ORIGINAL RESEARCH

Antimicrobial Susceptibility Pattern and Associated Factors of Pediatric Septicemia in Southern Ethiopia

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Gemechu Ameya¹ Temesgen Weldemedhin² Tsegaye Tsalla³ Feleke Gebremeskel⁴

¹Department of Medical Laboratory Sciences, Kotebe Metropolitan University, Addis Ababa, Ethiopia; ²Department of Public Health, Wolaita Sodo University, Wolaita Sodo, Ethiopia; ³Department of Medical Laboratory Sciences, College of Medicine and Health Sciences, Arba Minch University, Arba Minch, Ethiopia; ⁴School of Public Health, College of Medicine and Health Sciences, Arba Minch University, Arba Minch, Ethiopia

Correspondence: Gemechu Ameya Department of Medical Laboratory Sciences, Kotebe Metropolitan University, P.O. Box: 3268, Addis Ababa, Ethiopia Tel +251-91-783-7681 Email gemechuameya@gmail.com



Background: Septicemia is one of the major causes of morbidity and mortality in pediatric patients throughout the world. Drug-resistant pathogens are one of the major challenges to control. The study aimed to identify the major etiological agents, antimicrobial susceptibility pattern and associated factors of septicemia among pediatric patients in southern Ethiopia.

Methods: A cross-sectional study was conducted on pediatric patients. Blood samples were cultured and antimicrobial susceptibility testing was conducted by Kirby-Bauer disc diffusion techniques. Data were collected by pre-tested questionnaire to identify potential associated factors of septicemia. A bivariate logistic regression analysis was used and adjusted odds ratio with 95% CI at ≤ 0.05 level of significance was computed to determine the presence and strength of the association.

Results: Of 238 participants, 27 (11.3%) of them had a positive blood culture. *Staphylococcus aureus* (32.2%), coagulase negative Staphylococci (25%), and *Klebsiella pneumoniae* (14.3%) were the predominant isolates. The isolated bacteria showed high rates of resistance to amoxicillin, ceftriaxone, streptomycin and ampicillin. Multi-drug resistance (MDR) was observed in 82.1% of the isolates. Being infant [AOR=4.18, 95% CI, (1.3–13.0)], admission >10 days [AOR=5.54, 95% CI, (1.51–20.41)], burn [AOR=3.55, 95% CI, (1.02–12.38)] and wound cases [AOR=5.52, 95% CI, (1.50–20.34)] were associated with pediatric septicemia.

Conclusion: Gram positive bacteria were the predominant isolates and majority of isolates were MDR pathogens. Very young age, prolonged hospital stays, burn and wound cases were associated with pediatric septicemia. Establishing antibiotic stewardship is mandatory to minimize the high prevalence of drug resistance.

Keywords: antimicrobial susceptibility, multi-drug resistance, pediatric, septicemia

Introduction

Septicemia is caused by the presence of microorganisms within the bloodstream and their dissemination throughout the body with evidence of systemic responses towards those microbes with variable severity.¹ The introduction of bacteria to the bloodstream can be from the lungs, genitourinary tract, gastrointestinal tract, skin or soft tissue.² The pathogenesis of the case may vary depending on the virulence of the pathogen, the portal of entry, the susceptibility and response of the host, and the temporal evolution of the condition.² The predominant etiologic agents and their antimicrobial resistance rate may vary with geographic area and time period.³ This systemic dissemination of microbes can cause a life-threatening illness that gets

© 2020 Aneya et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms. bp and incorporate the Creative Commons Attribution — Non Commercial (unported, v3.0) License (http://creativecommons.org/licenses/by-nc/3.0/). By accessing the work you hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission foro Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, please see paragraphs 4.2 and 5 of our Terms (https://www.dovepress.com/terms.php). worse quickly from their spreading and releasing of toxins in the blood. As a result, septicemia requires rapid and aggressive diagnosis and antimicrobial treatment.⁴

Globally, 31.5 million cases occur each year and septicemia is the leading cause of death worldwide in the pediatric population resulting in an estimated 7.5 million deaths annually.^{5,6} In sub-Saharan African countries, septicemia is an important cause of illness and death in children, accounting for 30–70% of illness, making it a significant health problem.^{5,7} Septicemia is very common in children and can lead to complications such as shock, multi-organ failure and death.⁴

Multiple factors may be associated with pediatric sepsis. It may vary based on geographic area and health-care setup. Some of the identified associated factors of pediatric sepsis are presence of indwelling intravenous devices, use of steroids and immunomodulators, prolonged hospitalization, chronic antibiotic therapy, surgery, burns or bedsores and serious injuries.^{8–10} Underlying diseases such as chronic kidney disease,¹¹ hematologic malignancies (HMs),¹² immune suppressed individuals and HIV/ AIDS² can be associated with septicemia.

Multi-drug resistance in various bacterial pathogens has reached a pandemic level during the last two decades.¹³ This resistance pattern varies in accordance with geographic and regional location, health-care setup, and existing practice.^{14,15} The timely and appropriate use of antimicrobial drugs is very important and the only way to treat septicemia. However, antibiotics resistance is fast-growing problem in developing countries.⁷ а Inappropriate treatment of sepsis aggravates the disease which leads to death of patients and emergence of new drug- resistant strains. This has become a serious health problem with many economic and social impacts all over the world.¹⁵ The increasing frequency of antimicrobial resistance found in bloodstream infections is of great concern for African countries where access to care and broadspectrum antimicrobials is often limited.¹⁶ The infections caused by multi-drug resistant strains result in prolonged hospital stays, increased risk of death, and require treatment with more expensive antibiotics.¹⁶ Research conducted in Ethiopia revealed that drug resistance is increasing to commonly used antibiotics in the country.⁷

Only a few studies have been conducted to investigate the epidemiology and to provide viable alternative approaches in the management of septicemia in Ethiopia.^{1,5,7,15,17} No research has been conducted on pediatric septicemia in the study area. The aim of this

study is to identify the etiological agents and antimicrobial susceptibility patterns in septicemia and its associated factors among pediatric patients attending Arba Minch general hospital, southern Ethiopia.

Materials and Methods Study Design and Setting

An institutional based cross-sectional study design was conducted on pediatric patients attending the pediatric emergency, inpatient and outpatient departments of Arba Minch general hospital of southern Ethiopia. Arba Minch general hospital is in Arba Minch town, which is in Gamo zone, a regional state in in the south of the country. The hospital serves more than 1.5 million people.

Study Population

The study participants were sampled from pediatric patients in ranging in age from 1 month to 15 years old who were visiting the emergency, inpatient and outpatient departments of Arba Minch General Hospital during the data collection period (September 2018 to January 2019). Pediatric patient with clinical symptoms of septicemia were included in the study. Study participants who took antibiotic treatment within the last two weeks during the data collection time were excluded. Pediatric patients without care givers during the data collection period were also excluded from the study.

Sample Size Determination and Sampling Procedure

Sample size was determined using a single population proportion formula. The prevalence of septicemia and associated factors and multi-drug resistance proportion were considered to determine the maximum sample size. The maximum sample size was obtained from a study conducted in Addis Ababa, Ethiopia,¹ in which the hospital admission was 16.9% (p= 0.17), with margin of error (d= 0.05) and 95% confidence interval; the total sample size became 217. Then 10% non-respondent rate was added and the final sample size became 238.

A systematic random sampling technique was used to select the study participants. The skip interval was computed by taking the previous number of patients who visited the pediatric wards in a similar time frame. The k value was calculated by N/n formula; while N was the number of patients who visited the hospital in the similar period and n was the sample size. Then an integer between 1 to k was randomly selected to determine a random start point. Finally the sampling was done by every Kth interval.

Data Collection Procedures and Instrument

Semi-structured pretested questionnaire was used to collect data related to socio-demographic, socio-economic, clinical characteristics and potential associated factors after reviewing previous similar studies.^{1,5,7,15,17} Other relevant information such as previous antibacterial therapy, application of indwelling medical device, prolonged hospital stays, medical conditions and other factors for each patient were collected from patient charts. The data were collected by face to face interview.

Major Laboratory Examinations Blood Specimen Collection

The vein puncture site was disinfected with 70% alcohol and 2% tincture of iodine before collecting. A venous blood sample of 5 mL was collected for children older than 2 years and 2–3 mL for 2-year-old and younger patients. For each patent two samples were collected from different sites aseptically. The blood sample of each pediatric patient was collected before the beginning of antibiotic treatment.

Blood Culture

The collected blood samples were inoculated into blood culture bottles of tryptic soya broth (Oxoid, Hampshire, UK) by maintaining a minimum of 1:10 blood to broth ratio. The bottles were labeled with each patient's identification number, date and time of collection. The bottles containing specimens were transported to laboratory within 30 min at room temperature. Then blood culture bottles were incubated at 37°C and inspected daily for the presence of visible sign of microbial growth such as floccular deposits on the top of the layer, sub-surface turbidity, hemolysis, surface pellicle, gas production and/or coagulation of broth for 7 consecutive days. If any sign of microbial growth was observed in a blood culture bottle, sub-culture was done according to the method stated below.

Sub-Culture of Primary Blood Culture

For blood cultures that showed signs of microbial growth, sub-cultures were made onto blood culture plate (BAP), chocolate culture plate (CAP), and MacConkey & Mannitol salt agar (Oxoid, Hampshire, UK). The MacConkey & Mannitol salt agar plates were incubated in aerobic atmosphere whereas the blood and chocolate agar plates in candle jar at 37°C for 24–48 hours. Bacterial growths on the subcultured plate were isolated by their characteristic appearance such as colony morphology, effect on culture media, Gram reaction and other specific identification panels for biochemical reactions.

Bacteria Identification

The pure cultures of isolated organisms from the subculture plates were subjected to biochemical tests for further identification of the pathogen. The biochemical tests were done according to their Gram reaction. Gram negative organisms were identified using biochemical tests such as indole test, carbohydrate fermentation reaction in triple sugar iron agar, citrate utilization, urease test, motility test, lysine decarboxylase, lysine deaminase, and oxidase test. For Gram positive bacteria biochemical tests such as catalase, coagulase, and mannitol fermentation were used for identification according to methods described in CLSI.¹⁸

Antimicrobial Susceptibility Testing

Antimicrobial susceptibility testing for all bacterial isolates was done by disc diffusion assay on Mueller-Hinton agar plates according to Kirby-Bauer technique.¹⁸ Standard inoculums were adjusted to 0.5 McFarland standard and four drug disks were used in a single 110 mm plate.

The following antimicrobial disks were used in the given concentrations: ampicillin (AMP) (10 μ g), amoxicillinclavulanic acid (AUG) (30 μ g), ceftriaxone (CTR) (30 μ g), cephalothin (CLT) (30 μ g), ciprofloxacin (CPR) (5 μ g), chloramphenicol (CHL) (30 μ g), doxycycline (DOX) (30 μ g), erythromycin (ERY) (15 μ g), gentamicin (GEN) (10 μ g), kanamycin (K) (30 μ g), cefoxitin (CXT) (30 μ g), penicillin (PEN) (10 IU), streptomycin (STR) (10 μ g), tetracycline (TET) (30 μ g), trimethoprim-sulphamethoxazole (COT) (1.25/23.75 μ g) and amoxicillin (AMC) (30 μ g). The result was interpreted by measuring zone of inhibition as sensitive, intermediate, and resistant according to the standardized CLSI.

Quality Control

Quality control of the questionnaires was done before starting the actual data collection. Pre-test questionnaire was done on 5% of the sample size one week before data collection. The completeness of data was checked before data entry. Standard operating procedures were strictly followed throughout the laboratory analysis. Quality control of the culture media was done whenever a new batch of media was prepared. Visual inspections for cracks in media, unequal fill, hemolysis, evidence of freezing, presence of air bubbles and any sign of contamination were conducted before inoculating the culture media. Performance of the culture media was checked by standard known bacteria specifically *Escherichia coli* ATCC 25,922, *S. aureus* ATCC 25,923, *Enterococcus faecalis* (ATCC-29,212) and *Pseudomonas aeruginosa* (ATCC-27,853).

Data Processing and Analysis

Data were analyzed using SPSS version 20.0 software package. Data completeness and consistency were checked by running cross tabulation of each variable. A bivariate logistic regression was performed to show any association between independent variables and the outcome variable (septicemia). A variable with p-value ≤ 0.25 in the bivariate logistic regression were included in multivariable logistic regression analysis to identify variables that independently associated with the outcome variable. The presence and strength of association between independent variables and outcome variable were computed by adjusted odds ratio with 95% CI and Hosmer–Lemeshow test was used to test the fitness of model.

Ethical Approval and Consent to Participate

The study was conducted in accordance with the Declaration of Helsinki. An ethical clearance was obtained from Arba Minch University, College of Medicine and Health Sciences institutional ethical review board. Before data collection, a permission letter was obtained from the relevant departments of the Arba Minch general hospital. Prior to sample collection participants were informed clearly about the objective and procedure of the study. Written informed consent and assent was obtained from all participants and their care givers. Participation in the study was fully voluntary. All information obtained in this study was kept confidential at all levels and utilized only for the study. For positive findings, we communicated with the attending physician for appropriate treatment of the pediatric patients.

Results

Socio-Demographic Characteristics of Study Participants

A total of 238 pediatric patients participated in the study. Out of these participants, 120 (50.4%) were females. The mean age and standard deviation of the participants were 2.25-year-old \pm 1.024. The infants were slightly above one-quarter while children with age range from 1 to 4 years old were more than one-third of the participants. Underweight children (BMI < 18.5kg/m²) accounted for 18% of the participants. More than half of the study participants were urban dwellers (Table 1).

Out of all pediatric care givers about 29% of them were illiterate (unable to read or write). The majority of the care givers were governmental employees, followed by farmers. The monthly income of 56% of care givers was less than 1000 Ethiopian Birr (< USD 40). Nearly three out of four care givers had a family size of 5-9 while about 13% of them had 10 and more (Table 1).

Clinical Characteristics of Study Participants

Among study participants, slightly less than two-thirds of the patients attended the outpatients department while the others were admitted in pediatrics emergency and pediatrics inpatient ward. From all admitted pediatric patients, only 21 (24.7%) were admitted for more than 10 days at the time of data/sample collection. The length of hospital stay from the date of admission to blood sample collection was 1–21 days with a mean of 3 days admission. Usage of intravenous device during treatment was observed in 61 (25.6%) of the study participants and about 4.2% of the participants had used antibiotics for a prolonged time. Among the study participants, 2.5%were HIV/AIDS positive (Table 2).

Magnitude and Bacterial Isolates

Out of 238 study participants, 27 (11.3%) blood culture results were positive for different bacterial species. From the positive cultures 28 bacteria were identified and one of the blood cultures showed mixed growth. Gram positive bacterial species were the predominant (57.1%) isolate. The Gram positive bacteria isolated were *S. aureus* (32.2%) and coagulase negative Staphylococci (CONS) (25.0%). Among Gram negative bacteria *K. pneumoniae* (14.3%) and *E. coli* (10.7%) were predominantly isolatedfollowed by *Salmonella* species (7.1%), *Proteus mirabilis* (7.1%) and *P. aeruginosa* (3.6%) (Figure 1).

Factors Associated with Pediatric Septicemia

All socio-demographic and clinical factors of study participants were analyzed for their association to septicemia by

Variables	Frequency	Percent (%)
Sex		
Male	118	49.6
Female	120	50.4
Age (Year)		
<	66	27.7
I_4	83	34.9
5–9	53	22.3
10–15	36	15.1
Pediatric BMI		
<18.5kg/m ²	42	17.6
18.5–24.9kg/m ²	188	79
>25kg/m ²	8	3.4
Resident		
Urban	128	53.8
Rural	110	46.2
Educational status of care givers		
Illiterate	69	29
Primary education	51	21.4
Secondary education	77	32.4
Tertiary education	41	17.2
Occupational status care givers		
Government employee	46	19.3
Farmer	43	18.1
Merchant	41	17.2
Daily labor	40	16.8
Private organizations	40	16.8
House servant	17	7.2
Others*	П	4.6
Family Size		
≤4	25	10.5
5–9	182	76.5
≥ 10	31	13
Monthly Income of Parents (Birr)		
< 1000	133	55.9
≥1000	105	44.1

Table I Socio-Demographic & Socio-Economic Characteristics of

using bivariate analysis with backward logistic regression. Age group between 1 to 11 months had a statistically significant association to septicemia [AOR= 4.18, 95 CI, (1.34, 13.00)]. Participants who were admitted to hospital admission for more than 10 days were about 6 times more likely to develop septicemia [AOR= 5.54, 95 CI, (1.51-20.41)]. Patients with burn injuries were about 3.5 times more likely to develop septicemia compared with the other participants [AOR=3.55, 95 CI, (1.02-12.38)]. Participants with

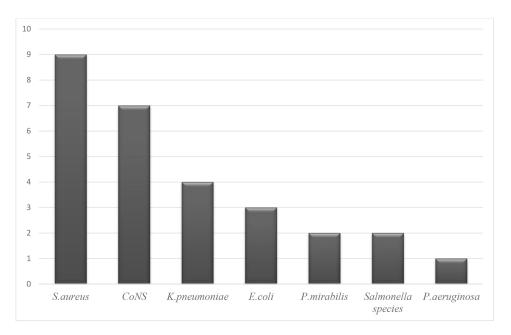
Variables	Frequency	Percent		
Hospital Admission				
Yes	85	35.7		
No	153	64.3		
Admission Date				
<10	64	75.3		
≥10	21	24.7		
Medical Procedures Usage				
Intravenous device	61	25.6		
Endotracheal tube	6	2.5		
Surgery	4	1.7		
Prolonged antibiotic usage				
Yes	10	4.2		
No	228	95.8		
Chronic Disease				
HIV/AIDS	6	2.5		
Others*	9	3.8		
Clinical Diagnosis				
Burns case	23	9.7		
Respiratory disease	47	19.7		
Febrile illness	87	36.6		
Wound case	17	7.1		
Urinary tract disease	17	7.1		
Gastrointestinal disease	21	8.8		
Skin disease	16	6.7		
Others**	11	4.1		

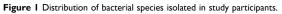
Notes: Others* includes diabetes mellitus, chronic liver disease, chronic kidney disease and asthmatic patient; Others** includes skin disease, heart disease, generalized swelling.

wounds were about 5.5 times more likely to develop septicemia compared with their counterparts [AOR=5.52, 95 CI, (1.50–20.34)] (Table 3).

Antimicrobial Susceptibility Patterns of **Bacterial Isolates**

Fifteen antimicrobial drug disks were used to check susceptibility pattern of Gram positive bacterial isolates. All Gram positive bacterial isolates showed resistance to ceftriaxone, high resistance to amoxicillin (93.8), ciprofloxacin, tetracycline and erythromycin. On the other hand, the lowest resistance rate was observed against cefoxitin, chloramphenicol and amoxicillin-clavulanic acid while resistance was not observed to cephalothin. More than 77.8% of S. aureus isolates were resistant to amoxicillin, ceftriaxone, doxycycline, tetracycline, erythromycin, ciprofloxacin, penicillin, ampicillin and trimethoprim-sulfamethoxazole. In contrast the





lowest resistance rate was observed for chloramphenicol (11.1%), and amoxicillin-clavulanic acid. From the total *S. aureus* isolates about 22.2% were methicillin resistant *S. aureus* (MRSA). Coagulase negative staphylococci isolates were highly resistant to ceftriaxone and amoxicillin, however the lowest resistance was observed to chloramphenicol, ampicillin, cefoxitin and doxycycline. All CONS

isolates showed no resistance to cephalothin and amoxicillinclavulanic acid (Table 4).

A total of 12 different antibacterial drug disks were used to examine susceptibility patterns of Gram negative bacterial isolates. All Gram negative isolates showed resistance to streptomycin and amoxicillin. High resistance was observed against cephalothin, ampicillin,

Variables	Positive N (%)	Negative N (%)	COR (95% CI)	AOR (95% CI)	P-value	
Age						
I–II months	17 (25.8)	49 (74.2)	4.80 (1.77, 12.99)	4.18 (1.34, 13.00)	0.014	
I-4 years	4 (4.8)	79 (95.2)	0.70 (0.19,2.58)	0.68 (0.17, 2.78)	0.591	
5–15 years	6 (6.7)	83 (93.3)	Ref	Ref	Ref	
Admission Date						
No admissions	11 (7.2)	142 (92.8)	Ref	Ref	Ref	
≤10	6 (9.4)	58 (90.6)	0.99 (0.34, 2.95)	0.81 (0.24, 2.67)	0.723	
>10	10 (47.6)	11 (52.4)	10.68 (3.78, 30.21)	5.54 (1.51, 20.41)	0.010	
Wound case						
Yes	7 (41.2)	10 (58.8)	7.04 (2.41, 20.50)	5.52 (1.50, 20.34)	0.010	
No	20 (9)	201 (91.0)	Ref	Ref	Ref	
Burn case						
Yes	8 (34.8)	15 (65.2)	5.50 (2.07,14.64)	3.55 (1.02, 12.38)	0.047	
No	19 (8.8)	196 (91.2)	Ref	Ref	Ref	
Usage of intravenous device						
Yes	9 (14.8)	52 (85.2)	1.53 (0.65, 3.61)	0.41 (0.09, 1.89)	0.253	
No	18 (10.2)	159 (89.8)	Ref	Ref	Ref	

 Table 3 Factors Associated with Pediatric Septicemia in Study Participants

Abbreviations: COR, crude odds ratio; AOR, adjusted odds ratio; CI, confidence interval; Ref, reference.

Antibiotics	S. aureus	; (%)		CONS (%)		Total (%	Total (%)			
	S	I	R	s	I	R	s	I	R		
CTR	0	0	100	0	0	100	0	0	100		
AMC	0	0	100	0	14.3	85.7	0	6.2	93.8		
AMP	22.2	0	77.8	71.4	14.3	14.3	43.8	6.2	50		
DOX	22.2	0	77.8	85.7	0	14.3	50	0	50		
CPR	11.1	11.1	77.8	14.3	42.9	42.9	12.5	25	62.5		
CHL	55.6	33.3	11.1	85.7	0	14.3	68.8	18.8	12.5		
CLT	88.9	11.1	0	85.7	14.3	0	87.5	12.5	0		
К	22.2	44.4	33.3	14.3	57.1	28.6	18.8	50	31.2		
TET	22.2	0	77.8	57.1	0	42.9	37.5	0	62.5		
GEN	44.4	22.2	33.3	42.9	28.6	28.6	43.8	25	31.2		
СХТ	77.8	0	22.2	85.7	0	14.3	81.2	0	18.8		
СОТ	22.2	0	77.8	71.4	0	28.6	43.8	0	56.2		
ERY	11.1	0	88.9	28.6	42.9	28.6	18.8	18.8	62.5		
PEN	0	22.2	77.8	0	71.4	28.6	0	43.8	56.2		
AUG	55.6	22.2	22.2	71.4	28.6	0	62.5	25	12.5		

Table 4 Antimicrobial Susceptib	vility Pattern of Gram-Positive Ba	acteria Isolated from Study Participants
Table 4 Anumicrobial Susceptio	Dility Fattern of Grani-Fositive Da	icteria isolated ironi study rarticipants

Abbreviations: CTR, ceftriaxone; AMC, amoxicillin; AMP, ampicillin; DOX, doxycycline; AUG, amoxicillin-clavulanic acid; CLT, cephalothin; CHL, chloramphenicol; CPR, ciprofloxacin; GEN, gentamicin; TET, tetracycline; K, kanamycin; PEN, penicillin; COT, trimethoprim-sulfamethoxazole; CXT, cefoxitin; ERY, erythromycin; S, sensitive; I, intermediate; R, resistance; MRSA, methicillin resistant *Staphylococcus aureus; MSSA*, methicillin sensitive *Staphylococcus aureus*.

trimethoprim-sulfamethoxazole, ciprofloxacin, doxycycline, tetracycline, amoxicillin-clavulanic acid and gentamicin. *K. pnemoniae* were resistant to amoxicillin, ampicillin, cephalothin, amoxicillin-clavulanic acid and streptomycin. *P. aeroginosa* isolates were only sensitive to gentamicin and intermediate to ciprofloxacin and doxycycline while the isolate was resistant to the rest of all the tested antibiotics. No resistance was observed to ciprofloxacin, doxycycline and trimethoprim- sulfamethoxazole by *E. coli* and *Salmonella* species while highresistance to amoxicillin, streptomycin and gentamicin was observed. *P. mirabilus* was highly resistant to amoxicillin, tetracycline, doxycycline and streptomycin but sensitive to ciprofloxacin, chloramphenicol, trimethoprim-sulfamethoxazole and ceftriaxone (Table 5).

From the total of 28 isolates tested for antimicrobial susceptibility tests, multiple drug resistance (MDR) was observed in 23 (82.1%). Among the MDR isolates, 12 (three-quarters) of the isolates were from Gram positive and 11 (91.7%) isolates were from Gram negative bacteria. All isolated *K. pnemoniae, E. coli, P. aeruginosa* and *P. mirabilis* were MDR from Gram negative bacteria. All *S. aureus* isolates and nearly half of CONS were MDR from Gram positive bacterial (Figure 2). In general, trimethoprim-sulfamethoxazole, ciprofloxacin and chloramphenicol were effective drugs against Gram negative bacteria. However, Gram negative isolates were highly

resistant to streptomycin, amoxicillin-clavulanic acid, ampicillin, amoxicillin and cephalothin. Cefoxitin, amoxicillin-clavulanic acid, cephalothin and chloramphenicol were effective drugs for Gram positive bacteria but ceftriaxone and amoxicillin were ineffective.

Discussion

The results of this study showed that the overall culture positivity rate of septicemia identified from pediatric patients was 11.3% [95% CI (7.9%, 16%)] which was similar with studies conducted in Trinidad $(10.1\%)^3$, Addis Ababa, Ethiopia (8.9% and 12.4%)^{1, 4}, Zanzibar (9.6%)¹⁹ and Uganda (14%).²⁰ The present study finding was also in line with global epidemiology (8.2%).²¹ Bacterial sepsis remains a major cause of pediatric morbidity and mortality despite advances in laboratory diagnosis and treatment.²²

A relatively higher rate of septicemia was reported compared with studies conducted in Mekelle, Ethiopia (7.4%),¹⁵ Nepal (7.28%,²³ 7.48%,²⁴ 7.7%²⁵), Europe and Australia/New Zealand.²¹ The reason of this variation may be due to study differences in the socio-demographic factors and health-care system. North America, Europe and Australia/New Zealand showed lower pediatric sepsis prevalence which may be related to good implementation of infection prevention, early identification of cases and effective treatment of identified cases. On the other hand,

Bacteria Isolates ((%)	Antibacterial Drugs											
		CTR	AMC	AMP	DOX	CPR	CHL	CLT	TET	GEN	сот	STR	AUG
K. pneumoniae	S	0	0	0	50	75	0	0	75	0	100	0	0
	I	50	0	0	25	25	75	0	0	50	0	0	0
	R	50	100	100	25	0	25	100	25	50	0	100	100
E. coli	S	0	0	0	100	100	66.7	0	66.7	0	100	0	0
	I	33.3	0	0	0	0	33.3	0	33.3	0	0	0	0
	R	66.7	100	100	0	0	0	100	0	100	0	100	100
P. mirabilis	S	100	0	0	0	100	100	0	0	0	100	0	100
	I	0	0	50	0	0	0	50	0	50	0	0	0
	R	0	100	50	100	0	0	50	100	50	0	100	0
Salmonella species	S	50	0	0	100	100	100	0	100	0	100	0	100
	I	50	0	50	0	0	0	50	0	0	0	0	0
	R	0	100	50	0	0	0	50	0	100	0	100	0
P. aeruginosa	S	0	0	0	0	0	0	0	0	100	0	0	0
	I	0	0	0	100	100	0	0	0	0	0	0	0
	R	100	100	100	0	0	100	100	100	0	100	100	100
Total	S	25	0	0	58.3	83.3	50	0	58.3	8.3	91.7	0	33.3
	I	33.3	0	16.7	16.7	16.7	33.3	16.7	8.3	25	0	0	0
	R	41.7	100	83.3	25	0	16.7	83.3	33.3	66.7	8.3	100	66.7

Table 5 Antimicrobial Susceptibility Pattern of Gram Negative Bacteria Isolated from Study Participants

Abbreviations: CTR, ceftriaxone; AMC, amoxicillin; AMP, ampicillin; DOX, doxycycline; AUG, amoxicillin-clavulanic acid; CLT, cephalothin; CHL, chloramphenicol; CPR, ciprofloxacin; GEN, gentamicin; TET, tetracycline; COT, trimethoprim-sulfamethoxazole; STR, streptomycin; S, sensitive; I, intermediate; R, resistance.

the findings of present study were lower than the study conducted in India (27%)²⁶ and Nigeria (16.4%).²⁷ The possible reason for the lower prevalence in our study may be differences in the socio-demographic and socioeconomic status of the participants and difference in the number of the study participants.

In the current study Gram positive organisms were more frequently isolated than Gram negative organisms. A similar finding was observed in the study conducted in Gondar, Ethiopia⁷ and in Cape Town, South Africa.¹⁶ Staphylococcus aureus was the most prevalent etiological agent for bloodstream bacterial infections and this finding

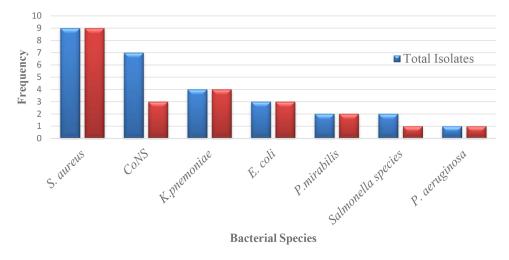


Figure 2 Multi-drug resistance isolates among identified bacteria from blood sample.

is comparable with other studies conducted in different areas.^{15,17,28} Coagulase negative staphylococci (CONS) was the other common cause of bacterial infection in the bloodstream which was consistent with other studies conducted in India, Egypt and Ethiopia.^{5,29,30} The high prevalence of CONS might be in association with an increasing burden of nosocomial infection in hospitalized patients. The pathogen was previously considered as a culture contaminant but is now recognized to be an important cause of bloodstream infections.^{31,32} In the present study *K. pneumoniae* was the predominant pathogen among Gram negative organisms followed *by E. coli*. Similar findings was observed in studies conducted in Ethiopia and Nigeria.^{1,7,27,33}

In the current study different factors were found to be associated with septicemia. Participants in the infant age group were about 4 times more likely to developsepticemia compared with children above 5 years old. This finding is in concordance with other research conducted in Nepal and the USA.^{22,34} The possible reason for this is the infants have less immunity compared with the older children to protect themselves from the invasion of pathogens. In addition to this the bodies of infants are less colonized by normal microbiota. The presence of wellestablished resident normal microbiota has a significant role in protection from pathogenic bacteria.³⁵

Prolonged hospitalization was the other factor associated with septicemia in the present study. The finding is in line with another study conducted in Ethiopia.¹ This may be associated with nosocomial infection. The associated factors to the burden of septicemia are increasing worldwide due to the high number of patients admitted to hospital for prolonged periods. Septicemia accounts for 10–20% of all nosocomial infections and is one of the common causes of mortality in the hospitalized patients.⁷ The high number of hospitalizations is due to increasing levels of chronic illness and to medical procedures such as surgery and intravenous devices which could be risk factors for infection.

Burn and wound cases had a statistically significant association with bloodstream infections in this study. Our finding of association of septicemia with burn cases was in agreement with studies conducted in the USA.^{10,34} Patients with wounds had a statistically significant association with bloodstream infection and this may be due to transepidermal water loss and blood flow from their initial wounding and in the later stages and post-operatively.³⁶ The main reason for this high burden of pediatric sepsis may be a lack of continuous evaluation or surveillance in the management system and also the likelihood of implementing internationally accepted management of cases in low income countries.^{5,23,37} Unavailability of laboratory setup for routine examination of blood culture and antimicrobial susceptibility tests is also another challenge to overcome to prevent the development of sepsis in wound and burn cases.^{4,26}

The antimicrobial resistance pattern was varied among isolated pathogens. In the present study all Gram positive organisms showed resistance for ceftriaxone. Similar findings were observed in studies conducted in Egypt and Nigeria.^{27,29} In the current study the majority of isolated organisms of Gram positive and Gram negative bacteria were resistant to amoxicillin and ampicillin, similar to studies conducted in Egypt and Ghana.^{28,29} Most of the Gram positive organisms were resistant to tetracycline and erythromycin in the present study. A similar finding was observed in a study conducted in Egypt.²⁹ Most of the Gram negative organisms were resistant to cephalothin, gentamicin, streptomycin and amoxicillin-clavulanic acid which showed similarity to studies conducted in Nigeria and India.^{30,33} The possible explanation for such high resistance might be uncontrolled use of antimicrobial drugs in the study area.

K. pneumoniae, E. coli, Salmonella species, *P. mirabilis* and *P. aeruginosa* showed 100% resistance for amoxicillin and streptomycin. This finding is in line with other studies conducted in Ethiopia.^{1,2} *P. aeruginosa* was only sensitive to gentamicin and the isolate was resistant to the other tested drugs in the present study. This finding was similar to Ethiopian and Indian studies.^{5,7,26} However, this study finding was different from a study conducted in Gondar, Ethiopia that showed *P. aeruginosa* was resistant to gentamycin.² The observed difference may be due to difference in the use of antibiotic drugs and the presence of microbiology laboratory.

In the current study multi-drug resistance was observed in 82.1% of isolates which was similar to other studies conducted in Ethiopia which showed resistance of 89.1%, 80% and 65%.^{1,5,17} The present study had a higher multi-drug resistance rate compared with several other studies.^{23,25,38} Multi-drug resistance was observed to be higher in Gram negative bacteria than Gram positive bacteria. This finding was in line with other studies conducted in Ethiopia and north India.^{1,38} All *S. aureus. K. pneumoniae, E. coli, P. aeruginosa* and *P. mirabilis* strains showed multi-drug resistance similar to studies conducted in Ethiopia and Nepal.^{1,5,25} The observed multi-drug resistance may be due to uncontrolled use of antimicrobial drugs in the study area. Treatment of bacterial infection without drug sensitivity test is the other possible reason for the observed high prevalence.

Conclusion

The study showed that Gram positive bacteria were the predominant etiologic agents in pediatric septicemia. *Staphylococcus aureus*, CONS, *K. pneumoniae* and *E. coli* were the commonest causative agents. Infants, prolonged hospitalization, burns, and wound cases were independently associated with pediatric septicemia. Gram positive bacteria were more resistant to ceftriaxone and amoxicillin while Gram negative bacteria were resistant to amoxicillin, ampicillin and streptomycin. High multi-drug resistance rates were observed in most of the isolates. Establishing hospital antibiotic stewardship is essential to minimize the high prevalence of drug resistance. Working on the identified associated factors is vital to minimize the observed high prevalence of pediatric septicemia.

Abbreviations

AIDS, acquired immunodeficiency syndrome; ATCC, American Type Culture Collection; BMI, body mass index; CLSI, Clinical Laboratory Standards Institute; *CONS*, coagulase negative Staphylococci; COR, crude odds ratio; MDR, multi-drug resistance; SPSS, Statistical Package for Social Sciences.

Author Details

GA (gemechuameya@gmail.com), TM (temewmn@gmail. com), TT (tsegayetsa@yahoo.com), FG (fgfpeace@gmail. com).

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Disclosure

The authors declare that they have no competing interests.

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