Butorphanol Inhibits the Malignant Biological Behaviors of Ovarian Cancer Cells via Down-Regulating the Expression of TMEFF1

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Purpose: An important issue with compounds for treating ovarian cancer is the development of drug resistance and side effects. Butorphanol is a synthetic opioid. Opioids have been shown to promote or prevent tumor growth and metastasis. This research aimed to reveal the affection of Butorphanol on the malignant biological behaviors of ovarian cancer cells.

Methods: Different concentrations of Butorphanol were used to treat ovarian cancer cell lines, ES-2 and SKOV3. Biological functions of cells were performed by CCK-8 assay, colony formation assay, apoptosis analysis, transwell assays and scratch assays. The differences in the transcriptome of the Butorphanol treated and negative control (NC) cells were analyzed by RNA-Seq.

Results: Butorphanol treatment significantly inhibited the viability, colony-forming, migration and invasion of ES-2 and SKOV3 cells compared to NC. Furthermore, Butorphanol treatment obviously induced the apoptosis of ES-2 and SKOV3 cells and regulated the expression of apoptosis-related proteins. Additionally, Butorphanol treatment significantly reduced the expression of p-AKT, p-mTOR and P70S6K without affecting the expression of AKT and mTOR in ES-2 and SKOV3 cells. Forty-four genes were identified to up-regulate its expression, while 17 genes were identified to down-regulate its expression in Butorphanol-treated cells. Among them, TMEFF1 was found to be significantly down-regulated in Butorphanol-treated cells. Additionally, the restoration of TMEFF1 expression complemented the inhibitory effect of Butorphanol treatment on cell proliferation and invasion.

Conclusion: In conclusion, Butorphanol is a compound with potential to treat ovarian cancer. TMEFF1 may play a key role in inhibiting the malignant proliferation and metastasis of Butorphanol treatment on ovarian cancer cells.

Keywords: Butorphanol, ovarian cancer, TMEFF1, RNA-Seq, proliferation

Introduction

Ovarian cancer is one of the most deadly gynecological malignancies in the world.¹,² About 70% of patients with ovarian cancer show peritoneal metastasis in early diagnosis.³ Although great progress has been made in the therapy of ovarian cancer in recent years, an important issue with compounds for treating tumors is the development of drug resistance and side effects.⁴,⁵ Therefore, there is an urgent need to identify improved anti-tumor drugs.

Butorphanol is a synthetic opioid that has partial agonist and partial antagonist activity to the µ type opioid receptors, as well as antagonist and partial agonist
activity at the 𝜅 opioid receptor. It can be used as a nasal spray to treat migraine and parenterally as a narcotic analgesic for moderate to severe pain or as an adjunct to general anesthesia. Opioids have been documented either to promote or to prevent tumor growth and metastasis.

TMEFF1 is a transmembrane protein consisting of a epidermal growth factor (EGF)-like region and two follistatin domains. One study finds the differential expression of TMEFF1 in a variety of tumors including brain cancer, pancreatic cancer and liver cancer. Exogenously overexpressing TMEFF1 inhibits the proliferation of brain cancer cell lines. In addition, differential methylation of TMEFF1 have been found in both breast and colon cancer. These studies indicate the potential role of TMEFF1 in cancer, but require more detailed research.

In present research, we found that Butorphanol treatment significantly inhibited the malignant biological behaviors of ES-2 and SKOV3 cells. Then, TMEFF1 was found to be significantly down-regulated in Butorphanol-treated cells. Additionally, the restoration of TMEFF1 expression complemented the inhibitory effect of Butorphanol treatment on cell proliferation and invasion.

Materials and Methods

Cell Culture and Treatment

Human ovarian cancer cell lines (ES-2 and SKOV3) and normal cervical cell line (HCK1T) were obtained from the Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China). The cells were maintained in RPMI 1640 culture medium (Gibco, Grand Island, NY) containing 10% FBS (Thermo Fisher Scientific, Waltham, MA) at 37 °C and 5% CO2 in an incubator. Butorphanol tartrate injection was purchased from Hengrui Medicine Co., Ltd. (Jiangsu, China). Cells were treated with different concentrations of Butorphanol (0, 2.5, 5, 10, 20, 30, 40, 50, 60, 80, 120, 200 μg/mL). The TMEFF1 cDNA sequence was cloned into the pcDNA3.1 vector. And recombinant plasmid was transfected into cells by Lipofectamine 2000 (Invitrogen, Carlsbad, CA).

CCK8 Assay

Five thousand cells were planted in each well of a 96-well plate, and incubated in different concentrations of Butorphanol. Then 10 μL CCK-8 solution (Dojindo, Kumamoto, Japan) was added into each well. This was followed by the incubation of the plate for 2 h at 37 °C. Finally, a microplate reader was used to measure the absorbance at 450 nm.

Colony Formation Assay

Five hundred cells were planted in each well of a 6-well plate, and incubated with Butorphanol for 10 days. Then colonies were fixed with 10% neutral formalin for 1 h and dyed with crystal violet (Beyotime, Haimen, China). The cells were photographed under a microscope (Olympus, Tokyo, Japan).

Transwell Assay

For cell migration, 2000 cells were seeded into the upper chamber of transwell chambers (Corning Incorporated, Corning, NY) and 200 μL of serum-free RPMI-1640 medium containing different concentrations of Butorphanol was added. The lower chamber was added with 600 μL of RPMI-1640 medium containing 10% FBS. After 24 h incubation at 37°C, the surface of each upper chamber was wiped gently with a cotton swab. Transwell chambers were fixed with 4% paraformaldehyde at room temperature for 30 min and stained with crystal violet for 30 min. The tumor cells infiltrating the filter membrane on the surface of the lower chamber were then counted under a microscope (Olympus) and the cell number is expressed as the average number of the cells in each field. For cell invasion, Matrigel (BD) was spread onto the upper wells of Transwell chambers and the other steps were consistent with cell migration experiments.

Scratch Assay

When the cell monolayer reached 90%, the microplate was scratched gently with a 10 μL pipette tip. After washing with PBS, medium was replaced with the serum-free culture medium containing difference concentration of Butorphanol. The width of scratch was observed under a microscope. After 24 h of culturing, the width of each scratch was measured under a microscope.

Detection of Cell Apoptosis by Flow Cytometry

5 x 10³ cells were cultured in each well of a 6-well plate, and incubated in different concentrations of Butorphanol for 48 h. Then, cells were harvested and resuspended in binding buffer. The cells were stained with 5 μL Annexin V-FITC plus 5 μL propidium iodide (BD Biosciences, Franklin Lakes, NJ). The stained cells were added of 400 μL PBS. The percentage of apoptosis was analyzed by using flow cytometry (Tree Star, Ashland, OR).
Western Blot
Treated cells were dissolved with RIPA buffer at 4°C for 10 min. The total protein was collected by centrifugation at 12,000×g for 30 min (4°C) and quantified using a bicinchoninic acid method (Beyotime, Shanghai, China). Proteins were separated using 10% SDS-PAGE and transferred to a PVDF membrane (EMD Millipore, Billerica, MA). After blocking with 5% fat-free milk for 1 h, each primary antibody was incubated overnight at 4°C. After washing with TBST, secondary antibody was incubated for 1 h at room temperature. Protein bands were developed using the ECL reagent (Thermo Fisher Scientific).

The primary antibodies used in this study were as follow: anti-Bcl2 (1:1000, ab32124), anti-Bax (1:1000, ab32503), anti-cleaved caspase3 (1:1000, ab13847), anti-GAPDH (1:5000, ab8245), anti-AKT (1:5000, ab8805), anti-p-AKT (1:5000, ab38449), anti-mTOR (1:5000, ab2732), anti-p-mTOR (1:5000, ab84400), anti-P70S6K (1:5000, ab131526) were purchased from Abcam. Anti-TMEFF1 (1:5000, T4829) was purchased from MERCK.

Statistical Analysis
The raw image data files obtained by high-throughput sequencing (Illumina HiSeq) were converted into raw data of sequencing sequences by CASAVA base calling analysis, and the results were stored in the FASTQ file. Bowtie2 was used to align clean reads to the reference sequence, and RSEM was used to calculate the expression levels of genes and transcripts. According to the DEseq2 method (based on the principle of negative binomial distribution), differentially expressed genes (DEG) were detected. According to the differential gene detection results, the heatmap map function in R software was used to perform hierarchical cluster analysis. The data were analyzed with the SPSS 22.0 software. Each experiment was repeated at least three times. The results were presented as mean ± SD. Student’s t-test was used for comparisons between two independent groups, and one-way ANOVA was used to compare differences among more than two groups. P < 0.05 indicated that the difference was statistically significant.

Results
Butorphanol Inhibits the Viability and Colony-Forming Ability of Ovarian Cancer Cells
To reveal the affection of Butorphanol on the malignant biological behaviors of ovarian cancer cells, the viability and colony-forming ability of ovarian cancer cells were firstly examined by CCK8 and colony formation assays. As shown in Figure 1A, the results of CCK8 assay showed the absorbance at 450 nm of ES-2 and SKOV3 cells which were treated with different concentrations of Butorphanol for 24 h. The IC 50 of Butorphanol on ES-2 cells was 31.23 μg/mL, while the IC 50 of Butorphanol on SKOV3 cells was 39.90 μg/mL. Therefore, these concentrations (30 μg/mL to ES-2 cells and 40 μg/mL to SKOV3 cells) were used in subsequent experiments. As shown in Figure 1B, Butorphanol treatment significantly inhibited the viability of ES-2 and SKOV3 cells compared into negative control (NC) group (P < 0.05). Moreover, the concentration of Butorphanol was safe for normal cervical cells (Supplementary figure 1). Similarly, Butorphanol treatment obviously reduced the colony-forming ability of ES-2 and SKOV3 cells compared into NC group (Figure 1C, P < 0.05).

Butorphanol Impedes the Migration and Invasion of Ovarian Cancer Cells
Furthermore, the affection of Butorphanol on the migration and invasion of ovarian cancer cells was measured by Transwell and scratch assays. As shown in Figure 2AC, Butorphanol treatment significantly inhibited the migration and invasion capacities of ES-2 and SKOV3 cells compared into NC group (P < 0.05).

Butorphanol Induces the Apoptosis of Ovarian Cancer Cells
Flow cytometry was used to detect the apoptosis of ovarian cancer cells. As shown in Figure 2D, the apoptotic rate was significantly increased after the Butorphanol treatment (P < 0.05). In addition, the expression of Bcl2 was decreased, while the expression of Bax and cleaved-caspase 3 was increased after the Butorphanol treatment (Figure 3A, P < 0.05).

Butorphanol Inhibits the Activation of AKT/mTOR Signaling Pathway in Ovarian Cancer Cells
Previous studies showed that AKT/mTOR signaling pathway is involved in various human diseases including cancers.13,14 Therefore, Western blot was utilized to measure the key proteins expression of AKT/mTOR signaling pathway. As shown in Figure 3B, Butorphanol treatment significantly reduced the expression of p-AKT, p-mTOR and P70S6K (P < 0.05) without affecting the expression of AKT and mTOR in ES-2 and SKOV3 cells.
Butorphanol Inhibits the Expression of TMEFF1 in Ovarian Cancer Cells

The differences in the transcriptome of the Butorphanol treated and NC cells were analyzed by RNA-Seq (Beijing Genomics Institute) (Figure 4A). qRT-PCR was used to confirm the accuracy of the analyze results (results not shown). The results of RNA-Seq performed that 44 genes were up-regulated, while 17 genes were down-regulated when cells were treated with Butorphanol (Figure 4B and C). Through literature search and
experimental verification, TMEFF1 gene was selected for the subsequent research.

As shown in Figure 4D, Butorphanol treatment significantly reduced the expression of TMEFF1, while co-transfection of TMEFF1 over-expression plasmid restored the TMEFF1 expression. Additionally, the restoration of TMEFF1 expression complemented the inhibitory effect of Butorphanol treatment on cell proliferation and invasion (Figure 4E and F).

Discussion

Globally, 240,000 women are diagnosed with ovarian cancer each year, and the 5-year survival rate is less than 45%, resulting in 150,000 deaths.15 Although the age-standardized ratio has stabilized or declined in most high-income countries, the age-standardized ratio has increased in many low- and middle-income countries. In addition, as life expectancy increases, the number of confirmed cases each year also increases. Therefore, there is an urgent need to identify new antitumor drugs.

Butorphanol is a competitive μ-receptor antagonist that provides analgesia through an agonist effect on κ-receptors.7 In the United States, Butorphanol is used to treat anesthesia, anesthesia aids, labor pain and pain, as a member of the drug class of narcotic analgesics. It will provide a new possibility for cancer treatment. In this study, the affection of Butorphanol on the viability of ovarian cancer cells was firstly detected by CCK8 assays. The IC 50 of Butorphanol on ES-2 cells was 31.23 μg/mL, while the IC 50 of Butorphanol on SKOV3 cells was 39.90 μg/mL. Furthermore, Butorphanol treatment significantly inhibited the proliferation, migration and invasion of ES-2 and SKOV3 cells. Additionally, Butorphanol treatment obviously induced the apoptosis of ovarian cancer cells by regulating the expression of Bcl2, Bax and
cleaved-caspase 3. These data reveal that Butorphanol could inhibit ovarian cancer progression. In addition, the concentrations of Butorphanol, which significantly inhibit the proliferation of cervical cancer cells, is safe for normal cervical cells. This demonstrates the possibility of Butorphanol in the treatment of cervical cancer. Although there is still a lack of pharmacological and pharmacokinetic experiments for Butorphanol.

We also found that Butorphanol treatment inhibited the activation of AKT/mTOR signaling pathway in ovarian cancer cells. The results of Western blot showed that Butorphanol treatment significantly reduced the expression of p-AKT, p-mTOR and P70S6K without affecting the expression of AKT and mTOR in ES-2 and SKOV3 cells. These data reveal that Butorphanol inhibits the malignant biological behaviors of ovarian cancer cells by regulating the AKT/mTOR signaling pathway, which has the potential to treat ovarian cancer. Butorphanol is a synthetic opioid. The sequencing results indicated that the cell lines (ES-2 and SKOV3) used in this study expressed opioid receptors. However, Butorphanol treatment did not cause changes in the expression of opioid receptors. The sequencing results showed that 44 genes were up-regulated its expression, while 17 genes were down-regulated its expression in Butorphanol-treated cells. The differential expression of these genes may play a key role in inhibiting the malignant proliferation and metastasis of Butorphanol treatment on ovarian cancer cells. Among them, TMEFF1 was found to be significantly down-regulated in Butorphanol-treated cells. Additionally, the restoration of TMEFF1 expression complemented the inhibitory effect of Butorphanol treatment on cell proliferation and invasion. Some recent studies find that some effects of opioids are not mediated by opioid receptors. For example, Morphine decreased matrix metalloprotease-9 (MMP-9) and increased tissue inhibitor of metalloproteases-1 (TIMP-1) in multiple experimental models, with consequences on tumor cell migration and invasion. Some studies show that opioids or their
Figure 4 Butorphanol inhibits the expression of TMEFF1 in ovarian cancer cells.

Notes: (A–C) The differences in the transcriptome of the Butorphanol treated and NC cells were analyzed by RNA-Seq. (D) Western blot assay was performed to examine the expression of TMEFF1. (E) CCK8 assay was used to examine cell viability of ES-2 and SKOV3 cells. (F) Cell migration and invasion were assessed by Transwell assay. *P<0.05, compared to NC group; #P<0.05, compared to Butorphanol group.

Abbreviations: negative control, NC; non-DEGs, non-difference expression genes.
metabolites can activate toll-like receptor 4 (TLR4), which is expressed on cancer cells and tumor-related cells and regulates metastasis signaling pathways. Although the sequencing results showed that Butorphanol treatment did not affect TLR4 expression. Therefore, we speculate that its effect does not seem to be related to opioid receptor and TLR4 activation.

Initial research showed that TMEFF1 is mainly expressed in the brain. Subsequent research found that TMEFF1 is widely expressed in a variety of tissues, and play different role. For example, TMEFF1 is highly expressed in normal brain tissues, and lowly expressed in brain tumor tissue. Exogenously overexpressing TMEFF1 inhibits the proliferation of brain cancer cell lines. Additionally, TMEFF1 is up-regulated in pancreatic and lung cancer cells. Hypermethylation of TMEFF1 is found in blood samples from patients of colon cancer. In TP53 mutant- and TP53 wild-type breast cancer samples, differences in methylation of CpG island of the TMEFF1 promoter are also found. Cancer-testis (CT) Ags are compelling targets for immunotherapy strategies, because they are abnormally expressed in malignant cells and not or limited expressed in somatic tissues, except germ cells. One study identifies TMEFF1 as a CT Ag candidate in multiple myeloma. These results indicate that the expression of TMEFF1 plays an important role in the occurrence and development of many types of tumors.

Conclusion
In conclusion, Butorphanol is a compound with potential to treat ovarian cancer. TMEFF1 may play a key role in inhibiting the malignant proliferation and metastasis of Butorphanol treatment on ovarian cancer cells.

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