

Impact of CD105 Flow-Cytometric Expression on Childhood B-Acute Lymphoblastic Leukemia

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Background: CD105 (Endoglin) is a receptor of the transforming growth factor-Beta (TGF- β) superfamily. It is expressed in angiogenic endothelial cells and is considered a powerful marker of angiogenesis and a potential main player in the pathogenesis of vascular diseases as well as tumor progression. CD105 expression was correlated with poor prognosis in many types of solid malignancies, however, its influence on hematological neoplasms is still an area of interest.

Purpose: To assess the flow-cytometric expression of CD105 in childhood B-acute lymphoblastic leukemia (B-ALL) and its relation to disease response after the induction chemotherapy.

Subjects and Methods: Eighty children newly diagnosed with B-ALL were screened for flow-cytometric expression of CD105 at time of diagnosis, then they were followed up to detect their response to induction therapy.

Results: CD105 was expressed in 41.2% of B-ALL patients. Higher expression of CD105 was observed in high and very high-risk groups. The multivariate analysis considered CD105 positivity as an independent prognostic marker for response to induction therapy. Values higher than 2.5 Specific fluorescence indices (SFIs) and 35% expression were sensitive predictors to induction failure.

Conclusion: CD105 can be considered as a potential prognostic marker for the detection of response to induction therapy in childhood B-ALL, and it can serve to optimize treatment decisions.

Keywords: B-acute lymphoblastic leukemia, endoglin, CD105

Introduction

Acute lymphoblastic leukemia (ALL) is a malignant disorder representing clonal expansion and arrest of maturation of lymphoid progenitor cells in the bone marrow, blood, and extramedullary sites.¹ The incidence of ALL generally follows a bimodal distribution, with its first peak occurring in childhood and the second around the age of 50.²

With proper risk stratification, ALL responds well to chemotherapy, yet early mortality during the induction phase of chemotherapy is not uncommon. Besides, the high relapse rate remains a major problem.³ Therefore, identification of new prognostic markers will not only aid in increasing the accuracy of patients' risk stratification but also will minimize the chances of relapse by optimizing therapy at the early stages of treatment.⁴

Flow-cytometry immunophenotyping is considered a potent technology used to identify cell membrane antigens.⁵ The identification of surface antigens on

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leukemic cells is essential for the assignment of the proper treatment plan and is also valuable for assessing prognosis and searching for applicable markers to detect minimal residual disease.⁶

CD105 (Endoglin) is a homodimeric transmembrane co-receptor that interacts with transforming growth factor-Beta (TGF- β) receptors type I and III, consequently adjusting angiogenesis by regulating proliferation, differentiation, and endothelial cell migration.⁷ It is expressed on the surface of endothelial cells, stromal cells, melanocytes, and different hematopoietic cells.^{8–10} It is highly expressed as well on the surface of activated vascular endothelial cells but not or weakly expressed on the normal quiescent vessels. Therefore, it has been proposed as a marker of angiogenesis.⁹ In contrast, lack of CD105 expression is diagnostic for hereditary hemorrhagic telangiectasia type –1, a disease characterized by multiple vascular malformations.¹¹

Buhring et al¹² reported that CD105 was not detectable on normal bone marrow CD34 positive hemopoietic precursor cells, mature T, B, natural killer, and myeloid cells, but was instead present on a subset of glycophorin A-positive mononuclear cells. In a subsequent study, Rokhlin et al¹³ demonstrated the existence of two CD105 positive populations in bone marrow: B-lineage precursor cells and pro-erythroblasts. The expression of CD105 on hemopoietic precursor cells serves as a mediator affecting quiescence and enables long-term repopulation.¹⁴

Regarding hematological malignancies, CD105 expression was reported in myelodysplastic syndrome,¹⁵ ALL,^{16,17} and acute myeloid leukemia (AML).^{16–19} CD105 positive blast cells exhibit higher leukemogenic activity when compared to their negative counterpart.²⁰

Cosimato, and his colleagues,¹⁶ extensively studied the expression of CD105 on blast cells of acute leukemia patients and reported that CD105 was expressed in the majority of B-ALL cases, and only the most immature types expressed this antigen, as it was consistently absent in “Burkitt-like” mature B-ALL without referring to its effect on patient’s prognosis. Also, Poręba et al¹⁷ reported that CD105 expression in patients with ALL was limited to a small group of patients and needs to be confirmed on a larger group. So, adequate information about its prognostic impact in B-ALL is still worth further evaluation.

The current work aimed to assess the CD105 flow-cytometric expression in B-ALL pediatric patients and its relation to disease response after the induction phase of chemotherapy.

Subjects and Methods

The current study was carried out on 80 children newly diagnosed with B-ALL referred to Hematology/Oncology Unit, Pediatric Department, Tanta University Hospitals, and Pediatric Oncology Unit, Tanta Cancer Center from June 2017 to August 2020.

Cases were diagnosed based on clinical presentation, complete blood count (CBC), bone marrow (BM) examination, morphological and cytochemical smears as well as immunophenotyping.

Risk stratification was done according to Children’s Oncology Group (COG) protocols.²¹ Cases were classified into three groups; standard-risk group (54 cases, 67.5%), high-risk group (17 cases, 21.25%), and very high-risk group (9 cases, 11.25%).

Initial investigations included; automated CBC on ERMA PCE-210N cell counter (Tokyo, Japan) with an examination of Giemsa-stained smears, Liver and renal function tests, and lactate dehydrogenase enzyme (LDH) on a fully automated chemistry analyzer (Konelab Prime 60i, Thermo-scientific, Vantaa, Finland), erythrocyte sedimentation rate (ESR), and cerebrospinal fluid (CSF) cytological examination. BM aspiration samples were evaluated through Giemsa-stained smears, and acute leukemia was diagnosed by the presence of $\geq 20\%$ blast cells. Philadelphia (Ph) chromosome was detected by fluorescence in-situ hybridization (FISH).

Immunophenotyping analysis was done on BM samples collected into EDTA-containing tubes using the four-color flow cytometry Becton Dickinson (BD) FACS Calibur instrument (Becton Dickinson, San Diego, California, USA), using the Cell Quest software (Becton Dickinson, version 3, verify software House Topsham, ME, USA). Before each run, calibrated beads provided by the manufacture were used to adjust the compensation of different fluorochromes. Mouse isotopic controls were used as negative controls to exclude autofluorescence. At least 10,000 events/tubes were acquired for each analysis. Blast cells were identified based on dim/intermediate CD45 Peridinin Chlorophyll Protein Complex (Per-CP) expression versus log side scatter characteristics (CD45/SSC gating strategy). The gated fluorescence dot plot was evaluated for positive cells using cursor position from the dot plot of isotypic controls. The internal negative control was checked using normal cells in the sample that lacked the antigen, while the internal positive control was checked using the normal cells in the sample that expressed the antigen.

B-ALL was diagnosed using the acute leukemia panel that included the following combinations of surface markers: CD45/CD14/CD117/CD34, CD45/HLA-DR/CD10/CD38, CD45/CD64/CD19/CD20, CD45/CD7/CD33, CD45/CD2/CD13, and the cytoplasmic markers anti TDT/anti MPO, anti CD79a and anti cyt μ , markers were supplied by Becton Dickinson (BD biosciences, Mountain View, California). B-ALL blast cells were identified by positive expression of CD19 in addition to CD10 and/or CD79a.

Immunophenotyping analysis of CD105 using CD105 fluorescein isothiocyanate (FITC) labeled monoclonal antibody, supplied by BD biosciences, Catalog number 561443 Clone 266 (RUO). The total leucocytic count (TLC) was adjusted to 10^6 cells/tube. Cells were incubated with 5 μ of CD45 Per-CP, and 10 μ of CD105 FITC in the dark, at room temperature, for 25 min. Red blood cells were then lysed with 1 mL of BD FACS lysing solution for 20 min before centrifugation, the cells were then washed twice with 0.5 mL of phosphate buffer saline (PBS), and suspended in 300 μ L of PBS to be ready for the flow cytometer acquisition. Blast cells were selectively gated using CD45/SSC strategy, and the percent of blast cells expressing CD105 was determined within this population using a cutoff value for positivity $>5\%$.¹⁷ Specific fluorescence indices (SFIs) were calculated by dividing median fluorescence intensity (MFI) of CD105 by MFI of negative isotype control, positive expression was defined as SFIs >1.5 ¹⁹ (Figures 1 and 2).

After being fully investigated at diagnosis, all the patients received induction chemotherapy, according to the protocol adopted by the treating centers; the Modified St Jude Children's Research Hospital (SJCRH) Total Therapy XV Protocol.²² At the end of induction therapy on day 28, all patients were reevaluated by CBC and BM samples. Remission was identified by the absence of peripheral blood blasts and BM blast cells less than 5%. Refractoriness to therapy was defined by the presence of greater than 5% BM blasts and/or CNS infiltration by leukemic cells. Those who did not achieve complete remission were reassigned to more intensified treatment protocols.²²

Statistical Analysis

Data were analyzed using IBM SPSS software package version 20.0. (Armonk, NY: IBM Corp). The Kolmogorov–Smirnov test was used to verify the normality of the distribution of variables. Comparisons

between groups for categorical variables were assessed using the Chi-square test [Fisher or Monte Carlo (MC)]. Student's *t*-test was used to compare two groups of normally distributed quantitative variables while the Mann Whitney test was used to compare two groups of non-normally distributed quantitative variables. For non-normally distributed quantitative variables more than two groups Kruskal Wallis test was used and followed by the Post Hoc test (Dunn's for multiple comparisons test). Pairwise comparison Spearman coefficient was used to correlate between quantitative variables. Univariate and multivariate logistic regression analyses were used to determine the related co-variables associated with poor response to induction therapy. The Receiver operating characteristic curve (ROC) was used to determine the diagnostic characteristics of the marker. The optimal cutoff value was assessed via the Youden index. The significance of obtained results was judged at the ≤ 0.05 level.²³

Results

This prospective study was conducted on eighty children newly diagnosed with B-ALL. They were 50 boys (62.5%) and 30 girls (37.5%) with a male to female ratio of 1.6:1, their ages ranged from 1 to 15 years with a median value 7.3 years.

At diagnosis, the mean and standard deviation (SD) of the hemoglobin (Hb) level was 7.9 ± 1.8 (gm/dl), blast percentage in peripheral blood (P.B) 38 ± 23.3 (%), and ESR 82 ± 15.3 (m/h). The median and range for the TLC was 20 (1–110) $\times 10^3$ /cmm, platelets count 65 (10–260) $\times 10^3$ /cmm, blast percentage in bone marrow 82.5 (50–98) %, LDH level 822 (340–3200) IU/L.

As regard immunophenotyping results, CD19 was expressed in all cases 80/80 (100%) of B-ALL, CD10 in 66/80 (82.5%) cases, CD34 in 71/80 (88.7%) cases, cytoplasmic μ in 12/80 (15%) cases, CD20 in 9/80 (11.2%) cases, with aberrant expression of CD33 and CD13 in 6/80 (7.5%) and 15/80 (18.7%) cases respectively. Based on these data, patients were classified according to EGIL classification as follows: pro-B-ALL (5 cases), common B-ALL (63 cases), pre-B-ALL (3 cases), and mature B-ALL (9 cases).

Positive expression of CD105 on BM samples was detected in 33/80 (41.2%) patients, and it ranged from 8–90%, 2–8 SFIs with a median value of 36% and 3 SFIs respectively, while negative expression was detected in 47/80 (58.8%) patients, and ranged from 0.5–5%,

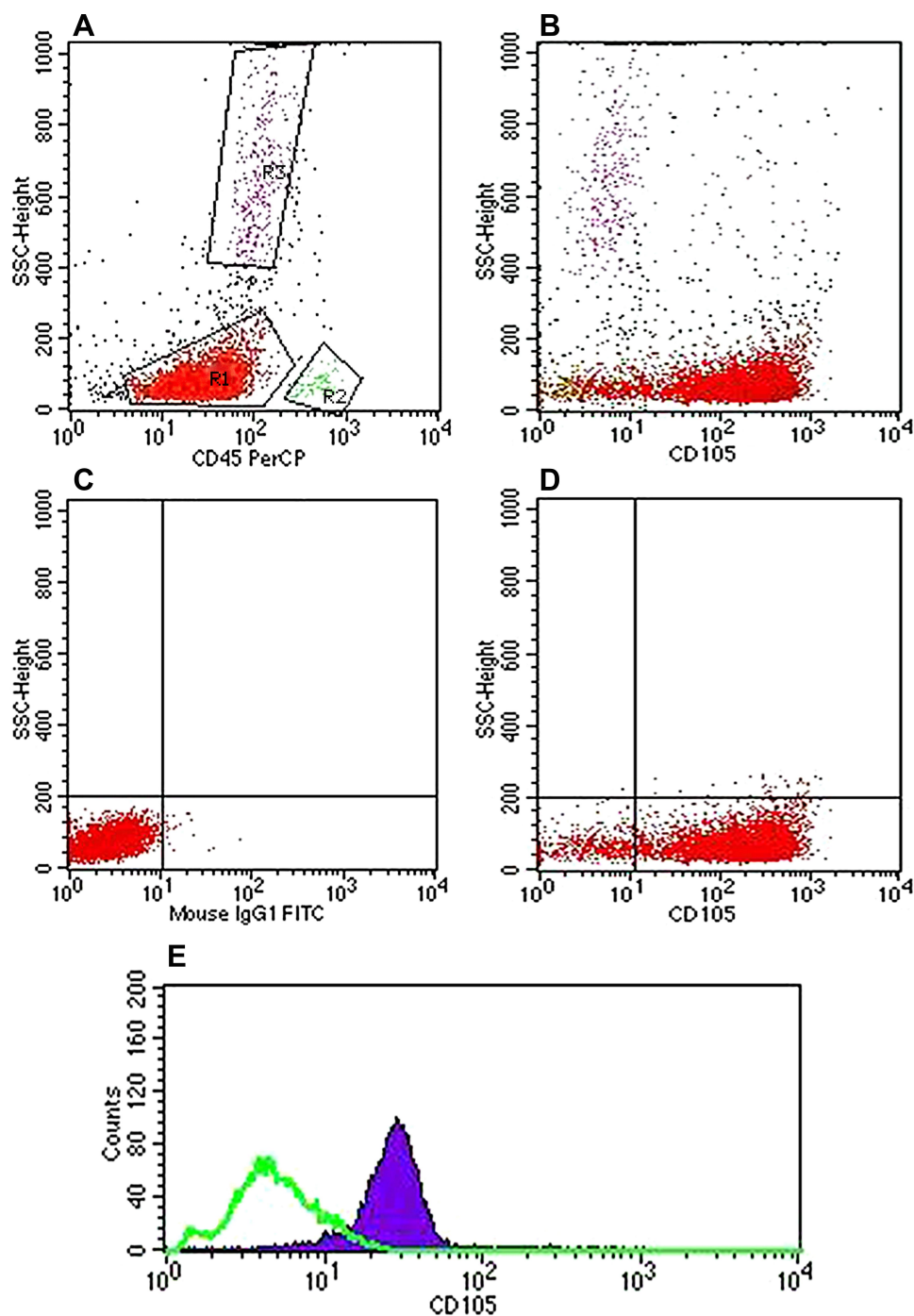


Figure 1 Flow cytometry analysis of CD105 positive case. **(A)** Dot plot showed gating using CD45/SSC strategy, blast cells (R1), normal lymphocytes (R2) and granulocytes (R3). **(B)** Scatter dot plot showing blast cells positive for CD105 expression in combination with normal lymphocytes and granulocytes negative for CD105 expression. **(C)** Dot plot showing the mouse IgG1 isotypic negative control. **(D)** Dot plot showing positive CD105 expression on gated blast cells. **(E)** Histogram showing positive CD105 (solid violet curve) versus negative control (green colored curve).

0.5–1.4 SFIs with a median value of 3% and 1 SFIs respectively.

CD105 expression in B-ALL subtypes was positive in (3/5) Pro B-ALL, (29/63) common B-ALL, (1/3) Pre B-ALL, while it was absent (0/9) in all cases of “Burkitt-like” mature B-ALL.

We found variations between CD105 percentage, SFIs and the patients’ risk stratification, with statistically significant difference between the standard-risk and the very high-risk groups (P -value=0.045, and 0.035 respectively) as shown in (Table 1). No statistically significant difference was observed between the CD105 positive and the

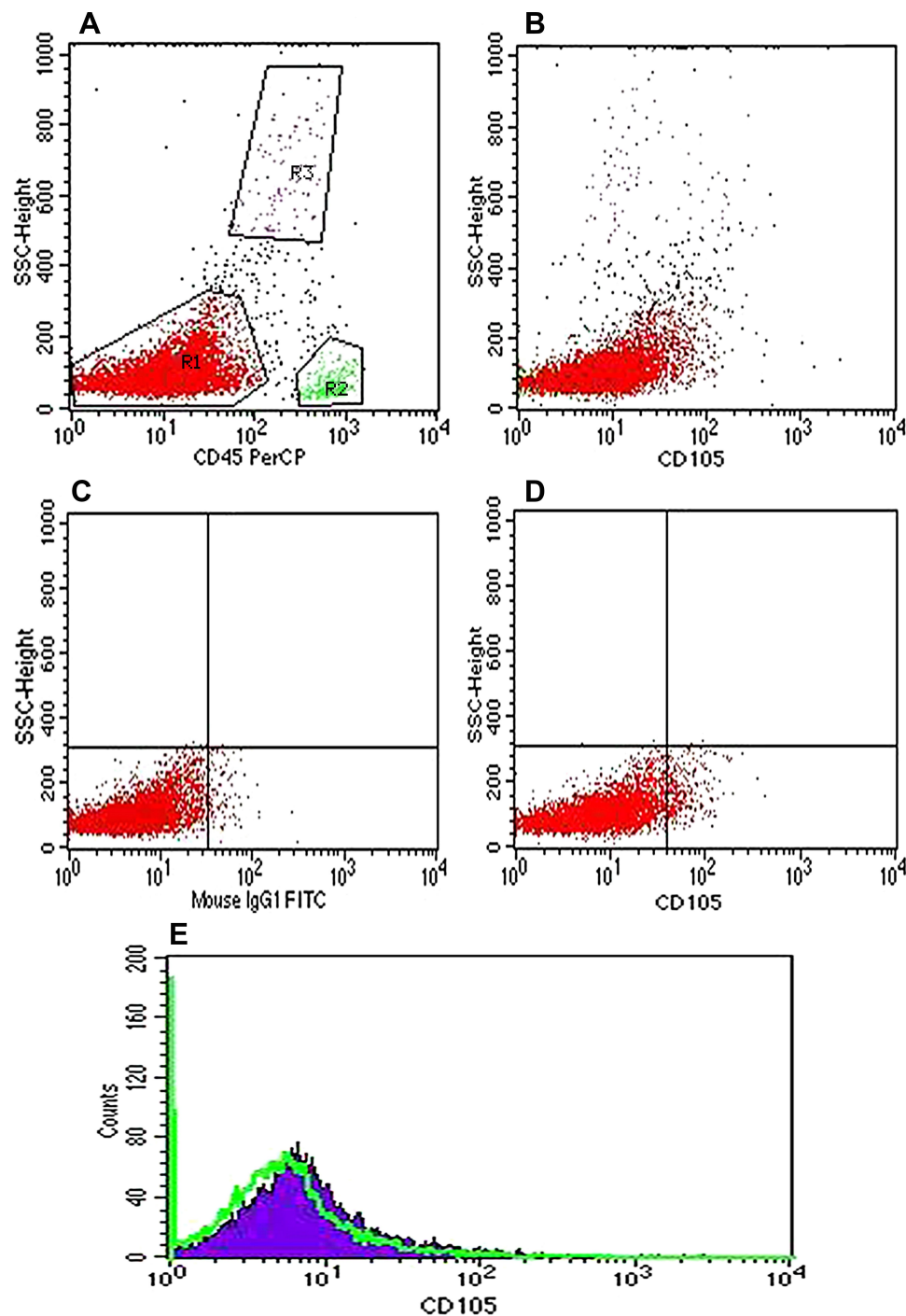


Figure 2 Flow cytometry of analysis of CD105 negative case. **(A)** Dot plot showed gating using CD45/SSC strategy, blast cells (R1), normal lymphocytes (R2) and granulocytes (R3). **(B)** Scatter dot plot showing blast cells, normal lymphocytes and granulocytes all negative for CD105 expression. **(C)** Dot plot showing the mouse IgG1 isotypic negative control. **(D)** Dot plot showing negative CD105 expression on gated blast cells. **(E)** Histogram showing negative CD105 (solid violet curve) versus negative control (green colored curve).

negative groups as regard clinical and laboratory parameters (P -value > 0.05) (Table 2).

CD105 expression had a weak positive correlation (<0.3) with age and TLC, LDH, and a weak negative correlation (<0.3) with CD20 as presented in (Table 3).

At follow up, 15 out of 80 patients were excluded from the statistical analysis (9 patients chose to complete induction at other centers, 3 of them were among the mature B-ALL group, the other 6 mature B-ALL patients were intentionally excluded as they received a different treatment protocol).

Table 1 The Pattern and Range of CD105 Expression in B-ALL Cases as Regards Risk Stratification

	Standard Risk (54 Patients)	High Risk (17 Patients)	Very-High (9 Patients)	P-value	Post Hoc Test
CD105 SFIs Median (Min. – Max.)	1.2 (0.5–6)	1.3 (0.5–6)	6 (1–8)	0.035*	P ₁ :0.239, P ₂ :0.013*, P ₃ :0.168
CD105% Median (Min. – Max.)	4 (0.5–60)	4 (2–60)	50 (1–90)	0.045*	P ₁ :0.169, P ₂ :0.022*, P ₃ :0.282

Notes: *Significant at $P \leq 0.05$. P₁: p-value for comparing between standard and high risk. P₂: p-value for the association between standard and very high. P₃: p-value for the association between high risk and very high.

Abbreviation: SFIs, specific fluorescence indices.

The remaining 65 cases were followed till the end of induction. Remission was achieved by 51/65 (78.4%) and 15 had CD105 positive expression. 10/65 cases (15.4%) were refractory to the first induction cycle, one cases; 36 of them had a negative expression of CD105 of them was CD105 negative while and 9 were CD105

Table 2 The Impact of CD105 Expression Pattern on B-ALL Patient's Characteristics

Parameters	CD105 Negative Group (47 Patients)	CD105 Positive Group (33 Patients)	P-value
Age (years) Median (Min. – Max.)	6.5 (1.5–14)	8.5 (1–15)	0.109
Hemoglobin (gm/dl) Mean \pm SD.	8.1 \pm 1.6	7.7 \pm 2	0.394
Total leucocytic count ($\times 10^3$/cmm) Median (Min. – Max.)	18 (1–110)	30 (1.7–96)	0.052
Platelets count ($\times 10^3$/cmm) Median (Min. – Max.)	57 (10–260)	70 (15–185)	0.384
Peripheral blood blasts (%) Mean \pm SD.	36.2 \pm 20.8	40.6 \pm 26.7	0.324
Bone marrow blasts (%) Median (Min. – Max.)	80 (50–98)	90 (59–98)	0.634
Lactate dehydrogenase (IU/L) Median (Min. – Max.)	787 (340–2300)	900 (444–3200)	0.095
Erythrocyte sedimentation rate (m/h) Mean \pm SD.	82.6 \pm 15.4	81.1 \pm 15.3	0.667
Sex Male Female	32 (68.1%) 15 (31.9%)	18 (54.5%) 15 (45.5%)	$\chi^2_{p=0.218}$
Central nervous system infiltration Absent Present	45 (95.7%) 2 (4.3%)	30 (90.9%) 3 (9.1%)	$\chi^2_{p=0.644}$
Philadelphia chromosome Negative Positive	46 (97.9%) 1 (2.1%)	28 (84.8%) 5 (15.2%)	$\chi^2_{p=0.077}$
Testicular infiltration Present Absent	2 (4.3%) 45 (95.7%)	4 (12.1%) 29 (87.9%)	$\chi^2_{p=0.224}$

Abbreviation: FE, Fisher exact.

Table 3 Correlation Between CD105 SFIs and CD105% with Different Parameters

	CD105 SFIs		CD105%	
	r_s	P-value	r_s	P-value
Age (years)	0.298	0.007*	0.293	0.008*
Hemoglobin (gm/dl)	-0.151	0.181	-0.105	0.354
Total leucocytic count ($\times 10^3$ /cmm)	0.261	0.019*	0.230	0.040*
Platelets count ($\times 10^3$ /cmm)	-0.039	0.731	-0.025	0.824
Peripheral blood blasts (%)	0.059	0.606	0.081	0.473
Bone marrow blasts (%)	0.113	0.320	0.189	0.092
Lactate dehydrogenase (IU/L)	0.236	0.035*	0.199	0.077
Erythrocyte sedimentation rate (m/h)	0.018	0.871	-0.036	0.754
CD19 (%)	0.041	0.721	0.018	0.874
CD20 (%)	-0.232	0.038*	-0.179	0.113

Notes: *Significant at $P \leq 0.05$. r_s : Spearman coefficient.

Abbreviation: SFIs, specific fluorescence indices.

positive expression. Four children died during the induction stage, one of them was CD105 negative and the other three were CD105 positive. Deaths were due to be sepsis with severe neutropenia in three children and intracranial hemorrhage with severe thrombocytopenia in one child.

The difference between the CD105 positive and the negative group regarding the response to therapy was highly significant (χ^2 14.383*^{MC}P-value <0.001).

To establish the effect of CD105 expression on response to induction therapy, logistic regression analyses were performed to detect the related covariables that can be associated with poor response to induction therapy. In univariate analysis, Ph chromosome positivity, and CD105 positivity were significantly associated with poor response

to therapy (P-value ≤ 0.05). In multivariate analysis, CD105 positivity remained the only significant independent factor associated with poor response to induction therapy (P-value ≤ 0.05). (Table 4).

As CD105 positivity was significantly correlated to poor response to induction therapy, we used the Receiver-operating characteristic (ROC) curve analysis to establish the most suitable cutoff value after exclusion of mature B-ALL cases, missed cases, and deaths. ROC curve analysis showed that CD105 expression >35% and SFIs >2.5% is highly significant to differentiate between good and poor responders to induction therapy with high diagnostic efficacy (AUROC 0.948 and 0.928 respectively) (Table 5) and (Figure 3).

Table 4 Univariate and Multivariate Analysis for the Parameters Affecting Response to Induction Therapy

	Univariate		Multivariate	
	P-value	OR (95% C.I)	P-value	OR (95% C.I)
Age (>8 versus. ≤ 8 years)	0.232	2.325 (0.582–9.283)		
Sex (female versus. male)	0.213	2.400 (0.605–9.522)		
Hemoglobin (>7.8 gm/dl versus. ≤ 7.8 gm/dl)	0.195	0.381 (0.088–1.640)		
Total leucocytic count (> 20×10^3 /cmm versus. $\leq 20 \times 10^3$ /cmm)	0.280	2.143 (0.538–8.540)		
Platelets count (> 65×10^3 /cmm versus. $\leq 65 \times 10^3$ /cmm)	0.865	1.125 (0.290–4.366)		
Peripheral blood blasts (>40% versus. $\leq 40\%$)	0.385	1.833 (0.468–7.187)		
Bone marrow blasts (>89% versus. $\leq 89\%$)	0.161	2.841 (0.659–12.239)		
Lactate dehydrogenase (>856 IU/L versus. ≤ 856 IU/L)	0.998	–		
Erythrocyte sedimentation rate (>85 m/h versus. ≤ 85 m/h)	0.683	0.750 (0.189–2.980)		
Central nervous system infiltration (present versus. absent)	0.635	1.778 (0.166–19.065)		
Philadelphia chromosome (positive versus. negative)	0.049*	12.500 (1.012–154.397)	0.110	14.820 (0.54–405.82)
CD105 (positive versus negative)	0.005*	21.600 (2.511–185.80)	0.007*	22.807 (2.34–221.84)

Notes: *Significant at $P \leq 0.05$. The number between brackets in columns I represent cutoff used was the median of the cases included.

Abbreviations: OR, odds ratio; C.I, confidence interval.

Table 5 Performance (AUC, Sensitivity, Specificity) for CD105 SFIs and CD 105% to Discriminate Between Poor Responder (n=10) and Good Responder (n=51)

	AUC	P-value	95% C.I	Cutoff	Sensitivity	Specificity	PPV	NPV
CD105 SFIs	0.928	<0.001*	0.800–1.057	>2.5	90.0	92.16	69.2	97.9
CD105%	0.948	<0.001*	0.867–1.029	>35	90.0	94.12	75.0	98.0

Note: *Significant at $P \leq 0.05$.

Abbreviations: AUC, area under the curve; C.I, confidence interval; NPV, negative predictive value; PPV, positive predictive value.

Discussion

The TGF- β co-receptor CD105 plays a major role in fetal, adult, and malignant angiogenesis. Previous studies reported the CD105 expression on tumor vessels of a variety of neoplasms.⁸ Although, CD105 is now well known to be expressed on malignant cells in various hematopoietic malignancies, data on its prognostic relevance in especially leukemias are still not adequate.¹⁹

CD105 positivity in the current study was present in 41.2% of cases diagnosed with B-ALL, this is lower than the values presented by Cosimato et al¹⁶ who reported positive CD105 expression in 68.4% of B-ALL cases, this difference can be attributed to the relatively small number of cases involved in our study. They also found that CD105 was absent in all cases of mature B-ALL, and expression levels were higher among the high and the very high-risk groups compared to the standard group, which is similar to our findings.

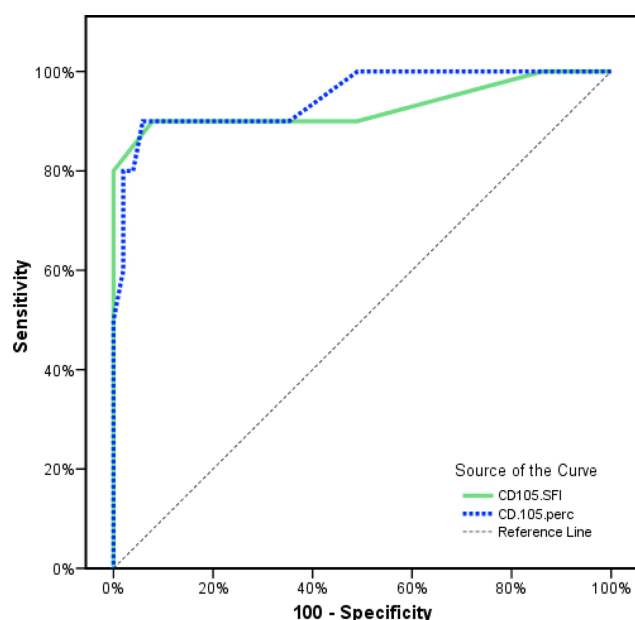


Figure 3 ROC curve for CD105 SFIs and CD105% to discriminate between poor and good responder.

Although the study of Poręba et al¹⁷ reported that no statistically significant correlation was present between CD105 expression and age, gender, LDH level, blast percentage in P.B or BM, we were able to detect a weak positive correlation with age, TLC, and LDH level and a weak negative correlation between CD105 SFIs and CD20 expression.

Patients with positive CD105 expression showed a higher rate of induction failure. Moreover, the multivariate analysis showed that CD105 positivity is an independent factor that associated with poor outcome to induction therapy. This highlights the importance of CD105 as a prognostic marker in B-ALL. This is comparable to what Kauer et al¹⁹ found in their study of CD105 in AML, where they correlated it to poor outcome and failure of response to chemotherapy, and they recommended the use of CD105 expression as a prognostic marker in AML, which can help optimize follow up and treatment decisions for AML patients. They attempted to explain the inferior outcome in CD105 positive AML patients by suggesting that CD105 contributes to dysregulation of TGF- β dependent and TGF- β independent signaling pathways and enhances angiogenesis which gives a better chance for survival of malignant cells with increasing the risk of minimal residual disease (MDR). Also, Xu et al²⁴ suggested that resistance to intensive therapy might be explained by the presence of hypoxia inducible factor 1 alpha induced multi-drug resistance transporters in CD 105 high AML blast cells.

To our knowledge, this is the first study that reported the impact of CD105 flow-cytometric expression on the response to induction therapy in B-ALL. Therefore, we established a cutoff value of > 35% and >2.5 SFIs as calculated by the ROC curve to identify patients who are at risk for induction failure that may require intensive therapy from the start to improve outcome.

Novel therapeutic targets for acute leukemia are urgently needed and successful treatment of acute leukemia remains a clinical challenge.²⁵ For ALL, although significant progress has been made in the last decade, in

cases of refractory or relapsed ALL, second-line chemotherapy has shown a poor effect, rarely resulting in long-term survival.²⁶ Thus, there is a critical need for new therapeutic options. Monoclonal antibodies are promising agents because they deliver their therapeutic effects with minimal toxicity.²⁷

CD105 is a promising target that can be used for tumor imaging and prognosis and it has therapeutic potential in patients with solid tumors and other neoplastic diseases with increased angiogenesis.¹⁸ Dourado and his colleagues,²¹ studied the use of monoclonal antibody TRC105 to prevent the engraftment of primary AML blasts and inhibit leukemia progression following disease establishment, but in B-ALL, TRC105 alone was ineffective due to the shedding of soluble CD105. However, in both B-ALL and AML, TRC105 synergized with reduced intensity myeloablation to inhibit leukemogenesis, indicating that TRC105 may represent a novel therapeutic option for B-ALL and AML. So, measurement of CD105 may be of great value not only to assess the prognosis but also in targeted therapy for B-ALL.

The results of our study support the important role of CD105 in leukemia progression and response to induction therapy which is one of the major contributors of B-ALL risk stratification.

The limitations of the present study included the relatively small number of patients included in this study, and the association between CD105 flow-cytometric expression and its soluble level was not investigated. Therefore, it is recommended to extend this research on a large patients cohort, investigate the coupled measurement of CD105 on blast cells and its soluble form in serum. Also, extend the follow-up duration to assess the impact of CD105 expression on the overall survival and disease-free survival, as well as studying the stability of the marker after therapy.

Conclusion

CD105 can be considered a potential marker for the prognosis of pediatric patients with B-ALL, as patients who showed expression higher than 35% and 2.5 SFIs were at higher risk for induction failure. So, CD105 may serve to optimize treatment decisions for B-ALL patients.

Data Sharing Statement

Data can be provided on request.

Ethics Approval and Consent to Participate

The study was approved by the local ethical committee of the Faculty of Medicine, Tanta University. Institutional Review Board (IRB) for human studies (Approval Code 34107/9/20). Our study conforms to provisions of the Declaration of Helsinki. Informed written consent was obtained from one of the parents or the legal guardian accompanying the child. All personal data were kept confidential and only scientific data are available for publication.

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Author Contributions

All authors made essential contributions to study design, analysis and interpretation of data; took part in drafting the article and revising it critically for important intellectual content; agreed to submit to the current journal, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

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Disclosure

The authors declare that they have no conflicts of interest.

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