

Down-Regulation of Circ_0032833 Sensitizes Colorectal Cancer to 5-Fluorouracil and Oxaliplatin Partly Depending on the Regulation of miR-125-5p and MSI1

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Shouchao Li
Sheng Zheng

Department of Anorectal Surgery,
Weifang People's Hospital, Weifang
261000, People's Republic of China

Background: 5-fluorouracil, leucovorin, and oxaliplatin (FOLFOX) is an effective chemotherapy for colorectal cancer (CRC) in clinic. It remains unclear regarding the effect of circular RNA (circRNA) circ_0032833 on regulating chemosensitivity in CRC.

Methods: Drug resistance analysis was performed by Cell Counting Kit-8 (CCK-8) assay. All RNA and protein levels were, respectively, measured via quantitative real-time polymerase chain reaction (qRT-PCR) and Western blot. Cellular colony capacity, apoptosis and metastasis were evaluated using colony formation assay, Annexin-FITC/PI flow cytometry and transwell migration/invasion assays. The molecular combination was notarized using dual-luciferase reporter and RNA immunoprecipitation (RIP) assays. The in vivo experiment was conducted via xenograft tumors in mice.

Results: Circ_0032833 was significantly up-regulated in FOLFOX-resistant CRC and associated with drug resistance. Knockdown of circ_0032833 could sensitize FOLFOX-resistant CRC cells to 5-fluorouracil and oxaliplatin. Circ_0032833 was a miR-125-5p sponge, and miR-125-5p overexpression was responsible for the effect of circ_0032833 knockdown on 5-fluorouracil and oxaliplatin sensitivities. Besides, miR-125-5p targeted Musashi1 (MSI1) to increase the susceptibility of 5-fluorouracil and oxaliplatin in FOLFOX-resistant CRC cells. We found that circ_0032833 generated the regulation on MSI1 by sponging miR-125-5p. Circ_0032833 down-regulation also promoted the 5-fluorouracil and oxaliplatin sensitivities partly through the miR-125-5p/MSI1 axis in vivo.

Conclusion: This study illuminated an unambiguous mechanism circ_0032833/miR-125-5p/MSI1 on regulating 5-fluorouracil and oxaliplatin sensitivities in FOLFOX therapy, maybe providing a deep insight of resistance formation and developing a novel strategy to enhance chemosensitivity in CRC.

Keywords: circ_0032833, colorectal cancer, chemosensitivity, miR-125-5p, MSI1

Introduction

Colorectal cancer (CRC) is the third most common cancer (10.2% of the total cases), closely following the lung cancer (11.6) and breast cancer (11.6%) for men and women.¹ 5-fluorouracil, leucovorin, and oxaliplatin (FOLFOX) has been developed as the first-line chemotherapy, especially in metastatic or advanced and recurrent CRC.²⁻⁴ But the resistance of 5-fluorouracil and oxaliplatin usually results in a poor response to FOLFOX chemotherapy.⁵ To improve the anti-tumor response

Correspondence: Sheng Zheng
Email Zhengsheng966@163.com

of FOLFOX therapy, knowing the occurrence mechanism of drug resistance and seeking effective biomarkers are imperative. As the well-known post-transcriptional regulators, non-coding RNAs (ncRNAs) have been recognized to mediate the chemoresistance of FOLFOX potentially.⁶

Non-coding circular RNAs (circRNAs) characterized by the specific covalent closed-loop without 5' and 3' free ends are stably enriched in many cancer cells as novel regulatory molecules.⁷ Recently, a number of differentially expressed circRNAs have been reported to be related to chemoresistant CRC.⁸ Hon et al have definitely identified that circ_0032833 was expressed with ectopic high level in FOLFOX-resistant CRC patient serum and cells.⁹ We will further explore the role of circ_0032833 on chemosensitivity in CRC.

MicroRNAs (miRNAs) mainly bind to 3' untranslated regions (3'UTRs) of pre-messenger RNA to function as regulators in multiple cancers.^{10,11} Han et al found that miR-552 played a pivotal role in drug response of liver tumor,¹² and abundant miRNAs have been used as diagnostic biomarkers for 5-fluorouracil chemoresistance in CRC.¹³ MiR-125 was a tumor inhibitor in CRC¹⁴ and its overexpression could promote the sensitivity of 5-fluorouracil in hepatocellular carcinoma cells.¹⁵ However, the effect of miR-125-5p on FOLFOX-resistant CRC is unclear.

Musashi1 (MSI1) is involved in various biological behaviors to regulate tumorigenesis, progression and chemoresistance.¹⁶ MSI1 was shown to enhance drug resistance in pediatric glioblastoma cells¹⁷ and non-small cell lung cancer cells.¹⁸ In addition, MSI1 could sensitize CRC cells to 5-fluorouracil.¹⁹ Here, the function of MSI1 on oxaliplatin sensitivity of CRC cells will also be addressed.

In different cancers, circRNAs can regulate gene expression via acting as "miRNAs sponges".²⁰ Thus, the relation among circ_0032833, miR-125-5p and MSI1 is set as another key point of our research.

Materials and Methods

Cell Culture and Establishment of FOLFOX-Resistant CRC Cells

HCT116 (a human CRC cell line) was bought from American Type Culture Collection (ATCC, Manassas, VA, USA), and maintained in McCoy's 5A medium (Sigma-Aldrich, St. Louis, MO, USA) complemented with 10% fetal bovine serum (FBS; Sigma-Aldrich) and

1% antibiotic solution (10,000 unit/mL penicillin and 10 mg/mL streptomycin; Sigma-Aldrich). Temperature of 37°C, CO₂ of 5% and 95% air were set as the standard conditions for HCT116 cell growth.

FOLFOX-resistant HCT116 (HCT116-R) cells were obtained by exposing the parental HCT-116 cells (HCT116-P) to 5-fluorouracil and oxaliplatin (5-FU and Oxa; Sigma-Aldrich) at clinically relevant doses as previously described,²¹ and finally cultivated in normal culture medium with a low dose of FOLFOX (5 μ M 5-FU and 0.125 μ M Oxa).

Drug Resistance Assay

Drug resistance was evaluated by the half inhibitory concentration (IC₅₀) via cell viability assay in HCT116-P and HCT116-R cell lines treated with 5-FU or Oxa. Firstly, 3×10^3 cells or transfected cells were seeded into each well of 96-well plates to incubate for 24 h, followed by another 48 h incubation in presence of different concentrations of 5-FU (1 μ M, 5 μ M, 10 μ M, 20 μ M, 40 μ M, 60 μ M, 80 μ M) and Oxa (1 μ M, 2 μ M, 5 μ M, 10 μ M, 20 μ M, 40 μ M, 60 μ M). Whereafter, cells were added with 10 μ L/well Cell Counting Kit-8 solution (Sigma-Aldrich) for 4 h, and the absorbance was determined by a microplate reader at 450 nm. The IC₅₀ value was obtained according to the drug concentration at 50% cell viability.

Clinical Tissue Samples

Clinical specimens were approved to be used for this research by the Ethical Committee of Weifang People's Hospital. Fifty CRC patients who have received FOLFOX therapy were finally subjected to surgical resection at Weifang People's Hospital. According to the Response Evaluation Criteria In Solid Tumors (RECIST) criteria: complete response (CR), partial response (PR), stable disease (SD) and progressive disease (PD), these 50 patients were classified into Responders (n=25, CR+PR) and Nonresponders (n=25, SD+PD) toward FOLFOX therapy. All patients signed the written informed consent and the collected tissues were conserved in liquid nitrogen.

RNA Extraction and Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR) Analysis

Our RNA extraction was performed using Trizol (Beyotime, Shanghai, China). FastKing one-step reverse transcription kit (TIANGEN, Beijing, China) was used for

complementary DNA (cDNA) synthesis and real-time PCR was implemented via FastFire qPCR PreMix (TIANGEN) in terms of the users' manuals. As for data analysis, the comparative cycle threshold method ($2^{-\Delta\Delta C_t}$) was used for calculating the relative expression levels. The used housekeeping genes were glyceraldehyde-3-phosphate dehydrogenase (GAPDH; for circ_0032833 or MSI1) and U6 (for miR-125-5p). All forward (F) and reverse (R) primer sequences were presented as below: circ_0032833: 5'-CATCAATGCTGAGGAGCAGA-3' (F) and 5'-GCCAAGGGCAGTACTCAGAA-3' (R); miR-125-5p: 5'-GGGTCCGAGGTATTCGCACT-3' (F) and 5'-TCCTGAGACCCTTTAACCTGTG-3' (R); MSI1: 5'-GC GACTGTCTGGACAGGAATTA-3' (F) and 5'-AGAGGGACACACAGAAGGGGGAT-3' (R); GAPDH: 5'-AGAAGGCTGGGGCTCATTTG-3' (F) and 5'-AGGGGC CATCCACAGTCTTC-3' (R); U6: 5'-CTCGCTTCGGCAGCACA-3' (F) and 5'-AACGCTTCACGAATTTGCGT-3' (R).

Western Blot

Cells in 24-well plates were washed with ice-cold phosphate buffer solution (PBS; Beyotime) and the protein was extracted from the harvested cell pellets using radio-immunoprecipitation assay buffer (RIPA; Beyotime) added with Phenylmethanesulfonyl fluoride (PMSF; Beyotime). Human tissues were cut into small pieces and sufficiently lysed in RIPA+PMSF solution, followed by the centrifugation at 12,000 g for 5 min to obtain the supernatant protein. Next, the separation of protein from cells or tissues was conducted by the electrophoresis on TruPAGE™ Precast Gels (12%, 10 × 10 cm, 12-well; Sigma-Aldrich) for 120 min and separated proteins were electrically moved onto the PVDF Transfer Membrane (0.2 μm; Beyotime). After immersing in 5% (w/v) non-fat milk (Beyotime) in tris-buffered saline with 0.1% Tween-20 (TBST) overnight, the membranes were incubated in primary antibodies diluted with TBST at room temperature for 4 h and then washed in TBST for three times (5 min/time). The membranes were incubated with diluted secondary antibody for 1 h and washed in TBST, then ECL Substrate Kit (Abcam, Cambridge, UK) and ImageLab software version 4.1 (Bio-Rad, Hercules, CA, USA) were applied for analyzing protein density. The antibodies (Abcam) and dilutability were as follows: anti-multidrug resistance-associated protein 1 (anti-MRP-1; ab234098, 1:1000), anti-myeloid cell leukemia-1 (anti-MCL-1; ab32087, 1:1000), anti-caspase-3 (ab44976,

1:1000), anti-N-cadherin (anti-N-cad; ab18203, 1:1000), anti-E-cadherin (anti-E-cad; ab15148, 1:1000), anti-MSI1 (ab21628, 1:1000), anti-GAPDH (ab9485, 1:2500) and Goat Anti-Rabbit IgG H&L (HRP) (ab205718, 1:5000).

Cell Transfection

The small interfering RNA (siRNA), short hairpin RNA (shRNA) and miRNA-related oligonucleotides were all purchased from GenePharma (Shanghai, China): siRNA targeting circ_0032833 (si-circ), shRNA targeting circ_0032833 (sh-circ), miR-125-5p mimic, miR-125-5p inhibitor and corresponding negative controls (si-NC, sh-NC, mimic NC and inhibitor NC). The overexpression vector of MSI1 (oe-MSI1) was bought from GENEWIZ (Suzhou, China). Cell transfection was then administrated using Lipofectamine3000 reagent (Invitrogen, Carlsbad, CA, USA) as per the provided instruction book.

Colony Formation Assay

3×10^2 cells in 2 mL complete media were seeded into the 6-well plates, and cultured in standard growth condition for two weeks. After macroscopic colonies were fixed and stained in 4% paraformaldehyde (Beyotime) and crystal violet (Beyotime), 2 mL PBS was exploited for washing out excess dye and the clonal number was counted.

Annexin V-FITC/PI Flow Cytometry

Annexin-FITC Apoptosis detection kit (Solarbio, Beijing, China) was used to examine apoptotic cells, referring to the producer's specification. Briefly, 100 μL cell suspension in 1 × Binding Buffer (1×10^5 cells) was pipetted into a 2-mL tube, followed by adding 5 μL Annexin V-FITC for 10 min and propidium iodide (PI) for 5 min in the dark. Afterwards, the tube was supplemented with PBS to 500 μL and the apoptotic cells were instantly detected via the flow cytometer (BD Biosciences, San Diego, CA, USA). The (Annexin V+/PI-) labelled cells were considered as early apoptotic cells and (Annexin V+/PI+) labelled cells were as late apoptotic cells. Then, cell apoptosis rate (apoptotic cells/total cells × 100%) was calculated.

Transwell Assay

A transwell chamber (Corning Inc., Corning, NY, USA) was used for migration determination while the matrigel (Corning Inc.)-coated chamber was applied to analyze invasion ability. Harvested cells were resuspended in medium without serum, and added to the upper chamber. The migrated or invaded phenomenon could occur after adding

the medium containing 10% FBS into the lower chamber. After cells passed into the lower chamber were fastened by 4% paraformaldehyde and dyed in crystal violet, the images of migrated and invaded cells were obtained by an inverted microscope (Olympus, Tokyo, Japan) and the number was counted.

Dual-Luciferase Reporter Assay

The wild-type (wt) sequences of circ_0032833 and MSI1 3'UTR were respectively cloned into the pGL3 luciferase empty vector (Promega, Madison, WI, USA) to construct circ_0032833 wt and MSI1 3'UTR wt luciferase plasmids. After the miR-125-5p binding sites in wt sequences of circ_0032833 and MSI1 3'UTR were mutated, the mutant-type (mut) sequences were used to obtain the novel plasmids circ_0032833 mut and MSI1 3'UTR mut. These constructed plasmids were respectively transfected into 293T cells (COBIOER, Nanjing, China) in Dulbecco's modified eagle medium (DMEM; Sigma-Aldrich), accompanied with miR-125-5p mimic or mimic NC. 48 h later, cells were collected and the relative luciferase activity (ratio of firefly and renilla) was examined via the dual luciferase reporter system (Promega).

RNA Immunoprecipitation (RIP) Assay

Magna RIP RNA-Binding Protein Immunoprecipitation Kit (Millipore, Billerica, MA, USA) was employed to analyze the interplay between circ_0032833 and miR-125-5p. After cell lysis in RIP lysis buffer for 1 h, cell suspension was incubated with A/G (anti-Ago2 or anti-IgG)-covered magnetic beads overnight at 4°C. The precipitated RNA-binding protein complexes were washed from magnetic beads, and qRT-PCR was used for circ_0032833 and miR-125-5p analysis following the protein removal and RNA acquisition.

In vivo Experiment

We purchased 24 BALB/c nude mice (6-week-old) from Shanghai SIPPR-BK Laboratory Animal Co. Ltd. (Shanghai, China) and randomly divided them into four groups: sh-NC+5-FU, sh-circ+5-FU, sh-NC+Oxa and sh-circ+Oxa. Mice were subcutaneously injected with 5×10^6 HCT116-R cells transfected sh-circ or sh-NC lentiviral vector. After 7 d, tumor volume ($\text{length} \times \text{width}^2 \times 0.5$) was measured by digital caliper and mice were subjected to intraperitoneal injection with 50 mg/kg 5-FU or 10 mg/kg Oxa twice a week. Then, tumor volume was recorded per 5 d, and all mice were euthanatized using CO₂ asphyxia

method at 32 d. The excised tumors were weighed and used for RNA or protein extraction, followed by the detection of related RNA and proteins. This experiment was administered based on the ratification from the Animal Ethical Committee of Weifang People's Hospital. Animal studies were performed in compliance with the ARRIVE guidelines and the Basel Declaration. All animals received humane care according to the National Institutes of Health (USA) guidelines.

Statistical Analysis

Our assays were all independently performed three times. Data expressed as mean $\pm$ standard deviation (SD) were carefully processed applying with SPSS 22.0 and Graphpad prism 7. The intergenic linear relationship was analyzed using Spearman correlation coefficient. Student's *t*-test and one-way analysis of variance (ANOVA) followed by Tukey's test were applied for difference comparison. Difference was statistically significant when *P* value <0.05.

Results

Through cell viability analysis, we found that IC₅₀ values of 5-FU (40 μ M) and Oxa (22 μ M) in HCT116-R cells were much higher than HCT116-P cells, confirming that HCT116-R cells were resistant to 5-FU and Oxa (Figure 1A). The qRT-PCR indicated that circ_0032833 expression was prominently increased in HCT116-R cells and clinical Nonresponders compared with HCT116-P cells and Responders (Figure 1B-C). Moreover, Western blot demonstrated that multidrug resistance-related proteins MRP-1 and MCL-1 were signally up-regulated in HCT116-R cells (Figure 1D) and Nonresponders (Figure 1E) in contrast with their respective control group. These data collectively suggested that circ_0032833 was closely related to drug resistance in FOLFOX-resistant CRC tissues and cells.

In HCT116-R cells, we used si-circ transfection to knock down the expression of circ_0032833 (Figure 2A). In si-circ-transfected HCT116-R cells, CCK-8 revealed that the viable cell number was overtly declined with the increase of 5-FU and Oxa, then the IC₅₀ values of 5-FU and Oxa were found to be lessened compared to si-NC-transfected HCT116-R cells (Figure 2B). Colony formation assay and flow cytometry showed that circ_0032833 knockdown triggered the decreased colonies (Figure 2C) and increased apoptosis rate (Figure 2D) in HCT116-R cells treated with 30 μ M 5-FU or Oxa. Determination of cell migration and invasion (Figure 3A-B) exhibited the

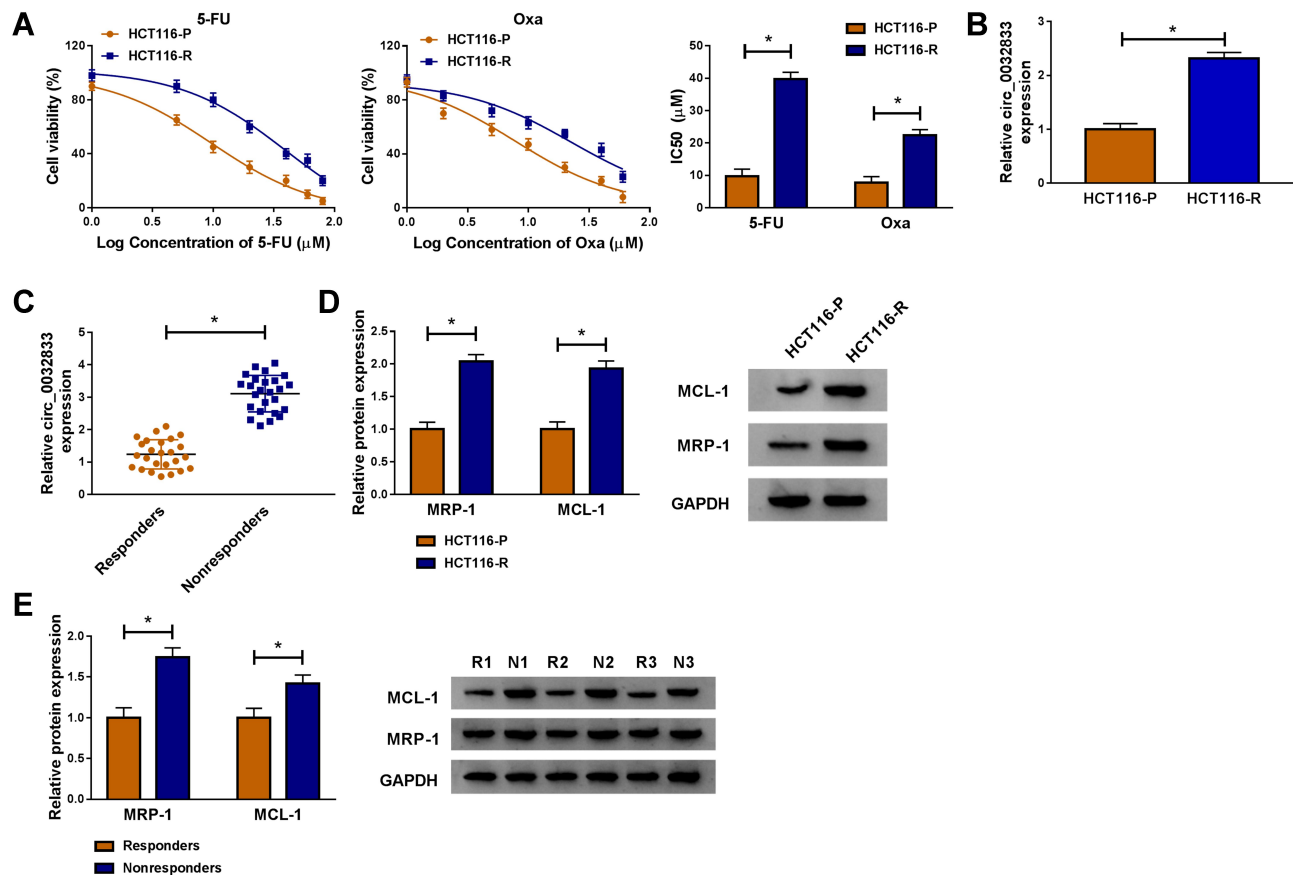


Figure 1 Circ_0032833 was overexpressed in FOLFOX-resistant CRC cells and associated with drug resistance. (A) CCK-8 assay was used for analyzing IC50 of 5-FU and Oxa in HCT116-P and HCT116-R cells. (B–C) Circ_0032833 was determined using qRT-PCR in HCT116-R cells (B) and nonresponsive tissues (C), as well as in their control HCT116-P cells and responders. (D–E) The multidrug resistance-related proteins were measured using Western blot in HCT116-R/P cells (D) and nonresponsive/responsive tissues (E). * $P < 0.05$.

obvious inhibition in si-circ group contrasted with si-NC group. By analyzing the related protein levels, we also discovered the si-circ transfection downregulated the protein expression of multidrug resistance-promoting markers (MRP-1 and MCL-1) and pro-metastasis marker (N-cad) but stimulated the levels of anti-metastasis marker (E-cad) and apoptosis positive marker (caspase-3) in 5-FU or Oxa-treated HCT116-R cells (Figure 3C). All these results proved that circ_0032833 down-regulation increased 5-fluorouracil and oxaliplatin sensitivities to inhibit the progression of FOLFOX-resistant CRC cells.

Starbase3.0 has predicted that 20 miRNAs had the binding sites for circ_0032833 and 3 miRNAs (miR-506-3p, miR-124-3p, miR-125-5p) that were downregulated in CRC were selected as the candidate targets for circ_0032833. Our screening assays indicated that three miRNAs were all downregulated in Nonresponders (n=5) compared to Responders (n=5) and their levels were significantly increased by knockdown of circ_0032833

(Supplemental Figure 1A–B). Thus, miR-125-5p with most obvious upregulation was chosen for further research. Then, we found the downregulation of miR-125-5p not only in HCT116-R cells but also in 25 Nonresponders, in comparison with HCT116-P cells and 25 Responders (Figure 4A–B). Interestingly, circ_0032833 was negatively correlated to miR-125-5p ($r = -0.5273$, $P = 0.0068$) in Nonresponsive tissues (Figure 4C). Starbase 3.0 presented the theoretical binding between the sequences of circ_0032833 and miR-125-5p (Figure 4D), and then we performed the dual-luciferase reporter assay in 293T cells to validate that miR-125-5p could interact with circ_0032833 wt not circ_0032833 mut (Figure 4E). As RIP analysis in Figure 4F, both circ_0032833 and miR-125-5p were largely captured by anti-Ago2 compared to anti-IgG, indicating the direct interaction between circ_0032833 and miR-125-5p.

Compared to si-NC transfection, si-circ transfection apparently heightened the relative level of miR-125-5p

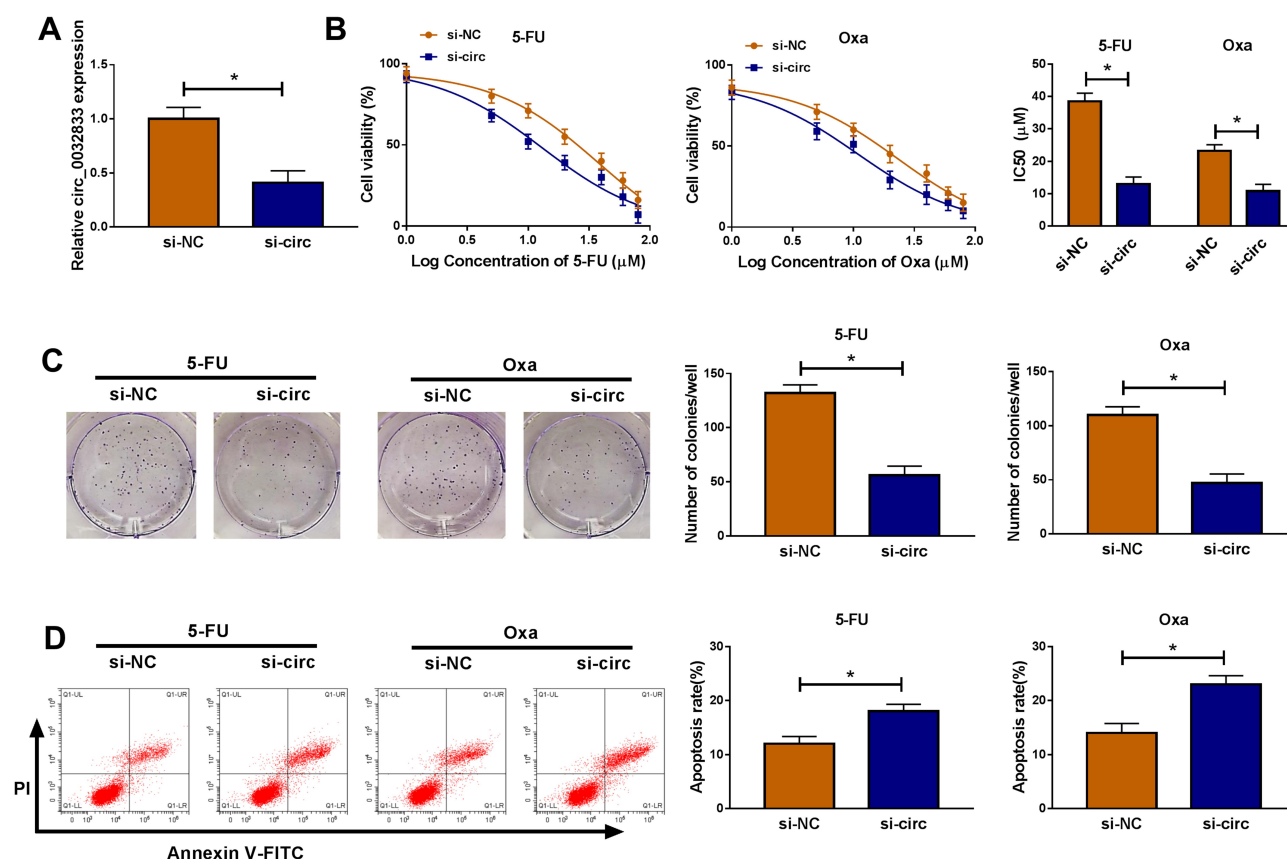


Figure 2 Down-regulation of circ_0032833 inhibited 5-fluorouracil and oxaliplatin resistance, colony formation but facilitated apoptosis in FOLFOX-resistant CRC cells. HCT116-R cells were transfected with si-NC or si-circ. (A) Knockdown efficiency of si-circ was assessed by qRT-PCR using si-NC as control group. (B) The examination of IC50 was conducted by CCK-8 assay in HCT116-R cells after treatment of different concentrations 5-FU or Oxa. (C–D) After treatment with 30 μM 5-FU or Oxa, colony formation and apoptosis of si-circ and si-NC groups were respectively assayed through colony formation assay and Annexin V-FITC/PI flow cytometry. * $P < 0.05$.

(Figure 5A). Given that circ_0032833 could act as a miR-125-5p sponge, we further researched their relation on regulating chemosensitivity in HCT116-R cells. As Figure 5B depicted, miR-125-5p inhibitor significantly abated the inhibitory effects of si-circ on IC50 of 5-FU and Oxa. By comparison with si-circ+inhibitor NC group, the colony formation capacity in si-circ+miR-125-5p inhibitor group was enhanced (Figure 5C) while cell apoptosis was restrained on the contrary (Figure 5D). Similarly, migrated and invaded cells were increased after co-transfection of si-circ and miR-125-5p inhibitor relative to alone si-circ transfection (Figure 5E–F). Western blot also testified that the si-circ-induced inhibition on resistance, metastasis and promotion on apoptosis were all relieved following the down-regulation of miR-125-5p in HCT116-R cells treated of 5-FU and Oxa (Figure 5G). Taken together, miR-125-5p inhibitor reversed the stimulative effects of circ_0032833 knockdown on 5-fluorouracil and oxaliplatin sensitivities.

MSI1 Was a Target of miR-125-5p

As for the target genes of miR-125-5p, many potential targets (>3000) were predicted by online Starbase3.0 and some genes (XIAP, FOXQ1, USP22, ABCC1, NOTCH-1, MSI1) have been reported to be implicated in the chemoresistance of CRC. The qRT-PCR results have indicated their upregulation in Nonresponders contrasted with Responders and Western blot showed that miR-125-5p mimic resulted in most evident downregulation of the protein expression of MSI1 (Supplemental Figure 2A–B). Then we further researched the target relation between miR-125-5p and MSI1 in CRC. Through the analysis of qRT-PCR and Western blot, MSI1 was identified as an up-regulated gene in HCT116-R cells (Figure 6A–B), as well as in Nonresponders (Figure 6C–D). Spearman correlation coefficient displayed a negative linear relationship ($r = -0.5689$, $p = 0.003$) between miR-125-5p and MSI1 mRNA levels (Figure 6E). In Starbase 3.0 online software, we found the hypothetical binding sites of miR-125-5p in the MSI1 3'UTR sequence (Figure 6F); subsequently, the

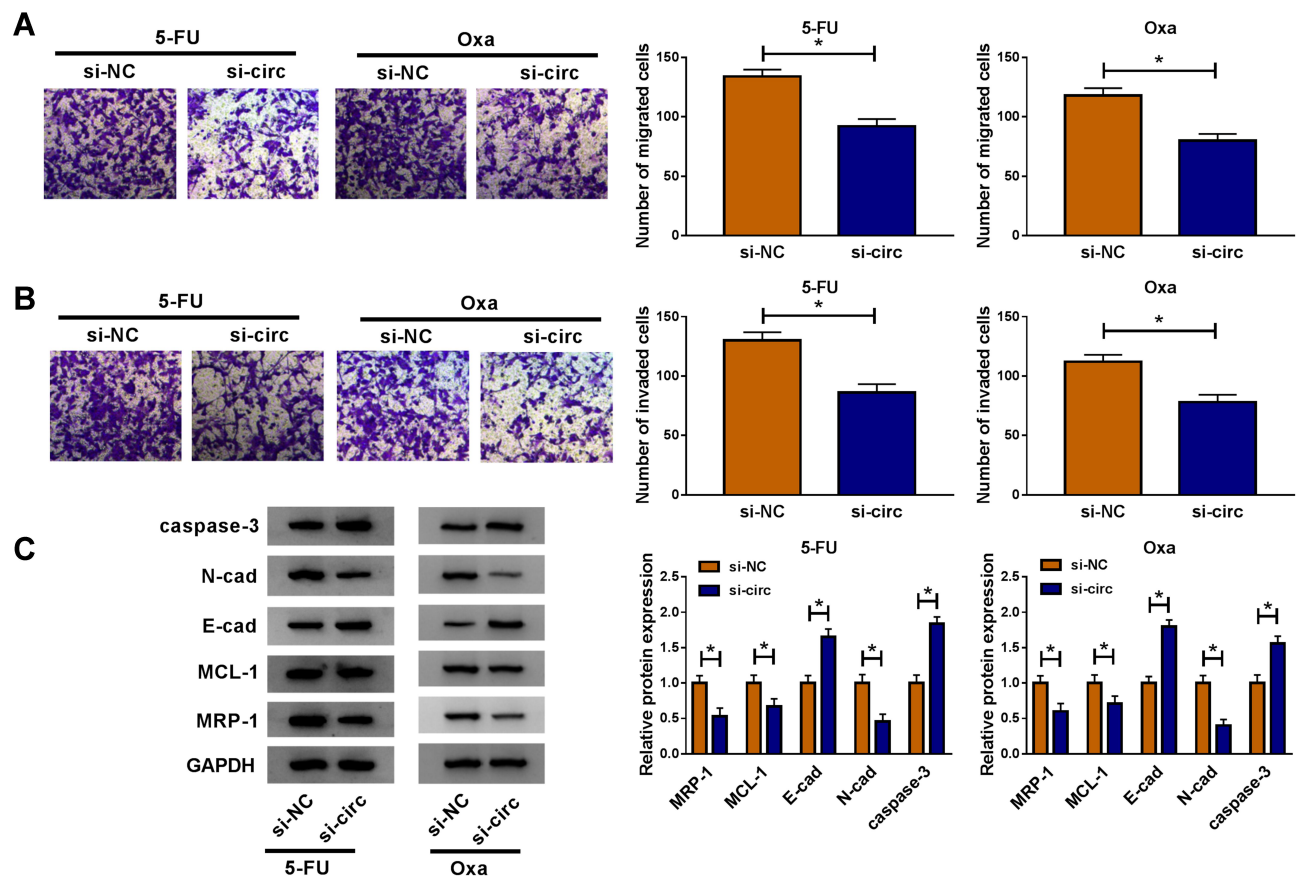


Figure 3 Circ_0032833 knockdown reduced resistance and metastasis but enhanced apoptosis in FOLFOX-resistant CRC cells. HCT116-R cells were transfected with si-NC or si-circ and treated with 5-FU or Oxa (respective 30 μ M). (A–B) Transwell assay was used to analyze cell migration (A) and invasion (B). (C) The resistance, metastasis and apoptosis-associated proteins were determined by Western blot. * $P < 0.05$.

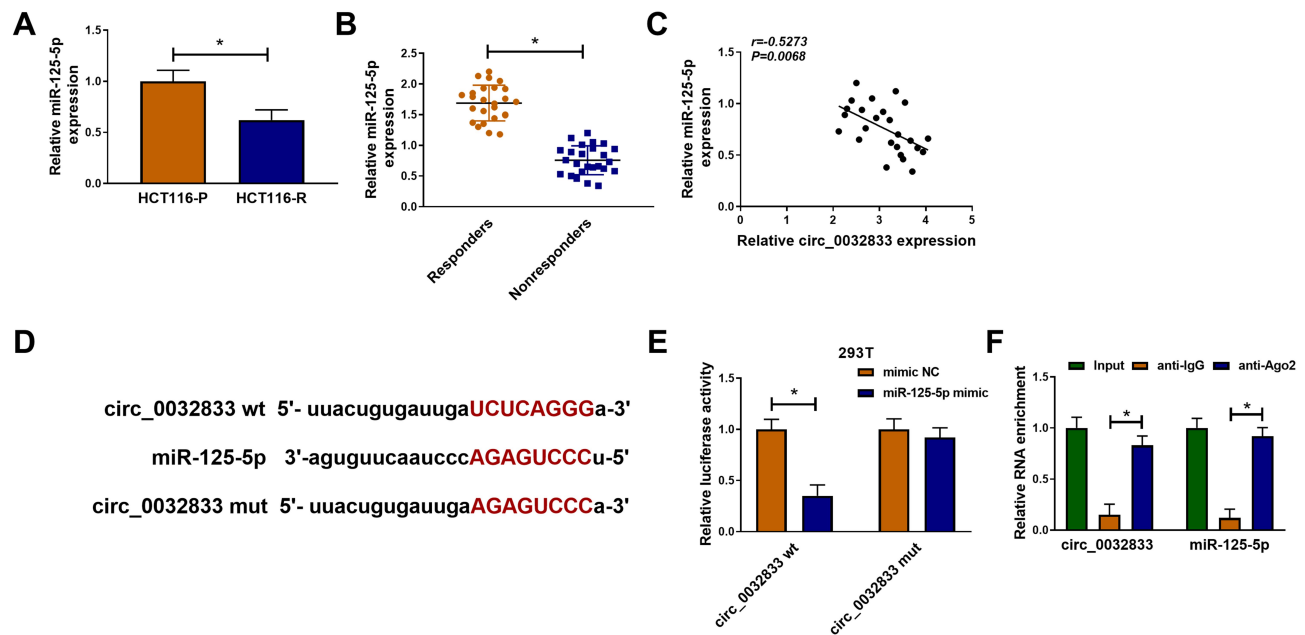


Figure 4 Circ_0032833 directly interacted with miR-125-5p. (A) The miR-125-5p detection was carried out by qRT-PCR in HCT116-P/R cells and responders/nonresponders. (C) Spearman correlation coefficient was used to explore the linear relationship between circ_0032833 and miR-125-5p. (D) Starbase 3.0 was exploited for the analysis of binding sites between circ_0032833 and miR-125-5p. (E–F) Dual-luciferase reporter and RIP assays were administrated for verifying the interaction between circ_0032833 and miR-125-5p. * $P < 0.05$.

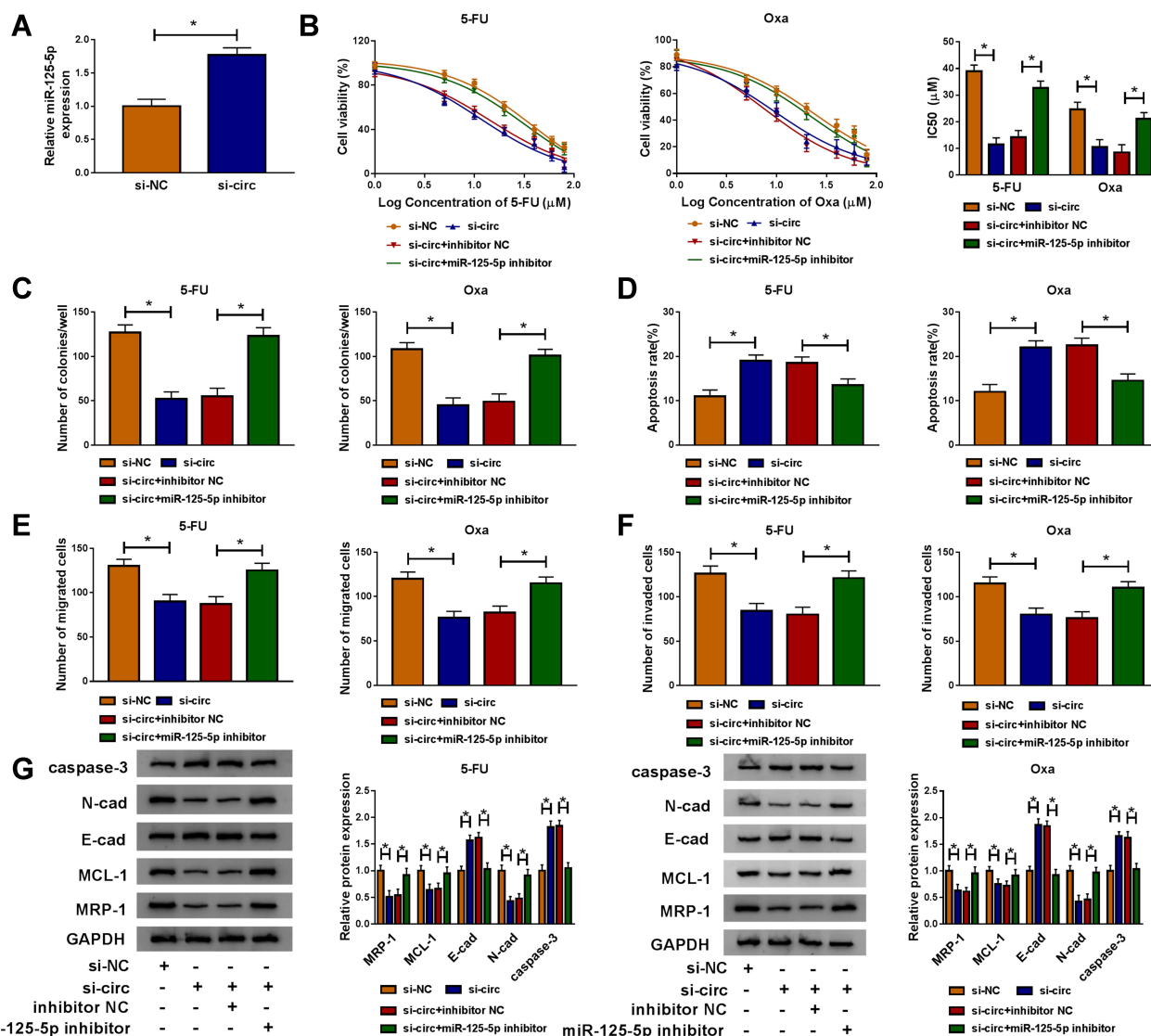


Figure 5 MiR-125-5p inhibitor counteracted the increase on 5-fluorouracil and oxaliplatin sensitivities caused by circ_0032833 knockdown in FOLFOX-resistant CRC cells. (A) The expression of miR-125-5p was examined via qRT-PCR in HCT116-R cells transfected with si-circ or si-NC. (B) CCK-8 was applied to measure IC₅₀ of 5-FU and Oxa in HCT116-R cells after transfection of si-NC, si-circ, si-circ+inhibitor NC or si-circ+miR-125-5p inhibitor. (C–F) In transfected (as Figure 5B) and treated (30 μM 5-FU or Oxa) HCT116-R cells, colony formation assay, Annexin V-FITC/PI flow cytometry and transwell assay were severally adopted for detecting cell colony ability (C), apoptosis (D) and migration/invasion (E–F). (G) Western blot was employed for related protein analysis. **P* < 0.05.

actual combination between miR-125-5p and MSI1 3'UTR was confirmed by the decrease of the relative luciferase activity in miR-125-5p mimic+MSI1 3'UTR wt co-transfection group (Figure 6G). Therefore, MSI1 can be used as a downstream target of miR-125-5p.

MiR-125-5p Worked as a Sensitizer of 5-Fluorouracil and Oxaliplatin in FOLFOX-Resistant CRC Cells via Targeting MSI1

By designing the four transfection groups: mimic NC, miR-125-5p mimic, miR-125-5p mimic+vector and

miR-125-5p mimic+oe-MSI1, the role of miR-125-5p on drug sensitivity and how it acted would be addressed. As shown in Figure 7A–B, miR-125-5p mimic repressed the mRNA and protein levels of MSI1 in HCT116-R cells while oe-MSI1 transfection promoted MSI1 expression of miR-125-5p mimic group in a reversible manner. CCK-8 assay indicated that the decreased IC₅₀ values of 5-FU and Oxa in miR-125-5p mimic group were elevated in miR-125-5p mimic+oe-MSI1 group, hinting that miR-125-5p inhibited 5-FU and Oxa resistance by targeting MSI1 (Figure 7C). In HCT116-R cells treated with 30 μM 5-FU or Oxa, we found that miR-125-5p overexpression led to the suppression of colony

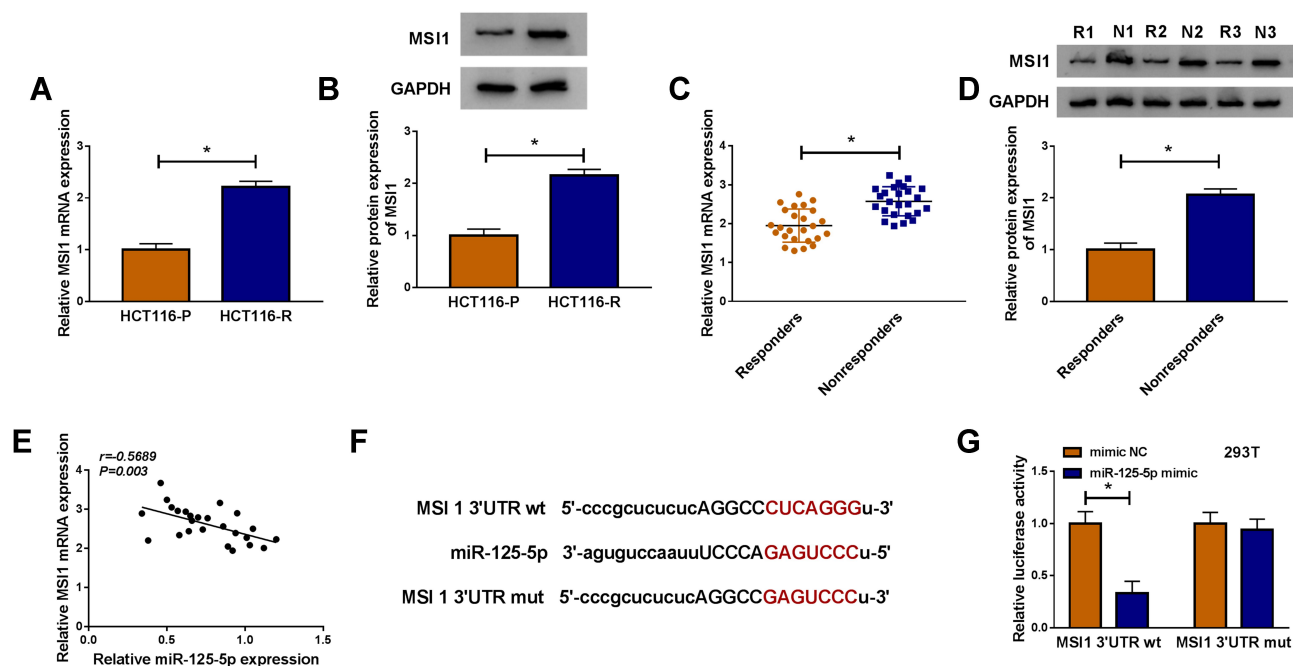


Figure 6 MSI1 was a target of miR-125-5p. (A–D) The qRT-PCR and Western blot were implemented to assay the mRNA and protein levels of MSI1 in HCT116-R cells (A–B) and Nonresponders (C–D) with HCT116-P cells and Responders as respective controls. (E) The correlation between miR-125-5p and MSI1 was evaluated by Spearman correlation coefficient. (F) The binding sites of miR-125-5p in 3'UTR of MSI1 was predicted via Starbase 3.0. (G) The actual binding between miR-125-5p and MSI1 was affirmed using dual-luciferase reporter assay. * $P < 0.05$.

formation ability (Figure 7D) and the increased apoptosis rate (Figure 7E) along with the blockage of migration/invasion (Figure 7F–G), which were all restored after the up-regulation of MSI1. Meanwhile, miR-125-5p mimic down-regulated the levels of MRP-1, MCL-1 and N-cad proteins but motivated the expression of E-cad and caspase-3, whereas the introduction of oe-MSI1 vector abrogated the miR-125-5p-induced resistance and metastasis obstruction as well as apoptosis promotion (Figure 7H). Above works manifested the role of miR-125-5p as a sensitizer of 5-fluorouracil and oxaliplatin in FOLFOX-resistant CRC cells by targeting MSI1.

Owing to the miR-125-5p link between circ_0032833 and MSI1, the reverted transfection was performed to analyze whether circ_0032833 could affect MSI1 via miR-125-5p. As Figure 8A–B illustrated, si-circ transfection evidently repressed the MSI1 mRNA and protein expression in HCT116-R cells, but the inhibitory effects were offset following the down-regulation of miR-125-5p. Indirectly, circ_0032833 could regulate MSI1 through directly binding to miR-125-5p.

Xenograft models were used for our experiment in vivo. After injection of HCT116-R cells and treatment of 5-FU, we found that tumor volume was reduced with

the increase of time (Figure 9A) and the finally excised tumors were lighter (Figure 9B) in sh-circ group by comparing with sh-NC group. The expression levels of circ_0032833 and MSI1 (mRNA and protein) were lower while miR-125-5p was higher in sh-circ group than these in sh-NC group (Figure 9C–D). Likewise, the interference of circ_0032833 expression also decreased tumor volume and weight (Figure 9E–F) by the miR-125-5p/MSI1 axis (Figure 9G–H) in Oxa-treated mice. Additionally, sh-circ introduction evoked the resistance inhibition (the down-regulated MRP-1 and MCL-1), metastasis impairment (the up-regulated E-cad and down-regulated N-cad) and apoptosis enhancement (the increased caspase-3) in tumors from 5-FU or Oxa-treated mice (Figure 9I). Also in vivo, circ_0032833 knockdown contributed to the 5-fluorouracil and oxaliplatin sensitivities via the miR-125-5p/MSI1 axis.

Discussion

Chemoresistance has become an intractable issue in clinical CRC treatment, which makes it increasingly important to facilitate chemosensitivity. In the current report, knockdown of circ_0032833 was proved to contribute to the sensitivity of CRC cells to 5-fluorouracil and oxaliplatin via regulating MSI1 expression by targeting miR-125-5p.

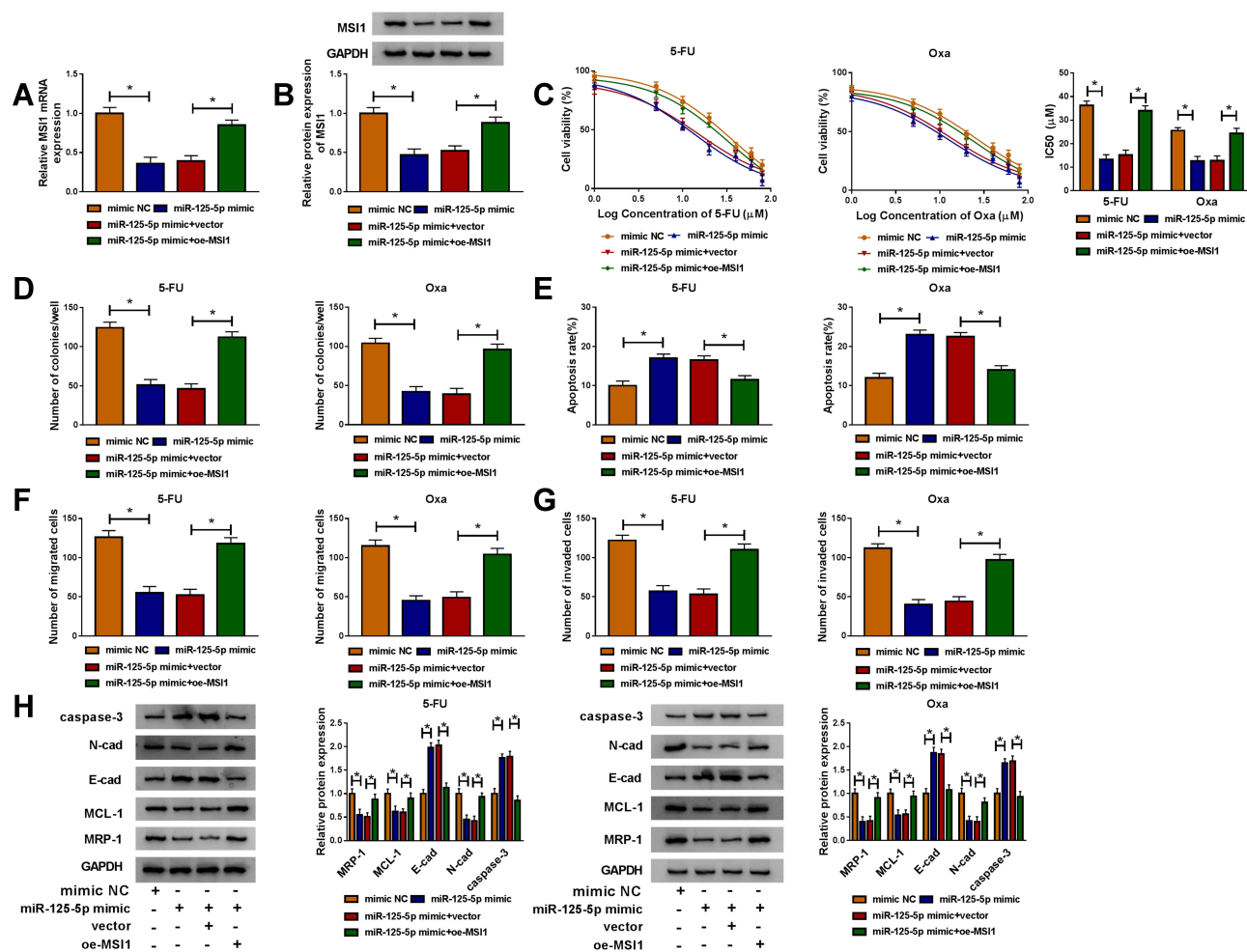


Figure 7 MiR-125-5p worked as a sensitizer of 5-fluorouracil and oxaliplatin in FOLFOX-resistant CRC cells via targeting MSII. Following transfection: mimic NC, miR-125-5p mimic, miR-125-5p mimic+vector or miR-125-5p mimic+oe-MSII was performed in HCT116-R cells. (A–B) MSII mRNA and protein levels were tested through qRT-PCR and Western blot. (C) After treatment of 5-FU or Oxa, IC50 was calculated by cell viability detection in CCK-8 assay. (D–G) After treatment with 30 μM 5-FU or Oxa, the examination of colony formation (D), apoptosis (E) and migration/invasion (F–G) was implemented by colony formation assay, Annexin V-FITC/PI flow cytometry and transwell assay. (H) The proteins associated with above cellular behaviors were assayed via Western blot. **P* < 0.05.

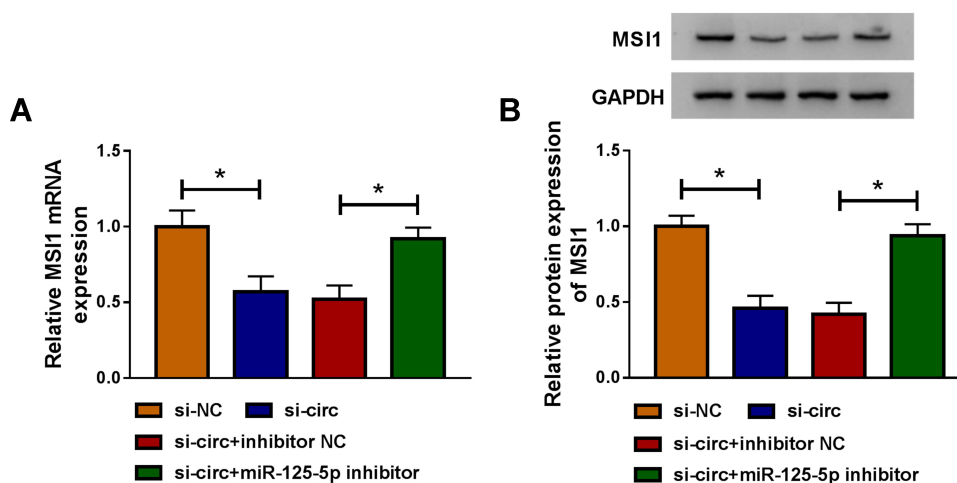


Figure 8 Circ_0032833 regulated MSII level by serving as a miR-125-5p sponge. (A–B) After HCT116-R cells were transfected with si-circ, si-circ+miR-125-5p inhibitor or matched controls, the measurement of MSII mRNA (A) and protein (B) expression was executed by qRT-PCR and Western blot. **P* < 0.05.

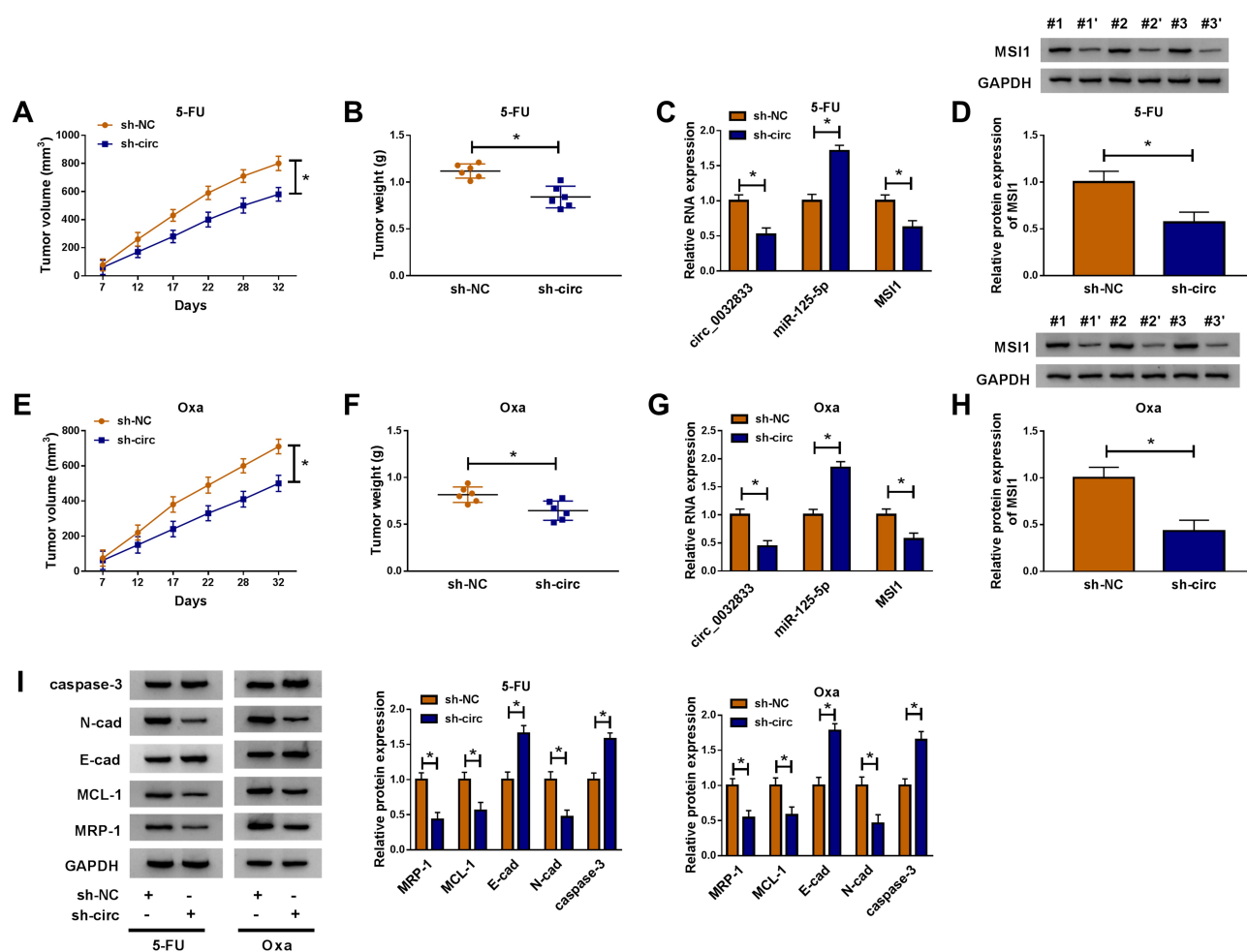


Figure 9 Inhibition of circ_0032833 contributed to the 5-fluorouracil and oxaliplatin sensitivities in vivo via regulating the miR-125-5p/MSI1 axis. (**A–B**) Tumor volume (**A**) and weight (**B**) were measured in sh-NC+5-FU and sh-circ+5-FU groups. (**C–D**) The qRT-PCR and Western blot were used to perform expression analysis of circ_0032833, miR-125-5p, MSI1 mRNA (**C**) and MSI1 protein (**D**) in sh-NC+5-FU and sh-circ+5-FU groups. (**E–F**) Tumor volume (**E**) and weight (**F**) of sh-NC+Oxa and sh-circ+Oxa groups were recorded. (**G–H**) The levels of circ_0032833, miR-125-5p and MSI1 (mRNA and protein) in sh-NC+Oxa and sh-circ+Oxa groups were assayed through qRT-PCR and Western blot. (**I**) Western blot was exploited for examining the cellular-related proteins in sh-NC+5-FU, sh-circ+5-FU, sh-NC+Oxa and sh-circ+Oxa groups. * $P < 0.05$.

CircRNAs are now in the spotlight as potential regulators in human cancers, also in resistance of cancer chemotherapy.²² Liu et al discovered that circHIPK3 expedited chemoresistance of gemcitabine in pancreatic cancer cells²³ and Yang et al clarified that the low circ-CDR1as had promotive effect on chemosensitivity in 5-fluorouracil-resistant breast cancer cells.²⁴ Microarray analysis of circRNAs affirmed their dysregulation in 5-fluorouracil-based chemoradiation-resistant CRC cells.²⁵ And the latest study has exhibited that circRNA ciRS-122 induced the generation of resistance in oxaliplatin-sensitive CRC cells.²⁶ Circ_0032833 was up-regulated in FOLFOX-resistant CRC cells by our qRT-PCR detection. Functionally, siRNA-mediated circ_0032833 interference notably decreased resistance both of 5-fluorouracil and

oxaliplatin along with cancer retardation by analyzing the cellular process. Thus, the repression of circ_0032833 could sensitize FOLFOX-resistant CRC cells to 5-fluorouracil and oxaliplatin in accordance with the above studies.

Subsequently, miR-125-5p was found to be down-regulated in FOLFOX-resistant CRC (tissues and cells) and negatively related to circ_0032833 level. Growing researches have demonstrated the miRNA sponge mechanism of circRNAs in all types of tumor cells. For instance, circ_0001564 acted as an oncogene to increase proliferation and inhibit apoptosis of osteosarcoma cells through sequestering miR-29c-3p;²⁷ circular RNA profiling proved circADAMTS13 as a miR-484 sponge to be an inhibitor in the progression of hepatocellular carcinoma cells;²⁸

circ_0000523 also was reported to regulate CRC development by relying on the sponge of miR-31.²⁹ It was similar that miR-125-5p also could be sponged by circ_0032833, and the chemosensitivity regulation of circ_0032833 knockdown on FOLFOX-resistant CRC cells was achieved by the elevation of miR-125-5p.

MSI1 was an overexpressed gene in CRC tissues and cells with FOLFOX-resistance through our expression analysis of qRT-PCR and Western blot. Like circ_0032833 and miR-125-5p, miR-125-5p also could directly target MSI1. miR-125-5p was identified as a great promoter in sensitizing CRC cells to 5-fluorouracil and oxaliplatin, by combining with the 3'UTR of MSI1 to block MSI1 expression. Moreover, MSI1 level was regulated by the circ_0032833-mediated miR-125-5p change here. Zhang et al have declared that circ_0063809 boosted the paclitaxel resistance in ovarian cancer via sponging miR-1252 to up-regulate FOXR2.³⁰ Sang et al showed that circ_0025202 enhanced tamoxifen efficacy of breast cancer cells through affecting the miR-182-5p/FOXO3a axis.³¹ Herein, circ_0032833 down-regulation regulated the miR-125-5p/MSI1 axis to elevate 5-fluorouracil and oxaliplatin efficacies of CRC not only in vitro but also in vivo.

Conclusion

In summary, the present study identified circ_0032833/miR-125-5p/MSI1 axis as a novel 5-fluorouracil and oxaliplatin resistance mechanism in CRC, and circ_0032833 as a diagnostic target for resistance generation. Promisingly, the efficiency of FOLFOX chemotherapy will be improved by increasing the 5-fluorouracil and oxaliplatin susceptibilities after knocking down circ_0032833.

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Disclosure

The authors declare that they have no conflicts of interest.

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