MiRNA-1225 Inhibits Osteosarcoma Tumor Growth and Progression by Targeting YWHAZ

Yubao Gong1
Zhengren Wei2
Jianguo Liu1
1Department of Orthopedics, The First Hospital of Jilin University, Changchun 130021, People’s Republic of China; 2Department of Pharmacology, Basic Medical School, Jilin University, Changchun 130021, People’s Republic of China

Introduction: Osteosarcoma is the most common bone tumor and is characterized by the presence of malignant mesenchymal cells produced in the bone stroma. MiRNAs are known to function as post-transcriptional negative regulators of gene expression. Emerging evidence showed that miR-1225-5P functions as a tumor suppressor in several types of cancers. The detailed mechanisms of which miR-1225-5P suppresses tumor growth are not fully understood. The objective of the present study was to test the hypothesis that miR-1225-5P inhibits osteosarcoma cell growth in vitro and tumor growth in vivo by targeting YWHAZ expression.

Methods: Real-time PCR and Western blot were carried out to test the expression of miR-1225-5P and YWHAZ in osteosarcoma cell lines. Luciferase assay was used to demonstrate whether miR-1225-5P targets YWHAZ 3′ UTR. To assess the function of miR-1225-5P in human osteosarcoma cell lines, gain-of-function and loss-of-function of miR-1225-5P were performed by transfecting miR-1225-5P mimic or miR-1225-5P inhibitor into osteosarcoma cell lines. Furthermore, cell cycle analysis was performed to elucidate the possible mechanisms of the action of miR-1225-5P and YWHAZ in human osteosarcoma cells. The potential therapeutic effect of miR-1225-5P was tested in human osteosarcoma xenograft mouse model, by intravenous injection of miR-1225-5P into nude mice. Tumor sizes were measured and lung metastasis was counted after the mice were sacrificed.

Results: The expression of miR-1225-5P was inversely correlated with the expression of YWHAZ in human osteosarcoma cell lines. Database search revealed that miR-1225-5P targeted YWHAZ 3′ UTR. Transfection of miR-1225-5P mimic downregulated YWHAZ expression, which was demonstrated by real-time PCR, Western blot and luciferase assay. Over-expression of miR-1225-5P reduced human osteosarcoma cell growth, migration and invasion by downregulating YWHAZ expression. Cell growth, migration and invasion were increased by inhibiting miR-1225-5P in human osteosarcoma cells. The inhibition of cell growth, migration and invasion was rescued by over-expression of YWHAZ in osteosarcoma cells. Cell cycle analysis revealed that miR-1225-5P inhibited G1/G0 phase exit. In vivo xenograft model demonstrated that miR-1225-5P inhibited in vivo osteosarcoma tumor growth and lung metastasis.

Conclusion: Our findings suggested that miR-1225-5P inhibits osteosarcoma cell growth in vitro and tumor growth in vivo by targeting YWHAZ. This study suggested that miR-1225-5P can serve as a potential therapeutic method for treating osteosarcoma.

Keywords: osteosarcoma, microRNA, miR-1225-5P, YWHAZ, metastasis

Introduction: Osteosarcoma is rare but is the most common histological form of primary bone cancer.1 Most osteosarcomas occur in children and young adults. Teens are the most
commonly affected age group, but osteosarcoma can develop at any age. Despite the advances of osteosarcoma treatment in recent years, metastasis remains a hurdle. The 10-year survival rate for patients with metastatic osteosarcoma is 24.0% compared with 65.8% for patients with local/regional disease.

YWHAZ, also known as Tyrosine 3-Monoxygenase/TPmt 5-Monoxygenase Activation Protein Zeta, or 14-3-3ζ, is a member of the 14-3-3 protein family. The 14-3-3 protein family is a group of highly conserved proteins. They are involved in many vital cellular processes such as metabolism, protein trafficking, signal transduction, apoptosis and cell cycle regulation. Emerging evidence shows that YWHAZ serves a critical role in many different types of tumor progression, such as prostate cancer, breast cancer, head and neck squamous cell carcinoma, gastric cancer, and lung cancer. Aberrant expression of YWHAZ in cancer cells is believed to serve a critical role in tumorigenesis. YWHAZ may inactivate tumor suppressors such as p53 and directly bind to PI3 kinase, promote Akt activation and enhance cell survival. Therefore, targeting YWHAZ as a potential cancer therapeutic method has been proposed.

MicroRNAs are a group of small non-codingRNA molecules that contain about 22 nucleotides, and function in RNA silencing and post-transcriptional regulation of gene expression. Numerous miRNAs have been found differentially expressed in osteosarcoma compared with normal bone tissue. MicroRNAs have been reported to inhibit cancer cell proliferation by targeting YWHAZ, such as miR-204-5P in esophageal squamous cell carcinoma, and miR-902 in epithelial ovarian cancer.

In this study, we found that miR-1225-5P negatively regulates YWHAZ expression in osteosarcoma cells and inhibits osteosarcoma cell growth in vitro and in vivo. These findings may provide a new avenue towards osteosarcoma treatment.

**Materials and Methods**

**Osteosarcoma Cell Lines and Culture**

All osteosarcoma cell lines (HOS, MG63, U2OS Saos-2 and hFOB1.19) used in this study were purchased from ATCC (ATCC.org). All the cells were cultured in RPMI-1640 medium (HyClone) containing 10% FBS (Gibco) at 37°C, 5% CO2.

**Cell Transfection**

HOS or U2OS cells were cultured as described above. Before transfection, the cells were plated in 6-well plates at the density of 5×10^5 cells/well and cultured at 37°C, 5% CO2 for 16 h. For miRNA transfection, the cells were transfected with irrelevant miRNA control (5'-UUCUCCGAACG UGUACGUTT-3', 100 pmol/well), miR-1225 mimic (5'- GUGGGUACGGCCCAGUGG GG-3', 100 pmol/well), or miR-1225 inhibitor (5'-CCCCCACUGGGCCGUC CCAC-3', 100 pmol/well) by using Lipofectamine 2000 (Life Technologies) and following manufacturer’s instructions. For Ywhaz over-expression, the cells were transfected with expression vector containing Ywhaz coding sequence, or pcDNA3 vector control (1.5 μg/well). Coding sequence of the Ywhaz gene in the expression vector was confirmed by sequencing (Supplemental data).

**RNA Isolation and Real-Time PCR**

To isolate total RNA, cells or tumor tissues were placed in 1 mL Trizol reagent (Invitrogen) and homogenized with Fluho homogenizer for 20 s. Total RNA was then isolated using Trizol reagent and following manufacturer’s protocol. First strand cDNA was synthesized using RevertAid First Strand cDNA Synthesis Kit (Fermentas) and following the kit protocol. The first strand cDNA samples were used as a template for quantitative real-time PCR to quantify the expression of mRNA by using SYBR green (Thermofisher) and gene specific primers (listed in Table 1).

**Cell Growth Assay with Cell Counting Kit-8 (CCK8)**

Transfected HOS or U2OS osteosarcoma cells were plated in 96-well plates at a density of 3000 cells/well in complete growth medium. The cells were cultured at 37°C, 5% CO2 for 12, 24, 48 and 72 h. CCK8 was diluted in serum-free medium (1:10). At each time point, 100 μL of diluted CCK8 was added to each well and incubated at 37°C, 5% CO2 for 1 h. OD 450 nm was measured in a plate reader (Pulang New Technology, Beijing).

**Wound Healing Assay**

Transfected HOS or U2OS osteosarcoma cells were plated in 6-well plates at a density of 8×10^5/well in complete growth medium and cultured until confluence. A gap in the cells was created by scratching the cells with a 1000 μL tip after removing the growth medium. The loose cells were washed away with phosphate buffered saline (PBS), and the cells were further cultured in complete growth medium for 12 and 24 h. After taking pictures of the gaps, the size of the gap was measured for each sample.
Transwell Cell Invasion Assay

In order to analyze the capability of the osteosarcoma cells to invade, transfected HOS or U2OS cells were plated in the upper chamber of the transwells (Corning) at a density of 6×10⁶/well in 300 μL 1% FBS medium, and the upper chamber inserted into the lower chamber containing 700 μL growth medium. The cells were cultured at 37°C, 5% CO₂ for 24 h. The low FBS medium was removed from the upper chamber. The cells in the lower chamber were washed with PBS and fixed with 4% formalin at room temperature for 10 min. The cells were then stained with 0.5% Crystal Violet at room temperature for 30 min and washed with PBS 3 times. The number of cells that had migrated into the lower chamber was scored by taking a picture of the stained cells under microscope and the number of cells counted in each picture.

Dual Luciferase Assay

The whole 3’ UTR sequence of YWHAZ gene (NCBI accession number: NM_00135699, DNA sequence is shown in the supplemental data) was synthesized and cloned into pGL3-promoter-Luc2 vector (Promega) to obtain pGL3-wt. A mutant 3’ UTR sequence (DNA sequence is shown in the supplemental data) was created to eliminate the putative miR-1225-5P binding site and cloned into pGL3-Promoter-Luc2 vector to obtain pGL3-mut. HOS osteosarcoma cells were plated in 6-well plates and cultured at 37°C, 5% CO₂ for 16 h. The cells were then co-transfected with combinations of (a) 1.5 μg of pGL3-wt, 20 ng of pGLI-TK (Promega), and 100 pmol of scramble miRNA; (b) 1.5 μg of pGL3-wt, 20 ng of pGLI-TK, and 100 pmol of miR-1225; (c) 1.5 μg of pGL3-mut, 20 ng of pGLI-TK, and 100 pmol of miR-1225, using Lipofectamine 2000 and following the kit protocol. The cells were washed with PBS and lysed with lysis buffer 48 h after transfection. Luciferase activity was analyzed with dual luciferase assay kit (Promega).

Western Blot Analysis

The cells on the plate were washed with ice cold PBS twice and lysed with radioimmunoprecipitation assay buffer (RIPA buffer). Tumor tissues were washed with ice cold PBS and 20 μg of each sample were homogenized with a homogenizer on ice. After centrifugation at 12,000 rpm, 4°C to remove the debris, protein concentration was measured with Pierce BCA Protein Assay Reagent A (Thermo Fisher Scientific US, Cat# 23223). Approximately 25 μg (from cells) or 50 μg (from tissue) of protein was loaded into each well and separated by sodium dodecyl sulphate-polyacrylamide gel electrophoresis in a 10% polyacrylamide gel and transferred to immunoblot nitrocellulose (NC) membranes (Millipore). The membranes were incubated with primary antibody against YWHAZ (Abcam, Cat# ab51129, 1:500 dilution) or GAPDH (Cell Signaling Technology, Cat# 5174, 1:1000 dilution), followed by incubation with horseradish peroxidase conjugated secondary antibody. ECL Western blot analysis was used to detect the binding of primary and secondary antibodies in the Tanon 5200CE Chemi-image system (Abclonal, Woburn MA, USA).

In vivo Tumor Model

All in vivo tumor model studies were approved by the Jilin University Institutional Animal Care and Use Committee, and followed the guidelines adapted from the “Animal Welfare Act” (the federal law governing the care and use of laboratory animals in the United States). HOS osteosarcoma cells were cultured in 100 mm tissue culture dishes until about 90% confluent and then were harvested from culture dishes with 0.05% Trypsin (Life Technology). The cells were re-suspended in PBS at a final concentration of 2×10⁷ cells/mL. Six-week-old female Balb/c athymic nude mice were obtained from Shanghai Experimental Animal Center of Chinese Academy of Sciences and were bred in a special pathogen-free (SPF) grade laboratory. The mice were randomly divided into 2 groups with 6 mice in each group. Each mouse received 100 μL of cells (2×10⁶ cells) injected subcutaneously. The mice were monitored closely every day. The tumor sizes were measured when they started to be visible and every 3 days thereafter. The volume of the tumor was calculated by using the formula:

<table>
<thead>
<tr>
<th>Gene Name</th>
<th>Forward Primer Sequence</th>
<th>Reverse Primer Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>miR-1225-5P</td>
<td>GGTGGTGACGGCAGGTATT</td>
<td>AGTCAGGGTGCGGAGTATT</td>
</tr>
<tr>
<td>U6</td>
<td>GTGCGCTCTGCGGACA</td>
<td>AACGCTCACGAAATTGGT</td>
</tr>
<tr>
<td>YWHAZ</td>
<td>ATGACGAGAAAGGGATG</td>
<td>AGAAGTGAAGGGCCAGAC</td>
</tr>
<tr>
<td>GAPDH</td>
<td>GGATTGTCTGGCAAGTCCCC</td>
<td>ATTTGGAAAGGCGAGGAG</td>
</tr>
</tbody>
</table>

Table I Real-Time PCR Primers Sequence
V = (4/3) × \pi × (L/2) × (L/2) × (D/2) (L means length and D means depth). Lung metastasis was examined by dissecting the whole lungs. Quantification of the metastasis was conducted by counting the number of tumors on the lung surface. After counting the metastasis tumor numbers in the lungs, the lungs were fixed in 10% formalin and embedded in paraffin for tumor histology.

**Cell Cycle Profile Analysis**
Transfected HOS or U2OS cells were trypsinized from culture dishes and washed with ice cold PBS twice. After removing PBS, the cells were fixed with ice cold 70% ethanol at 4°C for 24 h. The cells were then washed with PBS and stained with 20 μg/mL propidium iodide (Cell Cycle and Apoptosis Analysis kit, Beyotime) in 0.1% Triton-X 100/PBS with 0.2 mg/mL RNase A at a final concentration of 2×10^6 cells/mL at 37°C for 30 min. After staining, the cells were kept on ice and subject to FACS analysis (Beckman CytoFLEX flow cytometer). The cell cycle profile data were analysed using FlowJo software.

**Results**

**miR-1225-5P Targeted 3’ UTR of YWHAZ Gene and Inhibited YWHAZ Expression in Osteosarcoma Cells**
In multiple osteosarcoma cell lines, miR-1225-5P expression was detected by real-time PCR (Figure 1A). The level of expression varies from cell line to cell line. Interestingly, the expression of Yw haz gene on the mRNA level in the same set of cell lines was roughly conversely correlated to the expression of miR-1225-5P. For instance, among the 5 cell lines tested, the cell lines that express the highest and the lowest miR-1225-5P are HOS and hFOB1.19, respectively (Figure 1A), and the expression of Yw haz in HOS and hFOB1.19 cell lines were the lowest and the highest, respectively (Figure 1B). Western blot confirmed the finding of reverse-correlation of miR-1225-5P and YWHAZ expression on the protein level (Figure 1C). This finding indicates that miR-1225-5P can potentially be a novel negative regulator of YWHAZ expression. To confirm this hypothesis, we performed a database search ([www.mirbase.org](http://www.mirbase.org)) and found that miR-1225-5P potentially targets 3’ UTR of human Yw haz gene by alignment of sequences (Figure 1D). This was further validated by transfecting miR-1225-5P mimic into HOS osteosarcoma cells: increasing miR-1225-5P significantly inhibited Yw haz expression (Figure 1E). Inhibiting miR-1225-5P significantly increased Yw haz expression in U2OS osteosarcoma cells (Figure 1E). To further confirm that miR-1225-5P targets Yw haz gene 3’ UTR sequence, we constructed luciferase reporter plasmid by cloning 3’ UTR of Yw haz gene to 3’ of luciferase reporter gene. The luciferase reporter assay showed that miR-1225-5P downregulated luciferase activity by targeting Yw haz gene 3’ UTR (Figure 1F). Luciferase activity was not reduced when the miR-1225-5P targeting sequence was mutated (Figure 1F). These data indicate that miR-1225-5P is a novel negative regulator of Yw haz expression.

**MiR-1225-5P Inhibited Osteosarcoma Cells’ Growth and Migration in vitro**
Targeting Yw haz expression has been shown to inhibit other cancer cells’ proliferation and migration.20-22 Here we asked the question of whether miR-1225 inhibits osteosarcoma cells’ growth and migration by negatively regulating Yw haz expression. As observed above, HOS osteosarcoma cells transfected with miR-1225 mimic significantly reduced expression of Yw haz (Figure 1E). HOS osteosarcoma cells transfected with miR-1225 5P mimic showed significantly slower growth compared with the cells transfected with scramble miRNA (Figure 2A). Wound healing assay further showed that the miR-1225-5P mimic transfected cells migrate significantly slower than control cells transfected with scramble miRNA (Figure 2B and C). In order to address the question of whether miR-1225-5P inhibits osteosarcoma cells invasion, we used a transwell cell invasion assay. HOS osteosarcoma cells transfected with miR-1225-5P or scramble miRNA control were plated in the upper chamber of the transwell. After being cultured for 24 h, the cells that migrated through the transwell into the lower chamber of the plate were fixed, stained and the cell number was counted after imaging. As shown in Figure 2D and E, miR-1225-5P significantly inhibited osteosarcoma cells invasion. The cell number collected in the lower chamber from miR-1225-5P transfected cells was 18±4, which is about a 86% reduction compared with the control scramble miRNA transfection cells (116±6). These data indicate that miR-1225-5P inhibited osteosarcoma cells’ growth, migration, and invasion by downregulating Yw haz expression.

**Inhibiting miR-1225-5P Promoted Osteosarcoma Cells Growth and Migration**
Since increasing miR-1225-5P in osteosarcoma cells inhibited cell growth and migration, we further asked whether inhibiting miR-1225-5P will promote osteosarcoma cells’
growth and migration. Inhibiting miR-1225-5P significantly increased YWHAZ expression on the protein level in U2OS osteosarcoma cells (Figure 1E). To address this question, we transfected miR-1225-5P inhibitor into U2OS osteosarcoma cells with scramble inhibitor as a control. The U2OS cells transfected with miR-1225-5P grew significantly faster compared with the cells transfected with scramble (Figure 3A). Inhibiting miR-1225-5P increased U2OS cells’ wound healing capacity (Figure 3B and C). Inhibiting miR-1225-5P also promoted U2OS cell migration through transwells (Figure 3D and E). These data further proved that miR-1225-5P regulates osteosarcoma cells proliferation, migration, and invasion by targeting YWHAZ expression.

miR-1225-5P Regulated Osteosarcoma Cells Proliferation by Affecting G1/G0 Phase Exit

We further asked, what is the mechanism of miR-1225-5P in regulating osteosarcoma cell proliferation through YWHAZ? After HOS or U2OS cells were transfected with miR-1225-5P mimic or inhibitor, the cells were fixed and analyzed with cell cycle profile by DNA
As shown in Figure 4A and B, HOS transfected with miR-1225-5P mimic significantly increased percentage of cells in G1/G0 phase (68%) compared with those transfected with scramble (52%). However, inhibiting miR-1225-5P in U2OS cells significantly decreased the percentage of cells in G1/G0 phase (35%) compared with control (53%) (Figure 4C and D). These data indicate that miR-1225-5P regulates osteosarcoma cell proliferation by affecting G1/G0 phase exit via regulating YWHAZ expression.
Over-Expression of YWHAZ Attenuated the Inhibition Effect of miR-1225-5P in Osteosarcoma Cells’ Proliferation, Migration and Cell Cycle Progression

To further elucidate that miR-1225-5P regulates osteosarcoma cell proliferation and migration by targeting YWHAZ, we used YWHAZ gain-of-function strategy. HOS osteosarcoma cells were transfected with pCDNA3.1 expression vector containing Ywhaz coding sequence (Ywhaz/pCDNA) combined with miR-1225-5P mimic or scramble. HOS cells transfected with pCDNA3.1 empty vector combined with miR-1225-5P mimic or scramble were used as controls. Therefore, 4 groups of cells (Vector + Scramble, Vector + mimic, Ywhaz + Scramble, and Ywhaz + mimic) were analyzed for cell proliferation, migration, invasion, and cell cycle profile. Western blotting analysis confirmed over-expression of YWHAZ in YWHAZ + Scramble HOS cell group (Figure 5A). Western blotting analysis also confirmed downregulated YWHAZ expression in Vector + mimic HOS cell group (Figure 5A). In these 4 groups of HOS osteosarcoma cells, the expression level of YWHAZ correlated with the cell growth. Cells over-expressed with YWHAZ (YWHAZ + Scramble group) proliferated significantly faster than Vector + Scramble and Vector + mimic groups (Figure 5B). It was also further confirmed that knocking down YWHAZ in Vector + mimic group inhibited cell growth (Figure 5A and B). These data hinted that YWHAZ promotes osteosarcoma cell proliferation. In the wound healing assay, the gap sizes of YWHAZ + Scramble group were significantly smaller than Vector + Scramble and Vector + mimic groups (Figure 5C and D). In the transwell invasion assay, significantly higher numbers of cells were observed in the lower chamber in YWHAZ + Scramble group than in Vector + Scramble group (Figure 5E). Knocking down of YWHAZ in Vector + mimic group significantly inhibited cell invasion which confirmed our observation in Figure 2D. In the cell cycle profile analysis, YWHAZ + Scramble group cells had a significantly lower percentage of G1/G0 phase cells (30.16±1.54%) compared with the other 3 groups of cells and Vector + mimic group cells increased percentage of G1/G0 arrested cells (68.23±1.41%) compared with that of control Vector + Scramble group cells (50.88±2.49%) (Figure 5F and G). Of note, cotransfection of YWHAZ and miR-1225-5P mimic significantly reduced the effects of YWHAZ over-expression. These data indicated that miR-1225-5P negatively regulates osteosarcoma cells proliferation, migration, invasion, and cell cycle progression by (at least in part) targeting YWHAZ.

miR-1225-5P Inhibited Osteosarcoma Tumor Growth and Metastasis in vivo

Our finding that miR-1225-5P inhibited osteosarcoma cells’ proliferation, migration, and invasion by targeting YWHAZ expression and induced G1/G0 cell cycle arrest in vitro, leads us to further hypothesize that miR-1225-5P inhibits osteosarcoma tumor growth in vivo. To test this hypothesis, we established an in vivo tumor model by injecting HOS osteosarcoma cells subcutaneously in nude mice. The mice were intravenously injected with miR-1225-5P mimic or scramble miRNA daily for 3 weeks. The tumor sizes were measured every 3 days. The mice injected with miR-1225-5P mimic showed significantly slower tumor growth compared with those injected with scramble miRNA (Figure 6A). Thirty-three days after the osteosarcoma cell inoculation, the mice were sacrificed and the tumors were dissected. H&E staining confirmed that the dissected tumor tissues were osteosarcoma (data not shown). As shown in Figure 6B, the tumors from mice injected with miR-1225-5P mimic were significantly smaller compared with those injected with scramble miRNA (Figure 6B). Real-time PCR analysis and Western blot revealed that the expression of YWHAZ on both mRNA and protein levels were significantly lower in the tumors that received miR-1225-5P mimic IV injection compared with those that received scramble miRNA injection (Figure 6C and D), indicating that miR-1225-5P injection reduced YWHAZ expression in vivo. Lung metastasis of osteosarcoma was assayed by dissecting whole lungs and counting tumor numbers from the lungs of each mouse. After counting the lung metastasis, the lungs were fixed in formalin and embedded in paraffin. Paraffin sections and H&E staining of the lungs confirmed metastatic tumors in lungs (supplemental Figure S1). The mice injected with miR-1225-5P mimic showed a significantly lower number of tumors in the lungs compared with those injected with scramble miRNA (Figure 6E). These data indicate that miR-1225-5P not only inhibited tumor growth in vivo, but also inhibited metastasis by downregulating expression of YWHAZ in osteosarcoma.
Discussion

Osteosarcoma is the most common primary malignant of the bone. Patients with metastasis or relapse experience poor prognosis and low overall survival rate.\textsuperscript{2,3} The mechanisms of osteosarcoma malignancy and metastasis are not fully understood. Abnormal miRNA expression may be a possible part of malignant transformation mechanisms of osteosarcoma.\textsuperscript{17,24} Using data mining, Zhang et al. found that aberrant expression of miR-1225-5P in osteosarcoma and low expression was associated with poor prognosis.\textsuperscript{25} In the present study, we found that the higher expression of YWHAZ was reversely correlated with the expression of miR-1225-5P. We further
discovered that miR-1225-5P regulates YWHAZ expression by targeting YWHAZ 3′ UTR. In vitro evidence demonstrated that miR-1225-5P negatively regulates osteosarcoma cells’ growth, migration and invasion via negatively regulating YWHAZ expression in osteosarcoma cell lines.

Accumulating evidence shows that YWHAZ serves a role in many types of cancers and promotes cancer metastasis. Targeting YWHAZ has been suggested as an option for cancer treatment. YWHAZ was reported to be involved in cancer cell cycle regulation, especially in G1/G0 phase to S phase transition. Our in vitro evidence showed that inhibiting YWHAZ expression by overexpression of miR-1225-5P increased osteosarcoma cell G1/G0 arrest, inversely, over-expression of YWHAZ reduced G1/G0 cell percentage in osteosarcoma cell line. These data suggest that YWHAZ plays a critical role in osteosarcoma cells’ cell cycle regulation, namely G1/G0 phase to S phase transition.

MiR-1225-5P has been reported as a tumor suppressor in thyroid cancer, pancreatic cancer, glioblastoma, laryngeal cancer, and gastric carcinoma and osteosarcoma by targeting different genes and through different pathways. Zhang and colleagues’ in vitro work revealed that miR-1225-5P inhibits SOX9 expression in the osteosarcoma cell line and acts as a suppressor. In the present study, we found that miR-1225-5P negatively regulates YWHAZ post-transcriptional expression by targeting its 3′ UTR. Through inhibiting YWHAZ expression, miR-1225-5P inhibits osteosarcoma cells’ growth, migration and invasion. Our in vivo work further revealed that intravenous injection of miR-1225-5P inhibited tumor growth and
Figure 5 Overexpression of YWHAZ rescued inhibition of osteosarcoma cell growth, migration, and invasion caused by miR-1225-5P. (A) Western blot showed that overexpression of YWHAZ partially restored YWHAZ protein level in HOS osteosarcoma cells (lane 4). YWHAZ expression was knocked down by miR-1225-5P mimic (lane 2). (B) Over-expression of YWHAZ increased HOS cells growth (blue line and grey line, P<0.01 n=3), and rescued the inhibition of HOS cell growth (yellow line and red line, P<0.001 n=3) caused by miR-1225-5P mimic. (C) Quantification of HOS cells wound gap after 12 and 24 h culture. (D) Representative pictures of HOS cells wound gap after 12 and 24 h culture showed that over-expression of YWHAZ accelerated wound healing in the cell culture dishes. (E) Transwell invasion assay showed that inhibition of YWHAZ expression by miR-1225-5P mimic decreased HOS cells invasion and over-expression of YWHAZ increased HOS cells invasion. (F) Representative FACS histogram plots showed that YWHAZ over-expression decreased G1/G0 phase arrest induced by miR-1225-5P. (G) Average percentage of each phase in cell cycle (n=3 each group) showed that YWHAZ over-expression decreased G1/G0 phase arrest induced by miR-1225-5P. **Indicates P<0.01 in the indicated comparison, ***indicates P<0.001 in the indicated comparison.
To our knowledge, this is the first report that demonstrates miR-1225-5P inhibits osteosarcoma growth and metastasis in a mouse model.

Taken together, our present study revealed that miR-1225-5P negatively regulates YWHAZ expression in osteosarcoma cells, and inhibits osteosarcoma cells’ growth, migration and invasion in vitro. Moreover, our in vivo mouse model further proved that miR-1225-5P inhibits tumor growth and metastasis. This study provided evidence that miR-1225-5P could potentially be used as an osteosarcoma treatment therapy.

**Patient Consent for Publication**  
No patients are involved.

**Ethics Approval and Consent to Participate**  
This study does not contain any studies with human participants performed by any of the authors.
All in vivo tumor mouse model studies were approved by the Jilin University Institutional Animal Care and Use Committee, and followed the guidelines adapted from the “Animal Welfare Act” (the federal law governing the care and use of laboratory animals in the United States).

Author Contributions

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work.

Disclosure

The authors have no competing interests.

References


