5p on the Sensitivity of A549 and A549/DDP Cells to Cisplatin

Sensitivity to cisplatin in the different groups was measured as the IC₅₀ concentration after transfection. Growth inhibition rate of the A549 cells was 52.0% under the effect of cisplatin at an IC₅₀ concentration of 0.8 μg/mL but decreased to 37.6% after transfection with miR-195-5p-ASO. Moreover, inhibition rate of cell growth of A549 cells increased to 75.5% after transfection with shR-FGF2 alone, while the growth inhibition rate of the A549 cells after transfection of miR-195-5p-ASO and shR-FGF2 was 46.7% (Figure 4A). In the A549/DDP cells, growth inhibition rate after miR-195-5p transfection increased from 50.3% to 66.0% under the effect of cisplatin at an IC₅₀ concentration of 2.2 μg/mL. The growth inhibition rate of

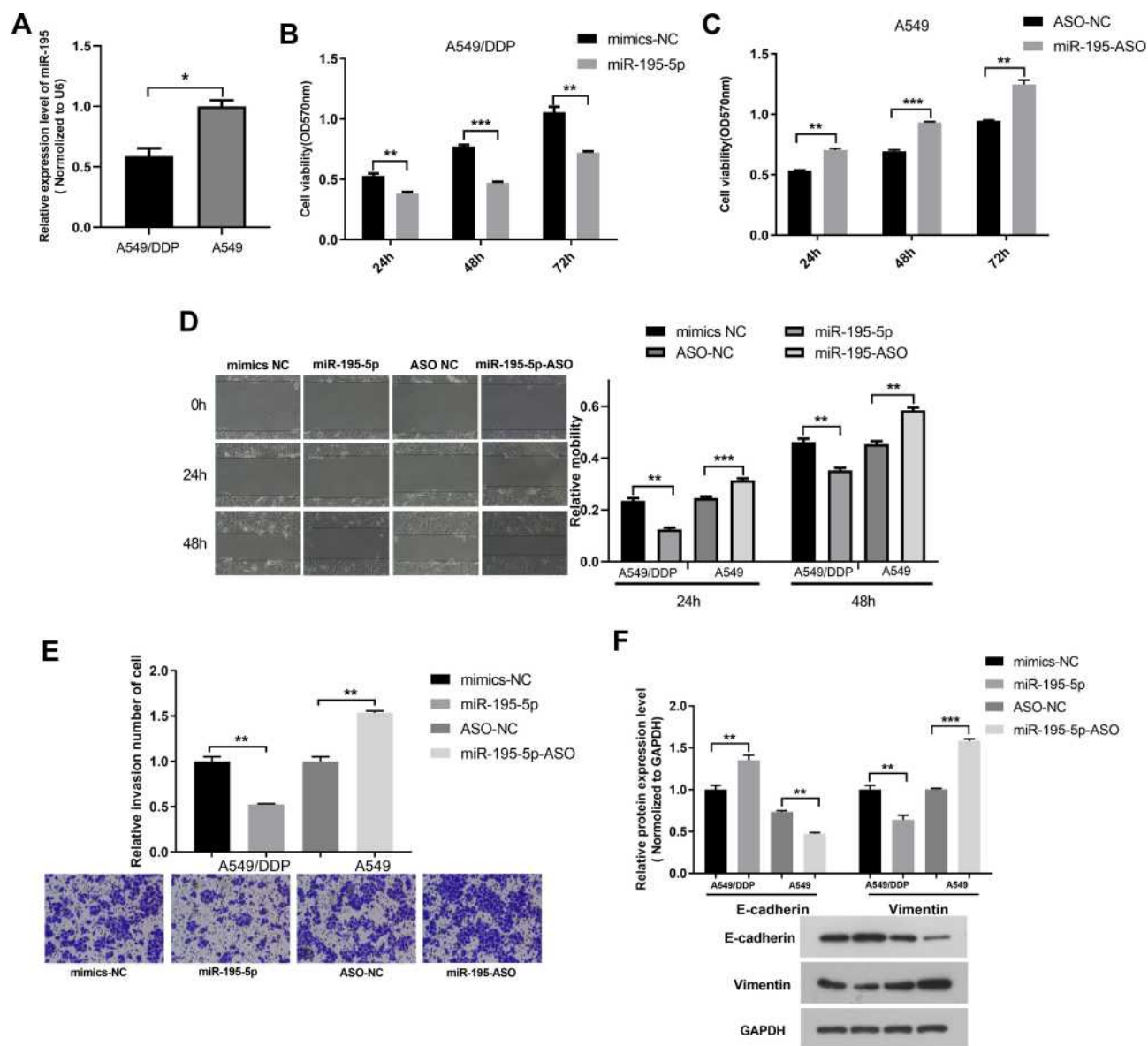


Figure 2 The effects of miR-195-5p on the cell function of the A549 and A549/DDP cells. **(A)** The miR-195-5p expression was examined in A549 and A549/DDP cells using RT-qPCR. **(B and C)** MTT assay was applied to determine the cell viability of A549/DDP and A549 cells. **(D)** Wound healing assays were measured in A549 and A549/DDP cells with miR-195-5p mimics or inhibitor. **(E)** Cell invasion analysis in cells with miR-195-5p mimics or inhibitor were detected. **(F)** Protein expression levels of EMT-associated markers were assessed by Western blotting and quantification was presented in bar graphs. All data represent mean \pm SD of three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

the A549/DDP cells after transfection with pcDNA3-FGF2 alone was 31.1%, while the growth inhibition rate of the A549/DDP cells after simultaneous transfection with the miR-195-5p and pcDNA3-FGF2 was 46.7% (Figure 4B).

Over-Expression of FGF2 Abolishes the Effects Induced by miR-195-5p in A549/DDP Cells

The effects of miR-195-5p overexpression on mobility, migration and invasion were significantly impaired when

FGF2 was overexpressed (Figure 5A–C). Furthermore, the expression of EMT markers in the miR-195-5p-expressing cells were restored to the normal levels after overexpression of FGF2 (Figure 5D). Conversely, knockdown of FGF2 expression in A549 cells abolished the promotion effect of miR-195-ASO on cell activity, migration, invasion, and EMT (Figure 5E–H). Collectively, these results indicate that the effects of miR-195-5p are probably due to the knockdown of FGF2.

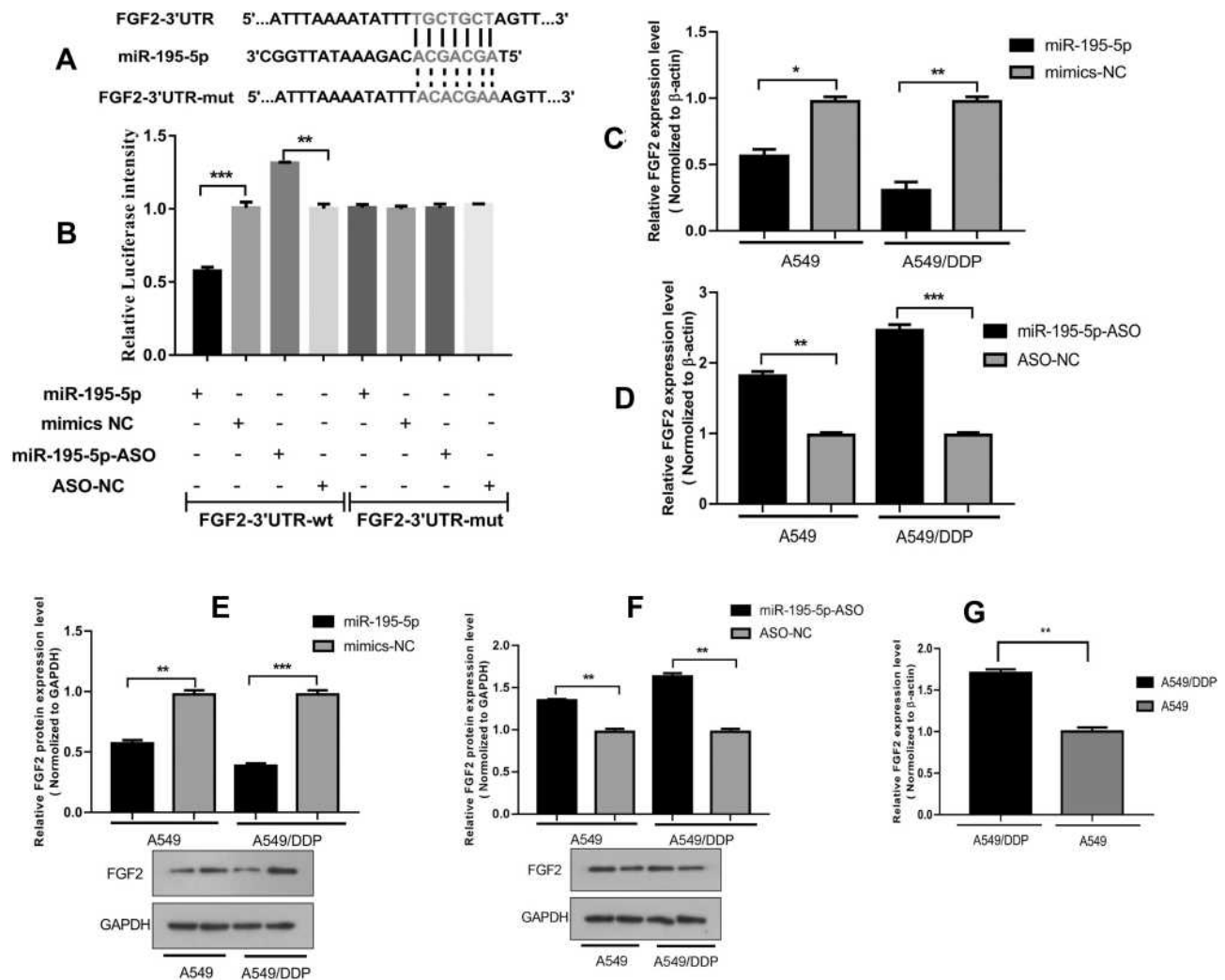


Figure 3 The miR-195-5p target gene prediction and verification. **(A)** The predicted miR-195-5p-binding sites in FGF2 mRNA using Targetscan 7.0 are shown. **(B)** A549 cells were transfected with PmiRGLO reporter vectors containing either wild-type or mutant FGF2 3'UTR. The relative Luciferase activity was determined after transfection. **(C and D)** The expression of FGF2 was analyzed in cells with miR-195-5p mimics or inhibitor. **(E and F)** The expression of FGF2 protein was detected by WB in cells with miR-195-5p mimics or inhibitor. **(G)** The difference expression of FGF2 was analyzed by RT-qPCR in A549 and A549/DDP cells. All data represent mean \pm SD of three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

miR-195-5p Overexpression Inhibit NSCLC

Tumor xenograft experiments was performed using A549/DDP cells transfected with miR-195-5p and A549 cells transfected with miR-195-5p-ASO. miR-195-5p overexpression inhibited and knockdown miR-195-5p promoted growth of NSCLC in vivo (Figure 6A–C). At the same time, tumor in the group injected with A549/DDP cells were larger than those injected with A549 cells. RT-qPCR and Western blot analysis showed that the protein and mRNA expressions of FGF2 in the tumor-forming tissues of the A549/DDP cells were significantly higher than those of FGF2 in the tumor-forming tissues of the A549 cells. Meanwhile, the miR-195-5p expression in the

tumor tissues of the A549/DDP cells was significantly lower than that in the tumor-forming tissues of the A549 cells. (Figure 6D–F).

Discussion

According to statistics by the National Cancer Institute of the USA, five-year overall survival was 23.6% in NSCLC⁶ with rates of approximately 55.6% for NSCLC without metastasis and 28.9% for NSCLC with regional lymph node metastasis. Once distant metastasis occurred, the five-year survival was only 4.5%.

One of the main reasons is drug resistance to chemotherapy, which is a complex process with many influencing factors, primarily individual genetic differences, the

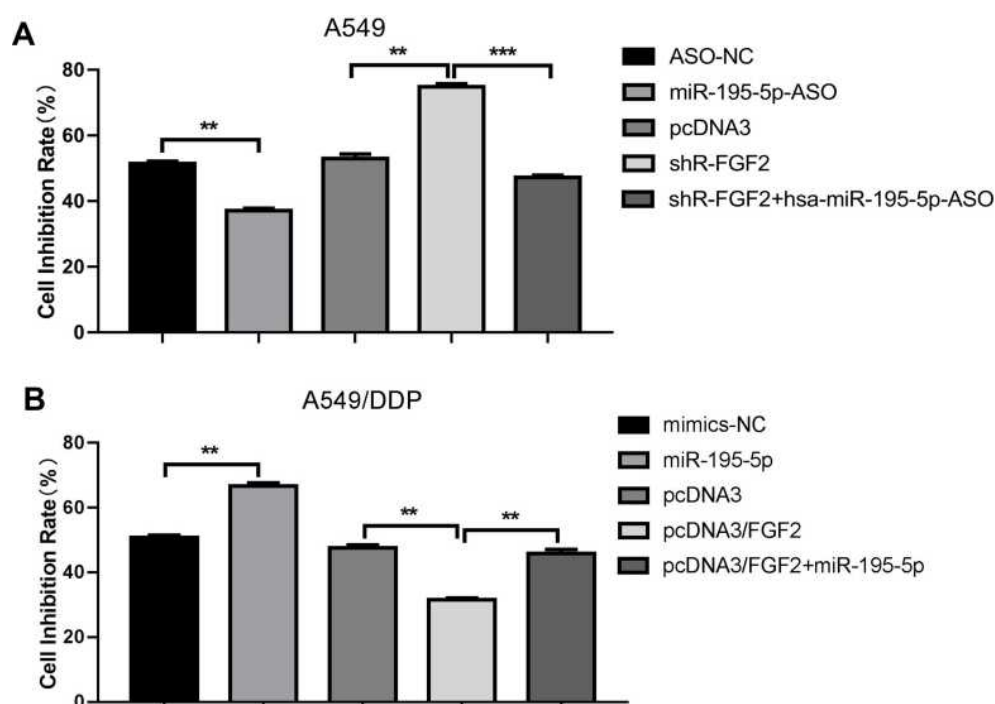


Figure 4 MiR-195-5p increased the sensitivity of A549 and A549/DDP cells to cisplatin by inhibiting the target gene FGF2. **(A and B)** An MTT assay tested the cells sensitivity of A549 and A549/DDP cells to cisplatin. All data represent mean \pm SD of three independent experiments. ** $p < 0.01$, *** $p < 0.001$.

tumor microenvironment, tumor stem cells, drug inactivation, reduced drug absorption, and altered metabolism of antitumor drugs. Some miRNAs can act on drug efflux pump molecules, apoptosis regulatory proteins, and cycle regulatory proteins to alter the drug-resistance of tumor cells.⁷ Cisplatin is an early developed and clinically mature drug for the treatment of LC and has remained the drug of choice for LC chemotherapy up to now. However, chemotherapy resistance limits the efficacy of cisplatin in clinical use. After entering the body, cisplatin dissociates in a low-chloride environment to form hydrated cations, which mainly act on DNA, forming covalent bonds with DNA and, thus, limiting DNA deconvolution and impeding DNA replication to inhibit the division of cancer cells and promote their apoptosis and necrosis.⁸

The miR-195 gene is a newly found micro RNA of the miR-15/107 family, which is homologous to miRNA-15, miRNA-16, miRNA-424, and miRNA-497 and shares the 5'core sequence of CGA CGA.⁹ The dysregulation of miR-195 will cause the development of many diseases; miRNA-195 was first found to be highly expressed in transgenic mice with cardiac hypertrophy and to cause progression from the pathological heart disease to heart failure.¹⁰

The high expression of miR-195 can inhibit tumorigenesis and progression and affect the sensitivity of chemotherapeutic agents for malignant tumors, a property that has been applied to drug development. Therefore, it was considered that propofol might induce miR-195 to inhibit the proliferation, migration, and invasion of gastric cancer cells.¹¹ Zuo et al investigated miR-195 together with long-chain non-coding 00485 (LINC00485) in cisplatin resistance and found that overexpression of miR-195 or silencing of LINC00485 enhances the sensitivity of lung adenocarcinoma cells to cisplatin.¹² These studies provided a theoretical basis for drug resistance studies in LC chemotherapy.

We found that the miR-195-5p expression in NSCLC resistant cells was significantly lower than that in the parental strain cells, while the chemotherapy sensitivity was decreased and the cell migration and invasion functions increased. Subsequently, we demonstrated that overexpression of miR-195-5p improved the chemosensitivity of cisplatin-resistant cells and reduced their migration and invasion functions. We found that miR-195-5p might play a role as an oncogene in LC cells and have an inhibitory effect on chemotherapy resistance to cisplatin. Afterward, by target gene prediction and confirmation, it was concluded that miR-195-5p might directly target downstream

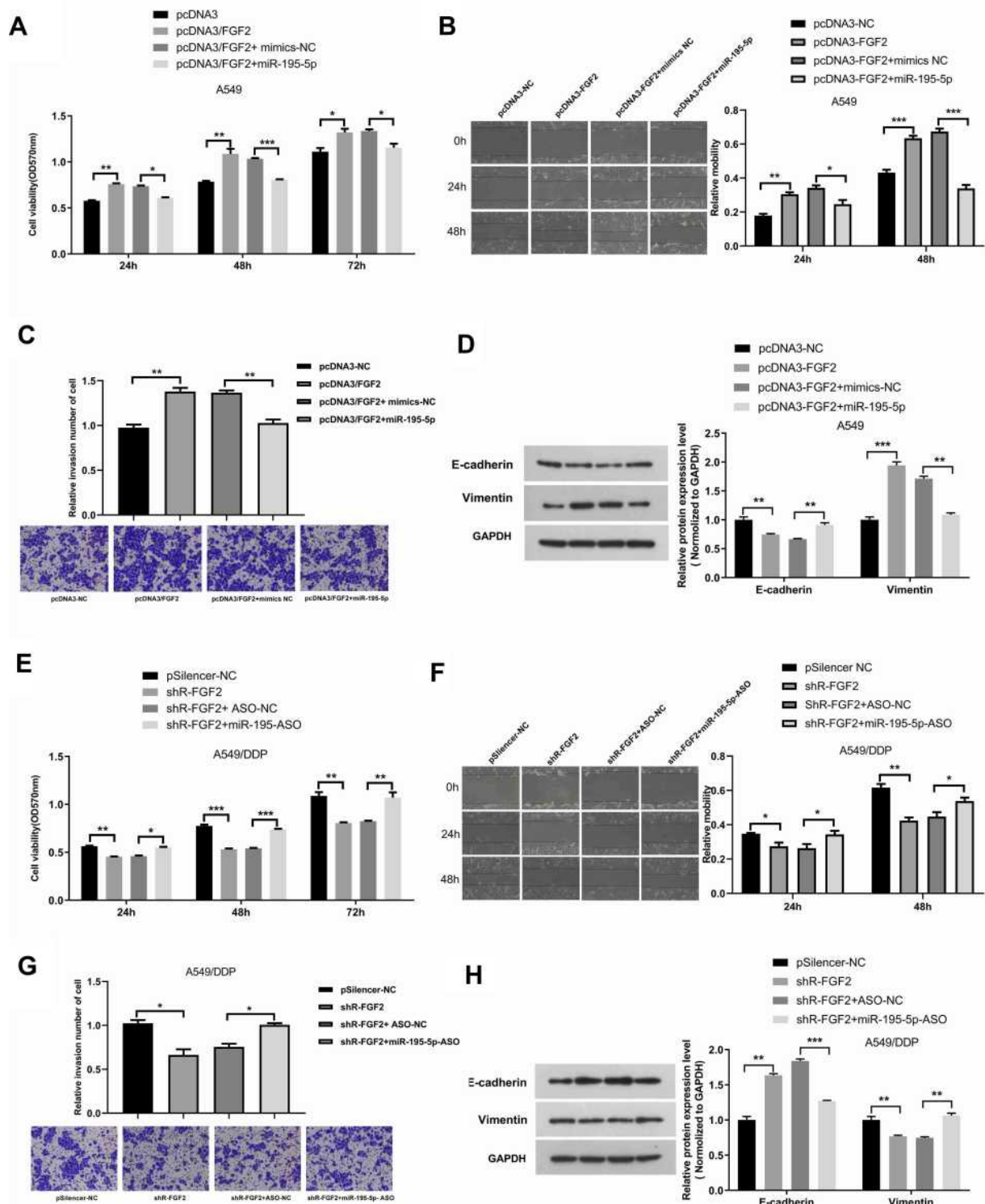


Figure 5 Over-expression of FGF2 abolishes the effects induced by miR-195-5p in A549/DDP cells. (A–C) Over-expression of FGF2 rescues the effect of miR-195-5p on cell viability (A), migration (B) and invasion (C) in A549/DDP cells. (D) After co-transfection with miR-195-5p and FGF2 in A549/DDP, Western blot assays were used to present the expression of EMT markers (E-cadherin and vimentin). (E–G) Knockdown of FGF2 rescues the effect of miR-195-5p on cell viability (E), migration (F) and invasion (G) in A549 cells. (H) After co-transfection with miR-195-5p-ASO and shR-FGF2 in A549, Western blot assays were used to present the expression of EMT markers (E-cadherin and vimentin). All data represent mean \pm SD of three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

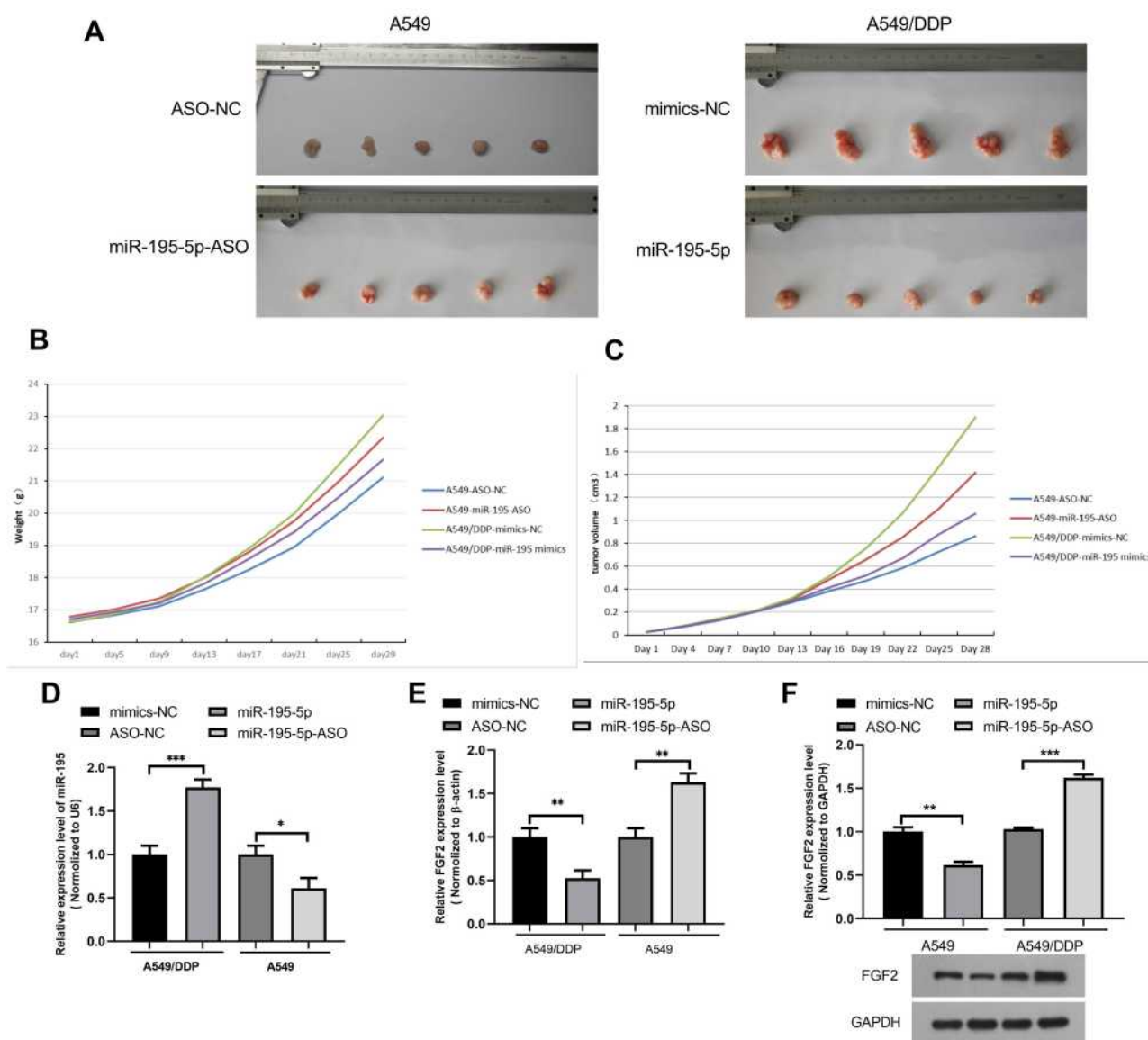


Figure 6 miR-195-5p over-expression inhibit NSCLC in vivo. **(A)** The mice were euthanized, and the tumors were isolated 5 weeks after implantation. **(B)** The body weight growth curve of nude mice was studied after cell injection. **(C)** Tumor growth curves of studied nude mice after cell injection was showed. **(D and E)** miR-195-5p expression level and FGF2 mRNA expression level of nodules in mice. **(F)** FGF2 protein expression level of nodules in mice. All data represent mean \pm SD of three independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

gene FGF2 to affect the resistance to cisplatin and migration invasion ability of LC cells.

Fibroblast growth factors are peptides secreted by the pituitary and hypothalamus as ligands for tyrosine kinase receptors (FGF receptors, FGFRs); FGFs and FGFRs are expressed in many nuclear tissues of epithelial and mesenchymal origin and are involved in the growth, differentiation, and angiogenesis of different organs during embryonic development. They also regulate important physiological behaviors such as proliferation, survival, and migration during human embryonic development, organogenesis, and tissue formation.¹³

In tumors, FGF2 is produced by the tumor cells or the surrounding stromal cells; it has a strong proliferation effect on fibroblasts and can promote the proliferation, migration, and invasion of tumor cells as well as affect endothelial cells to stimulate angiogenesis, which plays a promotional role in tumor growth. Clinical studies have found that FGF2 is overexpressed in malignant tumor tissues (such as prostate cancer, LC, esophageal cancer, and gynecological tumors) and is closely correlated with carcinogenic behaviors, including lymph node metastasis, invasion, migration, and prognosis.¹⁴ Moreover, increased expression of FGF2 can be detected in a variety of tumor

tissues and body fluids. Many researchers believe that FGF2 can be considered a tumor marker to predict and provide an early warning for changes in tumors.¹⁵

Regarding NSCLC, Huang et al suggested that FGF2 might be a target of miR-497-5p, which could reduce the invasion and metastasis of NSCLC by inhibiting FGF2.¹⁶ Wang et al found that FGF2 is a target gene of miR-646 in NSCLC and that miR-646 might reduce the proliferation and invasion ability of NSCLC by inhibiting FGF2.¹⁷ In the present study, the FGF2 expression significantly reduced in the A549/DDP cells with overexpression of miR-195-5p upon WB assay and PCR, while the FGF2 expression significantly elevated in the A549 cells with inhibited expression of miR-195-5p. Via the dual-luciferase reporter assay, FGF2 was confirmed to be the direct target gene of miR-195-5p, which could improve the chemosensitivity of cisplatin-resistant LC cells through FGF2 and inhibit migration and invasion.

Subsequently, it was also confirmed by WB assay that the inhibition of chemoresistance of miR-195-5p on FGF2 could be achieved by affecting EMT.¹⁸ The EMT process is involved in embryogenesis, wound healing, and fibrotic tissue changes and is an important biological process in which malignant tumor cells of epithelial origin acquire the ability to migrate and invade. With the occurrence of EMT in tumor cells, the secretion of intercellular junction protein and epithelial-like marker molecule E-cadherin decreases, while the expressions of mesenchymal cell phenotype N-cadherin, vimentin, and FSP-1 increase. This changes the cell morphology, causing the epithelial tumor cells to lose their polarity and basement membrane connection and adhesion and gradually break away from the binding between the epidermal cells, thus acquiring strong invasive and migratory properties.¹⁹ EMT leads to the tolerance of tumor cells to cytotoxic drugs and apoptosis regulation and is important in the development of cisplatin resistance in NSCLC.²⁰ Currently, extensive literature confirms that EMT and the drug resistance of tumor cells are closely linked and inseparable.²¹ It has been reported that CDDP, doxorubicin, and paclitaxel-resistant cell lines have the characteristics of EMT and a significantly increased cell invasion and metastasis ability compared with their sensitive parental cells. Moreover, cells that develop EMT show stronger drug resistance.²² Our results suggested that miR-195-5p could have inhibited the EMT signaling pathway in the A549/DDP cells to improve the cells' resistance to cisplatin. Moreover, it was also confirmed that the overexpression of FGF2 might have promoted EMT in the

A549/DDP cells, while miR-195-5p might have inhibited EMT by reducing the FGF2 expression. We also confirmed that miR-195-5p might have reversed the chemoresistance of the A549/DDP cells by inhibiting FGF2 expression, and miR-195-5p could have reversed the chemoresistance in the A549/DDP cells by inhibiting the expression of the target gene FGF2.

In summary, via cellular and nude mice assays, this study confirmed that there is a differential expression and difference in the cellular functions of miR-195-5p in cisplatin-resistant cells, and it was verified that the direct target gene is FGF2. We also confirmed that miR-195-5p could reverse the chemoresistance in A549/DDP cells by inhibiting the target gene, FGF2, to affect the development of EMT in the resistant cells. We considered that as an oncogene, miR-195-5p might have a potential for drug development and efficacy prediction in the reversal of chemoresistance in NSCLC, with promising practical applications.

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Disclosure

The authors reported no conflicts of interest for this work.

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