

Phytolacca dodecandra (Phytolaccaceae) Root Extract Exhibits Antioxidant and Hepatoprotective Activities in Mice with CCl₄-Induced Acute Liver Damage

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Background: The liver is a hub of metabolism and detoxification of substances. Since many redox reactions take place in the liver, it is prone to oxidative damage. Unlike conventional agents, botanicals act through several mechanisms in preventing oxidative damage. Among these *Phytolacca dodecandra* is the most commonly used agent in Ethiopian folk medicine.

Objective: To evaluate antioxidant and hepatoprotective activities of the 80% methanol extract of *P. dodecandra* root.

Methods: Male mice were divided into six groups and treated accordingly. Negative control was given 2% Tween 80, toxicant control administered with carbon tetrachloride (CCl₄), positive control treated with silymarin 100 mg/kg, and test groups were treated with 100, 200, and 400 mg/kg of the extract. Then, serum levels of alkaline phosphatase (ALP), alanine aminotransferase (ALT), aspartate aminotransferase (AST), gamma-glutamyl transferase (GGT), lactate dehydrogenase (LDH), total cholesterol (TC), triglyceride (TG), low-density lipoprotein (LDL) cholesterol, high-density lipoprotein (HDL) cholesterol, albumin, total protein, and bilirubin were determined. Determination of the change in body weight and liver weight, histopathologic examination of the liver, and in vitro and in vivo antioxidant assays were also carried out.

Results: The levels of ALP, ALT, AST, GGT, LDH, and bilirubin were significantly reduced, while albumin and total protein were significantly increased after treatment with *P. dodecandra* root extract at the doses of 200 and 400 mg/kg in CCl₄ intoxicated mice. Cholesterol metabolism and lipoprotein synthesis capabilities of the liver of mice were also returned to normal in the two doses. Besides, the 200 and 400 mg/kg doses were able to return the normal architecture and morphology of hepatocytes. Furthermore, the plant extract was found to scavenge free radicals in vitro and inhibit lipid peroxidation in vivo.

Conclusion: The results suggest that the 80% methanol extract of *P. dodecandra* root can be used for the management of liver disease.

Keywords: CCl₄, DPPH, hepatoprotective, mice, *Phytolacca dodecandra*, TBARS

Background

P. dodecandra L'Her also called “gopo berry or soapberry or African soapberry” in English,^{1,2} “Endod” in Amharic, “Handoodee” in Afaan Oromoo,³ and “Shibt” in Tigrigna,⁴ belongs to the family Phytolaccaceae. It is a climbing or scrambling dioecious, semi-succulent shrub, sometimes a liana with stems up to 10–20 m long and usually glabrous, trunks up to 35 cm in diameter, and with a taproot.² *P.*

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dodecandra is widely distributed across sub-Saharan Africa, Madagascar, Asia, and tropical America.^{2,5} All parts of *P. dodecandra* are widely used in Ethiopian traditional medicine. The leaves, roots, and fruits are used orally to treat anthrax, abdominal pain, itching,^{4,5} rabies, leech infestation, warts, herpes virus infection,⁶ liver disease, and helminthiasis.⁴ Several ethnobotanical studies showed that the roots,^{4,7–11} root barks,¹² leaves,¹³ and berries⁴ of *P. dodecandra* are used for the management of liver problems. *P. dodecandra* has several pharmacologically proven activities such as antirabies,¹⁴ abortifacient,¹⁵ analgesic and anti-inflammatory,¹⁶ antibacterial,¹⁷ antioxidant,¹⁸ anti-mycelial against *Histoplasma capsulatum*,¹⁹ molluscicidal,^{20–25} spermicidal,²⁶ antimalarial,²⁷ larvicidal against *Anopheles arabiensis*²⁸ and *Anopheles gambiae*,²⁹ and insecticidal against cabbage flea beetle.³⁰ Phytochemical analysis of the parts of *P. dodecandra* showed that it is found to contain terpenoids, phenolics,³¹ and saponins.^{32–34}

Materials and Methods

Chemicals and Reagents

Chemicals and reagents used were 1, 1-diphenyl-2-picrylhydrazyl (DPPH) (Chemos GmbH & Co. KG, Germany), distilled water and normal saline were obtained from EPHARM (Ethiopia), ammonium hydroxide, ascorbic acid, chloroform, ethanol, ferric chloride, glacial acetic acid, hydrochloric acid, isopropyl alcohol, methanol, n-butanol, and sulfuric acid were purchased from Sigma-Aldrich (Germany), acetic anhydride, bismuth nitrate, iodine, lead acetate, mercuric chloride, potassium hydroxide, potassium iodide, sodium hydroxide, thiobarbituric acid (TBA), and trichloroacetic acid (TCA) were purchased from Fisher Scientific (UK), eosin and hematoxylin were purchased from Santa Cruz Biotechnology, Inc. (USA), ether was obtained from Puyer BioPharma Ltd., (P.R.China), 10% formalin was obtained from Novochem Engineering (India), silymarin was purchased from Silybon-140, Micro Lab Limited (India), CCl₄, liquid paraffin, paraffin wax, and Tween 80 were purchased from Oxford Lab Fine Chem LLP (India), and xylene was obtained from scienTEST - bioKEMIX GmbH (Germany). All chemicals and reagents used were of analytical grade.

Plant Material

The roots of *P. dodecandra* were collected in May 2020 from Dessie, about 401 km north of Addis Ababa. Authentication and identification were done by a taxonomist and a voucher specimen (BGM01/2012) was

deposited. Then, roots were gently washed with tap water to remove dirt and cut into pieces manually. They were allowed to dry under shade for 2 weeks and finally pulverized into coarse powder using an electrical mill.

Experimental Animals

Both male and female Swiss albino mice aged 6–8 weeks and with a weight range of 25–35 g inbred in the animal house of the Department of Pharmacy, Wollo University, were used for the experiment. The animals were housed in polypropylene cages (6 mice per cage) under standard environmental conditions (25 ± 2°C, 55 ± 5% humidity, and 12 h/12 h light/dark cycle). The animals were allowed free access to tap water and laboratory pellet and acclimatized to laboratory conditions for 1 week before the commencement of the experiment.

Preparation of Plant Extract

Three hundred grams of the coarse powder of the roots of *P. dodecandra* were macerated for 3 days using 80% methanol as a solvent. Then, the fluid extract was filtered using muslin cloth and Whatman® grade 1 filter paper and the marc was re-extracted for the second and third times by adding another fresh 80% methanol. The extract was concentrated in a rotary evaporator (Buchi, Rotavapor R-210/215, Switzerland), freeze-dried in a lyophilizer (OPERON, OPR-FDU-5012, Korea), and stored in a chest freezer.

Acute Oral Toxicity Test

Female mice were used for the acute oral toxicity test of the 80% methanol extract of *P. dodecandra* roots following OECD-425 guidelines.³⁵ They fasted 3–4 h before and 1–2 h after administration of the 80% methanol extract of *P. dodecandra* root. First, a sighting study was performed to determine the starting dose. For this a female mouse was given a maximum dose (2000 mg/kg) of the 80% methanol extract as a single dose orally using oral gavage. Since no death was observed within 24 h, an additional four mice were used and dosed as mentioned above. Then, mice were observed continuously for 4 h at 30 min. intervals and then for 14 consecutive days with an interval of 24 h for the general signs and symptoms of toxicity (diarrhea, weight loss, tremor, lethargy, and paralysis), food and water intake, and mortality. Then, three dose levels were chosen for the extract: a middle dose; which is one-tenth of the maximum dose, a low dose; which is half of the middle dose, and a high dose that is twice the middle dose during the acute toxicity study.³⁵

Experimental Design

Mice were randomly assigned into 6 groups (three control and three test groups) comprising of six animals per group. Negative controls were given the vehicle used for reconstitution (2% Tween 80 in water (2%TW80)) for 6 consecutive days. Positive controls were treated with the standard drug, silymarin 100 mg/kg, for 6 consecutive days and CCl₄ (CCl₄ dissolved in an equal volume of liquid paraffin), 2 mL/kg, i.p.) on the 4th day. Toxicant controls were administered with 2%TW80 for 6 consecutive days and CCl₄ (2 mL/kg, i.p.) on the 4th day. Three test groups were treated with the 80% methanol extract of *P. dodecandra* root at the doses of 100, 200, and 400 mg/kg as PDRE100, PDRE200, and PDRE400, respectively, for 6 consecutive days. All the test groups were given CCl₄ (2 mL/kg, i.p.) on the 4th day. The route of administration for the test sample, the standard drug, and the vehicle was orally using oral gavage and the volume administered was 1 mL/100 g.^{36,37}

Hepatoprotective Activity

The hepatoprotective activity of 80% methanol extract of *P. dodecandra* root was determined following Sintayehu et al.³⁶ and Meharia et al.³⁷ The weight of each mouse was measured and recorded at the beginning (3–4 h of fasting) and the end of the experiment (on the 7th day) as initial and final body weight, respectively. After administration of respective treatment for 6 days as mentioned above in the “experimental design” section, mice were sacrificed under light ether anesthesia on the 7th day. Then, blood sample was collected from each mouse by cardiac puncture and transferred into heparinized sterile centrifuge tubes. The collected blood was centrifuged at 3000 rpm for 10 min and clear serum was separated. The separated serum was analyzed for the levels of biomarkers of liver function (levels of albumin, total protein, and total bilirubin) and chemistry (levels of ALP, ALT, AST, GGT, and LDH), and levels of TC, TG, HDL, and LDL using an automatic analyzer and commercial assay kits (Cobas Integra® 400 plus analyzer, Roche Diagnostics, Japan). Besides, the hepatoprotective activity of the 80% methanol extract of *P. dodecandra* root can be calculated as percentage protection which is; % protection = $\frac{(a-b)}{(a-c)} \times 100$ where a; is the mean value of the marker produced by hepatotoxin, b; is the mean value of the marker produced by toxin plus test material, and c; is the mean value produced by the vehicle control.^{37,38}

Liver Histopathological Examination

At the end of the experiment, all mice were sacrificed and their livers were harvested, weighed, and examined for microscopic pathology. The procedure was done as follows; individual liver samples were excised, washed with normal saline, and fixed in 10% formalin. Formalin-fixed tissue was washed in tap water. Then, it was dehydrated in serial ethanol and cleared in xylene. Finally, xylene cleared tissue was embedded in paraffin blocks. The paraffin blocks were sliced into sections of 5 µm in thickness and stained with hematoxylin and eosin. Histopathological evaluations were performed with a microscope (Olympus, Japan) and microscopic photographs were captured at 10x magnification.

Phytochemical Screening

Qualitative phytochemical investigation of the 80% methanol extract of *P. dodecandra* root was carried out to determine the presence of medicinally active secondary metabolites like alkaloids, cardiac glycosides, flavonoids, polyphenols, saponins, steroids, tannins, and terpenoids using standard methods.^{39–43}

Antioxidant Activity of *P. dodecandra* Root Extract

In vitro DPPH Radical Scavenging Activity

The in vitro DPPH free radical scavenging activity of the 80% methanol extract of *P. dodecandra* root was done following Sahu et al.⁴⁴ Two different stock solutions were prepared by dissolving 20 mg of DPPH in a liter of methanol and 250 mg of the 80% methanol extract of *P. dodecandra* root in the same liter of methanol. Then, the working solution was prepared by mixing 3.9 mL of DPPH solution with 0.1 mL of the sample solution. Blank solvent (methanol) was used to set the UV spectrophotometer (SHIMADZU UV-1800, Kyoto, Japan) at zero to cancel the solvent effect. DPPH solution was transferred into test tubes and the extract was added and followed by serial dilution. Then, the solution was incubated for 30 min at room temperature in a dark place and the absorbance was recorded at 517 nm using a UV spectrophotometer. Ascorbic acid was used as a reference standard and dissolved in methanol to make the stock solution with the same concentration.

The percent scavenging activity can be calculated as:

$$\% \text{ scavenging activity} = \frac{\text{Absorbance of control} - \text{absorbance of test sample}}{\text{Absorbance of control}} \times 100^{45}.$$

Analysis of Liver Lipid Peroxidation Thiobarbituric Acid Reactive Substance (TBARS) Assay

Thiobarbituric acid reactive substance (TBARS) assay was conducted according to the method of Oakes et al.⁴⁶ A molecule of malondialdehyde (MDA) forms an adduct with two TBA molecules to produce a red chromogen that absorbs ultraviolet-light at 532 nm. This assay was done by measuring the level of MDA spectrophotometrically. Primarily, 1 mL of the liver homogenate was mixed with 1 mL TCA (10% w/v) and centrifuged at 3000 rpm for 10 min. Then, 1 mL of TBA (0.67% w/v) was added to an equal volume of supernatant and incubated at 95°C for 60 min. The mixture was allowed to cool to room temperature in an ice bath for 10 min. One mL of n-butanol was added to precipitate the MDA-TBA complex. Then, n-butanol was removed and the precipitate was dissolved in water. Absorbance was recorded at 532 nm against an appropriate blank without the sample. The content of TBARS in the samples was calculated and the results were expressed as MDA equivalents in nmol/g of liver tissue.

Statistical Analysis

The results of the study are expressed as the mean \pm standard error of the mean (S.E.M). Statistical analysis of the data was performed with one-way analysis of variance (ANOVA) followed by Tukey post hoc multiple comparison test to determine the difference between the mean values of different groups. P values < 0.05 were considered statistically significant.

Ethical Consideration

Mice were handled according to the international animal care and welfare,⁴⁷ and the national institute of health guidelines for the care and use of laboratory animals.⁴⁸

Results

Acute Oral Toxicity Test

The acute oral toxicity test of the 80% methanol extract of *P. dodecandra* root indicated that it did not cause gross behavioral changes and mortality within 24 h and up to 14 days later. Therefore, the LD₅₀ was estimated to be greater than 2000 mg/kg in mice.

Hepatoprotective Activity

Effect of *P. dodecandra* Root Extract on Body Weight, Change in Body Weight, and Absolute and Relative Liver Weight of Mice

The effect of 80% methanol extract of *P. dodecandra* root on body weight, changes in body weight, and absolute and relative liver weight of mice is shown in Table 1.

PDRE100 treated mice did not show increment in the final body weight, whereas, PDRE200 and PDRE400 administered mice produced a significant increase in the final body weight ($p < 0.001$), as compared to the toxicant control (Table 1). PDRE200 and PDRE400 produced a significant decrement in the absolute and relative liver weight of mice ($p < 0.001$), as compared to the toxicant control. In contrast, the CCl₄-administered group resulted in significantly ($p < 0.001$) decreased final body weight of mice, whilst their absolute and relative liver weight significantly increased ($p < 0.001$) as compared to the negative control (Table 1).

Effect of *P. dodecandra* Root Extract on Liver Function of Mice

Table 2 presents the effect of *P. dodecandra* root extract on the synthesis and detoxification functions of the liver of mice. PDRE100 did not show significant alteration in all of the parameters shown in Table 2 below as compared to the toxicant control. On the other hand, PDRE200 and PDRE400 treated mice showed a significantly increased level of albumin ($p < 0.05$) and ($p < 0.001$), respectively, and total protein ($p < 0.01$), and a significantly decreased level of bilirubin ($p < 0.001$) as compared to the toxicant control (Table 2).

Effect of *P. dodecandra* Root Extract on Liver Chemistry of Mice

The 80% methanol extract of *P. dodecandra* root resulted in a significant decline in liver chemistry biomarkers of hepatocyte injury following hepatotoxicant administration (Table 3). Liver chemistry biomarkers of hepatocyte injury such as ALP, ALT, AST, GGT, and LDH significantly increased in CCl₄ treated mice by 187.7% ($p < 0.001$), 495.2% ($p < 0.001$), 555.6% ($p < 0.001$), 140.9% ($p < 0.001$), and 165.2% ($p < 0.001$), respectively (Table 3). In contrast, mice pre- and post-treated with 200 and 400 mg/kg of *P. dodecandra* root extract significantly decreased CCl₄ induced elevation in ALP by 83.4% and 96.0%, ALT by

Table 1 Effect of 80% Methanol Extract of *P. dodecandra* Root on Body Weight, Change in Body Weight, and Absolute and Relative Liver Weight of Mice

Group	Body Weight (g)		Change in Body Weight	Liver Weight (g)	Relative Liver Weight (%)
	Initial	Final			
2%TW80	27.54 ± 0.66	28.86 ± 0.74	1.32 ± 0.38	2.18 ± 0.09	7.61 ± 0.48
CCl ₄	28.42 ± 0.94	25.07 ± 0.90 ^{a####}	-3.35 ± 0.69 ^{a####}	4.26 ± 0.16 ^{a####}	17.00 ± 0.35 ^{a####}
Silymarin + CCl ₄	27.60 ± 0.71	29.92 ± 0.32 ^{b####}	2.32 ± 0.44 ^{b####}	2.20 ± 0.18 ^{b####}	7.35 ± 0.58 ^{b####}
PDRE100 + CCl ₄	27.50 ± 0.89	25.34 ± 1.16 ^{a####,c####,d###,e####}	-2.16 ± 0.56 ^{a####,c####,d###,e####}	4.02 ± 0.21 ^{a####,c####,d###,e####}	15.93 ± 0.68 ^{a####,c####,d###,e####}
PDRE200 + CCl ₄	28.20 ± 0.80	30.04 ± 0.83 ^{b####}	1.84 ± 0.17 ^{b####}	2.46 ± 0.19 ^{b####}	8.19 ± 0.53 ^{b####}
PDRE400 + CCl ₄	27.94 ± 0.96	30.26 ± 1.13 ^{b####}	2.32 ± 0.34 ^{b####}	2.24 ± 0.12 ^{b####}	7.46 ± 0.52 ^{b####}

Notes: Each value represents mean ± S.E.M; n=6; ^aAgainst 2%TW80; ^bAgainst CCl₄; ^cAgainst standard (silymarin); ^dAgainst PDRE200 + CCl₄; ^eAgainst PDRE400 + CCl₄; ^{###}p < 0.01, ^{####}p < 0.001.

Abbreviations: PDRE100, *Phytolacca dodecandra* root extract 100 mg/kg; PDRE200, *Phytolacca dodecandra* root extract 200 mg/kg; PDRE400, *Phytolacca dodecandra* root extract 400 mg/kg; Silymarin, silymarin 100 mg/kg; CCl₄, carbon tetrachloride 2 mL/kg; 2%TW80, 2% Tween 80 in water.

Table 2 Effect of *P. dodecandra* Root Extract on the Synthetic and Detoxifying Functions of Liver of Mice

Group	Albumin (mg/dL)	Total Protein (g/dL)	Bilirubin (mg/dL)
2%TW80	3.21 ± 0.35	5.62 ± 0.38	1.02 ± 0.42
CCl ₄	1.49 ± 0.20 ^{a###}	3.78 ± 0.43 ^{a###}	3.11 ± 0.51 ^{a####}
Silymarin + CCl ₄	3.14 ± 0.31 ^{b####}	5.40 ± 0.52 ^{b###}	1.07 ± 0.44 ^{b####}
PDRE100 + CCl ₄	1.88 ± 0.19 ^{a####,c####,d###,e###}	4.01 ± 0.61 ^{a###,c####,d###,e###}	2.57 ± 0.25 ^{a####,c####,d####,e####}
PDRE200 + CCl ₄	2.73 ± 0.32 ^{b#}	5.18 ± 0.17 ^{b###}	1.14 ± 0.39 ^{b####}
PDRE400 + CCl ₄	3.08 ± 0.15 ^{b####}	5.35 ± 0.35 ^{b###}	1.10 ± 0.13 ^{b####}

Notes: Each value represents mean ± S.E.M; n=6; ^aAgainst 2%TW80; ^bAgainst CCl₄; ^cAgainst standard (silymarin); ^dAgainst PDRE200 + CCl₄; ^eAgainst PDRE400 + CCl₄; [#]p < 0.05, ^{###}p < 0.01, ^{####}p < 0.001.

Abbreviations: PDRE100, *Phytolacca dodecandra* root extract 100 mg/kg; PDRE200, *Phytolacca dodecandra* root extract 200 mg/kg; PDRE400, *Phytolacca dodecandra* root extract 400 mg/kg; Silymarin, silymarin 100 mg/kg; CCl₄, carbon tetrachloride 2 mL/kg; 2%TW80, 2% Tween 80 in water.

88.7% and 93.4%, AST by 85.6% and 95.4%, GGT by 66.6% and 76.9%, and LDH by 89.4% and 91.8%, respectively (Table 3). As it is presented in Table 3, PDRE400 demonstrated nearly equal hepatoprotective activity to the standard drug (silymarin).

Effect of *P. dodecandra* Root Extract on Lipid Profile of Mice

The effect of 80% methanol extract of *P. dodecandra* root on TC, TG, HDL, and LDL is shown in Table 4. As it is presented in Table 4 mice treated with 100 mg/kg of *P. dodecandra* root extract did not show a significant alteration in the levels of TC, TG, and LDL as compared to the toxicant control. Whereas, mice treated with 200 and 400 mg/kg produced significantly increased levels of TC, TG, and LDL (p<0.001) as compared to the CCl₄ treated group. Similarly, the standard drug significantly increased the levels of TC, TG, and LDL (p<0.001) as compared to the toxicant control. On the contrary, neither the plant extract nor the standard drug significantly affected the HDL level as compared to the toxicant control (Table 4).

Effect of *P. dodecandra* Root Extract on Histology of Liver of Mice

The effect of *P. dodecandra* root extract on the histology of the liver of mice is presented in Figure 1. In this study, histological observations provided supportive evidence for biochemical analysis. As shown in Figure 1A below, mice treated with 2%TW80 revealed normal hepatic cells with intact cytoplasm, prominent nucleus and nucleolus, and well defined central vein. In stark contrast, the toxicant group exhibited cells with prominent infiltration, necrosis, cytoplasmic vacuolization, and dilation of blood vessels (Figure 1B). Administration of silymarin however returned inflamed and necrotized cells to their normal architecture (Figure 1C). In a similar manner to silymarin, the 80% methanol extract of *P. dodecandra* root protected the mice liver from CCl₄ mediated injury as it is observed in Figure 1 below. *P. dodecandra* root extract normalized the defects observed in the histopathology of the liver of mice treated with CCl₄ nearly to the level of the negative control group in its 200 mg/kg (Figure 1E) and 400 mg/kg (Figure 1F) doses. On the other hand, the liver section of

Table 3 Effect of *P. dodecandra* Root Extract on Liver Chemistry of Mice

Group	ALP (IU/L)	ALT (IU/L)	AST (IU/L)	GGT (IU/L)	LDH (IU/L)
2%TW80	94.61 ± 4.02	41.54 ± 3.12	32.86 ± 2.76	56.14 ± 3.69	214.55 ± 6.32
CCl ₄	272.18 ± 7.86 ^{a###}	247.26 ± 9.32 ^{a###}	215.42 ± 8.68 ^{a###}	135.23 ± 4.45 ^{a###}	568.92 ± 12.30 ^{a###}
Silymarin + CCl ₄	99.41 ± 4.09(97.3) ^{b####}	48.74 ± 4.26(96.5) ^{b####}	36.33 ± 3.27(98.1) ^{b####}	66.40 ± 2.97(87.0) ^{b####}	237.21 ± 8.42(93.6) ^{b####}
PDRE100 + CCl ₄	251.23 ± 4.28(1.8) ^{a####,c###,d####,e####}	226.28 ± 5.04(10.2) ^{a####,c###,d####,e####}	192.78 ± 6.84(12.4) ^{a####,c###,d####,e####}	127.81 ± 5.22(9.4) ^{a####,c###,d####,e####}	491.67 ± 10.06(21.8) ^{a####,c###,d####,e####}
PDRE200 + CCl ₄	124.09 ± 5.21(83.4) ^{b####,c###}	64.79 ± 4.89(88.7) ^{b####,c###}	59.15 ± 5.74(85.6) ^{b####}	82.58 ± 3.87(66.6) ^{b####}	252.11 ± 9.78(89.4) ^{b####}
PDRE400 + CCl ₄	101.71 ± 3.96(96.0) ^{b####}	55.12 ± 4.51(93.4) ^{b####}	41.26 ± 3.44(95.4) ^{b####}	74.38 ± 4.16(76.9) ^{b####}	243.61 ± 8.74(91.8) ^{b####}

Notes: Each value represents mean ± S.E.M; n=6; ^aAgainst 2%TW80; ^bAgainst CCl₄; ^cAgainst standard (silymarin); ^dAgainst PDRE200 + CCl₄; ^eAgainst PDRE400 + CCl₄; ^{###}p < 0.05, ^{####}p < 0.001.

Abbreviations: PDRE100, *Phytolacca dodecandra* root extract 100 mg/kg; PDRE200, *Phytolacca dodecandra* root extract 200 mg/kg; PDRE400, *Phytolacca dodecandra* root extract 400 mg/kg; Silymarin, silymarin 100 mg/kg; CCl₄, carbon tetrachloride 2 mL/kg; 2%TW80, 2% Tween 80 in water.

the mouse treated with 100 mg/kg of *P. dodecandra* root extract revealed disarrangement of hepatocytes, well-observed hepatic lesions, infiltration of inflammatory cells, and necrosis (Figure 1D).

Phytochemical Screening

The 80% methanol extract of *P. dodecandra* root was examined for the composition of medicinally active secondary metabolites and it was found to be positive for polyphenols, saponins, tannins, and terpenoids (Table 5). On the other hand, alkaloids, anthraquinone glycosides, cardiac glycosides, flavonoids, and steroids were not detected in the plant extract.

Antioxidant Activity of *P. dodecandra* Root Extract

DPPH Radical Scavenging Activity

The antioxidant activity of the 80% methanol extract of *P. dodecandra* root was evaluated by measuring its free radical scavenging power using DPPH free radicals and the data are presented in Figure 2. It was observed that the 80% methanol extract of *P. dodecandra* root had a concentration-dependent free radical scavenging activity. The calculated IC₅₀ values for the extract and standard were 7.4 µg/mL and 6.2 µg/mL, respectively. In a similar manner, the maximum percentage inhibition of free radicals by the extract and the standard were 94% and 99%, respectively, at a concentration of 20 µg/mL (Figure 2).

Analysis of Liver Lipid Peroxidation TBARS Assay

Figure 3 below shows the effect of *P. dodecandra* root extract on the inhibition of lipid peroxidation reaction in CCl₄ induced hepatotoxicity.

A significantly increased level of MDA (14.64 ± 1.26 nmol/g of the liver) was obtained in the CCl₄ intoxicated group as compared to the negative control (4.82 ± 0.46 nmol/g of the liver) (p<0.001). But PDRE200 and PDRE400 showed a significant decrement in the levels of MDA 8.43 ± 1.24 and 6.24 ± 1.16 nmol/g of liver tissue (p<0.01) respectively, as compared to the toxicant control. The MDA level of mice treated with 100 mg/kg of the 80% methanol extract of *P. dodecandra* root is significantly higher than that of the standard (10.22 ± 1.35 nmol/g of liver vs 6.78 ± 1.14 nmol/g of the liver) (p<0.01) (Figure 3).

Table 4 Effect of 80% Methanol Extract of *P. dodecandra* Root on Lipid Profile of Mice

Group	TC (mg/dL)	TG (mg/dL)	HDL (mg/dL)	LDL (mg/dL)
2%TW80	95.42 ± 3.18	62.39 ± 2.41	33.71 ± 1.48	42.90 ± 3.04
CCl ₄	52.61 ± 6.42 ^{a####}	31.47 ± 5.36 ^{a####}	29.62 ± 2.06	26.46 ± 5.13 ^{a####}
Silymarin + CCl ₄	93.83 ± 3.81 ^{b####}	58.26 ± 2.98 ^{b####}	32.54 ± 1.66	40.28 ± 2.62 ^{b####}
PDRE100 + CCl ₄	58.94 ± 6.46 ^{a####,c####,d###,e####}	37.80 ± 5.29 ^{a####,c####,d###,e####}	30.19 ± 1.80	28.72 ± 3.76 ^{a####,c####,d###,e####}
PDRE200 + CCl ₄	84.27 ± 2.87 ^{b####}	54.54 ± 4.60 ^{b####}	32.08 ± 2.24	36.39 ± 2.11 ^{b####}
PDRE400 + CCl ₄	91.63 ± 4.22 ^{b####}	57.65 ± 3.21 ^{b####}	32.36 ± 1.98	39.85 ± 2.19 ^{b####}

Notes: Each value represents mean ± S.E.M; n=6; ^aAgainst 2%TW80; ^bAgainst CCl₄; ^cAgainst standard (silymarin); ^dAgainst PDRE200 + CCl₄; ^eAgainst PDRE400 + CCl₄; ^{###}p < 0.01, ^{####}p < 0.001.

Abbreviations: PDRE100, *Phytolacca dodecandra* root extract 100 mg/kg; PDRE200, *Phytolacca dodecandra* root extract 200 mg/kg; PDRE400, *Phytolacca dodecandra* root extract 400 mg/kg; Silymarin, silymarin 100 mg/kg; CCl₄, carbon tetrachloride 2 mL/kg; 2%TW80, 2% Tween 80 in water.

Discussion

The liver has a crucial role in life because it performs a vast array of biochemical and metabolic functions, including synthesis and elimination of numerous substances.⁴⁹ Since it is the first port of call for most nutrients absorbed across the gut wall and site of oxidation of most compounds, it is highly susceptible to oxidative stress and related damage.⁴⁹ So, it is highly imperative to demonstrate the hepatoprotective activity of the plant extract in the presence of hepatotoxicant that causes oxidative damage.

P. dodecandra is the most commonly used plant in Ethiopian folk medicine for the treatment of liver

disease.^{4,7–11} Its hepatoprotective activity was investigated pharmacologically by using a hepatotoxic agent, CCl₄. Since CCl₄ is a widely used hepatotoxicant in an experimental model of liver injury.^{36,37} The highly reactive metabolites of CCl₄ such as trichloromethyl (CCl₃·) and trichloromethyl peroxy (CCl₃O₂·) radicals are the main players in hepatocyte injury. They deactivate antioxidant enzymes and thereby alter the antioxidant state of hepatocytes.^{50,51}

As per the traditional claim, the 80% methanol extract of *P. dodecandra* root was administered to mice in three different doses pre- and post-dose of CCl₄. As it is

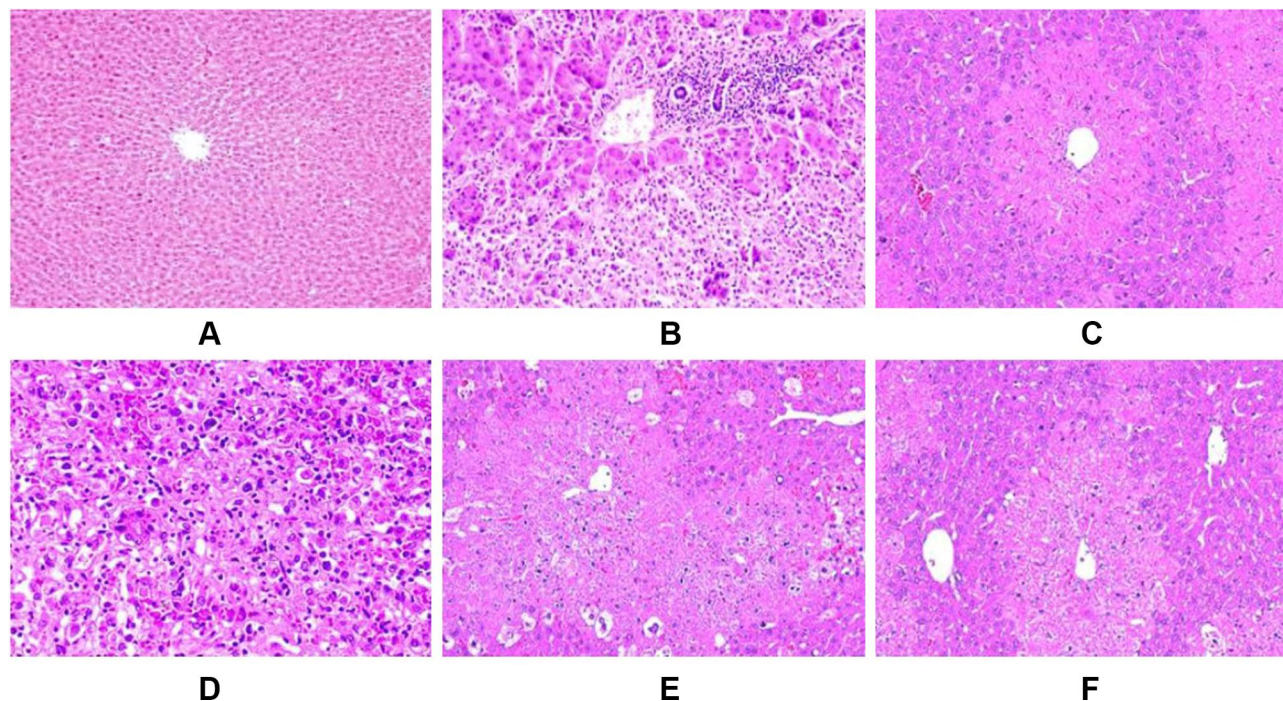


Figure 1 Microphotograph of hematoxylin and eosin (H & E)-stained sections of liver. Microscopic glass slides were prepared in 50μm intervals.

Notes: (A) negative control (received 2%TW80 only), (B) toxicant control (administered with CCl₄ only), (C) positive control (Silymarin + CCl₄), (D) PDRE100 + CCl₄, (E) PDRE200 + CCl₄, (F) PDRE400 + CCl₄.

Abbreviations: PDRE100, *Phytolacca dodecandra* root extract 100 mg/kg; PDRE200, *Phytolacca dodecandra* root extract 200 mg/kg; PDRE400, *Phytolacca dodecandra* root extract 400 mg/kg; Silymarin, silymarin 100 mg/kg; CCl₄, carbon tetrachloride 2 mL/kg; 2%TW80, 2% Tween 80 in water.

Table 5 Phytochemical Screening of 80% Methanol Extract of *P. dodecandra* Root

Phytochemical	Test Method	Observation	Result
Alkaloids	Bouchardat's reagent test Dragendorff's reagent test Mayer's reagent test Wagner's reagent test	Brown precipitate was not observed Red-orange precipitate was not formed Yellowish-white precipitate was not formed A red-brown precipitate was not formed	Absence of alkaloids Absence of alkaloids Absence of alkaloids Absence of alkaloids
Anthraquinone Glycosides	Borntrager's test Ammonium hydroxide test	Red color was not occurred Pink-red color was not formed	Absence of anthraquinone glycosides Absence of anthraquinone glycosides
Cardiac Glycosides	Keller-Kiliani test	No formation of brown-ring between layers	Absence of cardiac glycosides
Flavonoids	Alkaline reagent test Shinoda test	Yellow fluorescence was not present Red-pink color was not formed	Absence of flavonoids Absence of flavonoids
Polyphenols	Lead acetate test Ferric chloride test	A bulky white precipitate was formed Blue-green color was formed	Presence of polyphenols Presence of polyphenols
Saponins	Frothing test	A stable persistent froth was formed	Presence of saponins
Steroids	Liebermann-Buchard test	No formation of brown ring at the interface	Absence of steroids
Tannins	Ferric chloride test	Blue or purple or green precipitate was formed	Presence of tannins
Terpenoids	Salkowski test	A reddish-brown precipitate was formed	Presence of terpenoids

presented in Table 1, CCl₄ intoxicated mice had decreased final body weight and increased change in body weight and both absolute and relative liver weight as compared to the negative control. Proliferation, cytoplasmic vacuolization, infiltration of inflammatory cells, and deposition of collagen fibers in the hepatocytes are critical determinants for the survival of the liver from an injury.⁵¹ This was evidenced by the histopathology of the liver of mice given CCl₄ in which there was a profound disarrangement, inflammation, and infiltration of liver cells (Figure 1B). Therefore, up-regulation of hepatocyte activity in response to CCl₄ toxicity is likely to be the cause of the recorded increase in liver weight.

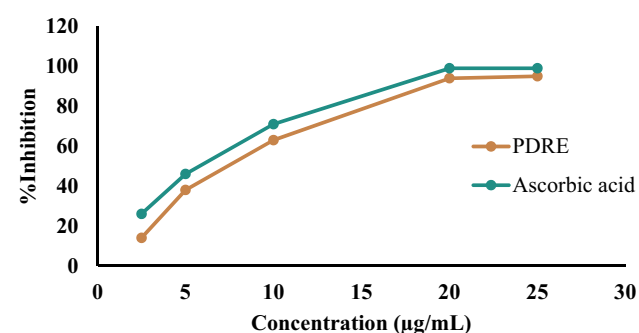


Figure 2 DPPH radical scavenging activity of *P. dodecandra* root extract. **Abbreviations:** PDRE, *Phytolacca dodecandra* root extract; Ascorbic acid, standard antioxidant.

In stark contrast, mice treated with *P. dodecandra* root extract at the doses of 200 and 400 mg/kg pre- and post-dose of CCl₄ were returned to their normal final body weight and absolute and relative liver weight as compared to the toxicant control (Table 1). Similarly, the hepatocyte architecture of mice treated with PDRE200 and PDRE400 was returned to normal (Figure 1E and F, respectively). On the other hand, *P. dodecandra* root extract administration to mice pre- and post-dose of CCl₄ normalized the liver's metabolic capability in a dose-dependent manner (Table 2). In the injured liver, there are depressed levels

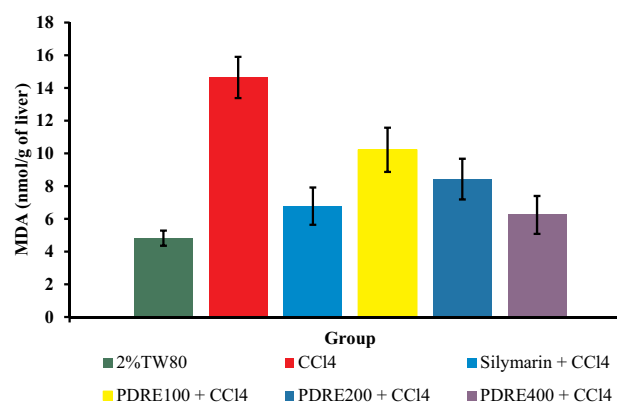


Figure 3 Effect of *P. dodecandra* root extract on MDA levels.

Notes: Each value represents mean \pm S.E.M; n=6.

Abbreviations: PDRE100, *Phytolacca dodecandra* root extract 100 mg/kg; PDRE200, *Phytolacca dodecandra* root extract 200 mg/kg; PDRE400, *Phytolacca dodecandra* root extract 400 mg/kg; Silymarin, silymarin 100 mg/kg; CCl₄, carbon tetrachloride 2 mL/kg; 2%TW80, 2% Tween 80 in water.

of total protein and albumin due to disturbances in carbohydrate, protein, and lipid metabolism or perturbed protein biosynthesis.⁵² The marked reduction of total protein and albumin synthesis and substantially increased bilirubin levels in the CCl₄ treated mice agreed with the early observations.^{36,37}

The liver plays an essential role in lipid metabolism, synthesis and oxidation of cholesterol, and synthesis of lipoproteins.⁵² However, mice intoxicated with CCl₄ showed significantly reduced levels of TC, TG, and LDL which is reasonably expected since cholesterol biosynthesis in the liver has been reduced. Whereas treatment with *P. dodecandra* root extract at the doses of 200 and 400 mg/kg returned the metabolic function of the liver as evidenced in the levels of TC, TG, LDL, and HDL, which were normalized as compared to the toxicant control (Table 4).

CCl₄ is known to damage liver cells via oxidation of membrane lipids and other cellular components.⁵¹ The damage of the hepatocyte membrane resulted in the release of cellular components and enzymes. The serum level of liver enzymes helps to diagnose and monitor liver disease or damage. Alteration in the structure and function of the liver resulted in a marked elevation in the levels of ALP, ALT, AST, GGT, and LDH.⁵³ An agent that returns the levels of these enzymes to normal is considered hepatoprotective since these enzymes are at a high level in plasma following hepatocyte injury.⁵³ According to this study, mice received *P. dodecandra* root extract at the doses of 200 and 400 mg/kg showed a significant reduction in the levels of liver enzymes (Table 3). This can be evidenced from the percent reduction of the levels of ALP, ALT, AST, GGT, and LDH for the two mentioned doses, where PDRE400 displayed a better activity than PDRE200 (96.0%, 93.4%, 95.4%, 76.9%, and 91.8% vs 83.4%, 88.7%, 85.6%, 66.6%, and 89.4%), respectively (Table 3). Probably *P. dodecandra* root extract might stabilize hepatocyte membranes and interrupt the release of enzymes from the liver into blood especially in the two larger doses. However, PDRE100 did not produce a visible effect in all biomarkers of hepatocyte injury. This is probably due to the lower dose might contain a low level of active principles and the other two doses might have enough concentration of active substances that can elicit the observed effect.

Results of histopathological examination served as compelling evidence in supporting the biochemical markers of liver injury. PDRE200 and PDRE400 treated mice

resulted in a marked decrease in the hepatocyte ballooning, cytoplasmic vacuolization, derangement of hepatocytes, and infiltration of hepatocyte membrane with inflammatory cells and collagen fibers (Figure 1E and F, respectively). Besides treatment with *P. dodecandra* root extract inhibits CCl₄ induced formation of lipid peroxides in the liver of mice. Since the toxicity mechanism of CCl₄ is peroxidation of membrane lipids after bio-activated to its toxic metabolites.⁵¹ In this study peroxidation of lipids was determined by measuring the levels of MDA, an end product of lipid peroxidation, in hepatic tissues. CCl₄ intoxication induced a marked increase in tissue MDA levels as compared with the negative control ($P < 0.001$; Figure 3). On the contrary, treatment with *P. dodecandra* root extract returned the increased MDA to its normal level. This showed that CCl₄ mediated oxidative damage is reversed by administration of the plant extract and probably prevents the liver from free radical-induced injury and subsequent pathological changes.

Though the active principles present in the *P. dodecandra* root extract are not identified, polyphenols, saponins, tannins, and terpenoids were found to have antioxidant and hepatoprotective activities via several mechanisms.

Qualitative phytochemical analysis of the 80% methanol extract of *P. dodecandra* root for the presence and absence of several medicinally active secondary metabolites was done and polyphenols, saponins, tannins, and terpenoids were found in the plant extract (Table 5). Probably phytochemicals present in the plant extract act individually or in synergy to produce the observed antioxidant, lipid peroxidation inhibition, and hepatoprotective effects.

Previous studies revealed that phenolic compounds have several antioxidant, anti-inflammatory, and immunomodulatory activities. They prevent oxidative induced cellular damage by inhibiting enzymes involved in the production of reactive oxygen species (ROS), through neutralizing ROS, and up-regulation of antioxidant defenses.^{55–57} Oxidative stress promotes inflammation via activation of genes and thereby the expression of several proteins (enzymes) involved in the inflammatory pathways.⁵⁸ Polyphenols also have an anti-inflammatory effect via inhibition of enzymes involved in the inflammatory process following membrane damage such as cyclooxygenase-2 (COX-2), lipoxygenase (LOX), and nitric oxide synthase (NOS).⁵⁹ In addition, they are also known for donating hydrogen atoms in the prevention of lipid peroxidation.⁶⁰

Tannins are also hepatoprotective secondary active metabolites either by their antioxidant activity (donate hydrogen atom or electrons, chelate metal ions, and interfere in the Fenton reaction)⁶¹ or via inhibition of apoptosis through increased expression of the anti-apoptotic protein Bcl-2.⁶² Besides, they also inhibit lipid peroxidation via inhibition of COX-2.⁶¹ Terpenoids and saponins are also natural hepatoprotective compounds.^{63,64} They inhibit the formation of ROS, scavenge free radicals, inhibit peroxidation of lipids, suppress prooxidants and increase antioxidants, up-regulate nuclear factor E2-related factor 2 (Nrf2)-antioxidant signaling,⁶⁵ and inhibit apoptosis through activating the mitogen-activated protein kinase (MAPK) signaling pathway.⁶⁶

To confirm the antioxidant activity of *P. dodecandra* root extract, DPPH free radical scavenging activity was carried out. In this assay, *P. dodecandra* root extract was observed to inhibit DPPH free radicals with the maximum value of 94% at the concentration of 20 µg/mL (Figure 2) and had a calculated IC₅₀ value of 7.4 µg/mL, which is nearly similar to the calculated IC₅₀ value of the standard antioxidant, ascorbic acid, i.e. 6.2 µg/mL. The presence of potent phytochemicals in the extract may be responsible for the observed antioxidant activity.

To sum up, the 80% methanol extract of *P. dodecandra* root possessed comparable antioxidant and hepatoprotective activities with the respective standards used in the study. Based on the results of this study *P. dodecandra* root extract showed a dose-dependent reduction in the biomarkers of liver damage and MDA level and increment in the metabolic functions of the liver. Even though the plant extract might have several mechanisms of hepatoprotective actions, lipid peroxidation inhibition and free radical scavenging activities are among the demonstrated mechanisms. Above all, the plant extract would be rewarded as safe based on the results of the acute oral toxicity study. Also, isolation and characterization of the specific hepatoprotective compound will be done in future studies by using HPLC/LC-MS techniques.

Conclusion

The results of serum biochemical analysis, histopathological investigations, and both in vitro and in vivo antioxidant studies provide evidence for the folkloric use of *P. dodecandra* for the management of liver problems. The middle and larger doses of *P. dodecandra* root extract produced remarkable hepatoprotective and antioxidant activities that were comparable to silymarin. The presence of medicinally active secondary metabolites in the plant extract may justify the observed hepatoprotective, lipid peroxidation inhibition, and in vitro free radical scavenging activities.

Abbreviations

ALP, Alkaline phosphatase; ALT, Alanine aminotransferase; AST, Aspartate aminotransferase; CCl₄, Carbon tetrachloride; DPPH, 1, 1-diphenyl-2-picrylhydrazyl; GGT, Gamma-glutamyl transferase; HDL, High-density lipoprotein; LDH, Lactate dehydrogenase; LDL, Low-density lipoprotein; MDA, Malondialdehyde; ROS, Reactive oxygen species; TBARS, Thiobarbituric acid reactive substance; TC, Total cholesterol; TG, Triglyceride.

Data Sharing Statement

All the necessary data and materials can be obtained from the corresponding author with a reasonable request.

Ethics Approval

The study was approved by the Ethical Review Board of the College of Medicine and Health Sciences of Wollo University.

Consent for Publication

The authors have consented to the publication of this manuscript.

Author Contributions

All authors made a significant contribution from the conception and design of the study to the execution, acquisition, analysis, and interpretation of data. Besides, they took part in drafting, revising, or critically reviewing the manuscript, gave final approval of the version to be published, have agreed on the journal to which the manuscript has been submitted, and agree to be accountable for all aspects of the work.

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The authors report no conflicts of interest for this work.

References

- Esser KB, Semagn K, Wolde-Yohannes L. Medicinal use and social status of the soap berry endod (*Phytolacca dodecandra*) in Ethiopia. *J Ethnopharmacol.* 2003;85(2-3):269-277. doi:10.1016/S0378-8741(03)00007-2
- Karunamoorthi K, Bishaw D, Mulat T. Laboratory evaluation of Ethiopian local plant *Phytolacca dodecandra* extract for its toxicity effectiveness against aquatic macroinvertebrates. *Eur Rev Med Pharmacol Sci.* 2008;12(6):381-386.
- Gijan M, Dalle G. Ethnobotanical study of medicinal plants in Nagelle Arsi District, West Arsi Zone of Oromia, Ethiopia. *J Nat Sci Res.* 2019;9(13):1-18.

4. Giday K, Lenaerts L, Gebrehiwot K, Yirga G, Verbist B, Muys B. Ethnobotanical study of medicinal plants from degraded dry afro-montane forest in northern Ethiopia: species, uses and conservation challenges. *J Herb Med.* 2016;6(2):96–104. doi:10.1016/j.hermed.2016.03.004
5. Worku AM. A review on significant of traditional medicinal plants for human use in case of Ethiopia. *J Plant Pathol Microbiol.* 2019;10(6):1–12.
6. Zenebe G, Zerihun M, Solomon Z. An ethnobotanical study of medicinal plants in Asgede Tsimbila district, Northwestern Tigray, northern Ethiopia. *Ethnobot Res Appl.* 2012;10:305–320. doi:10.17348/era.10.0.305-320
7. Birhan Y, Kitaw S, Alemayehu Y, Mengesha N. Ethnobotanical study of medicinal plants used to treat human diseases in Enarj Enawga district, East Gojjam zone, Amhara region, Ethiopia. *SM J Med Plant Stud.* 2017;1(1):1–9. doi:10.36876/smjmps.1006
8. Mekuanent T, Zebene A, Solomon Z. Ethnobotanical study of medicinal plants in Chilga district, Northwestern Ethiopia. *J Nat Remedies.* 2015;15(2):88–112. doi:10.18311/jnr/2015/476
9. Garedeu B, Bizuayehu B. A Review on Ethnobotanical study of traditional medicinal plants used for treatment of liver problems in Ethiopia. *Eur J Med Plants.* 2018;1–18. doi:10.9734/EJMP/2018/38153
10. Megersa M, Asfaw Z, Kelbessa E, Beyene A, Woldeab B. An ethnobotanical study of medicinal plants in Wayu Tuka district, east Welega zone of oromia regional state, West Ethiopia. *J Ethnobiol Ethnomed.* 2013;9(1):68. doi:10.1186/1746-4269-9-68
11. Teklehaymanot T. Ethnobotanical study of knowledge and medicinal plants use by the people in Dek Island in Ethiopia. *J Ethnopharmacol.* 2009;124(1):69–78. doi:10.1016/j.jep.2009.04.005
12. Wubetu M, Abula T, Dejenu G. Ethnopharmacologic survey of medicinal plants used to treat human diseases by traditional medical practitioners in Dega Damot district, Amhara, Northwestern Ethiopia. *BMC Res Notes.* 2017;10(1):157. doi:10.1186/s13104-017-2482-3
13. Teklehaymanot T, Giday M. Ethnobotanical study of medicinal plants used by people in Zegie Peninsula, Northwestern Ethiopia. *J Ethnobiol Ethnomed.* 2007;3(1):12. doi:10.1186/1746-4269-3-12
14. Admasu P, Deressa A, Mengistu A, Gebrewold G, Feyera T. In vivo antirabies activity evaluation of hydroethanolic extract of roots and leaves of *Phytolacca dodecandra*. *Glob Vet.* 2014;12(1):12–18. doi:10.5829/idosi.gv.2014.12.01.81150.
15. Namulindwa A, Nkwangu D, Oloro J. Determination of the abortifacient activity of the aqueous extract of *Phytolacca dodecandra* (L'Her) leaf in Wistar rats. *Afr J Pharm Pharmacol.* 2015;9(3):43–47. doi:10.5897/AJPP2014.4227
16. Nakalembe L, Kasolo JN, Nyatia E, Lubega A, Bbosa GS. Analgesic and anti-inflammatory activity of total crude leaf extract of *Phytolacca dodecandra* in Wistar Albino Rats. *Neurosci Med.* 2019;10(3):259–271. doi:10.4236/nm.2019.103020
17. Bernard N, Emmerance N, Ingabire A. Comparative bacterial inhibition by bioactive extracts from *Datura stramonium* and *Phytolacca dodecandra*. *Virol Res J.* 2018;2(1):1–5.
18. Itoku JB, Mbayi O, Bongo GN, et al. Phytochemical analysis and assessment of antibacterial and antioxidant activities of *Phytolacca dodecandra* L. Herit Leaf extracts (Phytolaccaceae). *Int J Biomed Eng Clin Sci.* 2019;5(3):31. doi:10.11648/j.ijbecs.20190503.11
19. Jaleta H, Ameni G, Arage M, Giday M, Girma M, Sori T. In vitro evaluation of the effects of selected plants on the growth of the mycelial form of *histoplasma capsulatum* variety *farciminosum* in Ethiopia. *J Equine Vet Sci.* 2020;91:103139. doi:10.1016/j.jevs.2020.103139.
20. Lemma A. Laboratory and field evaluation of the molluscicidal properties of *Phytolacca dodecandra*. *Bull World Health Organ.* 1970;42(4):597.
21. Kariuki S, Kariuki J, Mailu B, Muchiri D. *Phytolacca octandra* (L.), *Phytolacca dodecandra* (L'Herit) and *Balanites aegyptiaca* (L.) extracts as potential molluscicides of schistosomiasis transmitting snails. *J Med Plants Res.* 2016;10(44):823–828. doi:10.5897/JMPR2016.6237
22. Lemma A, Brody G, Newell GW, Parkhurst R, Skinner W. Studies on the molluscicidal properties of endod (*Phytolacca dodecandra*): I. Increased potency with butanol extraction. *J Parasitol.* 1972;58(1):104–107. doi:10.2307/3278251
23. Thiilborg ST, Christensen SB, Cornett C, Olsen CE, Lemmich E. Molluscicidal saponins from *Phytolacca dodecandra*. *Phytochemistry.* 1993;32(5):1167–1171. doi:10.1016/S0031-9422(00)95085-4
24. Madhina D, Shiff C. Prevention of snail miracidia interactions using *Phytolacca dodecandra* (L'Herit)(endod) as a miracidicide: an alternative approach to the focal control of schistosomiasis. *Trop Med Int Health.* 1996;1(2):221–226. doi:10.1111/j.1365-3156.1996.tb00030.x
25. Pezzuto JM, Swanson SM, Farnsworth NR. Evaluation of the mutagenic potential of endod (*Phytolacca dodecandra*), a molluscicide of potential value for the control of schistosomiasis. *Toxicol Lett.* 1984;22(1):15–20. doi:10.1016/0378-4274(84)90039-0
26. Stolzenberg S, Parkhurst R. Spermicidal actions of extracts and compounds from *Phytolacca dodecandra*. *Contraception.* 1974;10(2):135–143. doi:10.1016/0010-7824(74)90069-9
27. Adinew GM. Antimalarial activity of methanolic extract of *Phytolacca dodecandra* leaves against Plasmodium berghei infected Swiss albino mice. *Int J Pharmacol Clin Sci.* 2014;3(3):39–45.
28. Zeleke AJ, Shimo BA, Gebre DY. Larvicidal effect of endod (*Phytolacca dodecandra*) seed products against *Anopheles arabiensis* (Diptera: culicidae) in Ethiopia. *BMC Res Notes.* 2017;10(1):449. doi:10.1186/s13104-017-2792-5
29. Owiti YJ, Barack -O-OJ, Auma AC, India JJ, Pamela W-K, John VM. Larviciding potency of water and ethanol extracts of *Phytolacca dodecandra* (L'Herit) on *Anopheles gambiae* (Diptera: culicidae). *J Mosq Res.* 2015;5(2):1–6.
30. Raja N, Masresha G, Jemberie W. Insecticidal activity of *Phytolacca dodecandra* L. Herit (Phytolaccaceae) plant extracts against cabbage flea beetle *Phyllotreta cruciferae* Goeze (Coleoptera: chrysomilidae). *Am Eurasian J Sci Res.* 2015;10(5):325–331.
31. Matebie WA, Zhang W, Xie G. Chemical composition and antimicrobial activity of essential oil from *Phytolacca dodecandra* collected in Ethiopia. *Molecules.* 2019;24(2):342. doi:10.3390/molecules24020342
32. Perret C, Wolfender JL, Hostettmann K. LC/ES-MS analysis of triterpene glycosides: rapid estimation of the saponin content of dried berries of *Phytolacca dodecandra*. *Phytochem Anal.* 1999;10(5):272–278. doi:10.1002/(SICI)1099-1565(199909/10)10:5<272::AID-PCA470>3.0.CO;2-Z
33. Dorsaz AC, Hostettmann K. Further saponins from *Phytolacca dodecandra* L' Herit. *Helv Chim Acta.* 1986;69(8):2038–2047. doi:10.1002/hlca.19860690827
34. Domon B, Hostettmann K. New saponins from *Phytolacca dodecandra* L'Herit. *Helv Chim Acta.* 1984;67(5):1310–1315. doi:10.1002/hlca.19840670517
35. Guideline OO. 425: acute oral toxicity—up-and-down procedure. *OECD Guidelines Test Chem.* 2001;2:12–16.
36. Sintayehu B, Bucar F, Veeresham C, Asres K. Hepatoprotective and free radical scavenging activities of extracts and a major compound isolated from the leaves of *Cineraria abyssinica* Sch. Bip. exA. Rich. *Pharmacogn J.* 2012;4(29):40–46. doi:10.5530/pj.2012.29.6
37. Meharie BG, Amare GG, Belayneh YM. Evaluation of hepatoprotective activity of the crude extract and solvent fractions of *Clutia abyssinica* (Euphorbiaceae) leaf against CCl₄-induced hepatotoxicity in Mice. *J Exp Pharmacol.* 2020;12:137–150. doi:10.2147/JEP.S248677

38. Agrawal A, Rao M, Jasdanwala S, Mathur A, Eng M. Cephalixin induced cholestatic jaundice. *Case Rep Gastrointest Med*. 2014;2014:1–3. doi:10.1155/2014/260743
39. Maria R, Shirley M, Xavier C, et al. Preliminary phytochemical screening, total phenolic content and antibacterial activity of thirteen native species from Guayas province Ecuador. *J King Saud Univ Sci*. 2018;30(4):500–505. doi:10.1016/j.jksus.2017.03.009
40. Tiwari P, Kumar B, Kaur M, Kaur G, Kaur H. Phytochemical screening and extraction: a review. *Int Pharm Sci*. 2011;1(1):98–106.
41. Rohini MV, Padmini E. Preliminary phytochemical screening of selected medicinal plants of polyherbal formulation. *J Pharmacogn Phytochem*. 2016;5(5):277–282.
42. Hossain MA, AL-Raqmi KAS, AL-Mijizy ZH, Weli AM, Al-Riyami Q. Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown *Thymus vulgaris*. *Asian Pac J Trop Biomed*. 2013;3(9):705–710. doi:10.1016/S2221-1691(13)60142-2
43. Gul R, Jan SU, Faridullah S, Sherani S, Jahan N. Preliminary phytochemical screening, quantitative analysis of alkaloids, and antioxidant activity of crude plant extracts from *Ephedra intermedia* indigenous to Balochistan. *Sci World J*. 2017;2017:1–7. doi:10.1155/2017/5873648
44. Sahu RK, Kar M, Routray R. DPPH free radical scavenging activity of some leafy vegetables used by tribals of Odisha, India. *J Med Plants*. 2013;1(4):21–27.
45. Blois MS. Antioxidant determinations by the use of a stable free radical. *Nature*. 1958;181(4617):1199–1200. doi:10.1038/1811199a0
46. Oakes KD, Van Der Kraak GJ. Utility of the TBARS assay in detecting oxidative stress in white sucker (*Catostomus commersoni*) populations exposed to pulp mill effluent. *Aquat Toxicol*. 2003;63(4):447–463. doi:10.1016/S0166-445X(02)00204-7
47. Vogel HG. Guidelines for the Care and Use of Laboratory Animals. In: Vogel HG, editor. *Drug Discovery and Evaluation*. Heidelberg: Springer-Verlag Berlin Heidelberg; 2007:2023–2037. doi:10.1007/978-3-54070995-4_18.
48. National Research Council. *Guide for the Care and Use of Laboratory Animals*. National Academies Press; 2011. Available from: <https://grants.nih.gov/grants/olaw/guide-for-the-care-and-use-of-laboratory-animals.pdf>. Accessed November, 2020.
49. Brinklov S, Warrant E. Oilbirds. *Curr Biol*. 2017;27(21):R1145–R7. doi:10.1016/j.cub.2017.08.071
50. Boll M, Lutz W, Becker E, Stampfl A. Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites. *Z Naturforsch C*. 2001;56(7–8):649–659. doi:10.1515/znc-2001-7-826
51. Weber LW, Boll M, Stampfl A. Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. *Crit Rev Toxicol*. 2003;33(2):105–136. doi:10.1080/713611034
52. Ghadir MR, Riahi AA, Havaspour A, Nooranipour M, Habibinejad AA. The relationship between lipid profile and severity of liver damage in cirrhotic patients. *Hepat Mon*. 2010;10(4):285.
53. Dutta S, Chakraborty AK, Dey P, et al. Amelioration of CCl₄ induced liver injury in swiss albino mice by antioxidant rich leaf extract of *Croton bonplandianus* Baill. *PLoS One*. 2018;13(4):e0196411. doi:10.1371/journal.pone.0196411
54. Gupta A, Pandey AK. Chapter 3 - Plant Secondary Metabolites With Hepatoprotective Efficacy. In: Galanakis CM editor. *Nutraceuticals and Natural Product Pharmaceuticals*. Academic Press; 2019:71–104. <https://doi.org/10.1016/B978-0-12-816450-1.00003-9>.
55. Santos-Sánchez NF, Salas-Coronado R, Villanueva-Cañongo C, Hernández-Carlos B. *Antioxidant Compounds and Their Antioxidant Mechanism*. Antioxidants: IntechOpen; 2019:1–28.
56. Perron NR, Brumaghim JL. A review of the antioxidant mechanisms of polyphenol compounds related to iron binding. *Cell Biochem Biophys*. 2009;53(2):75–100.
57. Yahfoufi N, Alsadi N, Jambi M, Matar C. The immunomodulatory and anti-inflammatory role of polyphenols. *Nutrients*. 2018;10(11):1618. doi:10.3390/nu10111618
58. Hong S-Y, Roze LV, Linz JE. Oxidative stress-related transcription factors in the regulation of secondary metabolism. *Toxins*. 2013;5(4):683–702. doi:10.3390/toxins5040683
59. Yoon J-H, Baek SJ. Molecular targets of dietary polyphenols with anti-inflammatory properties. *Yonsei Med J*. 2005;46(5):585–596. doi:10.3349/ymj.2005.46.5.585
60. Ozgová Š, Heřmánek J, Gut I. Different antioxidant effects of polyphenols on lipid peroxidation and hydroxyl radicals in the NADPH-, Fe-ascorbate- and Fe-microsomal systems. *Biochem Pharmacol*. 2003;66(7):1127–1137. doi:10.1016/S0006-2952(03)00425-8
61. Amarowicz R. Tannins: the new natural antioxidants? *Eur J Lipid Sci Technol*. 2007;109(6):549–551. doi:10.1002/ejlt.200700145
62. Sobeh M, Mahmoud MF, Hasan RA, et al. Tannin-rich extracts from *Lannea stuhlmannii* and *Lannea humilis* (Anacardiaceae) exhibit hepatoprotective activities in vivo via enhancement of the anti-apoptotic protein Bcl-2. *Sci Rep*. 2018;8(1):1–16. doi:10.1038/s41598-018-27452-8
63. Krishnamurthy PT, Bajaj J, Sharma A, Manimaran S, Ravanappa PKB, Pottekad V. Hepatoprotective activity of terpenoids and terpenoid fractions of *Scoparia dulcis* L. *Orient Pharm Exp Med*. 2010;10(4):263–270.
64. Qu L, Xin H, Zheng G, Su Y, Ling C. Hepatoprotective activity of the total saponins from *Actinidia valvata* Dunn root against carbon tetrachloride-induced liver damage in Mice. *Evid Based Complement Alternat Med*. 2012;2012:1–13. doi:10.1155/2012/216061
65. Hussain T, Tan B, Liu G, et al. Modulatory mechanism of polyphenols and Nrf2 signaling pathway in LPS challenged pregnancy disorders. *Oxid Med Cell Longev*. 2017;2017:1–14. doi:10.1155/2017/8254289
66. Cui Y, Liu B, Sun X, et al. Protective effects of alfalfa saponins on oxidative stress-induced apoptotic cells. *Food Funct*. 2020;11(9):8133–8140. doi:10.1039/D0FO01797C

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