

Copper Nanoparticles Induce Oxidative Stress via the Heme Oxygenase 1 Signaling Pathway in vitro Studies

This article was published in the following Dove Press journal:
International Journal of Nanomedicine

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Purpose: The toxicity of copper nanoparticle (CuNP) exposure in the ovaries has attracted attention recently, but the precise molecular mechanism involved requires further investigation. We investigated the cytotoxicity of CuNPs in ovarian granulosa cells and the protective effect of heme oxygenase 1 (HO-1) against CuNP-induced damage.

Methods: Human ovarian granulosa cells (COV434) were treated with CuNPs, and cytotoxicity was evaluated using Cell Counting Kit-8 and flow cytometry assays. Oxidative stress was identified using biochemical markers of oxidation and anti-oxidation. The protein levels of mitogen-activated protein kinase 14 (MAPK14), phospho-MAPK14, nuclear factor erythroid 2-related factor 2 (Nrf2), and HO-1 were measured by immunoblotting. Subsequently, for oxidative stress parameter detection, the cells were pre-treated with hemin to induce HO-1 expression prior to CuNP treatment.

Results: Exposure to CuNPs decreased cell viability and the mitochondrial membrane potential, increased the apoptosis rate, and induced oxidative stress. Furthermore, hemin pretreatment induced HO-1 expression in cells, which partially reduced the accumulation of reactive oxygen species induced by CuNPs and increased the levels of antioxidant enzymes.

Conclusion: CuNPs exert cytotoxic effects on human ovarian granulosa cells by inducing oxidative stress, and may induce HO-1 expression via the MAPK14-Nrf2 signaling pathway. Moreover, HO-1 protects against oxidative stress induced by CuNPs.

Keywords: copper nanoparticles, cytotoxicity, oxidative stress, heme oxygenase 1

Introduction

Nanomaterials are artificial microparticles with maximum diameters of 100 nm. As relatively common metal nanomaterials, copper nanoparticles (CuNPs) have excellent physical and chemical properties.¹ CuNPs are currently used for drug delivery and antibacterial applications, as animal feed additives, as emerging environmental catalysts, in bioimaging, and in other modern technologies.²⁻⁴ With the rapid development and wide application of CuNPs, they have spread widely in the environment, increasing the risk of exposure of the general population. The toxicity of CuNPs to various living organisms has caused public concern.^{5,6} Numerous studies have shown that CuNPs have serious toxicological effects and induce severe damage to the liver, spleen, and kidney in vivo.⁷⁻⁹ Moreover, the brain and reproductive system are also target organs of toxicity.¹⁰⁻¹³

Small size is reported to be one of the reasons for the cytotoxic effects of nanoparticles.^{7,14-16} The cytotoxicity of CuNPs mainly comes from their ability

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to generate reactive oxygen species (ROS),^{17,18} which induce cell death. Previous studies have mainly focused on the cytotoxicity of CuNPs in the liver,¹⁹ kidney,²⁰ and spleen.²¹ However, the ovary has recently been reported to be a target organ for CuNP toxicity. Our previous studies show that CuNPs can migrate to the ovaries of rats and accumulate in large quantities, thereby causing various types of damage such as ovarian injury, follicular atresia, oxidative stress, and apoptosis.¹¹ The proliferation and secretory functions of granulosa cells are essential for germ cell development in animals. Granulosa cells nourish oocytes through gap junctions and regulate oocytes through paracrine.²² The ovaries of rats exposed to CuNPs exhibit increased lipid peroxidation and antioxidant system destruction.¹¹ Therefore, we speculate that the cytotoxicity of CuNPs in ovarian granulosa cells may be related to oxidative stress.

Oxidative stress is caused by an imbalance in the generation and removal of oxygen free radicals, which causes the accumulation of ROS and reactive nitrogen species in tissues or cells. Excessive ROS production destroys proteins, nucleic acids, and lipids and overloads the antioxidant defenses, thus affecting their normal physiological and biochemical functions and leading to cell death.^{23,24} ROS production and oxidative stress are closely related to apoptosis induced by CuNPs.¹⁷ Heme oxygenase 1 (HO-1) is an adaptive protein that plays an important role in protecting against oxidation and apoptosis.²⁵ Recent studies have explored the molecular mechanism whereby HO-1 protects cells against oxidation during heat stress by reducing the accumulation of ROS and activating the antioxidant response.²⁶ Studies have shown that HO-1 induction during oxidative stress is regulated by the mitogen-activated protein kinase 14 (MAPK14)/nuclear factor erythroid 2-related factor 2 (Nrf2) signaling pathway.²⁷

In this study, we speculated that an increase in HO-1 expression levels may be involved in the anti-oxidative stress response in granulosa cells exposed to CuNPs. Cell proliferation, the apoptosis rate, changes in mitochondrial membrane potential, and oxidative stress were assessed to explore the cytotoxicity of CuNPs in granulosa cells. Oxidative stress markers, including ROS, superoxide dismutase 2 (SOD2), and glutathione peroxidase (GSH), were analyzed after HO-1 induction to explore the cytoprotective properties of HO-1.

Materials and Methods

Preparation of CuNPs

CuNPs (100 nm) were purchased from Da Yu Medical Instrument Co. Ltd. (Wuhan, Hubei, China). These CuNPs were characterized in our previous study.¹¹ Before use, the CuNPs were suspended in medium and dispersed by sonication for 25 min to prevent aggregation. Different concentrations of the CuNP suspension were obtained by diluting with complete medium.

Cell Culture and Pharmacological Experiments

The human ovarian granulosa tumor cell line COV434 was obtained from Procell (Wuhan, China). COV434 cells were cultured in a 37°C incubator with humidified air containing 5% CO₂. The complete medium consisted of Dulbecco's modified Eagle medium, 15% fetal bovine serum (Gibco, Carlsbad, CA, USA), and 1% penicillin and streptomycin. To induce HO-1 expression, COV434 cells were pre-treated with 20 µmol/L hemin (MedChemExpress, Monmouth Junction, NJ, USA), an HO-1-specific activator, for 48 h.

Cell Viability Detection

Cell viability was determined using the Cell Counting Kit-8 (CCK-8) assay (Dojindo, Tokyo, Japan). COV434 cells in 96-well plates were treated with different doses of CuNP suspension with a density of 1×10^5 cells per well. After incubation for 12 or 24 h, the CCK8 detection solution was added, and cells were incubated for another 2 h. The absorbance value at 450 nm was measured using a multimode microplate reader (Synergy™ HTX; BioTek, Winooski, VT, USA). Relative cell viability was calculated using the following equation: % cell viability = $\frac{\text{absorbance value}_{(\text{CuNPs group} - \text{blank group})}}{\text{absorbance value}_{(\text{control group} - \text{blank group})}}$.

Apoptosis Detection

An FITC Annexin V and PI Apoptosis Kit (ABP Biosciences, Beltsville, MD, USA) was used to detect apoptotic cells. Briefly, COV434 cells (2.5 mL/well) were evenly plated in 6-well plates and then incubated with 150 µg/mL CuNPs for 12 h. The control group was not treated with CuNPs. After washing three times with phosphate-buffered saline, COV434 cells were resuspended in binding buffer containing annexin V-FITC and propidium iodide, and then incubated in the dark at 20–25°C for flow cytometry.

Mitochondrial Membrane Potential Detection

Flow cytometry with JC-1 (Beyotime, Shanghai, China) was used to measure mitochondrial membrane potential (MMP). COV434 cells at a density of 1×10^6 cells per wells were evenly plated in 6-well plates and then incubated with 150 $\mu\text{g/mL}$ CuNPs for 12 h. Cells were then collected, resuspended in JC-1 staining reagent, and placed in an incubator for 20 min. After washing with JC-1 buffer, the cell mixture was analyzed by flow cytometry.

Oxidative Stress Parameters

SOD activity and ROS levels were measured using commercially available kits (Nanjing Jiancheng, Jiangsu, China). Total SOD activity was determined according to the manufacturer's instructions. Intracellular ROS accumulation was measured using 2,7-dichlorofluorescein diacetate (DCFH-DA), as ROS oxidizes DCFH-DA into the green fluorescent compound dichlorofluorescein. Intracellular fluorescence was detected using a multimode microplate reader. The resulting fluorescence value was proportional to the intracellular ROS level. The levels of GSH and malondialdehyde (MDA) production and catalase (CAT) activity were quantified using commercially available kits (Solarbio, Beijing, China). The treated cells were collected, disrupted by ultrasonication, and centrifuged at $8000 \times g$ at 4°C for 10 min. The resulting supernatant was collected for analysis. The absorbance value of each sample was determined, and the results were calculated according to the manufacturer's instructions.

Western Blotting

Cells were fully lysed using radioimmunoprecipitation assay buffer (Servicebio, Wuhan, China) containing 2% protease inhibitors and phosphatase inhibitors (Beyotime). Total protein samples were obtained after centrifugation. The extracted proteins were separated by 8–12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (Beyotime), and transferred to 0.45 μm polyvinylidene fluoride membranes (Merck KGaA, Darmstadt, Germany). The membranes were blocked in 5% skim milk, incubated with primary antibodies overnight, and then incubated with secondary antibodies (horseradish peroxidase-conjugated IgG; Abclonal, Wuhan, China). The following primary antibodies were used: anti-phospho-MAPK14, -Keap1 and -NQO1 (Abclonal); anti-MAPK14 (Santa Cruz, Dallas, TX, USA); anti-SOD2 (Cell

Signaling Technology, Beverly, MA, USA); anti-Nrf2, -HO-1, and -GAPDH (Proteintech, Wuhan, China). The immunoblot signals were detected using a Super-Sensitive Chemiluminescence Substrate Kit (Biosharp, Beijing, China). The protein bands were analyzed using a chemiluminescent gel imaging system, with Image Lab software (ChemiDoc XRS; Bio-Rad, Hercules, CA, USA).

Statistical Analysis

Statistical analysis was performed using a one-way ANOVA, followed by least significant difference post hoc tests. SPSS 23.0 software was used for these calculations (IBM, Chicago, IL, USA). Data are presented as the means \pm SD. All experiments were repeated at least three times, and $p < 0.05$ was considered statistically significant.

Results

Cytotoxic Effect of CuNPs on COV434 Cells

To examine the cytotoxicity of CuNPs in granulosa cells, COV434 cells were exposed to culture media containing different concentrations of CuNPs or without CuNPs (control). The results were compared with those of the control group. We found that COV434 cell viability decreased with an increasing treatment concentration and time (Figure 1A). After 12 h of CuNP exposure, the effective dose resulting in a 50% inhibition of COV434 cell growth was 150 $\mu\text{g/mL}$.

CuNPs Induce Apoptosis and Decrease MMP

COV434 cell apoptosis was detected using a flow cytometer. The results from the CuNP group were compared to those from the control group. We observed a higher apoptosis rate in the CuNP group (Figure 1B). MMP, as measured by flow cytometry after JC-1-staining, decreased significantly in the CuNP group (Figure 1C).

CuNPs Induce Oxidative Stress

To investigate whether oxidative stress was involved in CuNP-induced cytotoxicity, relevant detection kits were used to measure biomarkers of oxidative stress in COV434 cells before and after CuNP exposure. As shown in Figure 2, the levels of the oxidation products ROS and MDA increased significantly, whereas the levels of antioxidant biochemical markers, including SOD, GSH, and CAT, decreased in the CuNP group.

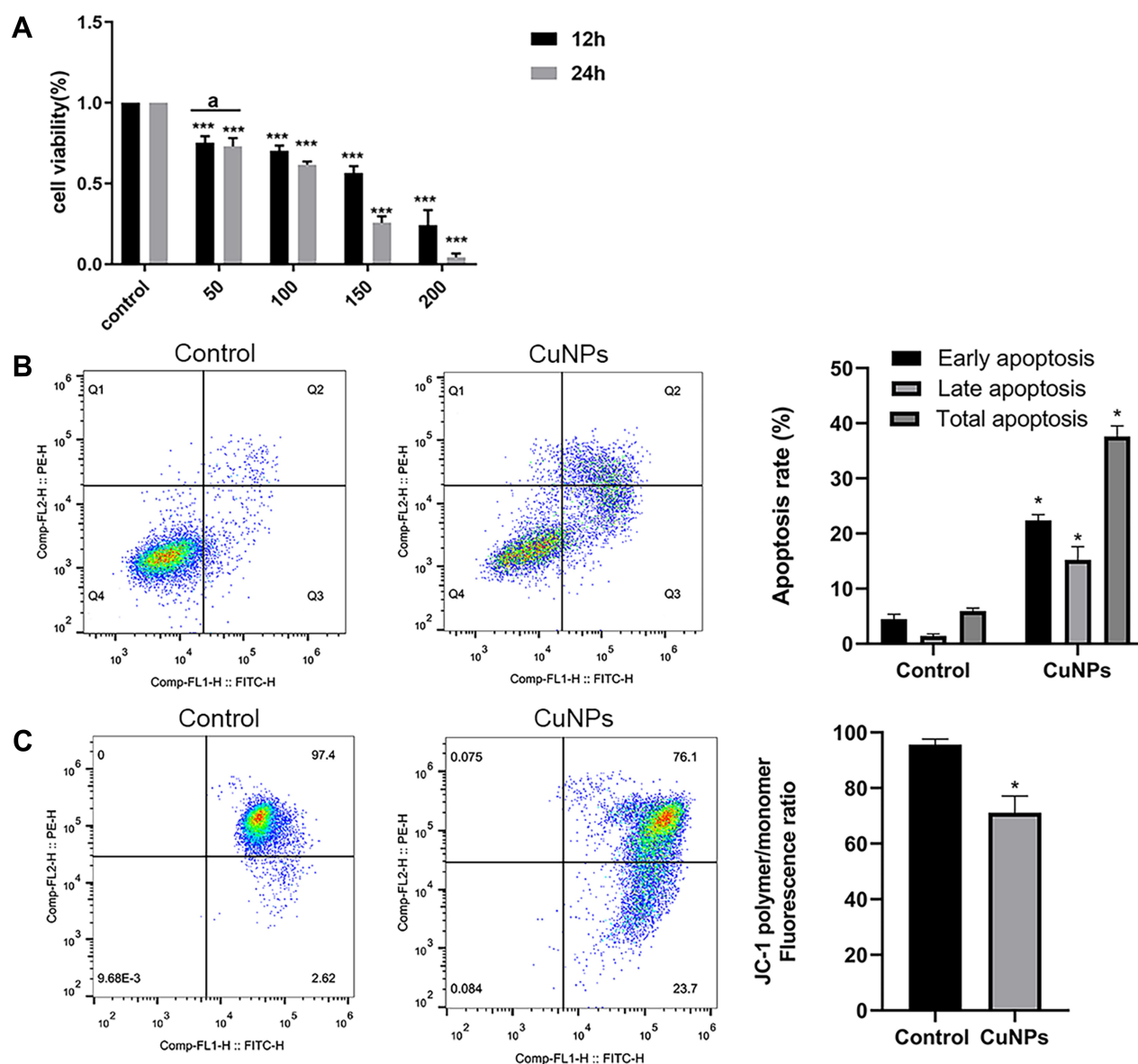


Figure 1 Cytotoxic effect of CuNP exposure in ovarian granulosa cells.

Notes: (A) The viability of COV434 cells after 12 and 24 h of treatment with different concentrations (0, 50, 100, 150, and 200 µg/mL) of CuNPs. (B) COV434 cell apoptosis before and after CuNP treatment. In the flow chart, Q3 indicates early apoptosis and Q2 indicates late apoptosis. (C) MMP in COV434 cells before and after CuNP treatment. ^aIndicates no difference between groups; **p* < 0.05, ****p* < 0.001.

Abbreviations: CuNPs, copper nanoparticles; MMP, mitochondrial membrane potential.

Effect of CuNPs on the Levels of MAPK14, Nrf2, NAD(P)H Quinone Dehydrogenase I (NQO1), and HO-1

To further explore the specific molecular events involved in the oxidative stress caused by CuNP exposure, members of the MAPK14 signaling pathway were analyzed by Western blotting. The expression of the unphosphorylated form of MAPK14 was constant,

whereas the phosphorylation of MAPK14 was significantly induced by CuNPs (Figure 3). Meanwhile, an increase in Nrf2 expression levels and a decrease in Keap1 expression levels were also detected. HO-1 and NQO1 are encoded by Nrf2 target genes and are well-characterized anti-oxidant proteins. CuNPs strongly induced HO-1 expression, but had no effect on NQO1 expression.

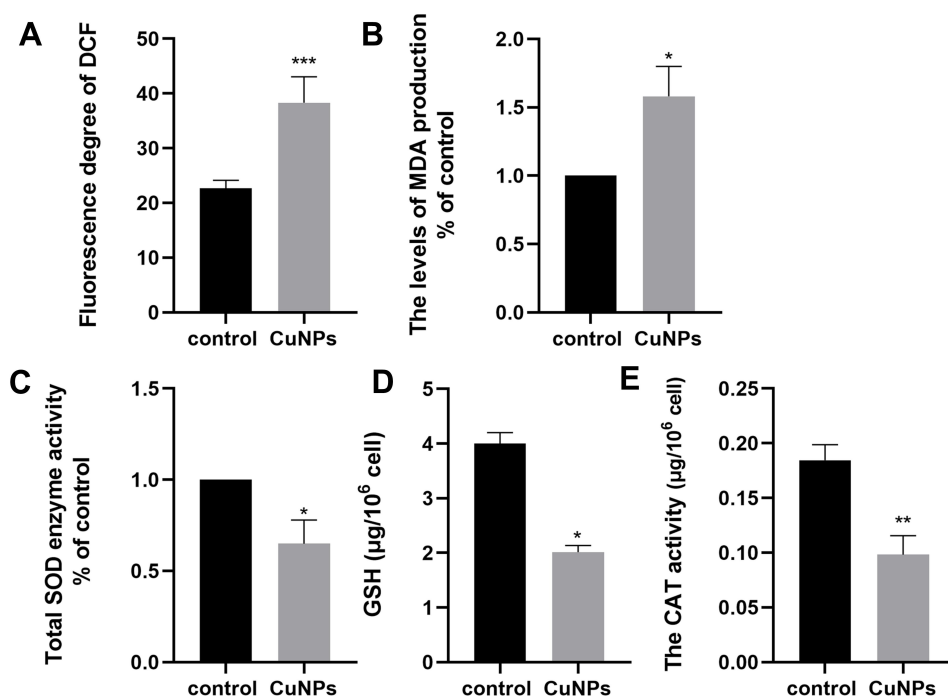


Figure 2 Effect of CuNP exposure on ROS (A), MDA (B), SOD (C), GSH (D), and CAT (E) in COV434 cells.

Notes: Compared to the control group: * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Abbreviations: ROS, reactive oxygen species; MDA, malonaldehyde; SOD, superoxide dismutase; GSH, glutathione peroxidase; CAT, catalase.

HO-1 Induction Attenuates ROS Generation and Increases Antioxidant Defenses in CuNP-Treated COV434 Cells

The effect of hemin on HO-1 expression was first determined by Western blotting (Figure 4A). Based on previous reports and the results of this study,^{26,28} we chose 20 μmol/L hemin as the optimal concentration for subsequent experiments. A significant decrease in ROS generation was detected in cells pretreated with hemin, compared to those treated with CuNPs only (Figure 4B). Meanwhile, HO-1 was induced and SOD2 protein levels were upregulated in the CuNPs+hemin group compared with the CuNPs group (Figure 4C). In addition, the GSH levels were increased in the CuNPs+hemin group (Figure 4D).

Discussion

Our previous experiments demonstrated that CuNPs may adversely impact rat ovaries.¹¹ Given limitations of animal studies, we further investigated the mechanism of CuNP-induced ovarian damage in vitro and explored possible protective mechanisms. Our results indicated that CuNPs have cytotoxic effects on COV434 cells in a dose- and time-dependent manner. We also observed an increase in the rate of apoptosis and a decrease in the MMP after

CuNP treatment. These results were consistent with those reported by Zhang et al, who showed that CuNPs can damage MMP and exert cytotoxicity in female mice via the mitochondrial apoptosis pathway.²⁹ Oxidative stress has been associated with the mitochondria-mediated apoptotic pathway and subsequent cell death.^{1,30}

Oxidative stress is a state of disturbance in the equilibrium between oxidative and antioxidative factors. It can be identified by analyzing various biochemical indicators that reflect the degree of destruction of the antioxidant defense system.³¹ To further understand the apoptosis of ovarian granulosa cells, we examined the intracellular levels of oxidative and antioxidative factors. CuNPs were found to significantly increase levels of the intracellular oxidation products, ROS and MDA, and to significantly destroy the antioxidant defenses, as indicated by reduced SOD and CAT activity and decreased GSH levels. These results suggest that the cytotoxicity of CuNPs to ovarian granulosa cells may be caused by oxidative stress.

MAPK signals are upstream of multiple stress-stimulation cascade events.^{32,33} Oxidative damage can activate MAPK14 phosphorylation,²¹ which in turn promotes the separation of Nrf2 from Keap1.²⁷ Nrf2 then translocates to the nucleus and binds to antioxidant response elements to activate various related genes, such

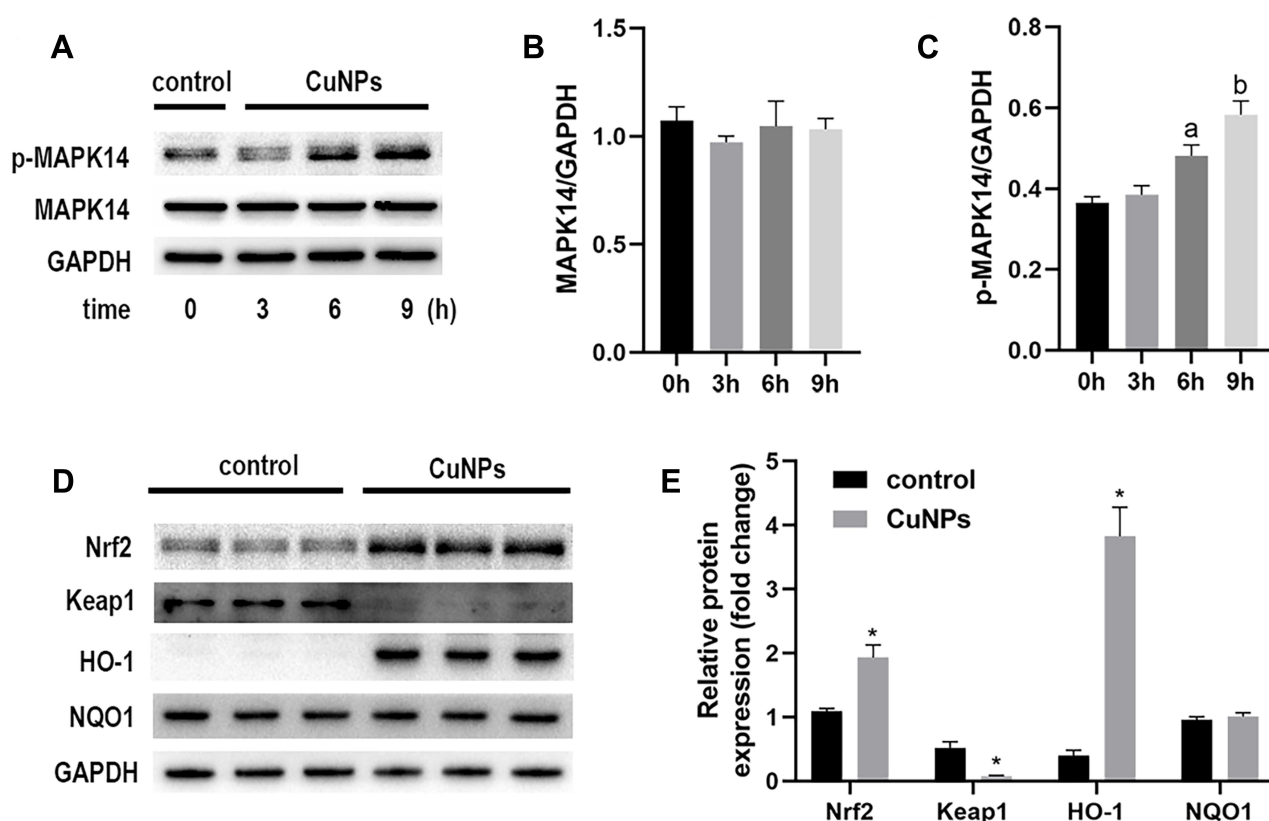


Figure 3 Levels of MAPK14, p-MAPK14, Nrf2, HO-1, and NQO1 proteins in COV434 cells exposed to CuNPs.

Notes: (A) Western blotting images of p-MAPK14 and MAPK14 proteins at different times of exposure to CuNPs. (B and C) Densitometric analysis of p-MAPK14 and MAPK14 protein bands. (D) Western blotting images of Nrf2, Keap1, HO-1, and NQO1 proteins. (E) Densitometric analysis of protein bands. * $p < 0.05$; ^{a,b}Indicate statistical significance, with the same letters indicating no difference between groups.

Abbreviations: MAPK14, mitogen-activated protein kinase 14; Nrf2, nuclear factor erythroid 2-related factor 2; HO-1, heme oxygenase 1; NQO1, NAD(P)H quinone dehydrogenase 1; CuNPs, copper nanoparticles.

as *HO-1* and *NQO1*.³⁴ The role of the MAPK14 signaling pathway and its related transcription factors in oxidative stress events caused by metal nanoparticles has attracted increasing attention. Some studies have demonstrated that the activation of Nrf2/HO-1 protects cells from AgNP- and SiO₂NP-induced oxidative stress. Moreover, TiO₂ nanoparticles and naked gold nanoparticles can activate MAPK14/Nrf2/HO-1 signaling.^{35–38} We also detected the phosphorylation of MAPK14 and upregulation of Nrf2/HO-1 in CuNP-treated COV434 cells. HO-1 and NQO1 are well-characterized antioxidant proteins that are regulated by activated Nrf2. CuNPs significantly induced HO-1 expression, whereas no changes in NQO1 protein levels were detected. These results may be due to the fact that we only tested protein levels. In addition, it is possible that an increase in NQO1 protein levels occurs later, or that activated Nrf2 does not regulate *NQO1* transcription in this model.

HO-1 protein is readily induced by heme, heat stress, hyperoxemia, heavy metals, ultraviolet rays, and other

stimuli.³⁹ HO-1 is currently recognized as a potential antioxidant that may be effective in protecting against oxidative stress.^{40,41} To explore the cytoprotective effects of HO-1 against oxidative stress due to CuNP-induced damage, hemin was used to induce HO-1 overexpression in vitro. We found that the induction of HO-1 increased antioxidant defenses and decreased ROS generation. These results indicate that HO-1 may protect granulosa cells against oxidative stress triggered by CuNPs. Therefore, it can be speculated that high levels of HO-1 expression play a stress-protection role in this model of CuNP-induced damage.

Notably, this study only examined the cytoprotective effect of high levels of HO-1 expression. However, there may be other protective pathways in this CuNP-induced damage model that remain to be determined in future studies. Further investigation is also required to determine whether enhanced rescue measures can reverse oxidative stress-induced injury and the optimal timing of the intervention to maximize protection from oxidative stress-induced damage.

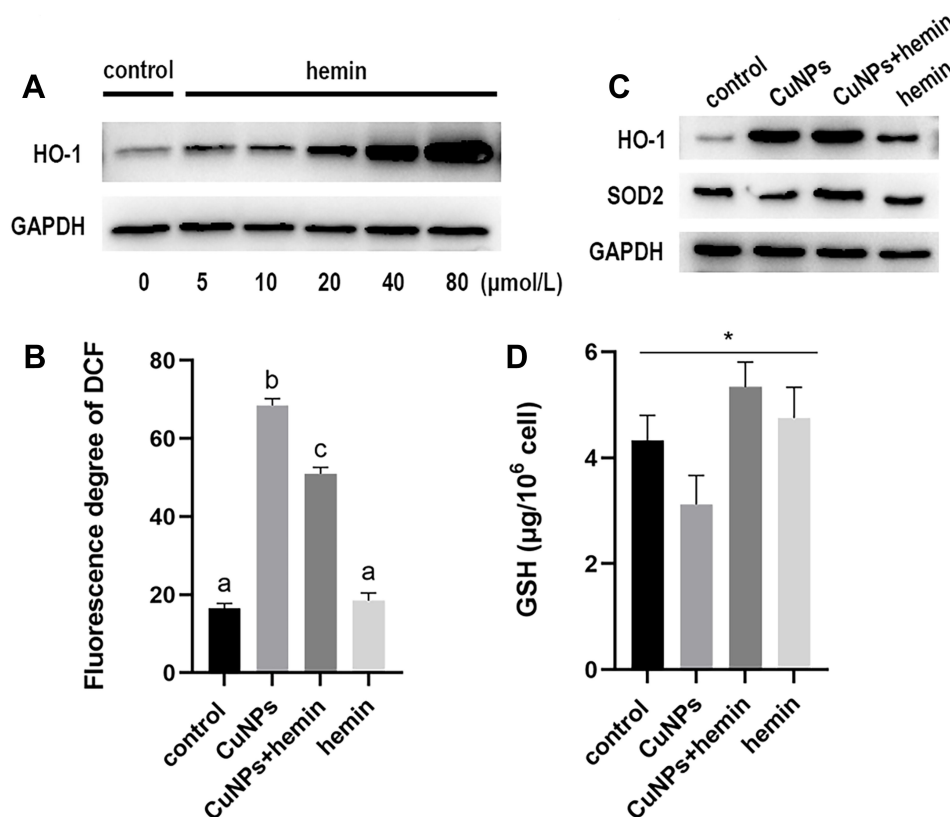


Figure 4 Effect of HO-1 induction in COV434 cells exposed to CuNPs.

Notes: (A) Western blotting images of HO-1 protein expression. COV434 cells were treated with different concentrations (μmol/L) of hemin for 48 h. (B) ROS levels in COV434 cells exposed to CuNPs before and after HO-1 overexpression. (C) Western blotting images of HO-1 and SOD2 proteins. (D) GSH levels in COV434 cells exposed to CuNPs before and after HO-1 overexpression. * $p < 0.05$; ^{a,b,c}Indicate statistical significance, with the same letters indicating no difference between groups.

Abbreviations: HO-1, heme oxygenase 1; CuNPs, copper nanoparticles; ROS, reactive oxygen species; SOD2, superoxide dismutase 2; GSH, glutathione peroxidase.

Conclusion

In summary, we have demonstrated that oxidative stress is one of the cytotoxic effects of CuNPs on ovarian granulosa cells, and that CuNPs may strongly induce HO-1 via the MAPK14-Nrf2 signaling pathway. The change of mitochondrial membrane potential, the increase of apoptosis rate and oxidative stress are the key mechanisms of cytotoxicity triggered by CuNPs, which can provide evidence for the study of CuNPs-induced damage. Moreover, HO-1 protects against CuNP-induced cytotoxicity by reducing oxidative stress. Therefore, the upregulation of HO-1 may help prevent and treat ovarian dysfunction caused by nanoparticles. We also realize the limitations of this study. Our results are only a preliminary exploration on the molecular mechanism and need to be further verified by in vivo models.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (No. 81971435)

Disclosure

The authors report no conflicts of interest in this work.

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