


O-GlcNAcylation Enhances NUSAPI Stability and Promotes Bladder Cancer Aggressiveness


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Objective: NUSAPI and O-GlcNAcylation were reported to be hyper-activated in many kinds of cancers and involved in the advanced progression of cancers. In bladder cancer, O-GlcNAc transferase (OGT) expresses in patients' urine samples, with no expression in healthy individuals, indicating O-GlcNAcylation might involve in the occurrence and development of bladder cancer. Therefore, the present study aims to investigate the effects of O-GlcNAcylation in bladder cancer and if it can regulate NUSAPI protein.

Materials and Methods: Western blot, immunohistochemistry, and PCR were used to evaluate the protein expression and mRNA level of NUSAPI; CCK-8 and flow cytometry used to evaluate the proliferation and inhibited the apoptosis of bladder cancer.

Results: The results showed that NUSAPI was highly expressed in bladder cancer cells and tissue samples. NUSAPI up-regulation significantly promoted the proliferation and inhibited the apoptosis of bladder cancer HT-1376 and T24 cells. Besides, the expression of O-GlcNAc was elevated in bladder cancer tissues and cells, and up-regulation of O-GlcNAc with GlcNAc and PuGNAc obviously increased NUSAPI protein expression and stability. Moreover, knockdown OGT significantly inhibited the proliferation and tumorigenesis and promoted the apoptosis of bladder cancer cells, confirmed by CCK-8, in vivo xenotransplantation, and flow cytometry, whereas these roles were impaired when NUSAPI was up-regulated.

Conclusion: Overall, our study makes clear that hyper-O-GlcNAcylation accelerates bladder cancer progression through promotion of NUSAPI expression and its stability.

Keywords: NUSAPI, O-GlcNAcylation, stability, bladder cancer, aggressiveness

Introduction

Bladder cancer is the most common malignant tumor among urogenital system carcinomas, with gradually increasing incidence in recent years influenced by factors such as smoking and environmental pollution worldwide.¹ In global male cancers, the incidence and mortality of bladder cancer rank sixth and ninth, respectively.² Radical cystoprostatectomy bilateral pelvic lymphadenectomy and urinary diversion are the gold standard for the treatment of male MIBC. However, patients often have distant recurrence after surgery, and the treatment effect is not satisfactory. Therefore, it is an important and even preferred way to fully understand and master the development mechanism of bladder cancer to find new effective therapeutic targets.

NUSAPI, first discovered in 2003, is a microtubule-binding protein mainly located in the nucleolus, with a molecular weight of about 55KD. In recent years, numerous studies have found that NUSAPI is highly expressed in a variety of kinds of tumors and participates in their development, including astrocytoma,³ renal cell carcinoma,⁴

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oral squamous cell carcinoma,⁵ prostate cancer,^{6,7} breast cancer,⁸ and so on. For instance, Gulzar et al⁹ found that the recurrence of tumor after prostatectomy was closely associated with the increased transcription level of NUSAP1. Besides, Gordon et al⁶ suggested that up-regulation of NUSAP1 significantly enhanced cell invasion and migration. Similarly, NUSAP1 was found to be over-expressed in 68.78% (152/221) of astrocytoma tissues, and the high level of it showed significant association with patients' poor survival, which also obviously promoted the invasion of astrocytoma cells.³ These findings all indicated that NUSAP1 might function as an oncogene in cancer. However, the expression pattern and effects of NUSAP1 in bladder cancer still remains unknown.

Glycosylation is a major post-transcriptional protein modification, which plays an important role in many biological processes. More than 50% of human proteins have been identified can be glycosylated,¹⁰ followed by enhanced stability.^{11,12} O-Linked β -N-acetylglucosamine (O-GlcNAc) is a main kind of glycosylation for cytosolic and nuclear proteins through covalent addition of a GlcNAc sugar moiety to the serine and/or threonine residues of proteins,¹³ which is regulated by O-GlcNAc transferase (OGT) and O-GlcNAcase (OGA).^{14–16} OGT transfers the GlcNAc moiety from the high-energy donor uridine 5'-diphosphate (UDP)-GlcNAc to substrate proteins, whereas OGA hydrolyzes O-GlcNAc from proteins. Dysregulation of key proteins O-GlcNAcylation has been demonstrated in the etiology of numerous varieties of tumors, including bladder cancer.¹⁷ However, the effects of O-GlcNAcylation in bladder cancer and if it can regulate NUSAP1 protein remain unclear.

Based on the above, one of the objectives of this study is to explore the function of NUSAP1 in bladder cancer progression; the other is to determine the effects of O-GlcNAcylation on NUSAP1 protein expression and stability, as well as to investigate the role of O-GlcNAcylation of NUSAP1 on the tumorigenesis of bladder cancer cells. Our study expects to further clarify the functions of O-GlcNAcylation and NUSAP1 in the occurrence and development of bladder cancer, hoping to provide new idea for the invention of antitumor drugs.

Materials and Methods

Patients and Tissue Samples

Twenty-five matched fresh bladder cancer tissues and their adjoining non-tumor tissues were all obtained from Shanghai Tenth People's Hospital, between January 2015

to January 2017. All histological samples were identified by three seasoned pathologists without knowing the clinical data. And the study involved human samples was approved by the Human Research Committee of Shanghai Tenth People's Hospital and had been proceeded in accordance with the Helsinki Declaration.

Immunohistochemistry (IHC)

Paraffin sections of tissues were sliced into sections of 4 μ m thickness by a slicing machine and processed just as follows: dewaxing and hydrating the tissues; incubation with 3% H₂O₂ for 10 min at room temperature; antigen repairing with Tris-EDTA; blocking with 5% goat serum (diluted in PBS); incubation overnight at 4°C with NUSAP1 primary antibody (1:50 dilution; No. PA5-24479, Thermofisher, MA, USA), followed by incubation with the Rabbit secondary antibody and washed with PBS. Chromogen 3, 3'-diaminobenzidine tetrachloride (Serva, Heidelberg, Germany) was used as a substrate. Harri's hematoxylin solution was used to stain the nucleus.

Cell Culture

Bladder cancer cell lines including HT-1376, 5637, and T24, and normal uroepithelium cell line SV-HUC-1 were all purchased from ATCC (VA, USA). HT-1376 and were grown in Eagle's Minimum Essential Medium (No. 30–2003, ATCC, VA, USA). Five thousand six hundred and thirty-seven cells were cultured in RPMI-1640 medium (Biological Industries, Kibbutz BeitHaemek, Israel), T24 cells were cultured in McCoy's 5a Medium (No. 30–2007, ATCC, VA, USA). The base medium for SV-HUC-1 cell line is ATCC-formulated F-12K Medium (No. 30–2004). The culture mediums for all cell lines contained 10% fetal bovine serum (FBS; Gibco, CA, USA), 100 U/mL of penicillin, and 100 μ g/mL of streptomycin. And cells were cultured at 37°C in a humidified incubator with air charge of 5% CO₂.

Cell treated with 25 mM PuGNac (Sigma, St Louis, MO, USA) and 4 mM GlcNAc (Sigma, St Louis, MO, USA) for 24 h to up-regulate O-GlcNAc level. Cells were treated with 0.1 mg/mL of Cycloheximide (Chx; Sigma, St Louis, MO, USA) for 1, 2, 4, 8, and 24 h to explore protein stability, all respectively.

Cell Transfection

The over-expressing plasmid of NUSAP1 (OE-NUSAP1) was synthesized by GenePharma (Suzhou, Jiangsu, China). The shRNAs (No. TR30023) targeting human OGT gene and their controls were purchased from

Origene (CA, USA). Lipofectamine 2000 obtained from Life Technologies (MA, USA) was used to perform cell transfection according to the manufacturer's instruction. For in vivo tumor formation assay, puromycin (for shRNA-OGT; 7 µg/mL) and G418 (for OE-NUSAP1; 4 µg/mL) were used to screen cells with stable transfection.

Real-Time Polymerase Chain Reaction (RT-PCR)

Total RNA was isolated from bladder cancer tissues and cell lines using Trizol Reagent (Invitrogen, Carlsbad, CA, USA) in accordance with the manufacturer's instructions. cDNA was generated using Reverse Transcription System (Promega, Thermofisher, MA, USA). RT-PCR was performed by using AccessQuick™ RT-PCR System (Promega, Thermofisher, MA, USA). Primer sequences used in this study were as follows: NUSAP1 (F): 5'-GTGTGGACCCTGACTCACAG-3', (R): 5'-CTTCTTGGTGCTCGTCTGGT-3'; OGT (F): 5'-CGGCTGCGTGTAGGATATGT-3', (R): 5'-GCCATCACC TTCCTCGGAA-3'; GAPDH (F): 5'-GAGAAGGCTGGG GCTCATTT-3'; (R): 5'-AGTGATGGCATGGACTGTGG-3'.

Western Blot Analysis (WB)

Proteins from cells and tissue samples were obtained by using RIPA Lysis Buffer (Beyotime Biotechnology, Shanghai, China), and the protein extraction process was proceeded in ice in accordance with the instructions. After quantified with bicinchoninic acid (BCA) Protein Assay Kit (Thermo Fisher Scientific, MA, USA) and denatured in 100°C for 10 min, protein samples from each group were separated on a 10% SDS-PAGE gel and transferred onto a PVDF membrane (Millipore, CA, USA). Then, the membranes were blocked by 5% nonfat milk for 1 h at room temperature and were incubated with the primary antibodies at 4°C overnight. Next day, the membranes were washed and incubated with the corresponding HRP-linked secondary antibodies (No. #7074, #7076, Cell Signaling Technology, CA, USA) at 1:10,000 dilution for 1 h at room temperature. The primary antibodies used for this study are anti-NUSAP1 (No. PA5-24479, Thermofisher, MA, USA) and anti-GAPDH (No. MA5-15738-BTIN, Thermofisher, MA, USA). The expression levels of proteins were measured by ECL (Millipore, CA, USA) and the ImageJ software (National Institutes of Health) was used for quantification after background subtraction.

Cell Counting Kit-8 (CCK-8) Assay

Cell proliferation was assessed by CCK-8 kit (Solarbio Life Sciences, Beijing, China) as described in the manufacturer's protocol.

Flow Cytometry with Annexin V/Propidium Iodide (PI) Staining

Flow cytometry with Annexin V/PI staining was used for cell apoptosis detection. Briefly, bladder cancer HT-1376 and T24 cells were grown in a 12-well plate. Forty-eight hours after the transfection, the cells were harvested and stained with PI and Annexin V (FITC) (Apoptosis Detection Kit, BD Biosciences, CA, USA) followed by detection with BD-FACS Canto™ II. The percentage of cell population undergoing early and late apoptosis was assessed by FlowJo7.6 software.

In vivo Tumor Formation Assay

Animal experiments were conducted in accordance with ethical standards (Guide to Ethical Review of Laboratory Animal Welfare -GB/T 35892-2018) and approved by Institutional Animal Care and Use Committee of Shanghai Tenth People's Hospital. HT-1376 and T24 cells with stable transfection of shRNA-OGT, OE-NUSAP1, sh-OGT+OE-NUSAP1, and control vectors were subcutaneously injected into the both flanks of 4-week BALB/c nude mice (Shanghai SLAC Laboratory Animal Co, Ltd. China), with 5 mice in each group. Twenty-six days after injection, the mice were euthanized and tumors were taken out to measure their weight.

Data Analysis

The data statistical analyses of this study were performed with SPSS 22.0 software. The data are expressed as means ± SD. The two-tailed Student's *t*-test and one-way ANOVA test were used to assess the difference between two groups or more. A value of *P* < 0.05 was considered statistically significant.

Results

NUSAP1 is Highly Expressed in Bladder Cancer Tissues and Cell Line

We firstly searched for Oncomine database to find the expression of NUSAP1 in bladder cancer, 48 cases of bladder tissue samples and 28 cases of superficial bladder cancer tissue samples were included. Results showed that NUSAP1 was highly expressed in bladder cancer tissues as compared

with that bladder tissues (Figure 1A). To confirm this, we also carried out IHC, WB, and RT-PCR to measure both the protein and mRNA levels of NUSAP1 in 25 bladder cancer tissues and the 25 paired adjacent non-tumor tissues, and bladder cancer tissues showed higher expression of NUSAP1 than the para-carcinoma tissues (Figure 1B–D). Similarly, compared with the normal uroepithelium SV-HUC-1 cells, the mRNA and protein levels of NUSAP1 in bladder cancer HT-1376, 5637, and T24 cells were significantly elevated (Figure 1E and F). These results confirm that

NUSAP1 is over-expressed in bladder cancer tissues and cells in both protein and mRNA levels.

Higher Expression of NUSAP1 is Closely Associated with Bladder Cancer Patients' Aggressiveness and Bad Outcome

We also evaluated the association of NUSAP1 expression pattern with patients' clinicopathological characteristics and prognosis. Thirty-two cases of bladder cancer patients

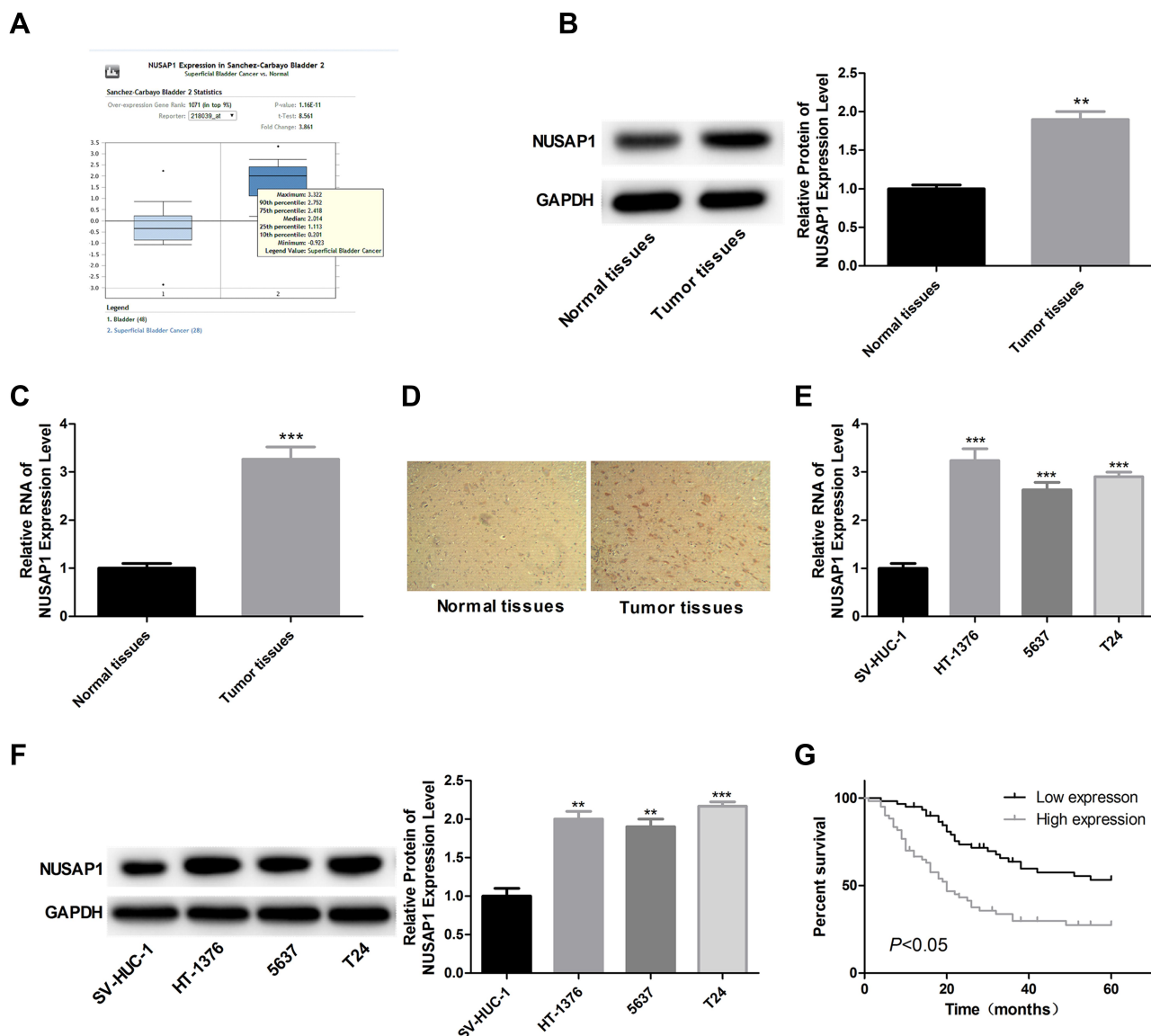


Figure 1 Overexpression of NUSAP1 in bladder cancer tissues and cells. **(A)** OncoPrint database showing the expression level of NUSAP1 in bladder tissues (n=48) and superficial bladder cancer tissues (n=28) ($P < 0.001$). **(B and C)** WB and RT-PCR was performed to determine the protein and mRNA levels of NUSAP1 in 25 paired bladder cancer tissues and tissues adjacent to cancer. One representative figure was shown. (n=25, $**P < 0.01$, $***P < 0.001$, tumor tissues group vs normal tissues). **(D)** IHC was performed to detect NUSAP1 expression using an-NUSAP1 antibody. One representative figure was shown. **(E and F)** WB and RT-PCR was performed to determine the protein and mRNA levels of NUSAP1 in normal uroepithelium cell line SV-HUC-1 and bladder cancer cell line HT-1376, T24 and 5637. (n=3, $**P < 0.01$, $***P < 0.001$, HT-1376, T24 and 5637 group vs SV-HUC-1 group). **(G)** Kaplan-Meier curve used to analysis the OS of bladder cancer patients with high (n=28) or low (n=32) expression of NUSAP1 ($P < 0.05$).

with NUSAP1 low expression and 28 cases with NUSAP1 high expression were chosen as the objects of our study. Statistical results indicate that patients with high NUSAP1 expression always showed larger tumors ($P=0.039$), higher occurrence rate of lymphonodus metastasis ($P=0.003$), more advanced TNM stage ($P=0.016$), and lower differentiation ($P=0.009$) (Table 1). Moreover, the Kaplan-Meier curves were built to assess the difference of OS in patients with NUSAP1 high or low expression, and results showed that OS was obviously shorter in NUSAP1 high expression patients than in patients with NUSAP1 low expression ($P<0.05$, Figure 1G).

Overexpression of NUSAP1 Promotes the Proliferation and Inhibits Apoptosis of Bladder Cancer Cells

Next, we explored the effects of NUSAP1 in the progression of bladder cancer in vivo. OE-NUSAP1 significantly up-regulated the mRNA and protein levels of NUSAP1 in HT-1376 and T24 cells (Figure 2A and B). CCK-8 assay performed to detect cell proliferation showed that overexpression of NUSAP1 significantly enhanced cell

proliferation in both HT-1376 and T24 cell lines, as compared with the control group (Figure 2C and D). Besides, flow cytometry analysis with Annexin V/PI staining indicated that cell apoptosis (total early and late apoptosis cells) was repressed from 8.12% to 3.64% in HT-1376 cells, and from 9.40% to 5.32% in T24 cells when up-regulation of NUSAP1 (Figure 2E). These results suggest that NUSAP1 functions as an oncogene in bladder cancer progression.

O-GlcNAc Promotes NUSAP1 Expression in Bladder Cancer

Then, to investigate the relationship of O-GlcNAcylation and NUSAP1 in bladder cancer, we firstly determined O-GlcNAc expression in 3 matched bladder cancer tissues and para-carcinoma tissues. In accordance with the expression of NUSAP1, WB results showed that compared with the normal tissues group, the expression of O-GlcNAc was increased in the bladder cancer tissues (Figure 3A). Similarly, O-GlcNAc expression was significantly increased in bladder cancer HT-1376, 5637, and T24 cells when compared with SV-HUC-1 cells (Figure 3B). Then, GlcNAc and PuGNAc were added to the culture medium to increase O-GlcNAcylation in both BT-1376 and T24 cells, results showed that the protein levels of O-GlcNAc and NUSAP1 were all elevated when adding GlcNAc and PuGNAc to the cells, especially in GlcNAc + PuGNAc group (Figure 3C and D).

O-GlcNAc Enhances the Stability of NUSAP1 Protein in Bladder Cancer

Subsequently, we assessed the stability of NUSAP1 protein after HT-1376 and T24 cells were treated with GlcNAc, PuGNAc, or GlcNAc + PuGNAc by using cycloheximide (Chx) to stop protein synthesis. And results showed that compared with the control group, treated cells with GlcNAc, PuGNAc or GlcNAc + PuGNAc obviously slowed down the degradation rate of NUSAP1 protein (Figure 4A). Moreover, we also measured the protein stability of NUSAP1 after O-GlcNAcylation was reduced by down-regulating OGT expression in HT-1376 and T24 cells. The expression of OGT was reduced about 50% when the cells were transfected shRNAs-OGT (Figure 4B), followed by reduced NUSAP1 stability (Figure 4C). Taken together, the above findings demonstrate that O-GlcNAc could slow down the degradation of NUSAP1 protein in bladder cancer cells.

Table 1 Expression and Clinical Significance of NUSAP1 in Bladder Cancer

Groups	Total Cases	Low Expression Cases	High Expression Cases	P value
Gender				0.929
Male	49	26	23	
Female	11	6	5	
Age/years				0.235
<60	43	25	18	
≥60	17	7	10	
Size/cm				0.022
≤3	22	16	6	
>3	38	16	22	
Lymphonodus metastasis				0.001
Yes	23	6	17	
No	37	26	11	
TNM				0.001
I-II	35	25	10	
III-IV	25	7	18	
Pathology				0.01
High	21	7	14	
Medium	25	13	12	
Low	14	12	2	

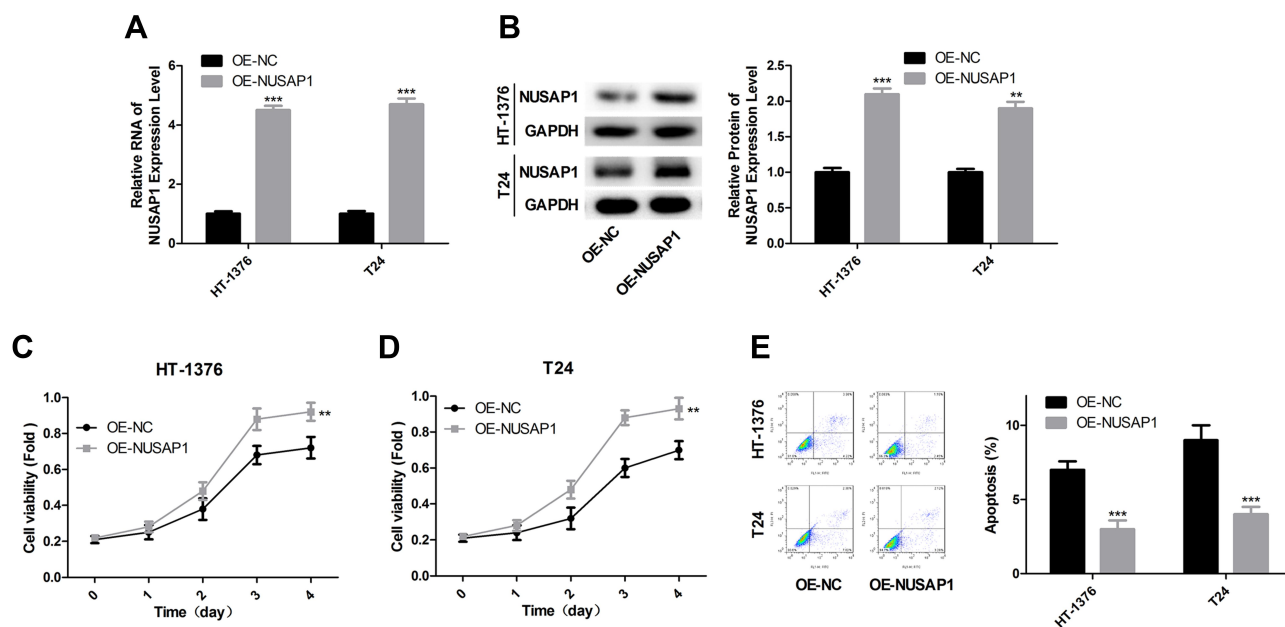


Figure 2 NUSAP1 promoted the proliferation and repressed the apoptosis of bladder cancer cells. **(A)** HT-1376 and T24 cells were transfected with OE-NUSAP1 and OE-NC, then A. 24 h after, RT-PCR was carried out to assess the mRNA level of NUSAP1 ($n=3$, *** $P<0.001$, OE-NUSAP1 group vs OE-NC group). **(B)** 48 h after, WB was used to evaluate the protein level of NUSAP1 ($n=3$, ** $P<0.01$, *** $P<0.001$, OE-NUSAP1 group vs OE-NC group). **(C and D)** CCK-8 analysis of the proliferation of HT-1376 and T24 cells after 0, 1, 2, 3 and 4 days of the transfection, respectively ($n=3$, ** $P<0.01$, OE-NUSAP1 group vs OE-NC group). **(E)** Flow cytometry with Annexin V/PI double staining was used to detect cell apoptosis 48 h after the transfection ($n=3$, *** $P<0.001$, OE-NUSAP1 group vs OE-NC group).

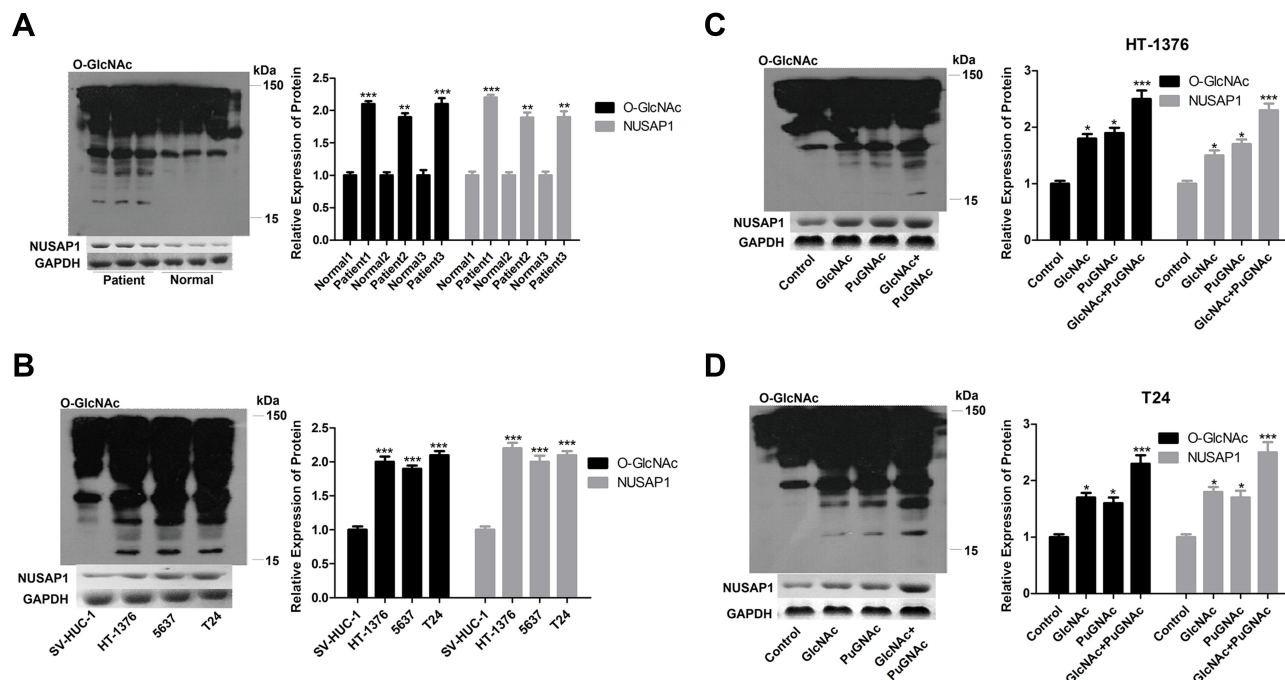


Figure 3 O-GlcNAc highly expressed in bladder cancers and increased NUSAP1 expression. **(A)** WB analysis of the expression of NUSAP1 and O-GlcNAc in 25 paired bladder cancer tissues and tissues adjacent to cancer. One representative figure was shown. ($n=25$, ** $P<0.01$, *** $P<0.001$, tumor tissues group vs normal tissues). **(B)** WB was performed to determine the protein expression of NUSAP1 and O-GlcNAc in normal uroepithelium cell line SV-HUC-1 and bladder cancer cell line HT-1376, T24 and 5637. ($n=3$, *** $P<0.001$, HT-1376, T24 and 5637 group vs SV-HUC-1 group). **(C and D)** HT-1376 and T24 cells were treated with PuGNAC (25 mM), GlcNAc (4 mM) or PuGNAC + GlcNAc for 24 h, then cells were collected for WB analysis to detect the expression of O-GlcNAc and NUSAP1 ($n=3$, * $P<0.05$, *** $P<0.001$, PuGNAC, GlcNAc or PuGNAC + GlcNAc group vs control group).

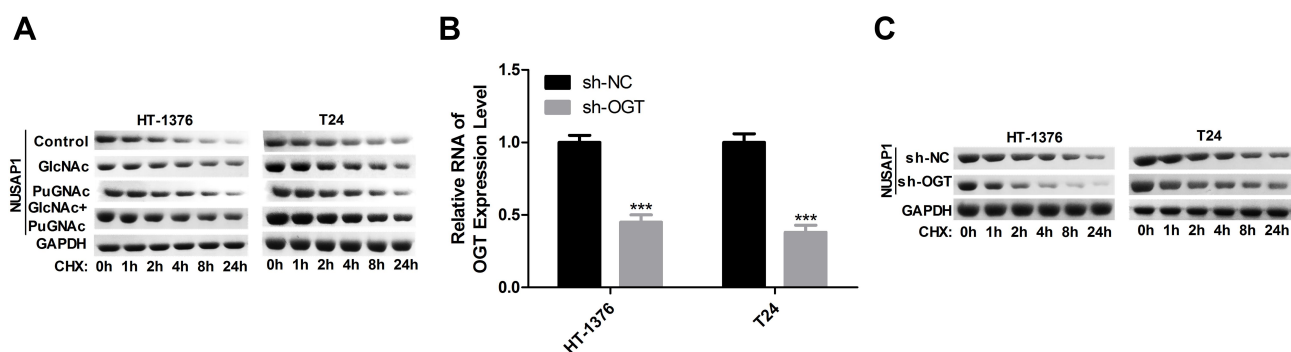


Figure 4 HT-1376 and T24 cells were treated with GlcNAc (4 mM), PuGNac (25 mM) or GlcNAc + PuGNac for 24 h, then Chx ((100 µg/mL) was added to stop protein synthesis. **(A)** Then cells were lysed and equal amounts of protein were separated to WB to measure NUSAP1 expression. **(B)** HT-1376 and T24 cells were transfected sh-OGT and sh-NC for 24 h, then cells were collected and analysis for RT-PCR to determine the mRNA level of OGT (n=3, *** $P < 0.001$, sh-OGT group vs sh-NC group). **(C)** WB analysis the expression of NUSAP1 after OGT was knocked down (n=3).

Knockdown of OGT Inhibits the Progression of Bladder Cancer Through Down-Regulating NUSAP1 Expression

In the end, we investigated the effects of OGT in the development of bladder cancer and if NUSAP1 involved in this process. Knockdown of OGT decreased the expression of O-GlcNAc (Figure 5A). To the opposite of NUSAP1 up-regulation, down-regulation of OGT significantly repressed cell proliferation (Figure 5B and C) and promoted cell apoptosis (Figure 5D), whereas these roles were abolished when NUSAP1 was overexpressed in HT-1376 and T24 cells (Figure 5B and D). In addition, tumorigenesis was significantly inhibited when HT-1376 and T24 cells were transfected shRNA-OGT when compared with the control group, which was reverse with NUSAP1 up-regulation, and tumorigenesis was significantly enhanced when T24 and HT-1376 cells were transfected sh-OGT and OE-NUSAP1 simultaneously (Figure 5E). These results indicated that down-regulation of OGT represses bladder cancer progression through down-regulating NUSAP1 expression (Figure 6).

Discussion

Recently, increasing evidence has manifested that hyper-O-GlcNAcylation is a general specialty for many kinds of cancers and promotes the transformation to more aggressive phenotypes, such as breast,¹⁸ ovarian,¹⁹ prostate,²⁰ colorectal,^{21,22} chronic lymphocytic leukemia,²³ and so on.²⁴ In bladder cancer, Rozanski et al¹⁷ reported that OGT mRNA is detectable in 51.7% of 176 urine samples come from bladder cancer patients, but not discoverable in 143 healthy individuals. They also demonstrate that the mRNA level of OGT is closely correlated with the poorly

differentiation (grade III, the most aggressive form) of cancer,¹⁷ suggesting that hyper-O-GlcNAcylation might involve in the occurrence and development of bladder cancer. However, the mechanism of O-GlcNAcylation remains largely unclear. In this study, we showed that O-GlcNAc level was elevated in bladder cancer cells and tissue samples, which could promote NUSAP1 expression and its protein stability and regulate the pathogenesis of bladder cancer in a NUSAP1 dependent manner.

NUSAP1 is identified as a protein related to cell cycle, and the mRNA and protein levels of which peak in G2 phase of mitosis, and drop rapidly after the split phase content in cells.²⁵ Besides, NUSAP1 expression is significantly elevated in the cells in proliferate stage; however, the high expression of NUSAP1 can result in the stagnation of cells in M phase, and inhibition of its expression would lead to spindle body cannot be formed and chromosome cannot be separated.^{26,27} NUSAP1 is highly expressed in breast cancer,²⁸ hepatocellular carcinoma,²⁹ and pancreatic cancer,³⁰ and functions as an oncogene in these cancers. To our best knowledge, the present study, for the first time, explored the expression of NUSAP1 and its effects on the occurrence and development of bladder cancer. We found that NUSAP1 was over-expression in bladder cancer tissues and cell lines, which high expression level was closely related to patients' aggressive clinical tumor phenotype and poor prognosis. Moreover, we observed that bladder cancer T24 and HT-1376 cells transfected with OE-NUSAP1 showed enhanced proliferation and decreased apoptosis when compared with the cells in the control group, suggesting that NUSAP1 promoted the progression of bladder cancer through inhibiting cell apoptosis and motivating cell growth.

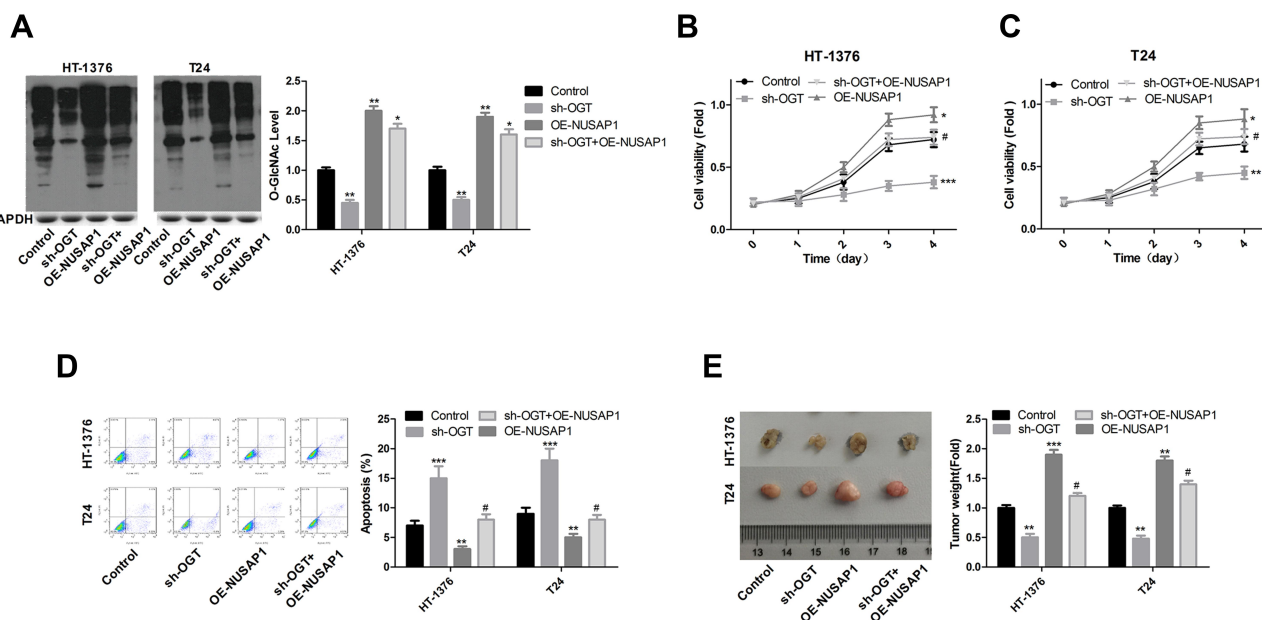


Figure 5 Down-regulation of OGT inhibited the proliferation and tumor formation and promoted the apoptosis of bladder cancer cells. HT-1376 and T24 cells were transfected control vector, OE-NUSAP1, sh-OGT and sh-OGT+OE-NUSAP1, then (A) WB analysis of the expression of O-GlcNAc after 48 h of the transfection. qPCR of the expression of O-GlcNAc (n=3, the experiment was repeated three times in each group; For HT-1376 ** $P<0.01$, sh-OGT group vs control group; ** $P<0.01$, OE-NUSAP1 group vs control group; * $P<0.05$, sh-OGT+OE-NUSAP1 group vs control group; For T24 ** $P<0.01$, sh-OGT group vs control group; ** $P<0.01$, OE-NUSAP1 group vs control group; * $P<0.05$, sh-OGT+OE-NUSAP1 group vs control group). (B and C) CCK-8 analysis of the proliferation of HT-1376 and T24 cells after 0, 1, 2, 3 and 4 days of the transfection, respectively (n=3, the experiment was repeated three times in each group. *** $P<0.001$, sh-OGT group vs control group; * $P<0.05$, OE-NUSAP1 group vs control group; $P>0.05$, sh-OGT+OE-NUSAP1 group vs sh-OGT group). (D) Flow cytometry with Annexin V/PI double staining was used to detect cell apoptosis 48 h after the transfection (n=3, the experiment was repeated three times in each group. For HT-1376 *** $P<0.001$, sh-OGT group vs control group; ** $P<0.01$, OE-NUSAP1 group vs control group; $P>0.05$, sh-OGT+OE-NUSAP1 group vs control group; For T24 *** $P<0.001$, sh-OGT group vs control group; ** $P<0.01$, OE-NUSAP1 group vs control group; $P>0.05$, sh-OGT+OE-NUSAP1 group vs control group). (E) In vivo tumor formation assay was used to evaluate cell tumorigenesis (n=3, For HT-1376 ** $P<0.01$, sh-OGT group vs control group; *** $P<0.001$, OE-NUSAP1 group vs control group; $P>0.05$, sh-OGT+OE-NUSAP1 group vs control group; For T24 ** $P<0.01$, sh-OGT group vs control group; ** $P<0.01$, OE-NUSAP1 group vs control group; $P>0.05$, sh-OGT+OE-NUSAP1 group vs control group).

We also observed that the expression level of O-GlcNAc was elevated in bladder cancer cells and tissue samples, just as NUSAP1, indicating that O-GlcNAcylation might play an important role in bladder cancer. A unique trait of energy metabolism in cancer cells is a change from oxidative phosphorylation to the less efficient glycolytic pathway (Warburg effect), leading to elevated glucose and glutamine uptake, indicating that the increased glucose and glutamine uptake by cancer cells might be related to increased O-GlcNAc levels.²⁴ Previous study has summarized the effects of O-GlcNAcylation on the expression and stability of proteins related to the modulation of cell proliferation, sustained angiogenesis, deregulated cellular energetics, invasion and metastasis, oxidative stress and metabolic stress and found that hyper-O-GlcNAcylation stimulation could increase the expression, stability, or activity of c-Myc, Cyclin D1, FoxM1, Snail, matrix Metalloproteinases (MMPs), OCT4, FoxO4, but reduced the expression, stability, or activity of E-cadherin, p27, phosphofructokinase1 (PFK1).²⁴ Similarly, we showed that stimulation of O-GlcNAcylation with PuGNac (an inhibitor of OGA), and GlcNAc (the substrate

of O-GlcNAcylation) up-regulated NUSAP1 expression and its protein stability in both HT-1376 and T24 cells.

Furthermore, we explored the effects of O-GlcNAcylation in the progression of bladder cancer and determined if NUSAP1 took part in this process mediated by O-GlcNAcylation. shRNA targeting to human OGT gene was used to knock down OGT expression in bladder cancer HT-1376 and T24 cells. Results showed that down-regulation of OGT significantly inhibited the growth and tumorigenesis and promoted the apoptosis of HT-1376 and T24 cells, but these effects were abolished when NUSAP1 was over-expressed, indicating that O-GlcNAcylation inhibition suppressed bladder cancer progression through down-regulation of NUSAP1.

Conclusion

In summary, this study demonstrates that hyper-O-GlcNAcylation and high NUSAP1 expression are observed in bladder cancer. O-GlcNAcylation promotes NUSAP1 expression and stability, and inhibition of O-GlcNAcylation suppresses bladder cancer progression

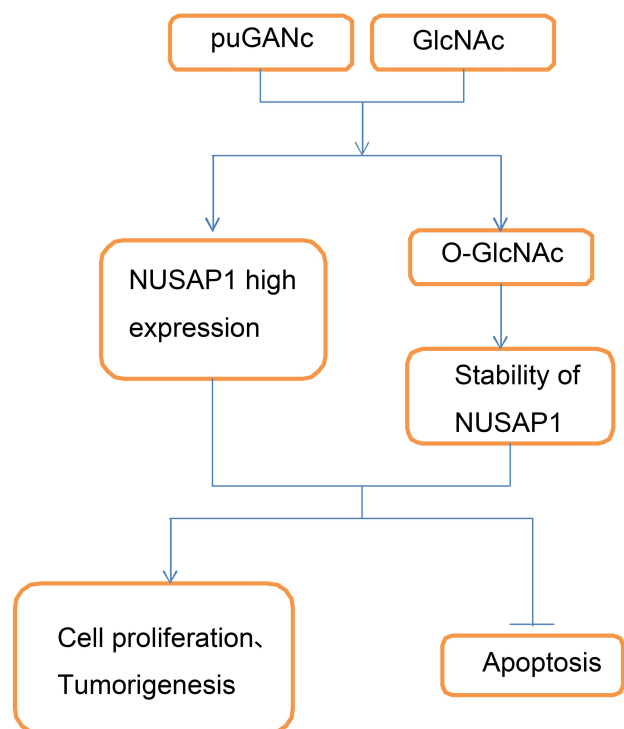


Figure 6 Schematic diagram of adverse effect of hyper-O-GlcNAcylation in the progression of bladder cancer. Up-regulation of O-GlcNAcylation level with PuGANc and GlcNAc treatment promotes the expression and stability of NUSAP1 protein, and accelerates the proliferation and tumorigenesis and represses the apoptosis of bladder cancer cells.

through down-regulation of NUSAP1. Our study proposes a comprehensive theory for the role of O-GlcNAcylation and NUSAP1 in bladder cancer.

Abbreviations

NUSAP1, nucleolar and spindle-associated protein 1; OGT, O-GlcNAc transferase; OGA, O-GlcNAcase.

Data Sharing Statement

The datasets generated in this study are available from WTZ on reasonable request.

Ethics Approval and Consent to Participate

The study was approved by the Ethics Committee at the Tenth People's Hospital of Shanghai. All patients who used the tissue in this study have signed informed consent forms.

Author Contributions

All authors contributed to data analysis, drafting or revising the article, have agreed on the journal to which the

article will be submitted, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors declare that they have no conflicts of interest for this work.

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