Clinical utility of voriconazole eye drops in ophthalmic fungal keratitis

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Abstract: Fungal keratitis is one of the major causes of ophthalmic mycosis and is difficult to treat. The range of common antifungal agents available for fungal keratitis remains inadequate and is generally associated with poor clinical outcomes. Voriconazole is a new generation triazole antifungal agent. Only marketed in systemic formulation and, with broad-spectrum activity and high intraocular penetration, voriconazole has demonstrated effectiveness against fungal keratitis. Systemic voriconazole, however, is not without side effects and is costly. Voriconazole eye drops have been prepared extemporaneously and used for the treatment of ophthalmic fungal keratitis. The current article sought to review the literature for evidence related to the effectiveness and safety of topical voriconazole and its corneal penetration into the aqueous humor of the eye. The voriconazole eye drops used are typically of 1% concentration, well tolerated by the eye, and are stable. Despite existing evidence to suggest that the eye drops are effective in the treatment of fungal keratitis, more studies are needed, especially in relation to using the eye drops as first-line and stand-alone treatment, preparation of higher concentrations, and optimal dosing frequency.

Keywords: voriconazole, fungal keratitis, eye drops, corneal penetration

Introduction
Corneal disease is second only to cataracts as the most common cause of blindness worldwide, resulting in more than 1.5 million new cases of vision loss annually.1 As a consequence of attention being directed towards the management of cataracts, especially in developing countries, strategies for the management of traditional infections that cause blindness have been neglected.2

Ophthalmic mycosis is emerging as a major cause of vision loss and morbidity, and can be life-threatening.3,4 Fungal keratitis is one of the major causes of ophthalmic mycosis,4 accounting for more than 50% of proven ophthalmic mycoses in some countries.6 Fungal keratitis is usually characterized by a corneal epithelial defect and inflammation of the corneal stroma. If untreated, fungal keratitis can lead to corneal scarring and vision loss.1

Fungal keratitis
The first description of fungal keratitis was in the late 1870s.7 Fungal keratitis is most common in tropical regions and developing countries, where it constitutes over 50% of keratitis.8 In South India, about 44% of corneal ulcers are caused by fungi. Although lower, the prevalence of fungal keratitis is still relatively high in other countries, being 17% in Nepal, 36% in Bangladesh, 38% in Ghana, and 35% in south Florida in the US.
In China, the incidence has been increasing in the past decade.9 By contrast, fungal keratitis generally accounts for only 1%–5% of the keratitis treated in developed countries and temperate regions, such as Britain and the northern US.9,10 This also applies to Australia, where the incidence of fungal keratitis at the Royal Victorian Eye and Ear Hospital (RVEEH) in Melbourne was reported at 5%. The RVEEH is a tertiary referral eye hospital responsible for the care of most serious corneal infections in a population of about five million across Victoria, southern New South Wales, and Tasmania.10

Etiology

Filamentous fungi were long considered as a major cause of fungal keratitis.11,12 Ophthalmic infections from these fungi are most commonly associated with agricultural and outdoor activities.13,14 Of the filamentous fungi, infections from *Fusarium* and *Aspergillus* species are the most common. While *Fusarium* species are particularly prevalent in crop plants,15 *Aspergillus* species are found in decaying vegetation and soil. *Aspergillus* is a contaminant in hospital air and has been involved in recent outbreaks of ocular infections in several hospitals.16,17 Keratitis caused by these filamentous fungi may involve any part of the cornea.18,19 Other less common keratitis-causative filamentous fungi include *Paecilomyces* and *Acremonium* species.20 *Paecilomyces* species have been shown to be resistant to most common sterilising techniques, including those applied during surgical procedures.21,22 *Acremonium* species can be isolated from a variety of common sources, and can be associated with severe eye infections.23,24

Dematiaceous fungi such as *Curvularia*, *Bipolaris*, and *Exserohilum* species have also been reported to cause fungal keratitis. After *Aspergillus* and *Fusarium* species, *Curvularia* and *Bipolaris* species are the third most common keratitis-causative fungi worldwide.20 *Curvularia*, *Bipolaris*, and *Exserohilum* species usually cause persistent, but low-grade ulcerations near the epithelial part of the cornea. These ulcerations, if not appropriately treated or if associated with topical steroid use, can develop into resilient infections involving the deeper layers of cornea.25–27 *Scedosporium* and *Lecythophora* species are also dematiaceous fungi that are known to result in very severe keratitis infections that often do not respond to medical therapies.25,28

Whilst filamentous and dematiaceous fungi are the common causes of fungal keratitis at a global level, yeasts are the major cause of fungal keratitis in developed countries.8 Yeasts are infrequent in tropical countries, characterized by major agricultural presence, which is associated with higher prevalence of other types of fungal keratitis, such as the filamentous fungi.20 Yeast infections have no geographical dominance and are most commonly caused by *Candida* species, especially *Candida albicans*.8,10,20 *Candida* keratitis predominantly occurs in the stromal layer of the cornea. It is associated with epithelial defect and distinct infiltration, and is slow in development.18 *Cryptococcus* species are another type of yeast that causes fungal keratitis, but less commonly than *Candida* species.20

Fungal keratitis can also be caused by zygomycetes fungi such as *Rhizopus* and *Mucorales* species,1,29,30 and other fungi such as *Cladosporium*, *Cylindrocarpon*, *Penicillium*, and *Chrysonilia* species.6,14,31–33 Keratitis due to these fungi, however, is very low in occurrence.

The incidence of different types of fungal keratitis in various areas and countries is shown in Table 1.

Risk factors

The general predisposing factors for fungal keratitis include ocular trauma, prolonged use of topical or systemic immunosuppressants, pre-existing corneal surface disease, underlying systemic disease (eg, diabetes mellitus), and contact lens wear.10,20,35

The significance of these factors, however, varies according to geographical area. For instance, in Melbourne, ocular trauma, chronic steroid use, and ocular surface disease were the most common risk factors,10 whilst the common risk factors in Philadelphia were ocular surface disease, contact lens wear, and topical steroids.33 In the southern US, however, trauma was generally identified as the major risk factor for fungal keratitis. A similar trend was also observed in Singapore and Bangladesh.36,37 In contrast, in the northern US, ocular trauma was reported as only a secondary risk factor for fungal keratitis.9

The type of predisposing risk factors relates to the type of causative fungi. For example, keratitis associated with ocular trauma is commonly caused by *Aspergillus*, *Fusarium*, and *Curvularia* species.14 The use of lawn trimmers was found to be associated with *Fusarium* and *Curvularia* keratitis,14,38 while the use of topical steroids was linked to *Candida*, *Aspergillus*, *Acremonium*, and *Curvularia* keratitis. Underlying chronic diseases were frequently related to keratitis caused by *Fusarium* and *Candida* species.14 *Candida* keratitis is common where traumatic keratitis is infrequent.20 Previous corneal ulceration resulting, for example, from previous keratitis or contact lens-related trauma, is a particular risk factor for *Candida* keratitis.18 Trauma by plant material, contaminated water, or immune suppression is a risk...
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factor for keratitis caused by Scedosporium apiospermum. Keratitis caused by Paecilomyces species has been reported following surgical procedures.

Treatment

The ultimate goal in the treatment of fungal keratitis is to conserve vision. This requires timely diagnosis of the infection and administration of the appropriate antifungal therapy. Patient with fungal keratitis can be treated with either medical or surgical therapy. Whilst surgical procedures are more effective in patients with acute corneal perforation, antifungal agents are still the major therapeutic option in fungal keratitis, whereby success depends on the agent’s ability to penetrate into the aqueous humor and achieve therapeutic levels. Currently, the range of antifungal therapies available for fungal keratitis remains inadequate. The antifungal agents that can be used in fungal keratitis are broadly divided into three main groups: polyenes (amphotericin B, natamycin, and nystatin), azoles (ketoconazole, miconazole, econazole, fluconazole, itraconazole, voriconazole, and posaconazole), allylamines (terbinafine) and echinocandins (caspofungin).

Amphotericin B has poor ocular penetration after intravenous (IV) administration and, hence, the administration of higher doses may be required to ensure adequate concentration of amphotericin B in the eye; however, IV administration of high-dose amphotericin B is known to cause severe renal toxicity, which can occur in up to 80% of patients. To minimize renal toxicity, low-dose amphotericin B is often used, which in many cases, results in suboptimal doses, especially when taking its poor ocular penetration into consideration. Amphotericin B eye drops are manufactured extemporaneously by hospital pharmacy departments. The most commonly prescribed concentration of the eye drops for fungal keratitis is 0.15%. Topical amphotericin B penetrates well into the stroma and can achieve sufficient concentrations against susceptible fungi; however, its penetration through the cornea with intact epithelium is poor. Whilst amphotericin B is active against Aspergillus and Candida keratitis, it has no activity against keratitis caused by Fusarium species.

Natamycin is the only commercially available topical antifungal preparation approved by the Food and Drug Administration for ophthalmic use. It is insoluble in water.

Table 1 Studies of the incidence of types of fungal keratitis

<table>
<thead>
<tr>
<th>Place</th>
<th>Number of patients</th>
<th>Study duration</th>
<th>Principal pathogen (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Melbourne, Australia</td>
<td>56</td>
<td>18 months</td>
<td>Candida albicans (37), Aspergillus fumigatus (17), Fusarium spp. (14)</td>
</tr>
<tr>
<td>Madurai, India</td>
<td>434</td>
<td>3 months</td>
<td>Fusarium spp. (47), Aspergillus spp. (16)</td>
</tr>
<tr>
<td>London, UK</td>
<td>65</td>
<td>13 years</td>
<td>Candida albicans (35), Candida parapsilosis (15), Fusarium solani (11), Aspergillus fumigatus (9)</td>
</tr>
<tr>
<td>Hyderabad, India</td>
<td>1352</td>
<td>10 years</td>
<td>Fusarium spp. (37), Aspergillus spp. (31)</td>
</tr>
<tr>
<td>Paraguay</td>
<td>45</td>
<td>1 year</td>
<td>Fusarium spp. (42), Aspergillus spp. (21)</td>
</tr>
<tr>
<td>Sri Lanka</td>
<td>66</td>
<td>2 years</td>
<td>Aspergillus spp. (25)</td>
</tr>
<tr>
<td>Florida, US</td>
<td>125</td>
<td>10 years</td>
<td>Fusarium spp. (68), Candida spp. (14), Curvularia spp. (9)</td>
</tr>
<tr>
<td>Bangladesh</td>
<td>142</td>
<td>11 months</td>
<td>Aspergillus spp. (37), Fusarium spp. (20), Curvularia spp. (18)</td>
</tr>
<tr>
<td>Ghana</td>
<td>199</td>
<td>N/A</td>
<td>Fusarium spp. (52), Aspergillus spp. (15), Lecytophora theobromae (9)</td>
</tr>
<tr>
<td>New Delhi, India</td>
<td>211</td>
<td>5 years</td>
<td>Aspergillus spp. (40), Fusarium spp. (14), Alternaria spp. (10)</td>
</tr>
<tr>
<td>Singapore</td>
<td>29</td>
<td>5 years</td>
<td>Fusarium spp. (52), Aspergillus flavus (17)</td>
</tr>
<tr>
<td>Houston, US</td>
<td>32</td>
<td>30 years</td>
<td>Curvularia senegalensis (30), Curvularia lunata (25)</td>
</tr>
<tr>
<td>Qingdao, China</td>
<td>108</td>
<td>4 years</td>
<td>Fusarium spp. (65), Aspergillus spp. (14), Candida spp. (9)</td>
</tr>
<tr>
<td>Nepal</td>
<td>405</td>
<td>2 years</td>
<td>Aspergillus spp. (47), Candida spp. (13), Fusarium spp. (12)</td>
</tr>
</tbody>
</table>

Abbreviations: spp., species; N/A, not available.
and is stable in 5% suspension.70 Natamycin is the standard of care in many countries, especially developed countries,9,10,20 and is initially administered as one drop every one or two hours.47 It adheres well to the cornea surface, is well tolerated, has good activity against Candida, Aspergillus, and Fusarium, and is routinely used for keratitis caused by filamentous fungi.9,20 This antifungal, however, has poor penetration into deeper structures of the eye and, hence, is generally effective against superficial infections that are not severe.9,48 In addition, only about 2% of the drug is bioavailable after topical application.49 The usefulness of topical natamycin is further complicated by the fact that it settles out on the cornea upon instillation and degrades easily.28

Nystatin is another polyene that can be used topically as a suspension in fungal keratitis. However, nystatin is rarely used clinically due to the availability of more potent polyene agents.90

Ketoconazole has a broad spectrum of activity, including against Aspergillus, Candida, and Fusarium species.28 It is available orally and, although it has demonstrated good tissue distribution after administration,46 it has not been used for fungal keratitis. Long-term administration of high-dose ketoconazole may result in impotence, gynecomastia, or alopecia, which is problematic considering the long-term nature of keratitis therapy.31 Topical 1% eye drops and suspension formulations of ketoconazole have been extemporaneously prepared and used for fungal keratitis. These have been reported to inhibit the progression of corneal fungal infections and were not associated with significant corneal toxicity.52,53

Miconazole has been used in patients with S. apiospermum orbital infections.20 It has a broad spectrum of activity, including against Aspergillus, Candida, and Scedosporium species.20 Systemic miconazole, however, is associated with significant toxicity and has resulted in undetectable concentrations in the cornea.20,54 Topical application of extemporaneously prepared 1% miconazole eye drops achieved high concentrations in the ocular tissues.54 The eye drops are generally well tolerated and are used as second-line therapy in fungal keratitis that is unresponsive to natamycin.54

Econazole has a broad spectrum of activity against filamentous fungi, and is effective against Fusarium keratitis.55 Topical application of 2% econazole appears to be as effective as 5% natamycin in fungal keratitis,56 but has been associated with ocular irritation.55

Oral and IV fluconazole are very safe, and penetrate very well into the corneal tissue.20,57,58 Whilst oral fluconazole is a commonly used agent for the treatment of fungal keratitis,10 topical application of 0.2% fluconazole solution is as effective as systemic fluconazole. Fluconazole, when applied topically, penetrates well into the cornea, is safe, and has been used successfully against fungal keratitis.59,61 A major limitation associated with fluconazole, however, is its narrow spectrum of antifungal activity. Fluconazole is inactive against Aspergillus and Fusarium species;52 although active against Candida species, it is less active against Candida glabrata and Candida krusei than against C. albicans.46

Although itraconazole is commonly associated with gastrointestinal side effects, it is considered relatively safe.53 It has activity against Candida and Aspergillus species; however, it is rarely used for the treatment of fungal keratitis.28 Itraconazole is inactive against Fusarium species10,28 but, more importantly, it has poor penetration into the cornea after systemic administration.64 Experimental use of topical itraconazole (1% solution) has been reported, but appears to demonstrate insufficient corneal penetration.65

Voriconazole, a more recent azole antifungal, is available commercially for systemic administration in the form of oral and IV formulations. It has an excellent broad spectrum of antifungal activity and is active against species that are known to be resistant to the other antifungal agents commonly used in fungal keratitis.28 Voriconazole is increasingly being used, orally in particular, against fungal keratitis. Oral voriconazole is highly bioavailable (96%) and has demonstrated good penetration into the different parts of the eye,66,67 with sufficient concentrations achieved to cover a wide range of keratitis-causative fungi.28 However, oral voriconazole can be associated with side effects as well as significant drug interactions.68 Topical voriconazole eye drops, manufactured extemporaneously and used in an off-label manner, have also been prescribed for the treatment of keratitis, with promising results.69 With topical administration, voriconazole demonstrated good penetration through the cornea into the aqueous humour,69 without compromising intraocular safety.70

Posaconazole has an excellent broad spectrum of activity and is as active as voriconazole, with added activity against zygomycetes.71 It is safe, with mild gastrointestinal side effects being the most common adverse events.72 Posaconazole was only recently introduced worldwide and, as such, studies on its ocular penetration are lacking. In a number of recent case reports involving the use of oral posaconazole alone as salvage therapy, or in combination with topical posaconazole, this antifungal agent demonstrated
success against fungal keratitis. The formulation used for the topical posaconazole was the same formulation used for the oral suspension (10 mg/0.1 mL).

Terbinafine is fungicidal against many molds, but only a few types of yeast. Despite its activity, its clinical efficacy and use are limited by its pharmacokinetic characteristics after the systemic administration. When used as 0.25% eye drops, however, it is as effective against filamentous mycotic keratitis as 5% natamycin, especially in cases with smaller and shallower ulcers. The eye drops are safe, but required longer durations of treatment when compared with other common topical therapies.

Caspofungin has significantly less systemic toxicity than azoles. Intravenously administered caspofungin does not penetrate well into the eye and, hence, it is not used for fungal keratitis. Nonetheless, in one recent case report, when administered topically (0.5%) as adjunctive therapy, caspofungin demonstrated clinical success against fungal keratitis. Caspofungin is safe, but lacks activity against Fusarium species.

Of the aforementioned antifungal agents, amphotericin B, natamycin, fluconazole, and miconazole have been used routinely to treat fungal keratitis for quite some time; however, poor corneal penetration after topical administration, poor ocular penetration after systemic administration, limited spectra of antifungal activity, and/or limited clinical success associated with these agents are major limitations and have rendered these therapies challenging and inadequate for fungal keratitis. The limited clinical success is particularly true with the topical use of these agents, as they often require co-administration of an additional systemic antifungal agent, which increases the risk of toxicity and is costly. This has led to consideration of using newer antifungal agents, such as voriconazole, posaconazole, and caspofungin, and/or in-house preparations of these agents as a means to overcome the shortcomings of the current therapies.

This review will focus on the use of voriconazole eye drops as a treatment for fungal keratitis.

**Topical voriconazole for fungal keratitis**

Voriconazole acts by inhibiting the synthesis of ergosterol in the fungal membranes and, ultimately, the growth of the microorganism (Figure 1). Voriconazole binds to the active site of the P450-dependent enzyme lanosterol 14-demethylase (CYP51 or Erg11p) and ligates the iron heme cofactor via a nitrogen atom. This results in depletion of ergosterol and the accumulation of 14-methyl sterols such as lanosterol, affecting the integrity and function of the fungal membrane. Voriconazole is a derivative of fluconazole with the addition of a methyl group to the propyl backbone and the replacement of a triazole moiety with a fluoropyrimidine group, which significantly increased the affinity of the compound for 14-demethylase and its potency to inhibit CYP51. Voriconazole concentrations needed for a 50% decrease in ergosterol synthesis (IC50) in fungi extracts of C. albicans and C. krusei are 2 and 20 µg/L, respectively, compared with 10 and 230 µg/L, respectively, with fluconazole. With these two fungi, voriconazole is considered to be a more potent inhibitor of CYP51 than fluconazole. Similarly, the IC50 for Aspergillus fumigatus is 0.48 with voriconazole against 4.8 for fluconazole.

Voriconazole is ideal for use in the treatment of fungal keratitis, as it has a broad spectrum of activity with low minimum inhibitory concentrations (MIC), as well as a high systemic intraocular penetration profile.

Voriconazole is potent against a wide spectrum of keratitis-causative fungi, namely, the most common pathogens C. albicans, C. parapsilosis, C. tropicalis, A. fumigatus, Aspergillus flavus, and Fusarium solani, and other less common pathogens from the Paecilomyces, Histoplasma, Scedosporium, Curvularia, and Acremonium species. The in vitro MICs of voriconazole against typical keratitis-causative fungi are shown in Table 2.

Although the MIC of voriconazole against Fusarium species is higher than that for other fungi, compared with other antifungal agents, voriconazole has the best activity against Fusarium species. In a study by Marangon et al., in which the in vitro susceptibility of common pathogens to voriconazole was compared with that for amphotericin B, fluconazole, itraconazole, and ketoconazole, voriconazole demonstrated the lowest MIC, as shown in Table 3. In addition, voriconazole was the only antifungal agent that demonstrated 100% antifungal activity against 541 different forms of fungal keratitis.
fungal isolates comprising Candida, Aspergillus, and Fusarium species (Figure 2).

Intraocular penetration of systemic voriconazole

In a prospective clinical study by Hariprasad et al,67 systemically administered voriconazole was demonstrated to achieve good penetration into the aqueous and vitreous humors of the human eye. Two 12-hourly 400 mg doses of voriconazole were administered to 14 patients with noninflamed eyes and attending elective surgery. The aqueous and vitreous humor samples were collected within three hours after drug administration. The mean measured plasma, aqueous, and vitreous voriconazole concentrations were 2.13, 1.13, and 0.81 µg/mL, respectively. The voriconazole concentration in the aqueous humor was 53% of the concentration obtained in the plasma, and was sufficiently high to be effective against most common fungi associated with fungal keratitis.

A similar outcome was reported by Nulens et al,87 where a case of S. apiospermum keratitis was successfully treated with oral voriconazole. The voriconazole concentration in the aqueous humor (1.8 µg/mL) was measured after 12 days of drug administration and was also 53% of the voriconazole concentration observed in the patient’s plasma (3.4 µg/mL). Surprisingly, however, and despite good intraocular penetration of oral voriconazole, the reported success of oral voriconazole against fungal keratitis may not necessarily translate into success against fungal infections occurring in the posterior part of the eye (eg, endophthalmitis). Indeed, while in the case reported by Nulens et al,87 oral voriconazole resulted in a voriconazole concentration in the aqueous humor that is sufficient to treat S. apiospermum keratitis, in a case report by Nochez et al,88 the voriconazole concentration in the vitreous humor, resulting from oral voriconazole, was not sufficiently high to manage successfully an endophthalmitis infection that is also caused by S. apiospermum. Concomitant administration of intravitreal voriconazole was required to achieve a successful outcome.88

Although it has good intraocular penetration, systemic voriconazole may result in side effects (including ocular events), complications, and interactions with concomitant medications.68 Whilst mostly reversible, these side effects may lead to the discontinuation of therapy.81 In addition, systemic voriconazole is very costly. The cheapest of its formulations (ie, oral voriconazole tablets) costs about AUS$3,000 (US$2,600) per month of therapy for fungal keratitis.61 When administered intravenously, it can cost up to AUS$11,400 (US$9,600) per month.89 Therefore, an efficient and economical strategy of using voriconazole for the treatment of fungal keratitis is highly desirable and would be invaluable in clinical practice.

![Table 2: In vitro minimum inhibitory concentrations (MIC90) with voriconazole](image)

<table>
<thead>
<tr>
<th>Organism</th>
<th>MIC90 (µg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Candida albicans</td>
<td>0.06</td>
</tr>
<tr>
<td>Candida parapsilosis</td>
<td>0.12–0.25</td>
</tr>
<tr>
<td>Candida tropicalis</td>
<td>0.25–&gt;16.0</td>
</tr>
<tr>
<td>Cryptococcus neoformans</td>
<td>0.06–0.25</td>
</tr>
<tr>
<td>Aspergillus fumigatus</td>
<td>0.50</td>
</tr>
<tr>
<td>Aspergillus flavus</td>
<td>0.50</td>
</tr>
<tr>
<td>Fusarium spp.</td>
<td>0.25–8</td>
</tr>
<tr>
<td>Fusarium solani</td>
<td>2</td>
</tr>
<tr>
<td>Paecilomyces lilacinus</td>
<td>0.50</td>
</tr>
<tr>
<td>Acremonium alabamensis</td>
<td>0.25</td>
</tr>
<tr>
<td>Blastomyces dermatitidis</td>
<td>0.25</td>
</tr>
<tr>
<td>Coccidioides immitis</td>
<td>0.25</td>
</tr>
<tr>
<td>Histoplasma capsulatum</td>
<td>0.25</td>
</tr>
<tr>
<td>Penicillium marneffei</td>
<td>0.03</td>
</tr>
<tr>
<td>Cuvularia spp.</td>
<td>0.06–0.25</td>
</tr>
<tr>
<td>Scedosporium spp.</td>
<td>0.5</td>
</tr>
<tr>
<td>Scedosporium apiospermum</td>
<td>0.5</td>
</tr>
</tbody>
</table>

**Abbreviation:** spp, species.

![Table 3: In vitro minimum inhibitory concentrations (MIC90) of common antifungals](image)

<table>
<thead>
<tr>
<th>Antifungal agent</th>
<th>Aspergillus spp. (µg/mL)</th>
<th>Candida spp. (µg/mL)</th>
<th>Fusarium spp. (µg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Voriconazole</td>
<td>0.5</td>
<td>0.016</td>
<td>2</td>
</tr>
<tr>
<td>Amphotericin B</td>
<td>2</td>
<td>0.5</td>
<td>&gt;16</td>
</tr>
<tr>
<td>Itraconazole</td>
<td>1</td>
<td>0.256</td>
<td>&gt;16</td>
</tr>
<tr>
<td>Fluconazole</td>
<td>&gt;256</td>
<td>0.5</td>
<td>&gt;256</td>
</tr>
<tr>
<td>Ketoconazole</td>
<td>4</td>
<td>0.032</td>
<td>&gt;16</td>
</tr>
</tbody>
</table>

**Abbreviation:** spp., species.
Eye drops and ophthalmic drug delivery

The topical administration of medications to the eye is a typical strategy for treating disorders of anterior eye structures, such as the cornea. Eye drops are the most common dosage form used, because they are an economical and efficient method of delivering drugs into the eye, and have four main advantages. Firstly, the drug effect is localized where it is needed and a minimal amount of the drug reaches the systemic circulation. Secondly, drug concentrations that are hard to achieve at the site via systemic administration, can be achieved via topical administration. Thirdly, topically administered drugs avoid hepatic metabolism. Lastly, topical administration is convenient, simple, and painless.

The corneal tissues comprise three major layers of cells, i.e., a lipophilic outermost layer called the epithelium, a hydrophilic middle layer called the stroma, and an innermost layer of single cells called the endothelium.

Given that the cornea comprises both lipophilic and hydrophilic substances, it represents an effective barrier against delivering both lipophilic and hydrophilic drugs into the eye. A lipophilic compound encounters minimal resistance in penetrating the outer epithelium of the cornea, but more resistance in infiltrating the stroma. The converse applies to hydrophilic compounds, which encounter more resistance to absorption from the epithelium and less by the stroma. As the corneal epithelium is the main and first barrier to drug absorption into the eye, it is not surprising that lipophilic compounds are more favorable for corneal absorption.

Formulation of voriconazole eye drops

Whilst lipophilic compounds (or drugs) have higher corneal permeability, they usually have limited aqueous solubility. As such, formulating eye drops for drugs with low aqueous solubility can be challenging. Voriconazole is a lipophilic compound with low solubility (0.061% at pH 7), and is unstable in aqueous environments. For the IV formulation of voriconazole to be feasible, the manufacturer encapsulated voriconazole with a β-cyclodextrin derivative in the form of lyophilized powder of cyclodextrin-voriconazole complex. This increases the solubility and stability of voriconazole in aqueous solutions, while maintaining its lipophilicity and high corneal permeability.

Cyclodextrins are a group of homologous cyclic oligosaccharides that, in complex formation with a drug, increase dissolution rate (solubility), aqueous stability, and/or bioavailability of the drug. Currently, voriconazole eye drops are not commercially available, and are aseptically manufactured by diluting the IV formulation of voriconazole. IV voriconazole (Vfend®, Pfizer) is available as a glass vial that contains a white lyophilized powder containing 200 mg voriconazole and 3,200 mg sulfobutyl ether β-cyclodextrin sodium. As per the voriconazole package insert, the powder is reconstituted with 19 mL of water for injection to produce a 20 mL aqueous voriconazole solution with a concentration of 10 mg/mL (1%). This voriconazole solution is what is typically being used as eye drops.

Pharmacokinetic data on the corneal penetration of topically administered voriconazole are lacking; however, a number of studies have suggested good penetration of voriconazole through the cornea into the aqueous humor.

Animal studies

Several studies have been performed to assess the penetration and tolerability of voriconazole eye drops in animals. In a study by Sponsel et al, topical voriconazole (5 or 10 μg/mL) was evaluated for efficacy against Paecilomyces lilacinus keratitis in 10 rabbits (10 infected eyes). Voriconazole demonstrated good and deep penetration into the rabbits’ eyes. The measured tissue concentrations in the cornea were sufficiently high (24.3 and 51 ng/mL with 5 and 10 μg/mL voriconazole, respectively), and the experimental keratitis was treated successfully.

Topical application of voriconazole eye drops was also investigated in a horse model by Clode et al, where voriconazole eye drops (0.5%, 1.0%, and 3% solutions) were administered to seven healthy horses (four eyes for each concentration). With the measured aqueous humor concentrations being 1.43, 2.35, and 2.4 μg/mL, respectively, topical voriconazole was shown to penetrate effectively through the cornea and achieve detectable levels.

It is important to recognize, however, that extrapolating penetration data from animal models to humans may not be reliable. Rabbits, for instance, have a very low blink rate and a large epithelial eye surface, which enhances the penetration of lipophilic and nonirritating drugs, such as voriconazole, into the cornea. In addition, while the drainage rate of eye drops from the ocular surface in rabbits is about 4 μL per minute, it is over twice as much in humans.

Studies in nonkeratitis patients

Three studies have investigated voriconazole penetration through the human cornea into the aqueous humor. Two of the studies investigated 1% voriconazole eye drops, and one study investigated 2% voriconazole eye drops.
In a prospective study by Vemulakonda et al, 86 13 patients scheduled for vitrectomy surgery were recruited to receive a two-hourly 1% voriconazole eye drop for 24 hours. Samples were taken 24 ± 14 minutes after the last dose. Topical voriconazole was well tolerated by the eye, and the mean measured voriconazole concentration in the aqueous humor was 6.49 ± 3.04 µg/mL, which is sufficiently high to be effective against common pathogens. The concentration, however, was a peak level concentration, because it was taken 24 minutes after a two-hourly dosing regimen (peak concentration is reached 20 to 30 minutes after eye drop administration). 91 While this study did not demonstrate that the two-hourly dosing regimen results in sustained and adequate therapeutic trough voriconazole concentrations in the eye, the ability of topically administered voriconazole eye drops to achieve high aqueous humor concentrations was demonstrated.

In another prospective study by Lau et al, 61 10 patients scheduled for anterior segment surgery were recruited to receive either a 1% voriconazole eye drop every six hours for three days, or every hour for four doses. The eye drops were well tolerated, but the aqueous humor concentrations achieved were not sufficiently high to be effective against all common pathogenic fungi. After the six-hourly and hourly dosing, the voriconazole concentrations in the aqueous humor were 0.94 ± 1.21 and 1.9 ± 1.12 µg/mL, with average sampling times of 2.1 ± 0.6 and 1.1 ± 0.5 hours after the last dose, respectively. If samples from the six-hourly regimen were to be taken six hours after the last dose (ie, at trough level), the concentrations measured would be even less than 0.94 µg/mL, suggesting that six-hourly dosing of 1% voriconazole eye drops may be ineffective. Samples taken after hourly dosing were collected approximately one hour after the last dose, representing trough level concentrations. Although the measured 1.9 µg/mL concentration is effective against Aspergillus and Candida keratitis, it is ineffective against other common types, such as Fusarium keratitis.

Al-Badriyeh et al investigated the penetration of 2% voriconazole eye drops, 101 with the hypothesis that increasing the concentration of voriconazole in the eye drops will increase the amount of voriconazole penetrating into the eye. Hourly 2% voriconazole eye drops were given to 13 human subjects four hours prior to elective anterior segment eye surgery. The mean voriconazole concentration in the aqueous humor was 1.67 ± 0.97 µg/mL, while the mean sampling time after the last eye drop administration was at 1.3 ± 0.3 hours. No side effects or toxicities were reported. The design of this study was similar to that of Lau et al, 61 in that both studies had identical numbers and frequencies of doses administered; trough voriconazole levels were measured; the same volumes (0.05 mL) of eye drops were administered at each dose, 102 and 0.01% benzalkonium chloride solution (a preservative) was used as a diluent for the preparation of the eye drops. It should be noted that the clinical studies by Lau et al, 61 and Al-Badriyeh et al, 101 are the only studies in the literature that involved the use of benzalkonium chloride solution. Benzalkonium chloride is a quaternary ammonium compound with a broad range of antimicrobial activity. 103 It is the most frequently used preservative in ophthalmic solutions, and its concentration ranges from 0.004 to 0.02%. 103 In addition to preventing microbial contamination, benzalkonium chloride is known to act as a corneal penetrating enhancer, promoting drug penetration through the strong corneal barrier. 104 The study by Al-Badriyeh et al found that the concentration of voriconazole in the aqueous humor resulting from the 2% voriconazole eye drops was not significantly different from that reported for the 1% solution, 61,101 suggesting that the penetration of voriconazole through an intact infection-free cornea is not concentration-dependent, at least for the concentration range studied. This appears to be counterintuitive to the hypothesis of the study but is consistent with data from the horse model by Clode et al, 99 in which the voriconazole level in the corneas of horses with fungal keratitis did not change when the concentration of the administered voriconazole eye drops was changed from 1% to 3%.

The studies by Vemulakonda et al, Lau et al, and Al-Badriyeh et al explored the penetration of voriconazole eye drops into the human aqueous humor using different dosage regimens and concentrations. 61,96, 101 However, it is important to recognize the major limitation of these studies, ie, that the eye drops were applied to noninfected eyes. It has been widely observed that corneal drug penetration will generally be enhanced with the destruction of the corneal epithelium. 105 For instance, the removal of the surface of the corneal epithelium is recommended to improve the penetration of topical amphotericin B. 100 On the other hand, in the rabbit model by Sponsel et al, 98 when the penetration of topical voriconazole into the infected eyes was compared with penetration into the noninfected eyes of the rabbits, it was found that the corneal concentration of voriconazole in the noninfected eyes (after topical administration) was higher than that in the infected corneas. However, in a recent case series by Thiel et al (where voriconazole concentrations in the aqueous humor, following the administration of voriconazole eye drops, were compared among patients with different degrees of corneal injuries) voriconazole concentrations in the infected eyes depended
neither on the size of the epithelial defect nor on epithelial removal. These, however, are preliminary findings, and the effect of corneal damage on the penetration of voriconazole into the human eye remains to be fully elucidated.

Studies in keratitis patients
To date, the penetration of topical voriconazole eye drops through the infected cornea in humans has only been reported twice, and in the form of case reports. In the case reported by Klont et al, the aqueous humor voriconazole concentration was measured after 13 days of topical 1% voriconazole, co-administered with oral voriconazole, for the treatment of a patient with *Fusarium* keratitis. The advantage of topical voriconazole was demonstrated by an aqueous humor concentration of 3.2 µg/mL, which was 160% of the voriconazole concentration in plasma (2 µg/mL). In the previously mentioned case series by Thiel et al, six patients, including five patients with fungal keratitis, received IV and topical voriconazole for the treatment of *Aspergillus* and *Candida* infections. The aqueous humor samples were collected at stages of therapy where voriconazole eye drops were used alone, yielding voriconazole concentrations ranging from 0.61 to 3.3 µg/mL. The results were highly variable, but provided support for the benefit of using voriconazole eye drops.

Efficacy
Although voriconazole concentrations were detected in the aqueous humor after topical administration of voriconazole eye drops, this may not necessarily correlate with efficacy in the clinical setting of fungal keratitis. Well-designed clinical studies of voriconazole eye drops in patients with active fungal keratitis are difficult to perform and, therefore, lacking. The difficulties in conducting such studies relate to the low incidence of fungal keratitis as well as the need for long treatment duration. In addition, in clinical settings, patients will mostly be receiving other antifungal therapies that will interfere with the outcomes measured.

Currently, evidence of the clinical efficacy of voriconazole eye drops in fungal keratitis is based solely on case reports. A review of the published literature identified nine reports on the use of voriconazole eye drops for the treatment of fungal keratitis. The case reports are summarized in Table 4.

In most of the reported cases, voriconazole eye drops were used in combination with systemic voriconazole, except for the case reports by Al-Badriyeh et al, where voriconazole eye drops were used as monotherapy. Voriconazole 1% eye drops were used in all cases, except in the case reported by Polizzi et al, where 2% voriconazole was used. Brief summaries of these cases are given below.

The first of these cases, reported by Reis et al, involved a 16-year-old girl diagnosed with keratitis caused by *F. solani* after swimming in a lake. After months of antifungal therapy, the fungal keratitis failed to respond. The patient was initially prescribed topical amphotericin B and fluconazole, followed by itraconazole at a later stage. These, however, had no effect on the infection and, hence, IV voriconazole followed by oral voriconazole was administered. A significant improvement was noticed, followed by resolution upon the addition of topical voriconazole to therapy, which was discontinued after eight weeks.

The case report by Klont et al also reported the use of 1% voriconazole eye drops in the treatment of *Fusarium* keratitis. A 23-year-old man with *F. solani* keratitis failed to respond to treatment despite initial topical amphotericin B and itraconazole. The patient was then prescribed, as salvage therapy, concomitant IV and topical voriconazole followed by oral and topical voriconazole. The treatment was ceased at week 6, with a successful outcome.

In the case report by Prats et al, a 19-year-old man was admitted with an incisive eye wound, with the cornea totally sectioned upon trauma. *S. apiospermum* keratitis was diagnosed. Upon failure of initial empirical antifungal therapy, systemic (IV and oral) and topical voriconazole were commenced. The infection resolved, and the eye did not have to be enucleated. This was the first case report where voriconazole was used for the treatment of *S. apiospermum* keratitis. Five months after the incident, a penetrating keratoplasty and chamber intraocular lens implantation was performed with a favorable visual outcome.

Jones et al demonstrated that voriconazole was effective in a 52-year-old woman diagnosed with *Aspergillus niger* keratitis. The patient was initially treated with topical amphotericin B, which was not effective. When the patient was switched to a combination of oral and topical voriconazole, the infection improved rapidly and resolved after five weeks.

In the first of the two cases reported by Tu et al, a 29-year-old man received oral and topical voriconazole for the treatment of trauma-induced *Fusarium* keratitis. In the second case, a 43-year-old woman received a combination of IV, topical, and intravitreal voriconazole for keratitis caused by *F. solani* that was associated with contact lens wear. In both of these cases, voriconazole was initially effective until it had to be discontinued because of severe
hepatotoxicity. Patients were then switched to posaconazole as salvage therapy.

In the two case reports by Al-Badriyeh et al.,
110,111 1% voriconazole was used as a stand-alone therapy. In one case report, a 54-year-old woman presented with a painful injected eye. Despite empirical therapy, symptoms persisted. Keratitis was later diagnosed and identified as a rare S. apiospermum keratitis. Primary antifungal therapy with natamycin 5% was not successful and was switched to 1% voriconazole eye drops. Vision improved in five days, and the corneal defect completely re-epithelialized in a week. In the other report by Al-Badriyeh et al.,
111 a 48-year-old presented with keratitis following exposure to dust, the cause of which was later identified as C. albicans. Despite empirical antibacterial therapy, the epithelial defect persisted. Primary antifungal therapy with 1% voriconazole eye drops was initiated. The corneal infiltrate resolved in two days, the epithelial defect was completely healed, and visual acuity was restored in seven days.

Polizzi et al reported the only case where 2% voriconazole eye drops were used.
106 A 56-year-old man developed corneal ulceration caused by F. solani upon accidental contact with vegetation. Corneal ulceration developed and a perforating keratoplasty was performed. Systemic and topical amphotericin B and fluconazole were administered initially, but the patient did not improve. A new abscess formed on the transplanted graft and a wound leak developed. Therapy was then switched to IV and oral voriconazole in combination with topical 2% voriconazole eye drops. The patient completely recovered after 20 days of treatment with voriconazole.

A number of important considerations are associated with the above cases. One is that the use of voriconazole eye drops was associated with successful outcomes in most cases of fungal keratitis. Whilst the voriconazole therapy appeared to

<table>
<thead>
<tr>
<th>Author</th>
<th>Patient (age, sex)</th>
<th>Pathogen</th>
<th>Voriconazole concentration</th>
<th>Treatment</th>
<th>Outcome</th>
<th>Systemic voriconazole</th>
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<tr>
<td>Reis et al</td>
<td>16, female</td>
<td>Fusarium solani</td>
<td>1%</td>
<td>Salvage</td>
<td>Success</td>
<td>Intravenous voriconazole 6 mg/kg twice daily on day 1, followed by intravenous</td>
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<td>voriconazole 4 mg/kg twice daily until day 1. Oral voriconazole 6 mg/kg twice daily</td>
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<td>was then given for eight weeks</td>
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<tr>
<td>Klont et al</td>
<td>23, male</td>
<td>Fusarium solani</td>
<td>1%</td>
<td>Salvage</td>
<td>Success</td>
<td>Intravenous voriconazole 6 mg/kg twice daily on day 1, followed by intravenous</td>
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<td>voriconazole 4 mg/kg twice daily for two weeks. Oral voriconazole 200 mg twice daily</td>
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<td>was then given for two weeks</td>
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<td>Prats et al</td>
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<td>Salvage</td>
<td>Success</td>
<td>Oral voriconazole 200 mg twice daily for six weeks</td>
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<td>Jones et al</td>
<td>52, female</td>
<td>Aspergillus niger</td>
<td>1%</td>
<td>Salvage</td>
<td>Success</td>
<td>Oral voriconazole 200 mg twice daily for five weeks</td>
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<td>Tu et al</td>
<td>29, male</td>
<td>Fusarium species</td>
<td>1%</td>
<td>Primary</td>
<td>Failure</td>
<td>Oral voriconazole 200 mg twice daily for five weeks</td>
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<td>43, female</td>
<td>Fusarium solani</td>
<td>1%</td>
<td>Primary</td>
<td>Failure</td>
<td>Intravenous voriconazole 200 mg twice daily on day 1, followed by oral</td>
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<td>voriconazole 200 mg twice daily for six weeks</td>
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<td>Al-Badriyeh et al</td>
<td>54, female</td>
<td>Scedosporium apiospermum</td>
<td>1%</td>
<td>Salvage</td>
<td>Success</td>
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<td>Candida albicans</td>
<td>1%</td>
<td>Primary</td>
<td>Success</td>
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<td>was then given for four weeks</td>
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fail in the cases reported by Tu et al,74 the failure was not due to lack of efficacy, but to severe side effects from systemically administered voriconazole, which required discontinuation of treatment. A second issue is that voriconazole eye drops were used as an adjunct to systemic voriconazole in most reported cases. Only Al-Badriyeh et al have demonstrated the clinical benefit of topical voriconazole when used alone as primary and salvage therapies.110,111 Thirdly, whilst the 2% voriconazole eye drops were effective,106 the advantage of using 2% compared with 1% voriconazole eye drops remains unknown. A fourth issue is that, in the reports by Al-Badriyeh et al,110,111 the voriconazole eye drops were prepared as a solution containing 0.01% benzalkonium chloride as preservative, whereas, in all other case reports, the eye drops were prepared in sterile water for injection. In these case reports, voriconazole eye drops were typically administered with a dosing frequency of one drop every 0.5, 1.0, or 2.0 hours,69,74,106–111 and the average duration of administration of the voriconazole eye drops as adjunct therapy (ie, one to two months) was similar to the average duration of administration of the voriconazole eye drops given as monotherapy (ie, one month). However, it is common for the administration of the eye drops to extend beyond the resolution of fungal keratitis, with a duration that is more related to local clinical practices rather than time to resolution.

Safety

The safety of voriconazole in the eye was first evaluated in a rat model by Gao et al,85 who demonstrated that intravitreal voriconazole concentration as high as 25 mg/mL did not result in any electroretinographic or histologic abnormalities in the rat retina. This, however, cannot be extrapolated as an evidence of the safety of voriconazole when topically applied to the human cornea.

In a stability study by Al-Badriyeh et al,112 changes in the pH of 1% and 2% voriconazole eye drops were followed for over three months, and were found to range between 6.02 and 6.27. This was consistent with the findings of the only other stability study of voriconazole eye drops, performed by Dupuis et al,97 where a pH of 7 was maintained for 1% voriconazole eye drops with a storage duration of 30 days. These pH values are usually well tolerated by the eye.112 It is unlikely, therefore, that any eye irritation resulting from the use of the voriconazole eye drops would be a consequence of low pH.

Indeed, in the above studies that evaluated the application of voriconazole eye drops to the eye of animals and humans, the drops were well tolerated, and associated with mild or no side effects. In the animal model by Sponsel et al,98 all rabbits responded well to the voriconazole eye drops with no apparent side effects. In the horse model by Clode et al,99 0.5% and 1% voriconazole eye drops resulted in no side effects, and only the higher 3% concentration was associated with ocular toxicity. In the clinical study by Vemulakonda et al,80 only two patients reported a mild transient stinging sensation on instillation of the 1% voriconazole eye drops. Visualization was excellent in all 13 patients. Of the 10 patients in the study by Lau et al,61 four patients reported one or two instances of mild stinging, and one patient reported sneezing after the initial dose. None of the patients in the clinical study by Al-Badriyeh et al reported any side effects with the 2% voriconazole eye drops.101 These outcomes were consistent with the outcome in another study by Lau et al that evaluated voriconazole penetration into the vitreous humor of the eye after topical application of the eye drops.113 Here also, no side effects were reported. Similarly, in all of the case reports of using topical voriconazole in fungal keratitis, the eye drops were well tolerated with no side effects reported.69,74,106–111

Systemic side effects resulting from the topical administration of the voriconazole solution should be negligible. In the case of the 2% eye drops (the highest concentration that has been reported in humans), each drop contains only 0.001 mg of voriconazole which, when compared with the standard systemic dose of 200 mg oral or IV voriconazole twice daily,68 is unlikely to result in systemic concentration that is high enough to induce side effects.

Stability of voriconazole eye drops

According to the stability study by Al-Badriyeh et al,112 1% voriconazole eye drops, prepared in sterile benzalkonium chloride 0.01% solution, were stable for at least 14 weeks when stored at 2–8°C, while 2% voriconazole eye drops, also prepared in sterile benzalkonium chloride solution, were stable for 16 weeks at 2–8, 25, and 40°C. This was consistent with the stability study by Dupuis et al,97 where 1% voriconazole eye drops, prepared in sterile water for injection, were stable for at least four weeks when stored at 4°C. Such long-term stability data will help minimize waste and is pivotal to facilitate the use of the eye drops in the outpatient setting.

Patient adherence and satisfaction

Given the observed efficacy, safety, and stability, voriconazole eye drops appear to be a good option in the treatment of fungal keratitis. However, non-adherence to the prescribed dosing regimen may pose a challenge to achieving the desired treatment outcome and, consequently, lead to progression of
the infection. The benefits of using voriconazole eye drops, especially in the outpatient setting, could be diminished due to poor patient adherence if the dosing regimen is intense (eg, hourly to two-hourly drops) and is for an extended duration. The outpatient setting is different from the situation in clinical studies, in which maximal compliance can be achieved because the eye drops are often administered by nursing staff in hospitals. To date, there are no published data available about patient adherence to voriconazole eye drops.

The appropriate dosing frequency of voriconazole eye drops is not clear. The voriconazole aqueous humor concentration measured by Vemulakonda et al (6.49 ± 3.04 mg/L), where 1% eye drops were administered every two hours for 24 hours, is more than three times the concentration reported by Lau et al (1.90 ± 1.12 mg/L), where one drop was administered every hour for four hours. Based on these data, it is tempting to assume that the higher concentration measured by Vemulakonda et al was a result of drug accumulation in the eye; however, the significant difference in the post-dose sampling time (0.4 hours versus 1.1 hours in the Vemulakonda et al and Lau et al studies, respectively) could have been the key factor behind the differences in the measured voriconazole level in the aqueous humor.

**Conclusion**

Voriconazole eye drops appear to be effective when used for the treatment of fungal keratitis caused by a variety of fungi, including *F. solani*, *C. albicans*, *S. apiospermum*, and *A. niger*. The eye drops are well tolerated in the reported clinical trials and case studies. In addition, stability data for the extemporaneously prepared eye drops are available, minimizing the manufacturing cost and wastage associated with the eye drops.

Case reports have shown that voriconazole eye drops are effective when used as adjunctive therapy or monotherapy, and as primary or salvage therapy for the management of ophthalmic fungal keratitis. However, the number of these case reports and, hence, their significance remains limited. Whilst large-scale, randomized, controlled studies are needed, such studies are difficult to conduct and, consequently, audits or large case series on the use of voriconazole eye drops as monotherapy are necessary to confirm current findings and help establish the extent of their effectiveness.

Increasing the concentration of voriconazole eye drops may lead to increased efficacy and/or reduced dosing frequency; however, the benefit of using concentrations greater than 1% has not been evaluated in patients with fungal keratitis beyond a single case report. Studies using intact corneas have suggested concentration-independent penetration of voriconazole through the cornea and, consequently, it appears that administering 2% over 1% voriconazole eye drops in fungal keratitis is unlikely to give any additional benefit. Although some studies have suggested that epithelial damage is not necessary for voriconazole penetration, future studies that evaluate the penetration of 2% versus 1% voriconazole eye drops in patients with fungal keratitis, despite being difficult to perform, will be important. Given that the optimal dosing of the voriconazole eye drops remains unknown, studies to investigate the extent of voriconazole clearance from the human eye after topical administration should be conducted to guide the dosing frequency.

Current literature has provided some evidence on the effectiveness of voriconazole eye drops for the treatment of fungal keratitis; however, more data are required before a definite conclusion regarding their utility is drawn.

**Disclosures**

The authors report no financial or other conflicts of interest in this work.

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