

AZINI-AS1, A Novel Oncogenic LncRNA, Promotes the Progression of Non-Small Cell Lung Cancer by Regulating MiR-513b-5p and DUSP11

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Background: Emerging researches have demonstrated that aberrantly expressed long non-coding RNAs (lncRNAs) have great significance in non-small cell lung cancer (NSCLC) progression. The aim of this study was to explore the role of lncRNA AZIN1 antisense RNA 1 (AZIN1-AS1) in NSCLC and the related mechanism.

Methods: Expressions of AZIN1-AS1 and miR-513b-5p in NSCLC samples were detected by qRT-PCR. NSCLC cell lines (H1299 and HCC827) were used in vitro assays. CCK-8 assay, EdU assay, wound healing test and Transwell assay were carried out to test the biological influence of AZIN1-AS1 on NSCLC cells. Subcutaneous xenotransplanted tumor model and tail vein injection model were established to test the role of AZIN1-AS1 in vivo. Interactions between AZIN1-AS1 and miR-513b-5p, miR-513b-5p and dual-specificity phosphatase 11 (*DUSP11*) were determined by bioinformatic analysis, qRT-PCR, Western blot, and luciferase reporter assay.

Results: AZIN1-AS1 was up-regulated in NSCLC cells and tissues, while miR-513b-5p was significantly down-regulated. Silencing of AZIN1-AS1 or overexpression of miR-513b-5p markedly inhibited proliferation, migration and invasion of NSCLC cells, while overexpression of AZIN1-AS1 or inhibition of miR-513b-5p functioned oppositely. Importantly, AZIN1-AS1 mediated the promotion of malignancy of NSCLC cells was reversed by miR-513b-5p mimics. What's more, AZIN1-AS1 could down-regulate miR-513b-5p via sponging it, and there existed a negative correlation between AZIN1-AS1 expression and miR-513b-5p expression in NSCLC samples. AZIN1-AS1 also enhanced the expression levels of *DUSP11*, which was proved as a target gene of miR-513b-5p. Further in vivo experiments showed that silencing of AZIN1-AS1 decreased tumor growth and metastasis, which was accompanied by overexpression of miR-513b-5p and inhibition of *DUSP11* in tumor tissues.

Conclusion: AZIN1-AS1 acts as a tumor promoter in NSCLC, which is ascribed to the regulation of miR-513b-5p and *DUSP11*.

Keywords: non-small cell lung cancer, AZIN1-AS1, miR-513b-5p, DUSP11

Introduction

Lung cancer is with the highest morbidity and mortality among cancers worldwide.¹⁻⁴ As the dominant pathologic type, non-small cell lung cancer (NSCLC) accounts for 80% of lung cancer.⁵ Although conventional treatment strategies (such as surgery, radiotherapy or chemotherapy) prolong the survival time of patients, the prognosis of NSCLC is still unsatisfactory, and the 5-year survival rate is less than 20%.^{6,7} It is necessary to figure out the molecular mechanism and look for new therapeutic targets of NSCLC.

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Long non-coding RNAs are transcripts with length exceeding 200 nucleotides and without the ability to encode proteins. Previous studies have found that lncRNAs can control gene expression at epigenetic, transcriptional and post-transcriptional levels. LncRNAs are key regulators in various biological activities, such as chromatin modification, intranuclear transport, genomic imprinting, X chromosome silencing, transcriptional interference and transcriptional activation.⁸ Recent studies have shown that abnormal expression of lncRNAs is associated with many human diseases, and they are involved in the growth, invasion, metastasis and relapse of cancers.⁹ LncRNA AZIN1 antisense RNA 1 (AZIN1-AS1) was located on human chromosome 8q22.3, and its role and mechanism in cancer are still unclear.

MicroRNAs (miRNAs) are small non-coding RNAs with a length of about 18–24 nucleotides. They regulate the post-transcriptional process via binding to the 3'UTR of target gene, thus resulting in inhibition of translation or degradation of the mRNA and being involved in the regulation of cellular proliferation, differentiation and apoptosis.¹⁰ Accumulating studies show that miRNAs participate in the tumor progression.¹¹ MiR-513b-5p originates from the precursor RNA transcribed from human chromosome Xq27.3. The expression of miR-513b-5p is remarkably down-regulated in testicular embryonic cancer, and it could impede the proliferation and metastasis of cancer cells by repressing interferon regulatory factor 2.¹² Our previous study demonstrated that miR-513b-5p was down-regulated in NSCLC tissues and it could significantly inhibit the proliferation, invasion, migration, and promote apoptosis of NSCLC cells by regulating mTOR signaling pathway.¹³ However, the mechanism of miR-513b-5p dysregulation in NSCLC remains unclear.

In this study, we demonstrated that the expression of AZIN1-AS1 was significantly up-regulated in NSCLC tissues. Gain- and loss-of-function experiments confirmed that AZIN1-AS1 could promote the proliferation, migration and invasion of NSCLC cells, suggesting that AZIN1-AS1 could play a carcinogenic role in the progression of NSCLC. To further explore the downstream mechanism, we found that overexpression of AZIN1-AS1 could up-regulate the expression of dual-specificity phosphatase 11 (*DUSP11*), playing the role of competitive endogenous RNA (ceRNA) to regulate the expression of miR-513b-5p. This study explored the role of AZIN1-AS1/miR-513b-5p/*DUSP11* axis in controlling NSCLC progression,

which provided new potential targets for NSCLC treatment.

Materials and Methods

Tissue Samples

This study obtained the approval of the Ethics Committee of Shanghai Pulmonary Hospital. Written informed consents were acquired from each participating patient and all protocols were conducted in accordance with the principles of the Declaration of Helsinki. Thirty-six patients with primary NSCLC underwent NSCLC resection in our hospital from May 2017 to May 2018 were selected in this study. In the control group, the specimens were from adjacent tissues of the same patient (at least 3 cm away from the surgical margin), and no cancer cells were found by pathological examination after the operation. All specimens were stored in liquid nitrogen at -196°C .

Cell Culture and Cell Transfection

Human normal lung epithelial cell lines BEAS-2B and NSCLC cell lines (HCC827, NCI-H460, A549, H1299) were purchased from American Type Culture Collection (Rockville, USA). These cells were cultured in RPMI-1640 medium (ThermoFisher Scientific, Shanghai, China) supplemented with 10% fetal bovine serum (FBS) (Invitrogen, Carlsbad, CA, USA), 100U/mL penicillin and 100mg/mL streptomycin (Invitrogen, Carlsbad, CA, USA), and cultured in an incubator at 37°C with 5% CO_2 . AZIN1-AS1 overexpression plasmid (pcDNA3.1-AZIN1-AS1), knockdown plasmid (carrying AZIN1-AS1 shRNA), *DUSP11* overexpression plasmid (pcDNA3.1-DUPS11), hsa-miR-153b-5p mimics and inhibitors, and corresponding control plasmids and miRNAs were obtained from GenePharma (Shanghai, China). The above vectors or miRNAs were transfected into cells using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. 1 $\mu\text{g}/\text{mL}$ puromycin (Beyotime, Shanghai, China) was added into the medium to obtain stable cell lines transfected with plasmids.

Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Total RNA in tissues and cells was extracted by TRIzol reagent (Invitrogen, Shanghai, China). miRNA was extracted with mirPremier[®] microRNA Isolation Kit (Sigma, St. Louis, MO, USA). Total RNA was retrieved by First Strand cDNA Synthesis Kit (Thermo Fisher

Scientific Inc., Rockford, IL, USA). In line with the manufacturer's instructions, qRT-PCR was conducted with Light Cycler Fast Start DNA MasterPlus SYBR Green I kit (Roche Diagnostics, Burgess Hill, UK) to analyze the expression of AZIN1-AS1 and GAPDH. TaqMan microRNA assay and TaqMan Universal Master Mix II (Applied Biosystems, Foster City, CA, USA) were used to analyze the expression of miR-513b-5p and U6. The primers were designed and synthesized by RiboBio Co., Ltd. (Guangzhou, China).

Cell Counting Kit-8 (CCK-8) Assay

HCC827 and H1299 cells in logarithmic growth phase were trypsinized, centrifuged and resuspended. After adjusting the cell density to 2×10^4 /mL, the cells were inoculated into a 96-well plate with 100 μ L cell suspension per well. After that, the 96-well plate was placed in an incubator for further culture. The absorbance of cells was measured in 24, 48, 72 and 96 hour, respectively. In brief, each well was incubated in the incubator for 1 hour with 10 μ L CCK-8 solution (Dojindo Molecular Technologies, Kumamoto, Japan). After the termination of the culture, the 96-well plate was placed in a microplate reader to determine the absorbance (OD value) of each well at 450 nm wavelength.

Ethynyl-Deoxyuridine (EdU) Assay

EdU kit (RiboBio Co., Ltd., Guangzhou, China) was also used to evaluate the proliferation of NSCLC cells. HCC827 and H1299 cell lines were seeded into 24-well plates (containing a cover slip in each well), respectively, and cultured for 24 hours, and then incubated with EdU (final concentration of 50 mmol/L) for 2 hours. Then, the medium was discarded and the cover slips were washed gently with PBS for 3 times. After that, the cells were fixed in 4% paraformaldehyde for 10 min. Then, the cells were incubated with fluorescent staining solution for 30 minutes, washed with methanol twice, incubated with $1 \times$ Hoechst 33,342 for 30 minutes. After the cover slips were washed with PBS for three times and sealed, the images were obtained under fluorescence microscope (Olympus, Tokyo, Japan). The percentage of EdU-positive cells was calculated.

Transwell Assay

Cells were collected and resuspended in serum-free RPMI-1640 medium. Then, cells were added into the upper compartment of Transwell chamber (8 μ m pore diameter, BD Biosciences, CA, USA) coated with Matrigel (BD,

San Jose, CA, USA), while the 10% FBS-containing medium was added in the lower compartment. After 24 h's culture, cells remaining in the upper compartment were wiped off with a cotton swab. Cells passing through the membrane were fixed with 4% polyformaldehyde and stained with crystal violet solution. At last, the cells were counted under a microscope.

Subcutaneous Xenotransplanted Tumor Model

Nude mice (6-week old) were purchased from the Laboratory Animal Center of Nanjing University. Cells in logarithmic growth phase from AZIN1-AS1 overexpression group or vector group, were trypsinized to prepare single-cell suspension. The cell density was adjusted to 2×10^7 /mL. One hundred μ L cell suspension was subcutaneously inoculated to the left side (AZIN1-AS1 overexpression group) and right side (vector group) of each mouse. The longest diameter (L) and shortest diameter (W) of the tumors were measured every 3 days, and the volume of tumors (V) = $0.5 \times L \times W^2$ was calculated. In the lung metastasis study, there were 10 mice in each group. 1×10^7 cells of AZIN1-AS1 overexpression group or vector group were injected into the tail vein of the mice. Two weeks later, the mice were killed by cervical decapitation. Pathological examination was used to quantify lung metastasis. All procedures were reviewed and approved by the Animal Study Committee of Shanghai Pulmonary Hospital, and in compliance with the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals and Animal Welfare Act.

Dual-Luciferase Reporter Assay

The wild-type (WT) AZIN1-AS1 sequence or the WT *DUSP11* 3'-UTR fragment containing predicted binding sites with miR-513b-5p was amplified and inserted into the pmirGLO dual-luciferase miRNA target expression vector (Promega Corp., Madison, WI, USA) to construct the report vector pmirGLO-AZIN1-AS1-WT or pmirGLO-*DUSP11*-WT. GeneArt Site-Directed Mutagenesis PLUS System (Thermo Fisher Scientific Inc., Rockford, IL, USA) was used to mutate the presumed binding site of miR-513b-5p with AZIN1-AS1 or *DUSP11* 3'-UTR. Mutant (MUT) AZIN1-AS1 or *DUSP11* 3'-UTR were inserted into pmirGLO dual-luciferase miRNA target expression vector to construct report vector pmirGLO-AZIN1-AS1-MUT or pmirGLO-*DUSP11*-MUT. The

corresponding reporter vectors and miR-513b-5p or NC mimics were co-transfected into HEK293 cells and the cells were cultured for 48 h. After that, the luciferase activity of the cells in each group was measured using the dual-luciferase reporter assay system (Promega, Madison, WI, USA).

Western Blot

RIPA lysate (containing 1% PMSF) was used to extract total proteins from different groups of cells. Protein concentration was determined by BCA method. Protein samples were separated by SDS-PAGE, and transferred to polyvinylidene fluoride (PVDF) membranes (Millipore, Bedford, MA, USA). The membranes were blocked with 5% skimmed milk for 1 h at room temperature, and then the primary antibody (anti-*DUSP11*, ab127299, 1:1000, abcam) was used to incubate the membrane at 4°C overnight. After washing the membranes with TBST buffer, horseradish peroxidase (HRP)-labelled secondary antibody (Beyotime, Shanghai, China) was added and the membranes were incubated at 37°C for 1 h. After washing the membranes with TBST buffer again, the signals were detected by ChemiLucent ECL Detection system (Millipore, Bedford, MA, USA).

Statistical Methods

Statistical software SPSS 17.0 (SPSS Inc., Chicago, IL, USA) was used to analyze all the experimental data in this study. The data were expressed as mean \pm standard deviation ($\bar{x} \pm s$). *t*-test was used to compare the differences between two groups. $P < 0.05$ was statistically significant.

Results

Expression and Clinical Significance of AZIN1-AS1 in NSCLC

To preliminarily detect the expression level of AZIN1-AS1 in cancers, we searched the expressions of AZIN1-AS1 in multiple cancer tissues using GEPIA database (<http://gepia.cancer-pku.cn/>) and in NSCLC tissues using StarBase databases (<http://starbase.sysu.edu.cn/>). As shown, AZIN1-AS1 expression was up-regulated in NSCLC tissues (Figure 1A and B). To further validate the high expression of AZIN1-AS1 in NSCLC, we detected the expression level of AZIN1-AS1 in NSCLC tissues and cell lines using qRT-PCR. The results showed that the expression levels of AZIN1-AS1 in NSCLC tissues and cell lines were increased significantly compared

with the control group (Figure 1C and D). Additionally, we examined the correlation between the expression level of AZIN1-AS1 and clinicopathological parameters of NSCLC patients. Patients were separated into AZIN1-AS1 high expression group and AZIN1-AS1 low expression group. The results suggested that the high expression of AZIN1-AS1 was significantly associated with an increase in tumor volume and the higher T stage (Table 1). These results hinted that AZIN1-AS1 was involved in the progression of NSCLC, and probably functioned as an oncogenic lncRNA.

The Effects of AZIN1-AS1 on Proliferation and Metastasis of NSCLC Cells in vitro and in vivo

To explore the function of AZIN1-AS1 in NSCLC, we successfully constructed AZIN1-AS1 knockdown and overexpression cell lines with NSCLC cell lines H1299 and HCC827, respectively (Figure 2A and B). CCK-8 and EdU experiments showed that compared with the control group, the proliferation of NSCLC cells was, respectively, enhanced and repressed after overexpression and knockdown of AZIN1-AS1 (Figure 2C–E). Wound healing test and Transwell test were used to detect cell migration and invasion. Compared with the control group, the migration and invasion abilities of tumor cells were significantly increased after up-regulating of AZIN1-AS1 in cells, while the knockdown of AZIN1-AS1 in cells had the opposite effect (Figure 2F and G). In addition, in vivo experiments with nude mice showed that overexpression of AZIN1-AS1 significantly promoted the growth of transplanted tumors (Figure 2H); pulmonary metastasis experiment showed that 7 out of 10 mice in AZIN1-AS1 overexpression group of had severe pulmonary metastasis, and the incidence was markedly higher than that in the control group (1 out of 10) (Figure 2I).

AZIN1-AS1 Can Target MiR-513b-5p

To explore the downstream molecular mechanism of AZIN1-AS1, we used StarBase to search for the potential targets of AZIN1-AS1. As shown in, AZIN1-AS1 contains a potential binding site of miR-513b-5p (Figure 3A). The levels of miR-513b-5p in NSCLC tissues and cells were further detected. The results of qRT-PCR indicated that the expression of miR-513b-5p was down-regulated in NSCLC tissues and cells (Figure 3B and C). Further experiments indicated that the expression levels of AZIN1-AS1 and miR-513b-5p in NSCLC tissues were negatively correlated (Figure 3D).

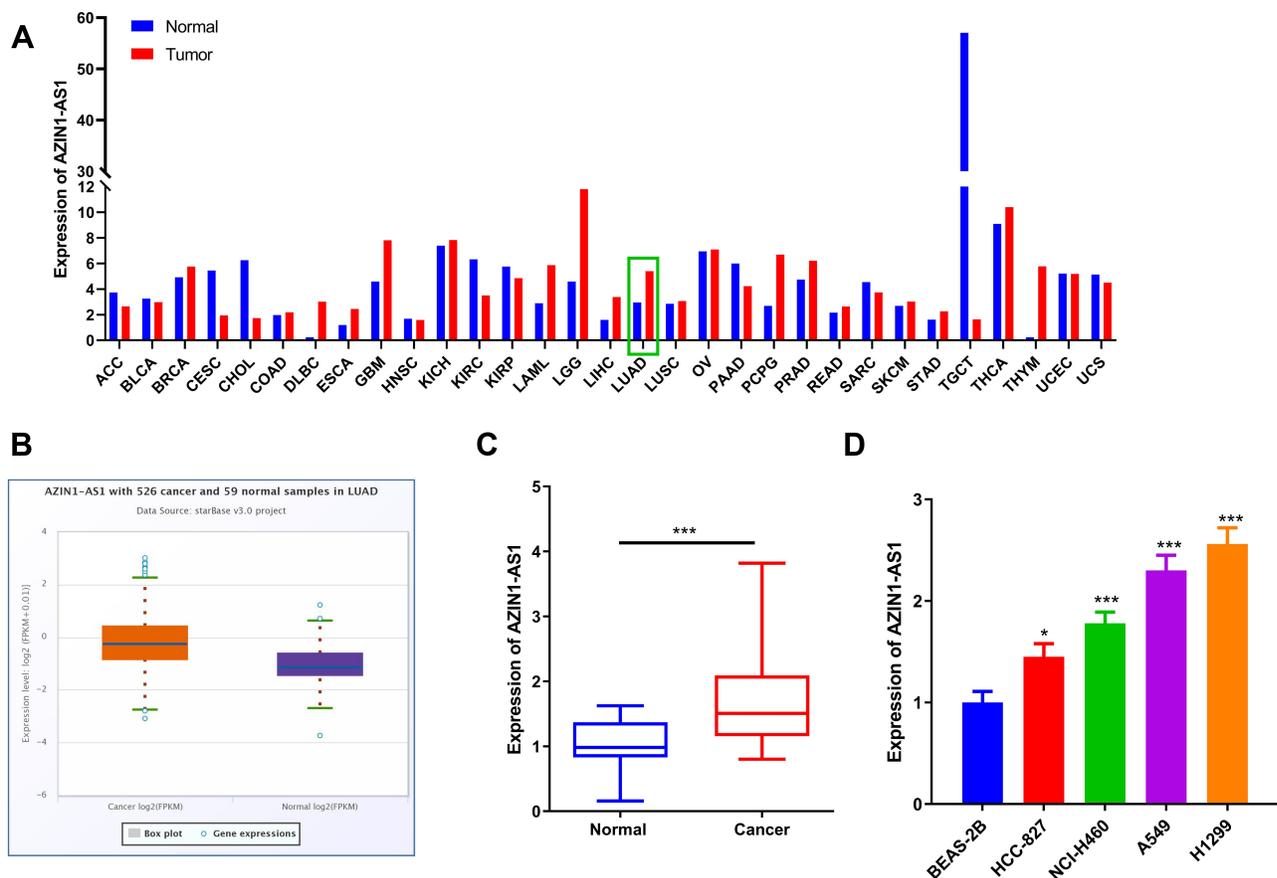


Figure 1 Expression characteristics of AZIN1-AS1 in NSCLC tissues and cell lines. **(A)** The expression levels of AZIN1-AS1 in cancer tissues and normal tissues was analyzed by GEPIA (TCGA data; LUAD: lung adenocarcinoma). **(B)** The expression levels of AZIN1-AS1 in normal tissues adjacent to cancer and NSCLC tissues in StarBase database. **(C)** The expression of AZIN1-AS1 in NSCLC tissues and normal tissues adjacent to cancer was detected by qRT-PCR. **(D)** The expression of AZIN1-AS1 in normal lung epithelial cell lines BEAS-2B and NSCLC cell lines (HCC-827, NCI-H460, A549 and H1299) was determined by qRT-PCR. *, ***Represent $P < 0.05$, and $P < 0.001$ respectively.

Moreover, the luciferase reporter assay showed that miR-513b-5p notably suppressed the luciferase activity of AZIN1-AS1-WT reporter, but had no significant effect on AZIN1-AS1-MUT reporter (Figure 3E). Next, we explored the regulatory effect of AZIN1-AS1 on miR-513b-5p. As shown, the expression of miR-513b-5p in the AZIN1-AS1 overexpression group was significantly lower than that in the control group, while the expression of AZIN1-AS1 knock-down could increase the expression level of miR-513b-5p in NSCLC cells (Figure 3F). In conclusion, these data demonstrated that AZIN1-AS1 interacted with miR-513b-5p and inhibited the expression of miR-513b-5p.

Regulatory Effects of MiR-513b-5p on Proliferation and Metastasis of NSCLC Cells in vitro

In our previous study, miR-513b-5p was proved to suppress the malignant phenotypes of NSCLC A549 and

H460 cell lines.¹³ To further validate the tumor-suppressive function of miR-513b-5p in NSCLC, in this study, we used H1299 and HCC827 cells to study the function of miR-513b-5p in NSCLC (Figure 4A and B). CCK-8 and EdU experiments showed that compared with the control group, the proliferation of H1299 cells transfected with miR-513b-5p mimics was significantly reduced, and the proliferation of HCC827 cells transfected with miR-513b-5p inhibitors was remarkably increased (Figure 4C–E). After investigating the effect of miR-513b-5p on the proliferation of cancer cells, we explored the effects of miR-513b-5p on the metastasis. As shown, the migration and invasion of the cells were reduced after transfection with miR-513b-5p mimics, while the transfection of miR-513b-5p inhibitors had the opposite effect (Figure 4F and G). These data further clarified that miR-513b-5p was associated with the malignant phenotypes of NSCLC and functioned as a tumor suppressor.

Table 1 Correlation Between LncRNA AZIN1-AS1 Expression and Clinical Features (n=36)

Parameters	Group	n	LncRNA TMPO-ASI Expression		P value
			Low=18	High=18	
Gender	Male	20	8	12	0.180
	Female	16	10	6	
Age (years)	≤65	11	7	4	0.278
	>65	25	11	14	
Tumor size	≤3cm	16	11	5	0.044*
	>3cm	20	7	13	
Differentiation	Low or Moderate	19	11	8	0.317
	High	17	7	10	
Lymphatic metastasis	Positive	22	9	13	0.171
	Negative	14	9	5	
T stage	I	9	7	2	0.001*
	II	12	9	3	
	III	15	2	13	

Note: *Presents the statistic difference less than 0.05.

DUSP11 is the Functional Target of MiR-513b-5p

To explore the mechanism of the role of miR-513b-5p in NSCLC, we selected a candidate target for miR-513b-5p in TargetScan database (http://www.targetscan.org/vert_72/). As shown, miR-513b-5p was complementary to the 3'UTR of *DUSP11*, suggesting that *DUSP11* might be a target gene for miR-513b-5p (Figure 5A). We further examined the expression of *DUSP11* in NSCLC tissues and cell lines and demonstrated that the expression levels of *DUSP11* in NSCLC tissues and cells were significantly higher than those of control group (Figure 5B and C). Dual-luciferase reporter assay showed that miR-513b-5p mimics could decrease the luciferase activity of luciferase reporter containing *DUSP11* 3'UTR-WT, but had no significant influence on the luciferase activity of *DUSP11* 3'UTR-MUT reporter (Figure 5D). In addition, qRT-PCR displayed that the expression levels of miR-513b-5p and *DUSP11* in NSCLC tissues were negatively correlated, while the expression levels of AZIN1-AS1 and *DUSP11* in NSCLC tissues were positively correlated ($P < 0.05$, Figure 5E and F). Western blot showed that the expression levels of *DUSP11* were down-regulated and up-regulated, respectively, after transfection of mimics and inhibitors of miR-513b-5p in NSCLC cells. What's more, as we expected,

after overexpression and knockdown of AZIN1-AS1 in NSCLC cells, *DUSP11* levels were up-regulated and down-regulated, respectively (Figure 5G and H). This indicated that AZIN1-AS1 could indirectly modulate the expression of *DUSP11*.

The Tumor-Suppressive Function of MiR-513b-5p Was Partly Dependent on Its Regulatory Function on DUSP11

To investigate the functional relationship between miR-513b-5p and *DUSP11*, *DUSP11* overexpression plasmids were co-transfected with miR-513b-5p into H1299 cells. As shown, the co-transfection of *DUSP11* overexpression plasmids did not affect the expression of miR-513b-5p in H1299 cells but significantly up-regulated the expression of *DUSP11* (Figure 6A and B). Next, CCK-8 assay, EdU assay, wound healing assay and Transwell assay were performed, and as expected, these functional experiments indicated that miR-513b-5p suppressed the malignant phenotypes of H1299 cells, which could be partially reversed by the co-transfection of *DUSP11* overexpression plasmids (Figure 6C–F). Collectively, these data validated that the tumor-suppressive function of miR-513b-5p was partly dependent on its regulatory function on *DUSP11*.

AZIN1-AS1 Affected the Proliferation and Metastasis of NSCLC Cells Through MiR-513-5p/DUSP11 Axis

To further verify the role of AZIN1-AS1 in NSCLC by regulating the expression of miR-513b-5p/*DUSP11*, we transfected miR-513b-5p mimics into HCC827 cells over-expressing AZIN1-AS1. We found that compared with AZIN1-AS1/NC group, AZIN1-AS1/miR-513b-5p mimics group had no significant change in AZIN1-AS1 level, while the expression of *DUSP11* was significantly decreased (Figure 7A–C). The results of CCK-8 and EdU assays indicated that the proliferation of HCC827 cells was significantly increased by AZIN1-AS1 overexpression, while this was partly reversed by the co-transfection of miR-513b-5p mimics (Figure 7D and E). Wound healing and Transwell assays showed that the migration and invasion abilities of cancer cells were enhanced by AZIN1-AS1 overexpression, but reversed by the co-transfection of miR-513b-5p mimics (Figure 7F and G). Collectively, the above results further validated the role of AZIN1-AS1/miR-513b-5p/*DUSP11* axis in NSCLC progression (Figure 7H).

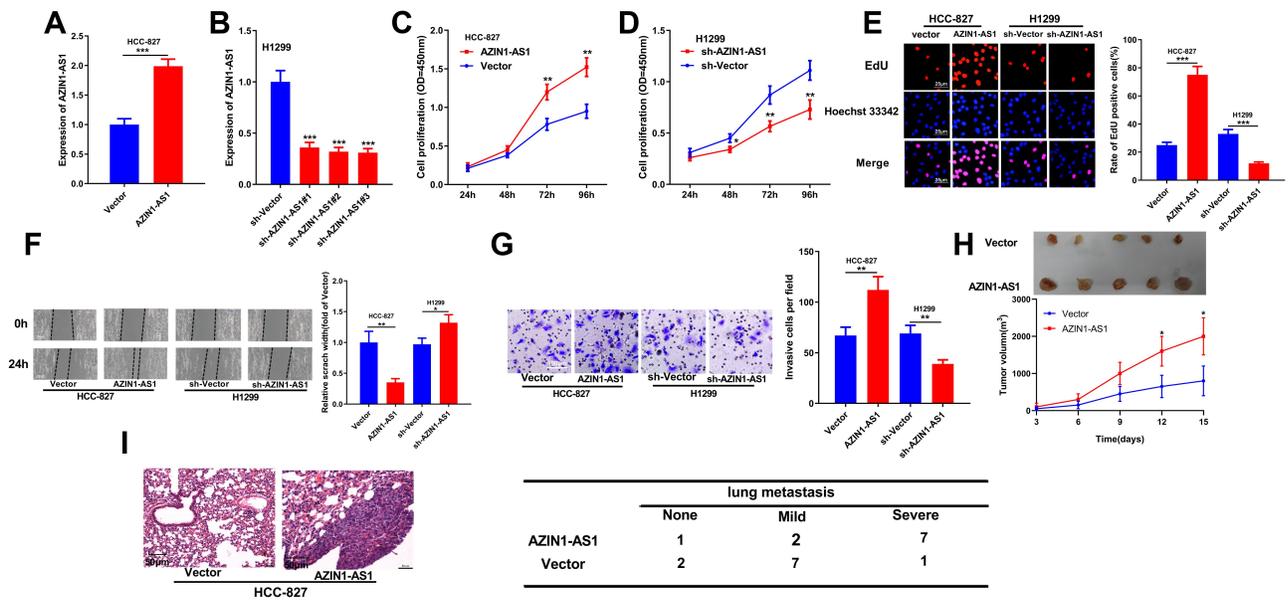


Figure 2 Effects of AZINI-AS1 on proliferation and metastasis of NSCLC cells in vitro and in vivo. (A) qRT-PCR was used to test the overexpression model of AZINI-AS1. (B) qRT-PCR was used to test the knockdown model of AZINI-AS1. (C and D) CCK-8 was used to determine the effects of overexpression and knockdown of AZINI-AS1 on cell proliferation. (E) EdU assay was used to determine the effects of overexpression and knockdown of AZINI-AS1 on cell proliferation. (F) Wound healing assay was used to determine the effects of overexpression and knockdown of AZINI-AS1 on cell migration. (G) Transwell assay was used to determine the effect of overexpression and knockdown of AZINI-AS1 on the invasive ability. (H) Subcutaneous tumorigenesis test with nude mice was used to determine the effect of knockdown of AZINI-AS1 on the growth of tumors in vivo. (I) The lung metastasis test was used to determine the metastatic ability of NSCLC cells in mice after knockdown of AZINI-AS1. *, **, ***Represent $P < 0.05$, $P < 0.01$ and $P < 0.001$ respectively.

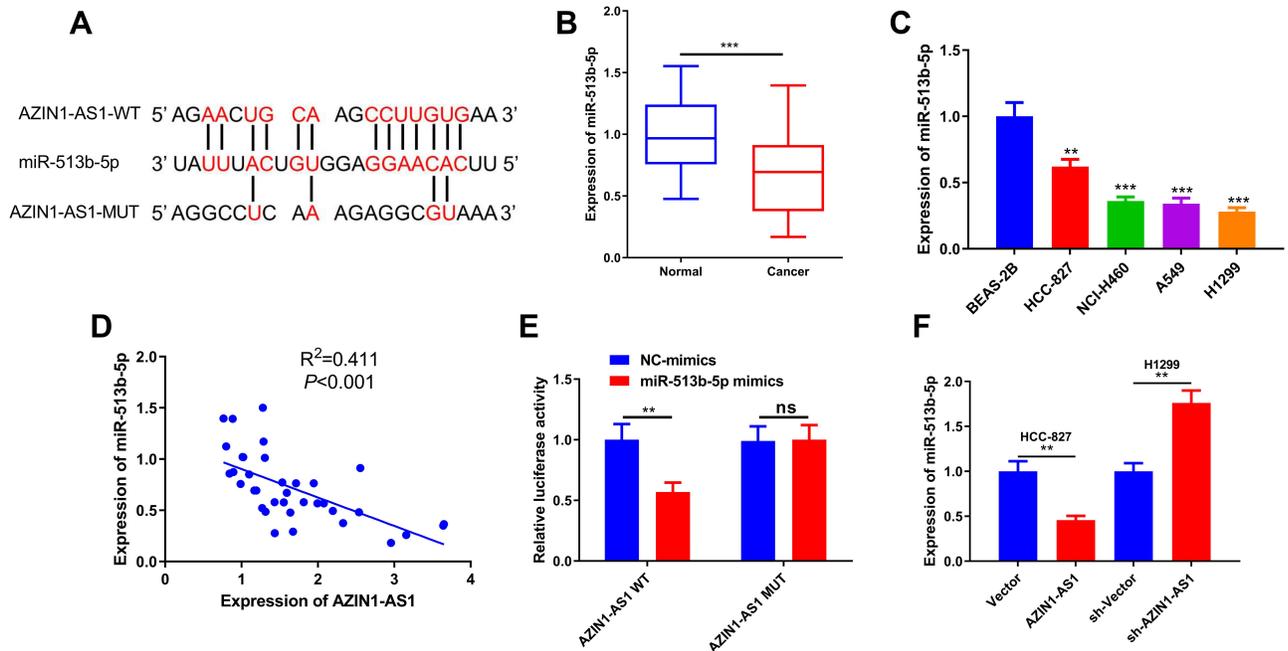


Figure 3 Interaction between AZINI-AS1 and miR-513b-5p in NSCLC cells. (A) AZINI-AS1 contained a potential binding site of miR-513b-5p. (B) qRT-PCR was used to detect the expression of miR-513b-5p in normal lung tissues and NSCLC tissues. (C) qRT-PCR was used to detect the expression of miR-513b-5p in normal lung epithelial cell lines and NSCLC cell lines. BEAS-2B was the control group. (D) Correlation analysis was used to analyze the relationship between AZINI-AS1 and the expression of miR-513b-5p in NSCLC samples. $R^2=0.411$, $P<0.001$. (E) Luciferase reporter assay was used to detect the binding between miR-513b-5p and AZINI-AS1-WT. (F) qRT-PCR was used to detect the expression of miR-513b-5p after AZINI-AS1 overexpression and AZINI-AS1 knockdown, respectively. *, ***Represent $P < 0.01$ and $P < 0.001$ respectively.

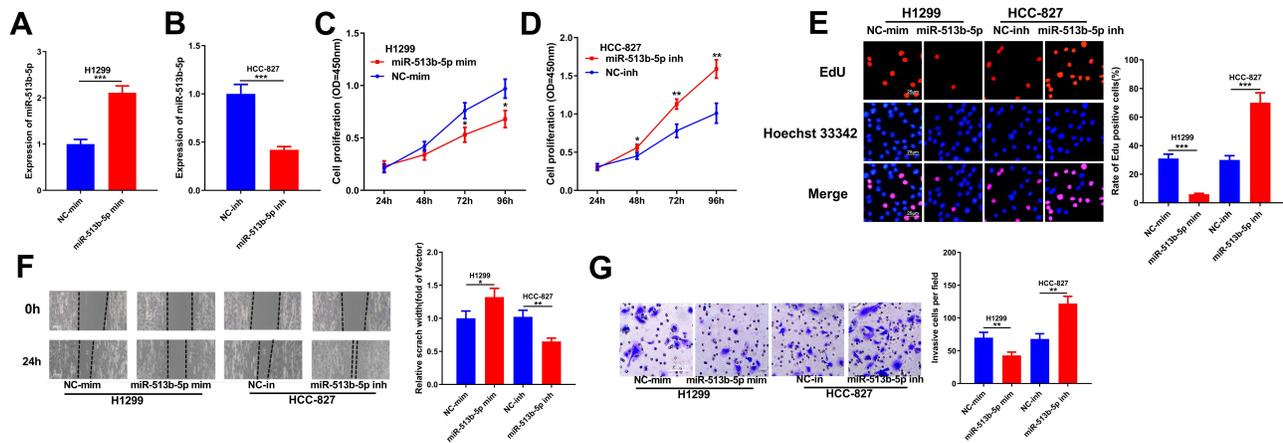


Figure 4 Effects of miR-513b-5p on proliferation and metastasis of NSCLC cells in vitro. (A) qRT-PCR was used to determine the transfection of miR-513b-5p mimics into H1299 cell line. (B) qRT-PCR was used to determine the transfection of miR-513b-5p inhibitors into HCC-827 cell line. (C and D) CCK-8 was used to determine the effects of miR-513b-5p mimics and inhibitors on the proliferation of NSCLC cells. (E) EdU assay was used to determine the effects of effects of miR-513b-5p mimics and inhibitors on the proliferation of NSCLC cells. (F) Wound healing assay was used to determine the migration of NSCLC cells transfected with miR-513b-5p mimics and inhibitors. (G) Transwell assay was used to determine the effects of miR-513b-5p mimics and inhibitors on the invasion of NSCLC cells. *, **, ***Represent $P < 0.05$, $P < 0.01$ and $P < 0.001$ respectively.

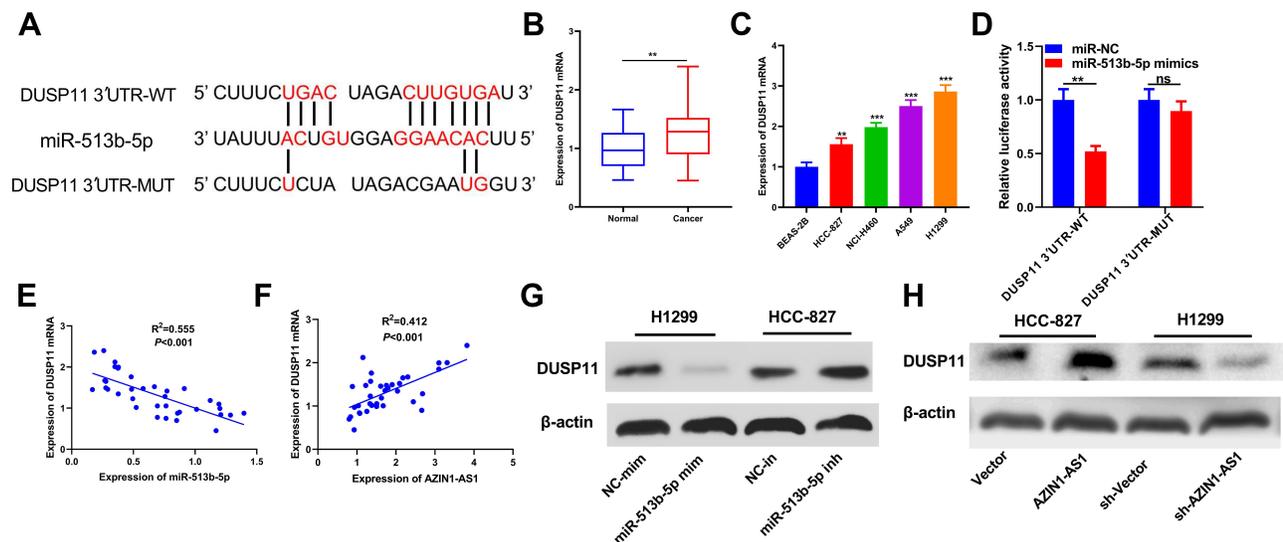


Figure 5 *DUSP11* was a target of miR-513b-5p in NSCLC cells. (A) miR-513b-5p contained a potential binding site with the 3'UTR of *DUSP11*. (B) qRT-PCR was used to detect the expression of *DUSP11* in normal lung epithelium and NSCLC tissues. (C) qRT-PCR was used to detect the expression of *DUSP11* in normal lung epithelial cell lines and NSCLC cell lines. BEAS-2B was the control group. (D) Dual-luciferase reporter assay was used to determine the binding of miR-513b-5p to *DUSP11*. (E) Correlation analysis was used to analyze the relationship between the expression of miR-513b-5p and *DUSP11*. $R^2=0.555$, $P<0.001$. (F) Correlation analysis was used to analyze the relationship between AZIN1-AS1 and *DUSP11* expression. $R^2=0.412$, $P<0.001$. (G) Western blot was used to detect the expression of *DUSP11* in NSCLC cells after overexpression and knockdown of AZIN1-AS1. (H) Western blot was used to detect the expression of *DUSP11* in NSCLC cells transfected with mimics or inhibitors of miR-513b-5p. **, ***Represent $P < 0.01$ and $P < 0.001$ respectively.

Discussion

Accumulating studies show that lncRNA can affect the progression of tumors.^{14–17} For example, in NSCLC, lncRNA XIST could enhance the chemoresistance of cancer cells by regulating autophagy;¹⁶ lncRNA PVT1 promoted the proliferation and inhibits apoptosis of NSCLC cells by targeting miR-195.¹⁷ In this study, we found that AZIN1-AS1 was up-regulated in NSCLC tissues and cells

compared with the control group. Additionally, its high expression in tumor tissues was associated with unfavorable pathological parameters. Furthermore, the biological function of AZIN1-AS1 in NSCLC was investigated. We observed that overexpression of AZIN1-AS1 could promote the proliferation and metastasis of NSCLC cells, whereas knockdown of AZIN1-AS1 could suppress these phenotypes. These results suggested that AZIN1-AS1 was

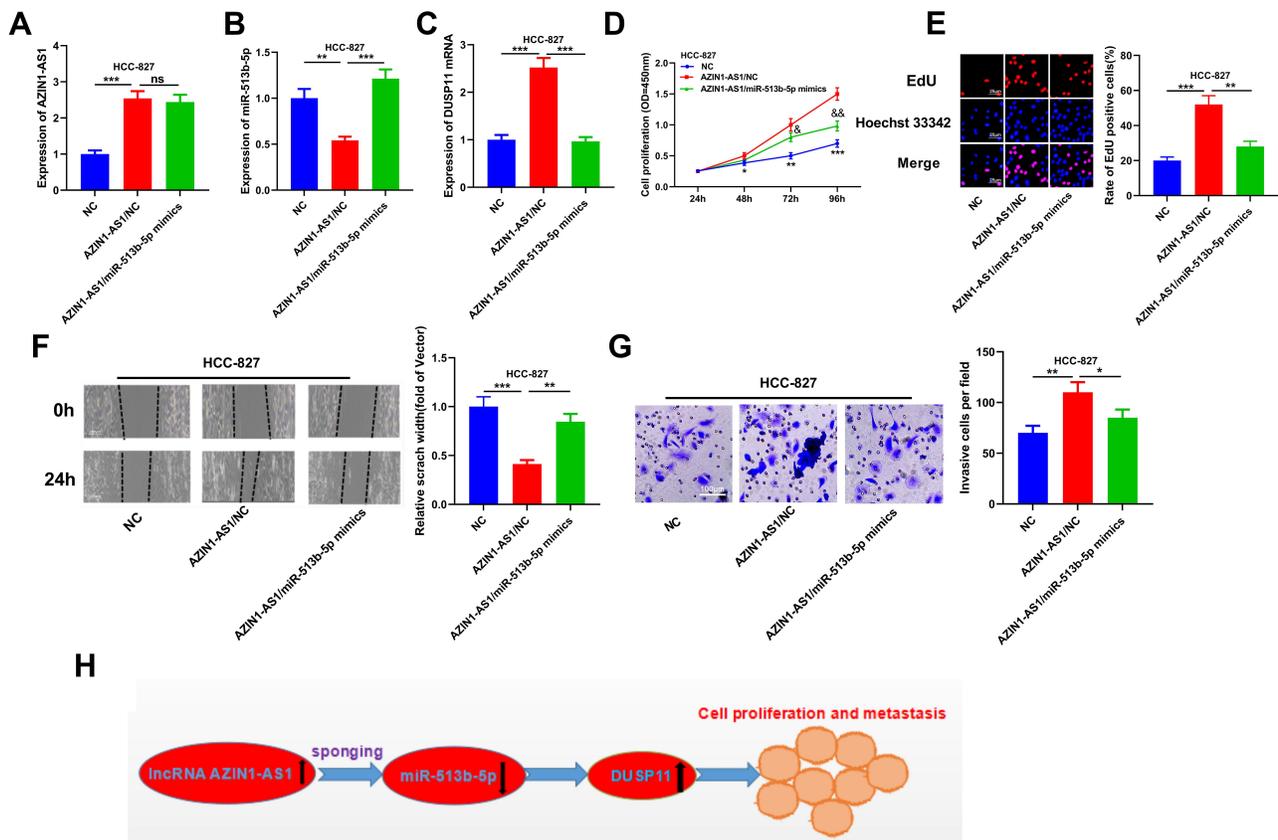


Figure 7 AZIN1-AS1/miR-513b-5p/*DUSP11* axis modulated the progression of NSCLC. **(A)** The level of AZIN1-AS1 in cells co-transfected with miR-513b-5p mimics and AZIN1-AS1 overexpression plasmids was detected by qRT-PCR. **(B)** The level of miR-513b-5p in cells co-transfected with miR-513b-5p mimics and AZIN1-AS1 overexpression plasmids was detected by qRT-PCR. **(C)** qRT-PCR was used to detect the level of *DUSP11* mRNA in cells co-transfected with miR-513b-5p mimics and AZIN1-AS1 overexpression plasmids. **(D)** CCK-8 method was used to detect the proliferation of HCC-827 cells. **(E)** The proliferation of HCC-827 cells was detected by EdU method. **(F)** Wound healing assay was used to detect the migration of HCC-827 cells. **(G)** The invasion of HCC-827 cells was detected by Transwell assay. **(H)** AZIN1-AS1 inhibited miR-513b-5p and up-regulated *DUSP11*, thus promoting proliferation and metastasis of NSCLC cells. *, **, ***Represent $P < 0.05$, $P < 0.01$ and $P < 0.001$ respectively. In figure D, & and &&Represent $P < 0.05$, $P < 0.01$ respectively, vs AZIN1-AS1/NC group.

an “AZIN1-AS1/miR-513b-5p/*DUSP11*” axis in NSCLC tumorigenesis.

In the following studies, other downstream miRNAs of AZIN1-AS1 needs to be identified. And it is necessary to explore whether the dysregulation of AZIN1-AS1 is associated with the overall survival time and disease-free survival time of NSCLC patients, which is quite important to evaluate its potential to be used as a biomarker. It is worth noting that *DUSP11* is reported to be a target of p53, and contributes to the proliferation inhibition of osteosarcoma cells line U2OS.²⁵ This study implies that the function of *DUSP11* is specific in different cancers, and its role in NSCLC requires further investigation.

In conclusion, our study demonstrates that AZIN1-AS1 is up-regulated in NSCLC tissues and cells, and it promotes the proliferation and metastasis of NSCLC cells by regulating the expression of miR-513b-5p/*DUSP11*. To our best knowledge, this is the first study to investigate the role of

AZIN1-AS1 in cancer biology and it is expected to provide new therapeutic targets for NSCLC patients.

Data Sharing Statement

All data generated or analyzed during this study are included in this published article.

Ethics Approval and Consent to Participate

The protocols of human samples collection were approved by the Ethics Committee of Shanghai Pulmonary Hospital. All participants provided written informed consent and the procedures were conducted in accordance with the principles of the Declaration of Helsinki. As for animal experiments, ethical and legal approval was obtained from Animal Study Committee of Shanghai Pulmonary Hospital prior to experiments. The protocols were in accordance with the National Institutes of Health Guide

for the Care and Use of Laboratory Animals and Animal Welfare Act.

Author Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

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Disclosure

The authors declare that they have no competing interests for this work.

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