

Up-Regulation of Tiam1 Promotes the Radioresistance of Laryngeal Squamous Cell Carcinoma Through Activation of the JNK/ATF-2 Signaling Pathway

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Purpose: Our previous study has revealed that T-lymphoma invasion and metastasis-inducing factor 1 (Tiam1) overexpression are significantly associated with aggressive behavior and poor prognosis in patients with laryngeal squamous cell carcinoma (LSCC). However, the influence of Tiam1 in the radioresistance of LSCC and its mechanism have never been elucidated.

Materials and Methods: Western blotting was used to confirm the relationship between Tiam1 and the JNK/ATF-2 signaling pathway. To explore the specific functions of Tiam1 and JNK/ATF-2 signaling pathway on the proliferation and apoptosis of LSCC after radiation, cloning formation assay and flow cytometry were conducted in vitro, and the experiments on a xenograft mouse model and TUNEL assay were performed in vivo.

Results: Western blotting indicated that Tiam1 can regulate the JNK/ATF-2 signaling pathway through the influence of the activity of JNK and ATF-2. Up-regulation of Tiam1 could promote proliferation and inhibit apoptosis of LSCC after radiation both in vitro and in vivo. Moreover, the down-regulation of the JNK/ATF-2 signaling pathway reduced the radioresistance of LSCC caused by Tiam1 up-regulation.

Conclusion: These results suggest that the up-regulation of Tiam1 expression can promote the radioresistance of LSCC through activation of the JNK/ATF-2 signaling pathway.

Keywords: JNK/ATF-2 signaling pathway, laryngeal squamous cell carcinoma, radioresistance, Tiam1, up-regulation

Introduction

Laryngeal squamous cell carcinoma (LSCC) is not a rare tumor occurring in the head and neck region, as it amounts to 6% of all cancer cases.¹ It comprises over 500,000 new cases and 200,000 deaths annually worldwide.^{2,3} The overall 5-year accumulative survival rate for patients with LSCC is unsatisfactory (50–70%).⁴ At present, the comprehensive treatment of LSCC includes surgery, radiotherapy, chemotherapy, and immunotherapy. Radiotherapy, as one of the most important treatments of malignant tumors, can cure some tumors. It plays a vital role in reducing postoperative recurrence and metastasis and improving the survival rate of patients.⁵ However, the curative effect of radiotherapy for some LSCC patients is currently disappointing. Radiotherapy resistance is one of the major factors compromising the radiotherapy effect on malignant tumors.⁶ Therefore, the recognition

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and identification of crucial proteins and the related signaling pathway associated with radiotherapy resistance are of great importance for the development of novel strategies in the prevention and treatment of patients with LSCC.

A gene designated T-lymphoma invasion and metastasis inducing factor 1 (Tiam1) was originally recognized as an invasion-inducing gene by proviral insertion combined with in vitro selection of invasive mouse T-lymphoma variants.⁷ Tiam1 can specifically activate the Rac signaling pathway, which mediates cellular growth, invasion, and metastasis.⁸ Tiam1 overexpression has been shown in a large number of tumors, including thyroid,⁹ nasopharyngeal,¹⁰ esophageal,¹¹ and renal carcinoma,¹² and colorectal cancer.¹³ Tiam1 has also been reported to have close correlations with apoptosis,¹⁴ migration, and invasion.^{15,16} Hence, we can conclude that Tiam1 overexpression is related to the malignant progression of tumors.

Studies have confirmed that the c-Jun N-terminal kinase (JNK)/activating transcription factor-2 (ATF-2) signaling pathway is closely related to invasion, metastasis, epithelial-mesenchymal transition, and apoptosis of malignant tumors.^{17–19} It has been proved that this pathway is relevant to the radioresistance of lung cancer cells.²⁰ Zhu et al's research findings revealed that Tiam1 can induce apoptosis by activating the Rac1/JNK signaling pathway.²¹ The Tiam1/Rac1/JNK signaling pathway plays a significant role in the apoptosis of malignant tumor cells.^{22,23} Consequently, Tiam1 may be an important regulator in the JNK signaling pathway.

Previous studies from our laboratory based on the analysis of clinical specimens illustrated that the overexpression of Tiam1 is significantly correlated with LSCC aggressive action and a poor outcome. We also confirmed that the up-regulation of Tiam1 could increase LSCC metastatic ability in vitro. Our previous experiment results demonstrated that Tiam1 expression may correlate with the tumorigenesis and development of LSCC and may be a beneficial therapeutic target for LSCC patients.²⁴ Nevertheless, there are relatively few reports on the role of Tiam1 on LSCC radioresistance. In an attempt to gain further insight into the radioresistance of Tiam1 in LSCC and its probable mechanism, here we report that the transfection of a Tiam1/C1199 plasmid to up-regulate Tiam1 expression in LSCC, and we observed changes in the proliferation, apoptosis and the expression of JNK/ATF-2 signaling pathway both in vitro and in vivo after radiation.

Materials and Methods

Cell Culture and Cell Transfection

The human TU686 cell line used in the current study was purchased from the Type Culture Collection of the Chinese Academy of Science (Shanghai, China). All cells were cultured in RPMI-1640 medium (Invitrogen, Carlsbad, CA, USA) containing 10% fetal bovine serum (FBS) and 100 IU/mL penicillin/streptomycin. The cells were grown in a 37°C humidified incubator with 5% CO₂.

The Tiam1 cDNA C1199 plasmid, a gift provided by Dr. John Collard (The Netherlands Cancer Institute, Amsterdam, Netherlands), which has the Tiam1 cDNA cloned as a *Bam*HI/*Xho*I fragment in the pcDNA3 plasmid containing a cytomegalovirus promoter and a hemagglutinin tag, was used for the stable transfection of Tiam1 up-regulation. The TU686 cells that transfected with Tiam1/C1199 cDNA and vector control plasmid were named Tiam1+ and Mock, respectively, as established and described in our previous study.²⁴

shRNA targeting JNK (TAGATGCATCTATTACCAG) and the nontargeting control (ATCTCGCTTGGGCGAGA GTAAG) cloned into a pGIPZ lentiviral plasmid were purchased from Dharmacon™. ATF-2 shRNA lentiviral particles (sc-29,205-V) were purchased from Santa Cruz Biotechnology. These shRNAs were transfected into TU686 and Tiam1+ cells, and stably expressed in these cells, which were termed JNK-, Tiam1+/JNK-, ATF-2-, and Tiam1+/ATF-2-. The TU686 cell transfected with the lentiviruses encoding the nontargeting control was termed LV/NC.

Western Blotting

All Western blotting analyses were performed as we described previously.²⁴ Briefly, total protein (40 µg/sample) was separated in SDS-PAGE (8%) and then electroblotted onto polyvinylidene difluoride membranes (Immobilon-P, Millipore, USA). The membranes were separately incubated with anti-Tiam1 mouse monoclonal antibody (sc-393,315, dilution 1:100) (Santa Cruz, CA, USA), rabbit monoclonal antibody against JNK (ab179461, dilution 1:1000) and phosphate (p)-JNK (ab124956, dilution 1:1000) (Abcam, Cambridge, UK), mouse monoclonal antibody against ATF-2 (sc-242, dilution 1:200), and p-ATF-2 (sc-8398, dilution 1:100) (Santa Cruz, CA, USA) at 4°C overnight. After washing three times, the membranes were blocked with HRP-conjugated Goat Anti-mouse IgG and Goat Anti-rabbit IgG for 1 h at room temperature. Then the membranes were incubated with 3, 3'-diaminobenzidine (Zhongshan GoldenBridge, Beijing, China) to visualize the bands. The results were obtained

using a Kodak film and quantified by software Quantity One (Version 4.6.2, Bio-Rad Technical Service Department, USA). The antibody against GAPDH (BM3874, dilution 1:2000) (Boster, China) was used as control. All assays were performed in triplicate.

Radiotherapy Treatment

TU686 cells were grown to 50% confluence and then divided into six groups. One was the control group, which did not receive any X-ray radiation (termed as TU686 only). The other groups received a total dose 10 Gy (2 Gy each time) X-ray radiation (termed TU686 RT, Mock RT, LV/NC RT, Tiam1+ RT, Tiam1+/JNK- RT, and Tiam1+/ATF-2- RT). The cells and xenograft animal tumors were exposed to X-ray radiation on a Varian Clinac 23 EX medical linear accelerator in a field sized of $35 \times 35 \text{ cm}^2$ with single energy of 6 MV X-ray and a dose rate of 500 MU/min; the source to skin distance was approximately 100 cm.

Plate Cloning Formation Assay

Cells were digested, and 700 cells/well of each group were inoculated in triplicate in 6-well plates. The medium was changed every other day. Cell culture was performed for ten days and was terminated when macroscopic colonies were found in plates. After washing with phosphate-buffered saline (PBS), fixation with 4% paraformaldehyde for 15 min was carried out. After that, the Giemsa solution was mixed, and staining was performed for 20 min. Clones containing > 50 cells were counted under an inverse microscope. Cloning formation rate (%) = (number of clones/number of inoculated cells) \times 100%.

Cell Apoptosis by Flow Cytometry

Cell apoptosis was analyzed by flow cytometry using the Annexin V-FITC Apoptosis Detection kit (Invitrogen, Carlsbad, CA, USA). The test was performed following the manufacturer's protocol. Cells were washed three times with ice-cold PBS, and the concentration was adjusted to $1 \times 10^6/\text{mL}$. Then they were mixed with 5 μL Annexin V-FITC and 5 μL PI. The reaction was carried out for 15 min at room temperature in the dark. After adding another 400 μL of 1 \times binding buffer, cell apoptosis was detected on a FACS Calibur cytometer (Becton-Dickinson). All experiments were carried out in triplicate.

LSCC Xenograft Mouse Model

The study was approved by the Institutional Animal Care and Use Committee, The Second Xiangya Hospital, Central South

University. It was conducted following the Institutional Guidelines and the Guide for the Care and Use of Laboratory Animals (NIH publication no. 85–23, revised 1996). A xenograft model in nude mice of human LSCC was established as described in Zhang et al.^{25,26} Briefly, 4-week-old athymic BALB/C nude male mice (about 20 g of body weight) were purchased from the Shanghai Laboratory Animal Center, Chinese Academy of Sciences, China, and housed under specific pathogen-free conditions. Each animal was injected with 8.0×10^6 of TU686 or Tiam1+ cells suspended in 0.4 mL of Hanks-buffered saline into the right armpit. The xenograft tumors were measured every four days, and all nude mice were killed 28 days after cell implantation. The primary tumors of the mice were collected, and tissue serial sections were formalin-fixed, paraffin-embedded, stained with hematoxylin-eosin, and studied on a light microscope.

TUNEL Assay

The TUNEL assay was performed to assess the changes of apoptosis-related factors by using the TUNEL fluorescence kit (Hoffman-La Roche Ltd.) to stain cells. Briefly, the formalin-fixed paraffin-embedded blocks of the xenograft tumors were deparaffinized in xylene and rehydrated in ethanol. The samples were pretreated with proteinase K (20 $\mu\text{g}/\text{mL}$) for 15 min. The TUNEL biotin marker (50 μL) was applied, and the sections were then incubated for 60 min. Finally, using a Confocal Laser Scanning Microscope (Leica TCS SP8, Leica Microsystems, Wetzlar, Germany), apoptosis was evaluated by counting the number of cells double-stained by PI and TUNEL.

Statistical Analysis

Statistical analysis was carried out using the SPSS 22.0 software package (SPSS, Inc., Chicago, IL). All quantitative results were expressed as mean \pm SD. Statistical differences between groups were compared using one-way analysis of variance (ANOVA). A p-value of < 0.05 was considered to be statistically significant.

Results

The Interplay Between Tiam1 and JNK/ATF-2 Signaling Pathway in LSCC

Stable clones overexpressing the Tiam1 were established, as mentioned before²⁴ (Figure 1A). Western blotting revealed that the JNK shRNA (Figure 1B) and ATF-2 shRNA (Figure 1C) were successfully transfected into

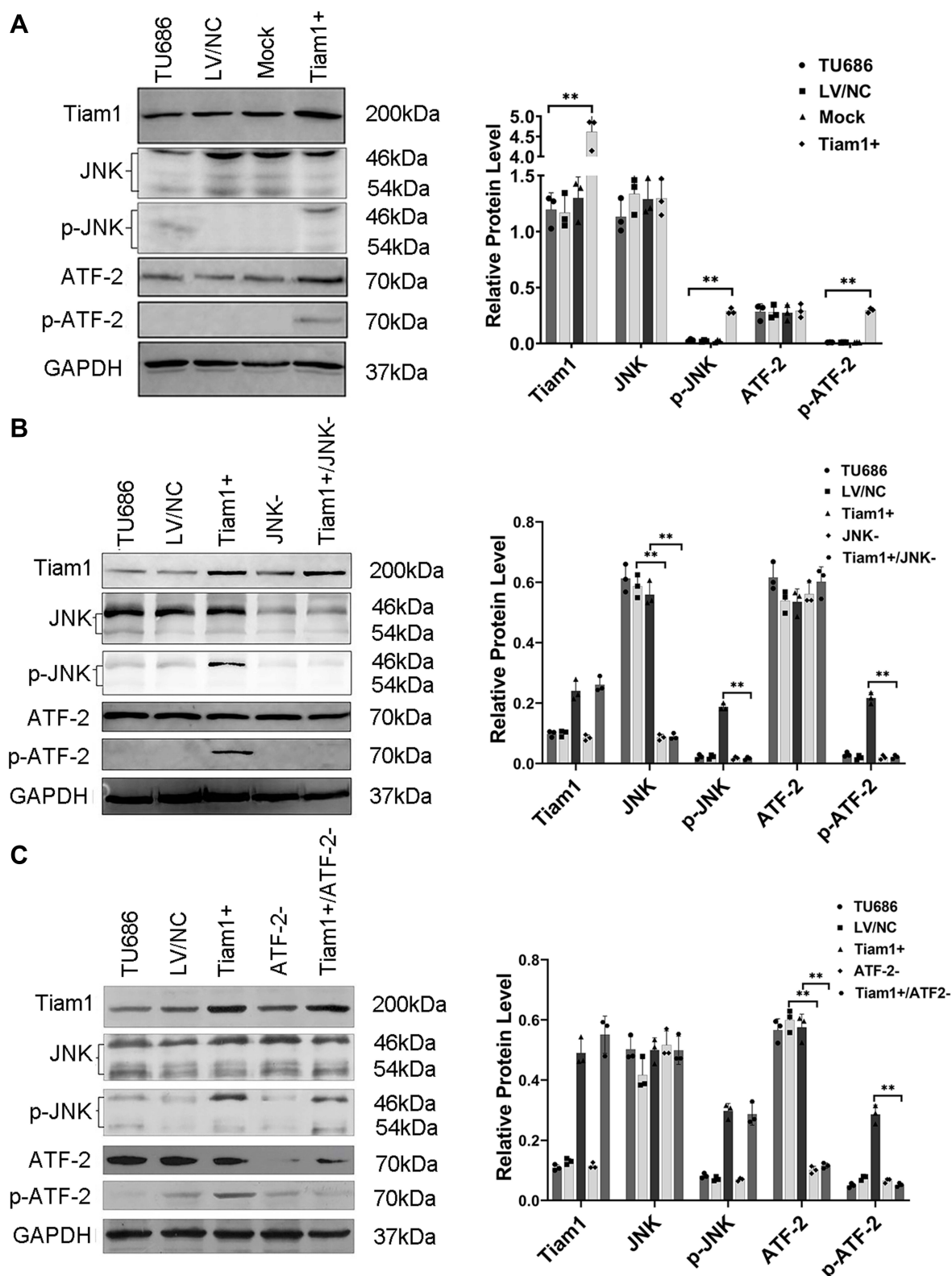


Figure 1 Up-regulation of Tiam1 and down-regulation of the JNK/ATF-2 signaling pathway in LSCC cells. Western blotting was used to confirm the relationship between Tiam1 and the JNK/ATF-2 signaling pathway.

Notes: (A) Up-regulation of Tiam1 and the expression of the JNK/ATF-2 signaling pathway were detected by Western blotting. (B, C) Down-regulation of the JNK/ATF-2 signaling pathway was detected by Western blotting. Data are shown as mean \pm SD. **P<0.01.

the TU686 and Tiam1+ cells. We obtained the following stable cell lines: Tiam1+ (TU686 cells transfected with the Tiam1/C1199 plasmid), Mock (TU686 cells transfected with vector control plasmid), LV/NC (TU686 cells transfected with a lentivirus encoding the nontargeting control), JNK- (TU686 cells transfected with JNK shRNA), ATF-2- (TU686 cells transfected with ATF-2 shRNA), Tiam1+/JNK- (Tiam1+ cells transfected with JNK shRNA), and Tiam1+/ATF-2- (Tiam1+ cells transfected with ATF-2 shRNA). Western blotting showed that the levels of p-JNK ($p<0.01$) and p-ATF-2 ($p<0.01$) were higher in Tiam1+ cells than those in the TU686 cells; however, the JNK ($p=0.874$) and ATF-2 ($p=0.512$) expression had no change. This suggests that Tiam1 may regulate the JNK/ATF-2 signaling pathway by influencing the activity of JNK and ATF-2 (Figure 1A). Furthermore, Western blotting results revealed that the Tiam1 expression ($p=0.794$) had no change in Tiam1+/JNK- and Tiam1+/ATF-2- cells compared with Tiam1+ cell (Figure 1B and C). In addition, the down-regulation of JNK and ATF-2 can reverse the phosphorylation of the JNK/ATF-2 signaling pathway (p-JNK, $p<0.01$, and p-ATF-2, $p<0.01$) caused by the Tiam1 up-regulation (Figure 1B and C). These results further show that Tiam1 is the upstream factor of the JNK/ATF-2 signaling pathway.

Up-Regulated Tiam1 Expression Promotes Proliferation and Inhibits the Apoptosis of LSCC Cells After X-Ray Radiation in vitro

To confirm the role of Tiam1 on the proliferation and apoptosis of LSCC cells after X-ray radiation in vitro,

a plate cloning formation assay and a flow cytometry analysis were performed, respectively. As shown in Figure 2, the cloning formation rate was decreased in TU686 RT cells compared with TU686 cells ($P<0.01$), but increased in Tiam1+ RT cells compared with the TU686 RT cells ($P<0.01$). This result reveals that the up-regulated Tiam1 can promote the proliferation of LSCC cells after radiation. In addition, the apoptotic rate of TU686 RT cells was increased compared with the TU686 cells ($P<0.01$), but the apoptotic rate of the Tiam1+ RT cells was significantly decreased compared with TU686 RT cells ($P<0.01$) (Figure 3). This result demonstrates that up-regulated Tiam1 can inhibit the apoptosis of LSCC cells after radiation. Therefore, we concluded that the up-regulation of Tiam1 can increase the radioresistance of LSCC.

Tiam1 Promotes the Radioresistance of LSCC Through the JNK/ATF-2 Signaling Pathway

To elucidate the mechanism of Tiam1 in the promotion of radioresistance of LSCC, the Tiam1+/JNK-, and Tiam1+/ATF-2- cells were also treated with X-ray radiation. The results demonstrated that the cloning formation rate was decreased in Tiam1+/JNK- RT and Tiam1+/ATF-2- RT cells compared with Tiam1+ RT cells (both $P<0.01$) (Figure 2), but the apoptotic rate of Tiam1+/JNK- RT and Tiam1+/ATF-2- RT cells were significantly increased (both $P<0.01$) (Figure 3). These results demonstrate that Tiam1 can regulate the JNK/ATF-2 signaling pathway and further influence the radiation sensitivity of LSCC.

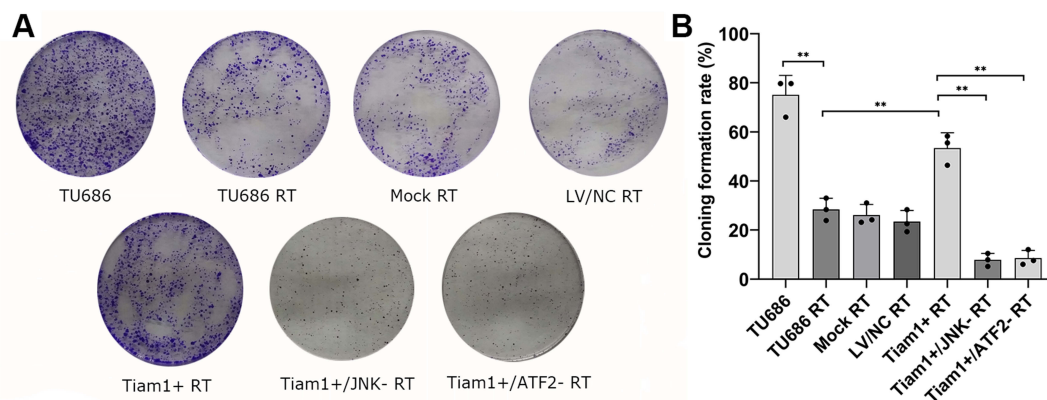


Figure 2 The effect of Tiam1 up-regulation and down-regulation of JNK/ATF-2 signaling pathway on the proliferation of LSCC cells after radiation.

Notes: (A) Cell proliferation was measured by the plate cloning formation assay. (B) The cloning formation rate in each group. Data are shown as mean \pm SD. ** $P<0.01$.

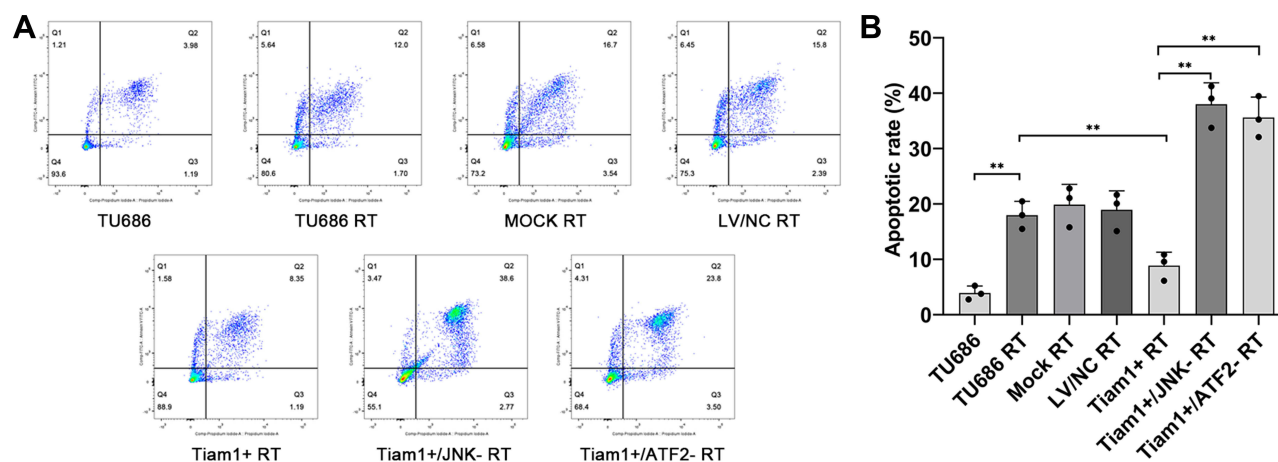


Figure 3 The effect of TiamI up-regulation and down-regulation of the JNK/ATF-2 signaling pathway on the apoptosis of LSCC cells after radiation.

Notes: (A) Cell apoptosis was measured by flow cytometry. (B) The apoptotic rate in each group. Data are shown as mean ± SD. **P<0.01.

Up-Regulation of TiamI Promotes the Proliferation and Inhibits the Apoptosis of LSCC Cells After X-Ray Radiation in vivo

An established LSCC xenograft mouse model was used to determine whether up-regulated Tiam1 could regulate the radiation sensitivity of LSCC in vivo. The experiment comprised three groups (five mice/group). Subsequently, TU686 (ten mice in total) and stable transfected Tiam1+ (a total of five mice) cells were injected subcutaneously into the right armpit. The TU686 (five mice) and Tiam1+ (five mice) groups received X-ray radiation (a total dose of 10 Gy, 2 Gy each time) 8 days after inoculation (termed TU686 RT and Tiam1+ RT groups, respectively). All the tumor formation was carefully monitored. After 28 days, all of the nude mice were sacrificed, and the tumor weights were measured. We used the TUNEL assay to test cell apoptosis in paraffin-embedded blocks

of the xenograft tumors. Moreover, to clarify the mechanism of Tiam1 on radioresistance of LSCC, Western blotting was performed to test the expression of the JNK/ATF-2 signaling pathway in the xenograft tumors.

As shown in Figure 4, the average tumor volume and tumor weight of mice in the TU686 RT group were lower than those in the TU686 group (without radiation) (P<0.01). However, the average tumor volume and tumor weight of mice in the Tiam1+ RT group were higher than that those in the TU686 RT group (P<0.01). This result reveals that the up-regulation of Tiam1 can promote the proliferation of LSCC cells after radiation. The TUNEL assay showed that the PI and TUNEL double-stained cells in the TU686 RT group were increased compared to the TU686 group (P<0.01). However, the percentage of PI and TUNEL double-stained cells in the Tiam1+ RT group was lower than that in the TU686 RT group (P<0.01) (Figure 5). This result

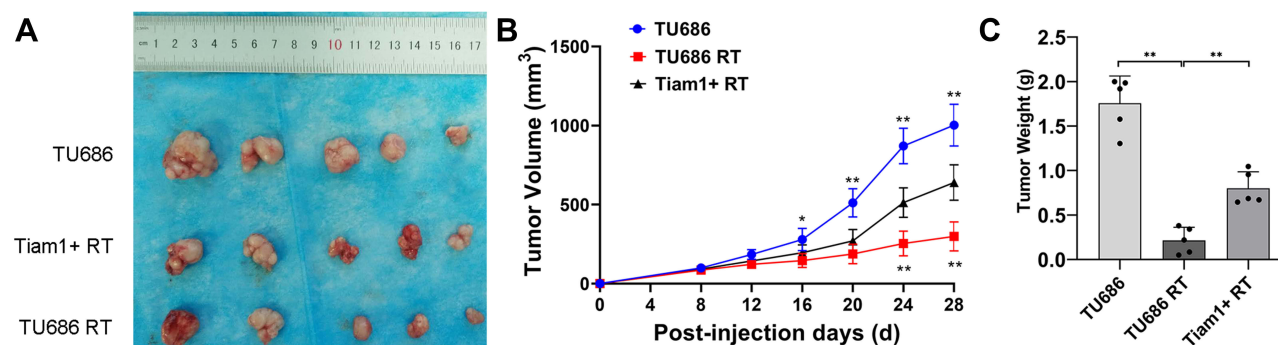


Figure 4 The effect of TiamI up-regulation on the proliferation of LSCC xenograft tumors after radiation.

Notes: (A) The primary tumors of killed nude mice at 28 days after inoculation with LSCC cells. (B) The dynamics of xenograft tumor growth in each group. (C) Quantitative measurement of all tumor weights. Data are shown as mean ± SD. *P<0.05, **P<0.01.

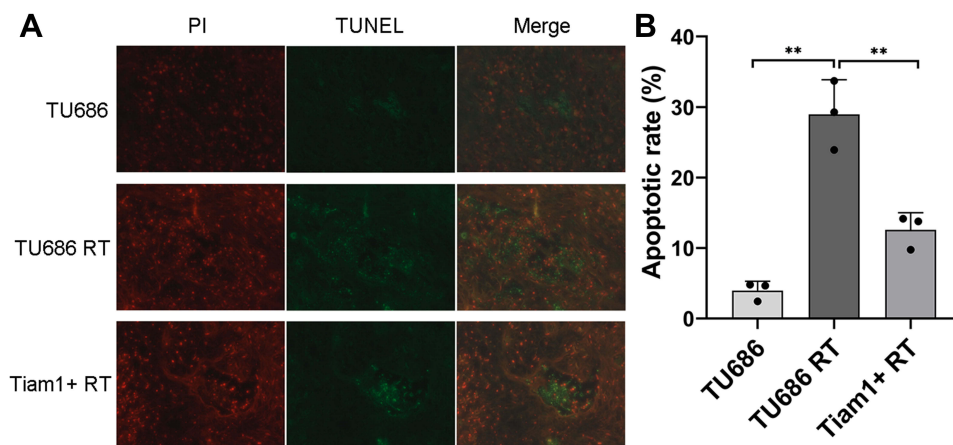


Figure 5 The effect of Tiam1 up-regulation on the apoptosis of LSCC xenograft tumors after radiation.

Notes: (A) The apoptosis of xenograft tumors was measured by the TUNEL assay. (B) The apoptotic rate in each group. Data are shown as mean \pm SD. ** $P < 0.01$.

demonstrates that up-regulated Tiam1 can inhibit the apoptosis of LSCC cells after radiation. Finally, Western blotting revealed that the expression of Tiam1 ($p < 0.01$), p-JNK ($p < 0.01$), and p-ATF-2 ($p < 0.05$) was increased in Tiam1+ RT group compared to the two other groups (Figure 6). Hence, the results of the in vivo and in vitro experiments are consistent. Consequently, we further demonstrated that the up-regulated Tiam1 expression could increase the radiation resistance of LSCC through the JNK/ATF-2 signaling pathway in vivo.

Discussion

Tiam1 was originally identified as an invasion and metastasis gene by Habets et al.⁷ Like all members of the Ras superfamily, the Rho-like small guanosine triphosphate

(GTP) binding proteins function as molecular switches, cycling between an inactive GDP-bound state and an active GTP-bound state. The cycle is catalyzed by Guanine nucleotide exchange factors (GEFs), which positively stimulate GTP binding proteins in response to multiple signaling pathways.⁸ As the specific GEFs for the Rho GTPase, the oncogenic effects of Tiam1 is a consequence of activation of the Rac signaling pathway.²⁷ Tiam1/Rac signaling is attributed to a stimulation of the c-Jun N-terminal kinase pathway, p38 mitogen-activated protein kinase pathway, and extracellular signal-regulated kinase pathway,^{28–30} which are thought to correlate with the regulation of gene expression at transcription. In addition, Tiam1 participates in oncogenic transformation³¹ and modulates changes in the actin cytoskeleton, cell migration,^{32–34} cell polarity,³⁵ and

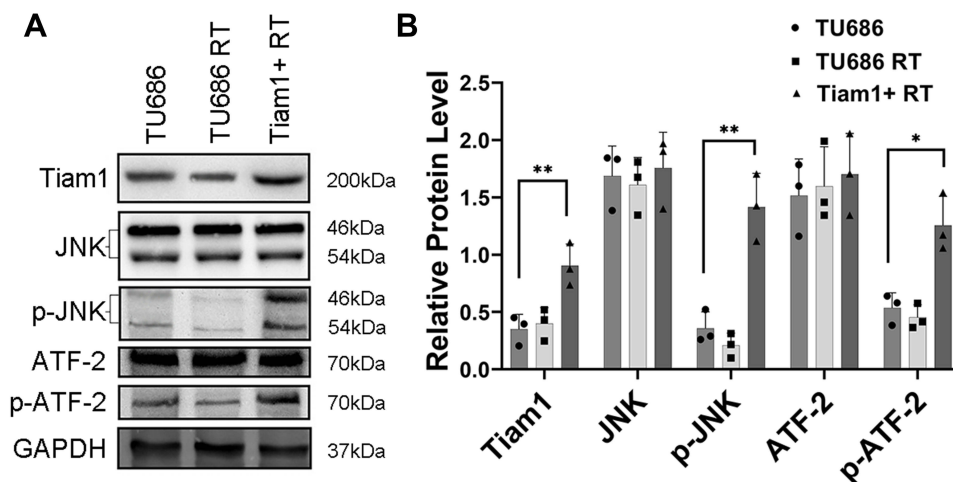


Figure 6 The expression levels of Tiam1 and the JNK/ATF-2 signaling pathway in xenograft tumors of TU686, TU686 RT, and Tiam1+ RT groups (A).

Notes: Data are shown as mean \pm SD (B). * $P < 0.05$, ** $P < 0.01$.

tumor microenvironment,³⁶ which are considered to be related to the tumor progression.

As we mentioned before, up to now, several studies have investigated the relationship between Tiam1 overexpression and multiple types of cancer.^{9–13} With respect to LSCC, our previous study focused on clinical tissue specimens had confirmed that overexpression of Tiam1 is significantly correlated with the LSCC aggressive action and poor outcome.²⁴ This suggests that Tiam1 may play a vital role in the progression of LSCC. Moreover, radiotherapy is one of the most important treatment methods for LSCC. Hence, we infer that Tiam1 may have an impact on the radiotherapy sensitivity of LSCC. However, to the best of our knowledge, there have been few published studies on the role of Tiam1 in tumor radioresistance. Consequently, to gain further insight into the radioresistance of Tiam1 in LSCC, in the current study, we up-regulated Tiam1 in cell lines of LSCC, and we observed the changes in cell proliferation and apoptosis after radiation both *in vitro* and *in vivo*.

In the present study, we found that the cloning formation rate was increased in Tiam1+ RT cells compared with TU686 RT cells. This result reveals that the up-regulation of Tiam1 can promote the proliferation of LSCC cells after radiation. In addition, the apoptotic rate of Tiam1+ RT cells was decreased compared with the TU686 RT cells. This result demonstrates that the up-regulation of Tiam1 can inhibit the apoptosis of LSCC cells after radiation. In the *in vivo* experiment, we also found that the average tumor volume and tumor weight of mice in the Tiam1+ RT group were higher than those in the TU686 RT group. This result further confirms that the up-regulation of Tiam1 can promote the proliferation of LSCC cells after radiation. The TUNEL assay showed that the PI and TUNEL double-stained cells in the Tiam1+ RT group were decreased compared to the TU686 RT group. This result also certifies that the up-regulated Tiam1 can inhibit the apoptosis of LSCC cells after radiation. Consequently, we demonstrated that the up-regulation of Tiam1 expression could increase the radioresistance of LSCC both *in vitro* and *in vivo*.

As we mentioned before, the JNK/ATF-2 signaling pathway is participating in the cell apoptosis, invasion, metastasis and radioresistance of malignant tumors.^{17–20} Concerning the relationship between Tiam1 and the JNK/ATF-2 signaling pathway, as far as we know, there have been several reports confirming that Tiam1 can induce

apoptosis through the activation of the Rac1/JNK signaling pathway.^{21–23} Therefore, we infer that Tiam1 can probably regulate the JNK/ATF-2 signaling pathway and further influence the radioresistance of LSCC. Then, we down-regulated the expression of the JNK/ATF-2 signaling pathway to test our hypothesis and elucidate the mechanism of Tiam1 on the promotion of radioresistance of LSCC.

In the present study, we first explored the interplay between Tiam1 and JNK/ATF-2 signaling pathway in LSCC by using Western blotting. The results showed that the levels of p-JNK and p-ATF-2 expression were increased after Tiam1 up-regulation; however, the JNK and ATF-2 expression had no change. In addition, Western blotting results revealed that the Tiam1 expression had no change after JNK or ATF-2 down-regulation. Hence, these results show that Tiam1 is the upstream factor of the JNK/ATF-2 signaling pathway. Furthermore, Tiam1 may regulate the JNK/ATF-2 signaling pathway by influencing the activity of JNK and ATF-2.

Subsequently, in the *in vitro* experiment, we found that the cloning formation rate was decreased in Tiam1+/JNK-RT and Tiam1+/ATF-2- RT cells compared with the Tiam1 + RT cells, but the apoptotic rate of Tiam1+/JNK- RT and Tiam1+/ATF-2- RT cells was significantly increased. These results reveal that the down-regulation of the JNK/ATF-2 signaling pathway can reduce the radioresistance of LSCC caused by Tiam1 up-regulation. Similarly, in the *in vivo* experiment, Western blotting revealed that the expression of Tiam1, p-JNK, and p-ATF-2 was increased in the xenograft tumors of the Tiam1+ RT group compared to the two other groups. Consequently, we further validated that Tiam1 can influence the radioresistance of LSCC through the JNK/ATF-2 signaling pathway.

Conclusion

In summary, our present study indicates that Tiam1 up-regulation can promote the radioresistance of LSCC through the activation of the JNK/ATF-2 signaling pathway, which suggests that Tiam1 and the JNK/ATF-2-mediated signaling are valuable therapeutic targets to improve the efficacy of radiotherapy for patients with LSCC.

Abbreviations

Tiam1, T-lymphoma invasion and metastasis-inducing factor 1; LSCC, laryngeal squamous cell carcinoma; JNK, c-Jun N-terminal kinase; ATF-2, activating transcription factor-2.

Data Sharing Statement

The data that support the findings of this study are available from the corresponding author Dr Xinming Yang upon reasonable request.

Ethics Approval and Informed Consent

The study was approved by the Institutional Animal Care and Use Committee, The Second Xiangya Hospital, Central South University. It was conducted following the Institutional Guidelines and the Guide for the Care and Use of Laboratory Animals (NIH publication no. 85-23, revised 1996).

Author Contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

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Disclosure

The authors have no conflicts of interest to disclose.

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