

Acoustic Waves in Axonal Membrane and Caveolins are the New Targets for Pain Treatment with High Frequency Ultrasound

This article was published in the following Dove Press journal:
Journal of Pain Research

Ilja Kruglikov 

Scientific Department, Wellcomet
GmbH, Karlsruhe, Germany

Abstract: Reciprocal interaction between electrical and mechanical waves observed in axonal membrane during its excitation leads to a paradigm shift in pain research making the uncoupling of electro-mechanical signals an interesting target in pain treatment. This uncoupling can be realized either through direct disturbance of the mechanical surface waves in axonal membrane or through shifting of the thermodynamic state of this membrane far from its phase transition point. Both effects can be effectively realized through application of the very high frequency ultrasound waves. Additional target for application of ultrasound in pain treatment is the caveolin-1, which is abundantly present in Schwann cells as well as in the non-axonal tissues. Both targets demonstrate frequency-dependent reactions, thus making a very high frequency ultrasound a promising treatment modality in pain treatment.

Keywords: chronic pain, axonal membrane, mechanical surface wave, caveolin, ultrasound

Introduction

For a long time, there was a broad consensus that the origin of peripheral pain is mainly connected to generation of action potentials (APs) and their propagation along the axonal membrane. APs were believed to have a pure electrical origin well described by a classical Hodgkin-Huxley model. According to this model, depolarization of the axonal membrane beyond a threshold potential leads to its local excitation and generation of spikes demonstrating some typical features such as all-or-none behavior, threshold stimulation and solitary character. Since AP generation was mainly connected to the functioning of the ion channels in axonal membrane, the primary attention in anesthesia was paid to the treatment methods influencing operation of proteins in these structures. This model remained predominant for a long time even despite the obvious contradiction with another cornerstone of anesthesiology – the Meyer-Overton correlation – stating that the effectiveness of applied anesthetics is positively connected to their lipophilicity. This correlation was observed over a very broad interval of solubilities and indicated that not the ion channels but the lipid components of the neuronal membrane should be the main site for anesthetic action.

However, some time ago it was revealed that axons during their excitation generate not merely electrical but also mechanical waves, which can propagate along the axonal membrane with a remarkably low attenuation. Electrical and mechanical waves interact with each other producing a reciprocal electro-mechanical excitation in axonal

Correspondence: Ilja Kruglikov
Email i.kruglikov@wellcomet.de

membrane. This finding caused a paradigm shift in neurostimulation and pain treatment – from a straightforward inhibition of electrical signals in axons through application of external electrical stimulation to more comprehensive treatment methods connected with “uncoupling” of electro-mechanical signals in axonal membrane. One of these methods is the application of ultrasound (US) waves, which received much attention during the recent past because of intensive research in the field of ultrasonic neuromodulation.¹

While US was considered as a standard method for peripheral pain treatment, it is well-known that its applications demonstrate mixed outcomes even in the same pain conditions. Here we re-analyze the possible reasons for this diversity considering the new axonal targets for US revealed during recent years.

Generation and Propagation of Acoustic Waves in Axonal Membrane

Early investigations demonstrated that AP transmission modulates the viscosity of axonal membrane and generates an additional pressure of up to 10 mPa in axoplasm.^{2,3} Much later, it was reported that the development of AP is accompanied by a modulation of the axonal diameter, which temporal course is very similar to that of AP.^{4,5} These observations indicated that excitation of axonal membrane does not have a pure electric origin and should involve some mechanical and conformational changes. Such assumption led to the development of different electro-mechanical models describing mechanical surface waves induced in axonal membrane during AP propagation as well as an induction of APs by application of mechanical forces to this membrane.^{4,6–8} These surface waves have a much lower attenuation than the normal bulk waves, which allows them to propagate far away from their place of origin.

It was also demonstrated that the non-linear surface acoustic waves can propagate in artificial lipid membranes if the thermodynamic state of these membranes is close to a corresponding phase transition point (melting from gel to liquid phase).⁹ Under physiological conditions, the state of axonal membrane is indeed quite close to this point and thus surface acoustic waves can be easily stimulated by mechanical, thermic and chemical factors, which makes the initiation and propagation of the surface acoustic waves in these structures to a fundamental phenomenon.

Among others, it was reported that such waves can be induced by electrical stimulation⁷ and by a local threshold acidification of the interface.¹⁰

This means that mechanical surface waves can be induced both in the native and artificial membranes and have the universal mechanism of generation. Discovery of these waves had significant impact on the theory and practical applications in the field of central and peripheral neurostimulation. At the same time, it also allowed another insight into the axonal-US interaction between axons and US in pain treatment.

Radiation Force as the Main Factor in Direct US Modulation of AP

While the early research concerning the application of the focused US of different frequencies to the peripheral nerves revealed that increasing frequency causes an enhanced modulation of AP generation in axonal membrane, this phenomenon was then generally connected to the thermic effect of US and explained by a stronger absorption of the high frequency waves in the tissue. Recently, it was shown by different authors that US can modulate AP generation in neurons both in vitro and in vivo within a broad range of much lower intensities (0.1–10.0 W/cm²).¹¹ This modulation occurred without any significant increase of the temperature, thus clearly indicating the primary role of mechanical effects in this phenomenon. Mechanical effects of US can be connected either with development of cavitation in the tissue or with a constitution of radiation pressure on the membranes. Investigation of the ganglion cell spiking activity by application of US with frequencies of 0.5–43.0 MHz revealed that the strength of axonal modulation increases with US frequency.¹² Also relevant to this discussion, the voltage-clamp experiments with lipid bilayers demonstrated that US with frequencies of 1 MHz and 43 MHz induce similar on/off currents.¹¹ Since the probability of cavitation quickly decreases with increasing US frequency, the radiation forces induced by US in axonal membrane must be the main factor responsible for this modulation.

Acoustic radiation force on the cell surface depends on different physical parameters and significantly varies for the targets of different geometries, e.g. spherical cells and cylindrical axons. This force also depends on the relationship between the wavenumber of the US wave (k) and the axonal radius (R), where $k = 2\pi f/c$, whereby f and c are the US frequency and velocity, respectively. Taking the typical

axon radius of 1 μm and the US velocity in the soft tissue of 1.500 m/s, condition $kR \ll 1$ will fulfill for $f \ll 240$ MHz and thus for all typical US frequencies applied for neuronal stimulation. In case of an US wave directed along the axis of a cylindrical axon, radiation force induced in axonal membrane will in this approximation be linearly proportional to the US frequency, f .¹³

The fluorescent investigations of the lipid membranes subjected to US pulses revealed sufficient conformational changes in these structures, which were especially pronounced as the membrane was thermodynamically in the vicinity of its phase transition point.¹⁴ It is well-known that near this point the heat capacity and compressibility of the lipid phase reach their maximum values. Since the US-induced radiation force is proportional to the adiabatic compressibility of the target,¹³ mechanical pressure induced by US in axonal membrane will also reach its maximum if the system is near its phase transition.

Altogether, US waves can induce mechanical perturbation in axonal membrane generating a propagating mechanical surface wave, which in its turn can cause generation of AP. This mechanism is actually considered an important pathway in neurostimulation.¹ On the other hand, mechanical stress induced in axonal membrane by acoustic radiation force can shift its thermodynamic state far away from a phase transition point, which will also suppress the AP generation. This should especially be the case if the US wave can produce a sufficient thermo-mechanical stress near axonal membrane. As we discussed previously, such stress quickly increases with increasing US frequency.¹⁵

Whereas a direct modulation of AP in axons is an important direct mechanism of US-axon interaction, another target connected with expression of caveolins in cell membranes should also be of significant importance at least in some types of pain.

Caveolin-1 as a Target for an Indirect Modulation of Pain

Caveolin-1 (Cav-1) is a principal structural component of the caveolae plasma membrane invaginations enriched in cholesterol and sphingolipids, which are present in different types of cells. Cav-1 is involved in rapid adaptation to cellular volume modifications, in multiple signal transduction processes as well as in the processes of endo- and exocytosis. Cav-1 demonstrates reciprocal interaction with oxidative stress: whereas induction of Cav-1 attenuates the

oxidative stress induced by H_2O_2 ,¹⁶ application of oxidative stress modulates Cav-1 expression in a dose-dependent and cell-type specific way.^{17–19} This effect is directly connected to pain sensation: as it was demonstrated in mouse models, antioxidants attenuate constriction of the sciatic nerve indicating that oxidative stress should be directly involved in development of neuropathic pain.²⁰

Caveolins were long believed to not be directly involved in neuronal signaling. The main reason for this was that these proteins were found only in glial cells and synapses, but not in axonal membranes.²¹ However, the rapid propagation of action potentials along axonal fibers of motor and sensory neurons is enabled only through their myelination provided by Schwann cells wrapping the axons. Importantly, Cav-1 is highly abundant in the abaxonal myelin membranes of the Schwann cells and loss of Cav-1 compromises the functioning of these cells as well as decreases nerve regeneration after injury. On the other hand, Cav-1 expression significantly increases during myelination and strongly decreases after axotomy.²²

Schwann cells react to oxidative stress induced by H_2O_2 with apoptotic cell death. This effect can be strongly attenuated by extracellular induction of the heat shock protein 72 (Hsp72).²³ Such Hsp72-mediated reduction of oxidative stress-induced injuries in Schwann cells has neuroprotective effect on peripheral nerves increasing their stress tolerance.²⁴ Remarkably, this protein demonstrates positive and almost linear correlation with Cav-1 level in human and mouse skin, indicating that induction of Hsp72 should lead to enhanced expression of Cav-1.²⁵

Additionally, nociceptive pain is connected to a local inflammation in affected tissue and it is widely accepted that this pathway involves prostaglandin E_2 (PGE_2). PGE_2 is a cyclooxygenase (COX) product and a lipid mediator, and inhibitors of COX-1/COX-2 were reported to effectively suppress the inflammatory pain through reduction of PGE_2 expression.²⁶ Remarkably, COX-2 co-localizes and physically interacts with Cav-1.²⁷ Since the Cav-1-mediated suppression of COX-2 reduces PGE_2 expression,²⁸ Cav-1 can be considered as a potential target in treatment of inflammation-induced nociceptive pain.

Moreover, different transient receptor potential ion channels (TRPs) are known to be involved in pain sensation, among others, TRP ankyrin 1 (TRPA1). This TRP is expressed in nociceptive sensory neurons and epithelial cells, where it plays a key role in detection of chemical, thermal and mechanical noxious stimuli.^{29–31} TRPA1 is

a common pathway for pro-nociceptive agonists generated in various pain conditions and thus a peripheral application of TRPA1 antagonists must be an effective approach for attenuation of primary hyperalgesia.³² In mice, TRPA1 channel is regulated by the local lipid environment and it was recently suggested that targeting lipid-TRPA1 interactions may be a strategy for the treatment of pain and neurogenic inflammation.³¹ Indeed, the amplitudes of the responses to such stimuli as cold and lipopolysaccharide (a toxic byproduct of bacterial lysis), were significantly lower in cells treated with the cholesterol scavenger methyl β -cyclodextrin deteriorating the lipid-TRPA1 interaction and reducing Cav-1 content in plasma membrane in a dose-dependent manner.³¹ TRPA1 is expressed not only in nociceptors but also in Schwann cells. Whereas silencing of TRPA1 in nociceptors attenuated mechanical allodynia but did not affect the macrophage infiltration and oxidative stress, which led to development of pain sensation in injured area, silencing of these channels in Schwann cells reduced both allodynia and neuroinflammation.³³ Hence, it can be strongly supposed that modification of Cav-1 expression in Schwann cells can modulate the pain sensation.

Further, mechanical trauma of the soft tissue causing appearance of edema in the injured area is often accompanied by pain and inflammation. Pathophysiologically, edema is connected to excessive accumulation of the fluid in the tissue caused by capillary leakage. The fluid retention can take place intracellularly or extracellularly and leads to a local constriction of the nerves providing pain sensation followed by degeneration of myelinated axons. Cav-1 is strongly expressed in endothelial cells and regulates fluid exchange between circulation and peripheral tissues. Indeed, Cav-1^{-/-} mice demonstrate pronounced abnormalities in vasoconstriction and vasorelaxation responses³⁴ as well as a microvascular hyperpermeability, which indicates that a microvascular leakage is mediated by Cav-1.³⁵ This effect was observed not only in an injured soft tissue but also in pulmonary and brain edema.^{36,37} Overexpression of Cav-1 was reported to attenuate edema preventing degradation of intercellular junctions;³⁷ hence, Cav-1 can serve as a target in early stages of edema development.

Cav-1 is also involved in development of diabetic peripheral neuropathy. Diabetic Cav-1 KO mice exhibit much more severe decrease of motor nerve conduction velocity as well as of mechanical and thermal sensitivity than their non-diabetic counterparts.³⁸ It is well-known

that myelinated Schwann cells undergo degeneration in diabetic neuropathy. Suppression of Cav-1 enhanced neurotrophin-induced neuronal demyelination and this effect was not connected to neuronal loss since the axons remained intact.³⁹ Since induced expression of endogenous Cav-1 provides recovery of myelination, a proper Cav-1 modulation can be used to improve the neuropathic pain.

Osteoarthritis is generally connected to development of inflammation and chronic pain. Peroxisome proliferator-activated receptor gamma (PPAR γ) demonstrates protective effect in osteoarthritis *in vivo*;⁴⁰ on the other hand, PPAR γ and its agonists are the known positive regulators of Cav-1 in different types of cells.^{41,42} Such interaction corresponds to a generally positive effect of Cav-1 induction in inflammatory conditions⁴³ and indicates that Cav-1 modulation can be a target in osteoarthritic pain.

Last but not least, muscle injury is a well-known source for acute and chronic pain. The process of muscle regeneration is substantially connected to modulation of Cav-1 expression in undifferentiated myogenic precursor (satellite) cells.⁴⁴ These cells quickly proliferate and fuse with damaged fibres after muscle injury. Under normal conditions, satellites cells are quiescent: they overexpress Cav-1 producing cellular senescence and cycle arrest, whereas suppression of Cav-1 induces their cycling. To initiate muscle regeneration, satellite cells must migrate to the wounded area shortly after injury. Simultaneously with this migration, Cav-1 level must be transiently downregulated. Recover of Cav-1 expression normally takes place three days after injury, which is a characteristic time for differentiation of precursors into myotubes. However, in a case of a high basal level of Cav-1 expression (e.g. in transgenic mice with overexpression of Cav-1) myogenic precursors are not able to reduce Cav-1 to a sufficiently low level allowing their migration, which can lead to impaired muscle regeneration.⁴⁴ From here, Cav-1 must also be an interesting target in muscle regeneration, and suppression of endogenous Cav-1 during the early stages of an acute muscle injury or in the case of delayed muscle recovery should provide improved healing.

US as a Local Modulator of Cav-1 Level in the Tissue

Caveolae are linked to the actin cytoskeleton and reorganization of this intracellular network can strongly modify their surface density.⁴⁵ Dependent on the amplitude of mechanical deformation, both stiffening and softening

(fluidization) of the cytoskeleton can occur.^{46,47} Whereas mechanical forces at a frequency of 1 Hz fluidize the cytoskeleton and modify the microdomain structure of the plasma membrane at strains of about 10%, application of mechanical forces at frequency 1 MHz reduces the critical strain to about 10^{-5} .⁴⁸ Application of higher US intensity and higher US frequencies induces higher levels of strain in cells and thus can strongly modify the caveolae,⁴⁹ indicating that US of higher frequencies (very high frequency US, VHF-US) should be more effective in pain treatment. Of note, as discussed above, US waves demonstrate frequency-dependent modulation of generation and propagation of APs in axonal membrane. Also, US-induced expression of Hsp72, which expression proceeds almost parallel to expression of Cav-1,²⁵ is strongly frequency-dependent and increases with increasing US frequency.⁵⁰

Analysis of expression profiles of US-induced genes revealed that the *CAVI* gene was indeed strongly upregulated after application of a low intensity US.⁵¹ At the same time, stress-dependent modulation of Cav-1 demonstrates a biphasic behavior: a low-level stimulation generally provides an enhanced Cav-1 expression whereas application of a strong mechanical stress will cause Cav-1 degradation.⁵² Additionally, US waves of different frequencies can induce very distinct values of mechanical stress in the cells, which can either properly modulate, not modulate or false modulate the local expression of Cav-1. Recent research also revealed that US can stimulate the mechanosensitive channels, thus directly influencing neurons and other excitable cells, and that the probability of such excitation strongly depends on the protocol of US application.⁵³ This can be an important reason for the mixed treatment outcomes observed after application of US for pain treatment. US intensity and frequency as well as the initial status of Cav-1 expression should be the main parameters defining direct and indirect interactions of US waves with axons and thus the observed pain release outcome.

Some Applications of VHF-US for Pain Treatment

From above, VHF-US should be effective in treatment of different types of peripheral pain. Applications of VHF-US with frequencies of 10 MHz and higher in different inflammatory and hyperproliferative conditions were discussed in our recent publications.^{46,47,54–57} Here we

shortly discuss some special clinical applications of VHF-US for pain reduction. All applications were provided by the emitter LDM[®]-MED (Wellcomet GmbH, Karlsruhe, Germany).

Effect of VHF-US in the form of dual US waves of 3 MHz and 10 MHz mixed in a special LDM[®] (Local Dynamical Micro-massage) mode on wound healing and pain reduction was investigated in a single-center, prospective pilot study on 10 patients with chronic venous leg ulcers.⁵⁸ In this mode, ultrasound waves are applied as short quickly oscillating pulse trains (in this study as 5 ms of 3 MHz and 5 ms of 10 MHz with spatial average temporal average ultrasound intensity (SATA) of 0.5–1.0 W/cm²). All patients demonstrated complete wound closure and a significant reduction of self-reported pain of at least three points measured on the 5-point numerical rating scale (NRS) just after 3–4 treatments ($p < 0.01$). The effect of VHF-US technology on the pain in patients treated with injection lipolysis (IL) was investigated in a randomized, single-center, contralateral, controlled pilot study on seven female healthy subjects.⁵⁹ Pain is a well-known side-effect of IL predominantly connected to a strong inflammatory reaction in the tissue. Patients were treated with LDM[®] (5 ms of 3 MHz and 5 ms of 10 MHz with SATA of 1.0 W/cm²) one time before and 10 times after injection lipolysis. Pain was assessed by patients using the 5-point NRS. Combinational IL + LDM[®] treatment provided not only a significant improvement of the fat tissue reduction compared to a pure IL, but also a significant reduction of pain ($p < 0.01$). A retrospective study on 58 patients who underwent breast reconstruction after mastectomy who complained about post-operative pain, which was not controllable with medication, was presented in Ahn et al.⁶⁰ Patients were treated with LDM[®] (10 MHz, SATA of 1.0 W/cm² and 1.5 W/cm² for 2 min each, then LDM[®] 3/10 MHz with SATA of 1.0 W/cm² and 2.0 W/cm² for 3 min each) every day or every other day. The degree of self-reported pain and discomfort with contracture were assessed by patients using the 10-point NRS. NRS score for pain was reduced from about 6.1 (pre-treatment) to about 3.8 (post treatment) ($p < 0.001$), corresponding NRS values for discomfort were 6.50 and 4.29, respectively ($p < 0.001$). Further, in a remarkable case report, the patient with radiation fibrosis and chronic pain induced by post-mastectomy radiotherapy was treated with LDM[®] (10 MHz, SATA of 1.0 W/cm² and 1.5 W/cm² for 2 min each, then LDM[®] 3/10 MHz with SATA of 1.0 W/cm² and 2.0 W/cm² for 3

min each).⁶¹ Treatment begun about 3 years after radiotherapy. After the second LDM[®] treatment, patient reported a significant pain reduction, which decreased from 6 to 2 on the 10-point NRS. These examples demonstrate that VHF-US can indeed be effectively applied for treatment of different types of pain.

Conclusions

Recent results obtained in a rapidly growing field of ultrasound neurostimulation demonstrated the important role of mechanical surface waves in generation and propagation of action potentials. Reciprocal interaction between electrical and mechanical waves observed in axonal membrane during its excitation leads to a paradigm shift in the pain treatment making the “uncoupling” of electro-mechanical signals an interesting target in pain treatment. This uncoupling can be realized either through direct disturbance of the mechanical surface waves in axonal membrane or through shifting of the thermodynamic state of this membrane far from its phase transition point. Whereas the first effect can be of a pure mechanical nature, the second one will demand application of the thermo-mechanical stress. Both effects can be effectively realized through application of the very high frequency ultrasound waves. Additional target for application of ultrasound in pain treatment is the caveolin-1, which is abundantly present in Schwann cells as well as in the non-axonal tissues. Recent research demonstrated that ultrasound can significantly modulate caveolin-1 expression in frequency-dependent manner, thus regulating the processes involved in inflammation, building of edema and injury repair in the affected area. Both targets demonstrate frequency-dependent reaction, thus making a very high frequency ultrasound a promising treatment modality in pain treatment. Further research will be needed to elucidate the peculiarities of ultrasound interaction with these new treatment targets.

Abbreviations

AP, action potentials; Cav, caveolin; COX, cyclooxygenase; Hsp, heat shock protein; IL, injection lipolysis; LDM, local mechanical micro-massage; NRS, numerical rating scale; PGE₂, prostaglandin E₂; PPAR γ , peroxisome proliferator-activated receptor gamma; TRPA1, receptor potential ion channel ankyrin 1; VHF-US, very high frequency ultrasound; US, ultrasound.

Disclosure

ILK is the managing partner of Wellcomet GmbH. Wellcomet GmbH provided support in the form of salaries for ILK, but did not have any additional role in decision to publish or preparation of the manuscript. The commercial affiliation of ILK with Wellcomet GmbH does not alter the adherence to all journal policies on sharing data and materials. The author reports no other conflict of interest in this work.

References

- Blackmore J, Shrivastava S, Sallet J, Butler CR, Cleveland RO. Ultrasound neuromodulation: a review of results, mechanisms and safety. *Ultrasound Med Biol*. 2019;45(7):1509–1536. doi:10.1016/j.ultrasmedbio.2018.12.015
- Tasaki I, Carnay L, Watanabe A. Transient changes in extrinsic fluorescence of nerve produced by electric stimulation. *Proc Natl Acad Sci USA*. 1969;64(4):1362–1368. doi:10.1073/pnas.64.4.1362
- Terakawa S. Potential-dependent variations of the intracellular pressure in the intracellularly perfused squid giant axon. *J Physiol*. 1985;369(1):229–248. doi:10.1113/jphysiol.1985.sp015898
- Heimburg T, Jackson AD. On the action potential as a propagating density pulse and the role of anesthetics. *Biophys Rev Lett*. 2007;2(1):57–78. doi:10.1142/S179304800700043X
- Yang Y, Liu XW, Wang H, et al. Imaging action potential in single mammalian neurons by tracking the accompanying sub-nanometer mechanical motion. *ACS Nano*. 2018;12(5):4186–4193. doi:10.1021/acsnano.8b00867
- Rvachev MM. On axoplasmic pressure waves and their possible role in nerve impulse propagation. *Biophys Rev Lett*. 2010;5(2):73–88. doi:10.1142/S1793048010001147
- El Hady A, Machta BB. Mechanical surface waves accompany action potential propagation. *Nat Commun*. 2015;6(1):1–7. doi:10.1038/ncomms7697
- Engelbrecht J, Peets T, Tamm K. Electromechanical coupling of waves in nerve fibres. *Biomech Model Mechanobiol*. 2018;17(6):1771–1783. doi:10.1007/s10237-018-1055-2
- Shrivastava S, Kang KH, Schneider MF. Collision and annihilation of nonlinear sound waves and action potentials in interfaces. *J R Soc Interface*. 2018;15(143):20170803. doi:10.1098/rsif.2017.0803
- Fichtl B, Shrivastava S, Schneider MF. Protons at the speed of sound: predicting specific biological signaling from physics. *Sci Rep*. 2016;6(1):22874. doi:10.1038/srep22874
- Prieto ML, Oralkan Ö, Khuri-Yakub BT, Maduke MC, Phillips W. Dynamic response of model lipid membranes to ultrasonic radiation force. *PLoS One*. 2013;8(10):e77115. doi:10.1371/journal.pone.0077115
- Menz MD, Ye P, Firouzi K, et al. Radiation force as a physical mechanism for ultrasonic neurostimulation of the ex vivo retina. *J Neurosci*. 2019;39(32):6251–6264. doi:10.1523/JNEUROSCI.2394-18.2019
- Wei W, Thiessen DB, Marston PL. Acoustic radiation force on a compressible cylinder in a standing wave. *J Acoust Soc Am*. 2004;116(1):201–208. doi:10.1121/1.1753291
- Shrivastava S, Cleveland RO, Schneider MF. On measuring the acoustic state changes in lipid membranes using fluorescent probes. *Soft Matter*. 2018;14(47):9702–9712. doi:10.1039/c8sm01635f
- Kruglikov IL. Modeling of the spatiotemporal distribution of temperature fields in skin and subcutaneous adipose tissue after exposure to ultrasound waves of different frequencies. *AIP Adv*. 2017;7(10):105317. doi:10.1063/1.4997833

16. Suchaoin W, Chanvorachote P. Caveolin-1 attenuates hydrogen peroxide-induced oxidative damage to lung carcinoma cells. *Anticancer Res.* 2012;32(2):483–490.
17. Zou H, Stoppani E, Volonte D, Galbiati F. Caveolin-1, cellular senescence and age-related diseases. *Mech Ageing Dev.* 2011;132(11–12):533–542. doi:10.1016/j.mad.2011.11.001
18. Mougeolle A, Poussard S, Decossas M, et al. Oxidative stress induces caveolin 1 degradation and impairs caveolae functions in skeletal muscle cells. *PLoS One.* 2015;10(3):e0122654. doi:10.1371/journal.pone.0122654
19. Chen Y-H, Lin -W-W, Liu C-S, Su S-L. H2O2 induces caveolin-1 degradation and impaired mitochondrial function in E11 podocytes. *Mol Med Rep.* 2017;16(5):7841–7847. doi:10.3892/mmr.2017.7497
20. Khalil Z, Liu T, Helme RD. Free radicals contribute to the reduction in peripheral vascular responses and the maintenance of thermal hyperalgesia in rats with chronic constriction injury. *Pain.* 1999;79(1):31–37. doi:10.1016/s0304-3959(98)00143-2
21. Stern CM, Mermelstein PG. Caveolin regulation of neuronal intracellular signaling. *Cell Mol Life Sci.* 2010;67(22):3785–3795. doi:10.1007/s00018-010-0447-y
22. Mikol DD, Scherer SS, Duckett SJ, Hong HL, Feldman EL. Schwann cell caveolin-1 expression increases during myelination and decreases after axotomy. *Glia.* 2002;38(3):191–199. doi:10.1002/glia.10063
23. Luo X, Tao L, Lin P, Mo X, Chen H. Extracellular heat shock protein 72 protects schwann cells from hydrogen peroxide-induced apoptosis. *J Neurosci Res.* 2012;90(6):1261–1269. doi:10.1002/jnr.22810
24. Guzlova I, Kislyakova K, Moskaliova O, et al. In vitro studies show that Hsp70 can be released by glia and that exogenous Hsp70 can enhance neuronal stress tolerance. *Brain Res.* 2001;914(1–2):66–73. doi:10.1016/s0006-8993(01)02774-3
25. Black AT, Hayden PJ, Casillas RP, et al. Regulation of Hsp27 and Hsp70 expression in human and mouse skin construct models by caveolae following exposure to the model sulfur mustard vesicant, 2-chloroethyl ethyl sulfide. *Toxicol Appl Pharmacol.* 2011;253(2):112–120. doi:10.1016/j.taap.2011.03.015
26. Kawabata A. Prostaglandin E2 and pain—an update. *Biol Pharm Bull.* 2011;34(8):1170–1173. doi:10.1248/bpb.34.1170
27. Liou JY, Deng WG, Gilroy DW, Shyue SK, Wu KK. Colocalization and interaction of cyclooxygenase-2 with caveolin-1 in human fibroblasts. *J Biol Chem.* 2001;276(37):34975–34982. doi:10.1074/jbc.M105946200
28. Rodriguez DA, Tapia JC, Fernandez JG, et al. Caveolin-1-mediated suppression of cyclooxygenase-2 via a β -catenin-Tcf/Lef-dependent transcriptional mechanism reduced prostaglandin E2 production and survivin expression. *Mol Biol Cell.* 2009;20(8):2297–2310. doi:10.1091/mbc.e08-09-0939
29. Mickle AD, Shepherd AJ, Mohapatra DP. Nociceptive TRP channels: sensory detectors and transducers in multiple pain pathologies. *Pharmaceuticals.* 2016;9(4):72. doi:10.3390/ph9040072
30. Khan A, Khan S, Kim YS. Insight into pain modulation: nociceptors sensitization and therapeutic targets. *Curr Drug Target.* 2019;20(7):775–788. doi:10.2174/138945012066619013114244
31. Startek JB, Talavera K. Lipid raft destabilization impairs mouse TRPA1 responses to cold and bacterial lipopolysaccharides. *Int J Mol Sci.* 2020;21(11):3826. doi:10.3390/ijms21113826
32. Koivisto A, Jalava N, Bratty R, Pertovaara A. TRPA1 antagonists for pain relief. *Pharmaceuticals.* 2018;11(4):117. doi:10.3390/ph11040117
33. De Logu F, Nassini R, Materazzi S, et al. Schwann cell TRPA1 mediates neuroinflammation that sustains macrophage-dependent neuropathic pain in mice. *Nat Commun.* 2017;8(1):1–16. doi:10.1038/s41467-017-01739-2
34. Razani B, Engelman JA, Wang XB, et al. Caveolin-1 null mice are viable but show evidence of hyperproliferative and vascular abnormalities. *J Biol Chem.* 2001;276(41):38121–38138. doi:10.1074/jbc.M105408200
35. Schubert W, Frank PG, Woodman SE, et al. Microvascular hyper-permeability in caveolin-1 (-/-) knock-out mice: treatment with a specific NOS inhibitor, L-NAME, restores normal microvascular permeability in Cav-1 null mice. *J Biol Chem.* 2002;277(42):40091–40098. doi:10.1074/jbc.M205948200
36. Maniatis NA, Kardara M, Hecimovich D, et al. Role of caveolin-1 expression in the pathogenesis of pulmonary edema in ventilator-induced lung injury. *Pulm Circ.* 2012;2(4):452–460. doi:10.4103/2045-8932.105033
37. Choi KH, Kim HS, Park MS, et al. Overexpression of caveolin-1 attenuates brain edema by inhibiting tight junction degradation. *Oncotarget.* 2016;7(42):67857. doi:10.18632/oncotarget.12346
38. McGuire JF, Rouen S, Siegfried E, Wright DE, Dobrowsky RT. Caveolin-1 and altered neuregulin signaling contribute to the pathophysiological progression of diabetic peripheral neuropathy. *Diabetes.* 2009;58(11):2677–2686. doi:10.2337/db09-0594
39. Yu C, Rouen S, Dobrowsky RT. Hyperglycemia and downregulation of caveolin-1 enhance neuregulin-induced demyelination. *Glia.* 2008;56(8):877–887. doi:10.1002/glia.20662
40. Fahmi H, Martel-Pelletier J, Pelletier JP, Kapoor M. Peroxisome proliferator-activated receptor gamma in osteoarthritis. *Mod Rheumatol.* 2011;21(1):1–9. doi:10.1007/s10165-010-0347-x
41. Llaverias G, Vázquez-Carrera M, Sánchez RM, et al. Rosiglitazone upregulates caveolin-1 expression in THP-1 cells through a PPAR-dependent mechanism. *J Lipid Res.* 2004;45(11):2015–2024. doi:10.1194/jlr.M400049-JLR200
42. Werion A, Joris V, Hepp M, et al. Pioglitazone, a PPAR γ agonist, upregulates the expression of caveolin-1 and catalase, essential for thyroid cell homeostasis: a clue to the pathogenesis of hashimoto's thyroiditis. *Thyroid.* 2016;26(9):1320–1331. doi:10.1089/thy.2015.0625
43. Kruglikov IL, Scherer PE. Caveolin as a universal target in dermatology. *Int J Mol Sci.* 2020;21(1):80. doi:10.3390/ijms21010080
44. Volonte D, Liu Y, Galbiati F. The modulation of caveolin-1 expression controls satellite cell activation during muscle repair. *FASEB J.* 2005;19(2):237–239. doi:10.1096/fj.04-2215fje
45. Echarri A, Del Pozo MA. Caveolae—mechanosensitive membrane invaginations linked to actin filaments. *J Cell Sci.* 2015;128(15):2747–2758. doi:10.1242/jcs.153940
46. Kruglikov IL, Scherer PE. Caveolin-1 as a pathophysiological factor and target in psoriasis. *NPJ Aging Mech Dis.* 2019;5(1):1–7. doi:10.1038/s41514-019-0034-x
47. Kruglikov IL, Scherer PE. Caveolin-1 as a target in prevention and treatment of hypertrophic scarring. *NPJ Regen Med.* 2019;4(1):1–7. doi:10.1038/s41536-019-0071-x
48. Mizrahi N, Zhou EH, Lenormand G, et al. Low intensity ultrasound perturbs cytoskeleton dynamics. *Soft Matter.* 2012;8(8):2438–2443. doi:10.1039/C2SM07246G
49. Samandari M, Abrinia K, Mokhtari-Dizaji M, Tamayol A. Ultrasound induced strain cytoskeleton rearrangement: an experimental and simulation study. *J Biomech.* 2017;60:39–47. doi:10.1016/j.jbiomech.2017.06.003
50. Sontag W, Kruglikov IL. Expression of heat shock proteins after ultrasound exposure in HL-60 cells. *Ultrasound Med Biol.* 2009;35(6):1032–1041. doi:10.1016/j.ultrasmedbio.2008.12.011
51. Yang Q, Nanayakkara GK, Drummer C, et al. Low-intensity ultrasound-induced anti-inflammatory effects are mediated by several new mechanisms including gene induction, immunosuppressor cell promotion, and enhancement of exosome biogenesis and docking. *Front Physiol.* 2017;8:818. doi:10.3389/fphys.2017.00818
52. Sinha B, Köster D, Ruez R, et al. Cells respond to mechanical stress by rapid disassembly of caveolae. *Cell.* 2011;144(3):402–413. doi:10.1016/j.cell.2010.12.031
53. Kubanek J, Shukla P, Das A, Baccus SA, Goodman MB. Ultrasound elicits behavioral responses through mechanical effects on neurons and ion channels in a simple nervous system. *J Neurosci.* 2018;38(12):3081–3091. doi:10.1523/JNEUROSCI.1458-17.2018

54. Kruglikov IL. Very high frequency ultrasound. *Hautarzt*. 2015;66(11):829–833. doi:10.1007/s00105-015-3676-z
55. Kruglikov IL. Sehr hochfrequenter Ultraschall in der ästhetischen Medizin und Chirurgie. *J Ästh Chir*. 2018;11(3):124–129. doi:10.1007/s12631-018-0140-9
56. Kruglikov IL, Zhang Z, Scherer PE. Caveolin-1 in skin aging. From innocent bystander to major contributor. *Ageing Res Rev*. 2019;55:100959. doi:10.1016/j.arr.2019.100959
57. Kruglikov IL, Scherer PE. Caveolin-1 as a possible target in the treatment for acne. *Exp Dermatol*. 2020;29(2):177–183. doi:10.1111/exd.14063
58. Kruglikov IL, Kruglikova E. Dual treatment strategy by venous ulcers: pilot study to dual-frequency ultrasound application. *J Cos Dermatol Sci Appl*. 2011;1(4):157–163.
59. Tausch I, Kruglikov I. The benefit of dual-frequency ultrasound in patients treated by injection lipolysis. *J Clin Aesthet Dermatol*. 2015;8(8):42–46.
60. Ahn KH, Lee SJ, Park ES, Park YG. Satisfaction with the effect of local dynamical micro-massage therapy on the pain and discomfort after breast reconstruction surgery. *Med Laser*. 2020;9(1):39–43. doi:10.25289/ML.2020.9.1.39
61. Choi YS, Park ES. Application of dual-frequency ultrasound to radiation-induced fibrosis in a breast cancer patient. *Med Laser*. 2017;6(2):86–89. doi:10.25289/ML.2017.6.2.86

Journal of Pain Research

Dovepress

Publish your work in this journal

The Journal of Pain Research is an international, peer reviewed, open access, online journal that welcomes laboratory and clinical findings in the fields of pain research and the prevention and management of pain. Original research, reviews, symposium reports, hypothesis formation and commentaries are all considered for publication. The manuscript

management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/journal-of-pain-research-journal>