

Paeonol Inhibits Cell Proliferation, Migration and Invasion and Induces Apoptosis in Hepatocellular Carcinoma by Regulating miR-21-5p/KLF6 Axis

This article was published in the following Dove Press journal:
Cancer Management and Research

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Background: Hepatocellular carcinoma (HCC) is one of the most common tumors with high mortality. MicroRNAs (miRNAs) were reported as crucial markers for the diagnosis of HCC. Paeonol exerted many pharmacological effects on tumor progression. This study aimed to elucidate the underlying molecular mechanism of paeonol in HCC progression.

Methods: Cell viability was determined by Cell Counting Kit-8 (CCK-8) assay. Cell apoptosis was examined by flow cytometry. The levels of Cyclin D1, cyclin-dependent kinase 4 (CDK4), B-cell lymphoma-2 (Bcl-2) and Bcl-2 associated X protein (Bax) were detected by Western blot assay. Cell migration and invasion were assessed by transwell assay. The levels of matrix metalloproteinase-2 (MMP2) and matrix metalloproteinase-9 (MMP9) were measured by Western blot. The expression of miR-21-5p and kruppel-like factor 6 (KLF6) was detected by quantitative real-time PCR (qRT-PCR) or Western blot assay, respectively. Dual-luciferase reporter assay was performed to analyze the interaction between miR-21-5p and KLF6. The enrichment of miR-21-5p was determined by RNA pull-down assay. Xenograft assay was conducted to analyze tumor growth in vivo.

Results: The results demonstrated that cell viability of Hep3B and Huh-7 cells was inhibited, while cell apoptosis was promoted after treatment with paeonol. Transwell assay indicated that cell migration and invasion were blocked in paeonol-treated cells. Moreover, miR-21-5p expression was markedly decreased in paeonol-treated cells and its knockdown suppressed cell viability, migration and invasion, but contributed to cell apoptosis. MiR-21-5p targeted KLF6 and its silencing prominently elevated KLF6 level. Furthermore, the restoration experiment determined that miR-21-5p and KLF6 were antagonisms on cell viability, apoptosis, migration and invasion. Also, paeonol abated the decrease in KLF6 level caused by miR-21-5p up-regulation. Besides, paeonol suppressed tumor growth in vivo.

Conclusion: Paeonol impeded cell viability, migration and invasion and triggered apoptosis by regulating miR-21-5p/KLF6 axis in HCC cells. Xenograft assay confirmed that paeonol inhibited tumor growth through miR-21-5p/KLF6 axis in HCC in vivo.

Keywords: hepatocellular carcinoma, paeonol, miR-21-5p, KLF6

Introduction

Liver is a metabolic organ of the body and plays the role of detoxification, storage of liver sugar, and synthesis of secretory proteins.¹ Liver diseases, including hepatitis, cirrhosis, hepatapostema and hepatocellular carcinoma, are regarded as major obstacles to the health of people worldwide.² Hepatocellular carcinoma (HCC) is one of the most common cancers in the world,^{3,4} causing about 750,000 deaths worldwide each year,⁵ and China accounts for half of all deaths. Limited

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treatment choices are the main cause of high mortality.⁶ As previously reported, there are many factors that contributed to HCC, such as alcohol drinking, hepatic toxin, hepatitis virus infection, and gene modification.⁷ According to the Barcelona-Clinic-Liver-Cancer (BCLC) staging system, HCC patients usually go through four stages: early subclinical stage, subclinical stage, metaphase stage and late stage.⁸ The stage of cancer directly determines the mortality rate of HCC patients. Early HCC patients can even be completely cured by hepatectomy or liver transplantation.⁹ For patients with HCC for more than five years, the survival rate was less than 14%.¹⁰ Hence, the diagnostic biomarkers for early HCC are urgently needed for the treatment and prevention of HCC.

Paeonol (2'-hydroxy-4'-methoxyacetophenone) is extracted from the herb *Pycnostelma paniculatum* K.S and *Paeonia suffruticosa* Andrews.¹¹ In recent years, paeonol has attracted worldwide attention due to its antineoplastic activity. In vitro experiments demonstrated that paeonol could cause the death of malignant tumor cells.¹² Also, paeonol plays a vital role as an anti-inflammatory, anti-oxidant agent in HCC.^{13,14} Although the pharmacological activity of paeonol has been extensively proposed, the mechanism of paeonol in the progression of HCC remains unclear.

MicroRNAs (miRNAs) are a family of non-coding RNAs and act as vital regulators in many physiological and biological processes, including cell viability, apoptosis, migration and invasion.^{15–17} Mature miRNAs regulate gene expression by the complementary match with the 3'-untranslated region (3'-UTR) of the target mRNA.¹⁸ A large number of miRNAs were identified to be involved in the diagnosis and therapy of HCC.^{19–21} MiR-21-5p, as a crucial regulatory factor in tumorigenesis, was proved to play a potential role in metabolism and apoptosis of HCC.²² The previous study indicated that miR-21 up-regulation enhanced disease progression in HCC patients.²³ Moreover, the aberrant expression of miR-21-5p made it a prognostic biomarker for HCC.^{24,25} Moreover, miR-21 promoted cell migration and invasion via regulating KLF5 in HCC.²⁶ However, the molecular mechanism of miR-21-5p in HCC needs to be further investigated.

Kruppel-like factor 6 (KLF6) is a member of the sp1/KLF transcription factor family and acts as a tumor suppressor gene in human cancers. KLF6 is widely expressed in human tissues and plays a pivotal role in cell cycle, apoptosis and differentiation.²⁷ He et al reported that KLF6 blocked cell proliferation and cell cycle in HCC

cells.²⁸ Besides, KLF6 expression was significantly down-regulated in HCC.²⁹ These data implied that KLF6 was associated with HCC progression and might be an essential target for HCC treatment. Nevertheless, the function of KLF6 in paeonol-treated HCC cells and the network of paeonol/miR-21-5p/KLF6 in HCC remain further research.

Our research was designed to explore the role of paeonol in HCC and the molecular mechanism of paeonol in cell viability, apoptosis, migration and invasion in vitro and in xenograft model in vivo.

Materials and Methods

Cell Culture and Paeonol Treatment

HCC cell lines (Hep3B and Huh-7) were provided by the Institute of Basic Medical Sciences of Peking Union Medical College (Beijing, China). Cells were maintained in Dulbecco's Modified Eagle Medium (Gibco, Carlsbad, CA, USA) supplementing with 10% fetal bovine serum (Hyclone, Logan, UT, USA). All cells were cultured in a constant temperature incubator at 37°C with 5% CO₂. Paeonol was purchased from the National Institutes for Food and Drug Control (Beijing, China). For different concentrations of paeonol (50 µM, 100 µM, 200 µM, 400 µM, 800 µM), varying amounts of normal saline were used to dissolve paeonol. HCC cell lines were cultivated in medium with various concentrations of paeonol for 24 h.

Cell Transfection

MiR-21-5p inhibitor, miR-21-5p mimic and their corresponding negative controls (NC inhibitor and NC mimic), small interference RNA (si-RNA) against KLF6 (si-KLF6) and si-RNA against NC (si-NC) were all obtained from Genepharma Co. Ltd (Genepharma, Shanghai, China). These oligos were transfected into Hep3B and Huh-7 cells with Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA, USA). After incubation for 24 h, cells were collected for further experiment.

Cell Viability Assay

Six groups of Hep3B cells and six groups of Huh-7 cells were prepared in 96-well plates, each well containing approximately 10,000 cells. Six concentrations of paeonol (0 µM, 50 µM, 100 µM, 200 µM, 400 µM, 800 µM) were transferred into cells, respectively. Then, the cells treated with paeonol were cultured for 24 h at 37°C with 5% CO₂. Subsequently, 10 µL of cell counting kit-8 (CCK-8) solution (Beyotime,

Shanghai, China) was added into cells. After incubation for 4 h, the absorbance was detected by a microplate reader (Bio-Rad, Hercules, CA, USA) at 450 nm. The value of absorbance is proportional to the number of cells.

Flow cytometry. For cell apoptosis assay, the Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) apoptosis detection kit (BD Biosciences, San Jose, CA, USA) was used to analyze cell apoptosis. HCC cells were harvested after treatment with paeonol or miRNA inhibitor for 1 day. The cells were then resuspended with binding buffer and 5 μ L Annexin V-FITC and PI solution. Dark incubation for 10 min, the apoptosis cells were determined by flow cytometer (BD Biosciences). For cell cycle assay, Hep3B and Huh-7 cells were incubated with paeonol (50 μ M or 800 μ M). Then, cells were resuspended in 200 μ L PBS containing 10 μ L PI and incubated at room temperature in the dark for 15 min. The cell cycle was monitored by the flow cytometer (BD Biosciences) with CellQuest Software (Becton–Dickinson).

Western Blot Assay

The cells were lysed in lysis buffer (Beyotime) and then subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) gel electrophoresis to separate proteins by molecular weight. Next, the proteins were blotted onto polyvinylidene fluoride (PVDF) membranes (Millipore, Billerica, MA, USA). Then, the membranes were treated with 5% non-fat milk for 1 h. Next, the membranes were incubated with primary antibodies against Cyclin D1 (ab16663; 1/100; Abcam, Cambridge, MA, USA), CDK4 (ab108357; 1/5000; Abcam), Bcl-2 (ab32124; 1/1000; Abcam), Bax (ab32503; 1/4000; Abcam), MMP2 (ab37150; 1 μ g/mL; Abcam), MMP9 (ab38898; 1/1000; Abcam), KLF6 (ab241385; 1/1000; Abcam) and GAPDH (ab8245; 1/2000; Abcam) overnight at 4°C. Next, the membranes were interacted with horseradish peroxidase-conjugated secondary antibody (ab6721; Abcam) with 1/3000 dilution at room temperature for 2 h. Finally, the protein bands were visualized by enhanced chemiluminescence chromogenic substrate (Beyotime) and measured by Image Lab software (Bio-Rad).

Cell Migration and Invasion Assays

Cell migration and invasion were detected by transwell assays. First, about 40,000 cells were seeded in the upper chambers (Corning, New York, NY, USA) and incubated at 37°C for 24 h. Then, the cells through the membrane were incubated with 0.5% crystal violet (Sigma, St. Louis, MO, USA). Finally, a microscope (Olympus, Tokyo,

Japan) was used to observe the migrated and invaded cells. For cell invasion assay, the upper chamber was coated by Matrigel (Becton Dickinson, Franklin Lakes, NJ, USA) prior to inoculation, and cell migration assay was not required.

Quantitative Real-Time PCR (qRT-PCR)

Total RNA of Hep3B and Huh-7 cells was extracted by RNA extraction Kit (TransGen Biotech, Beijing, China). TransScript miRNA First-Strand cDNA Synthesis Kit (TransGen Biotech) was utilized for the synthesis of qRT-PCR templates. TransStart Top Green qPCR SuperMix (TransGen) was used to perform qRT-PCR. The primers for miR-21-5p and internal control were purchased from Sangon Biotech (Sangon, Shanghai, China). All data were calculated by $2^{-\Delta\Delta C_t}$ method. The sequences of primers for miR-21-5p and U6 small RNA were listed as follows: U6: Forward: 5'-GCGCGTCGTGAAGCGTTC-3', Reverse: 5'-GTGCAGGGTCCGAGGT-3'; miR-21-5p: Forward: 5'-T GCGCTAGCTTATCAGACTGAT-3', Reverse: 5'-CCAGT GCAGGGTCCGAGGTATT-3'.

Dual-Luciferase Reporter Assay

TargetScan online database was used to predict the downstream gene of miR-21-5p. To confirm the relationship between miR-21-5p and KLF6, the sequences of KLF6 3'-UTR harboring wt-KLF6 or mut-KLF6 were amplified and cloned into the psiCHECK-2 vector (Promega, Madison, WI, USA), respectively. The luciferase reporter plasmids were constructed by Hanbio (Hanbio, Shanghai, China). Hep3B and Huh-7 cells were co-transfected with miR-21-5p mimic or NC mimic and wt-KLF6 or mut-KLF6 plasmids, and the luciferase activity was determined by dual-luciferase reporter kit (Promega). All experiments were repeated at least three times.

RNA Pull-Down Assay

Biotin-labeled negative control (Bio-NC) and KLF6 (Bio-KLF6) were purchased from Sangon (Sangon). Hep3B and Huh-7 cells were cultivated at 37°C overnight and then transfected with Bio-KLF6 or Bio-NC for 8 h. After transfection, cells were washed by PBS buffer and lysed using cell lysis buffer. The Streptavidin-Dyna beads were added into cell lysates to isolate RNA bound to Bio-KLF6. Finally, miR-21-5p enrichment in Hep3B and Huh-7 cells was detected by qRT-PCR.

Xenograft Experiment

BALB/c mice (5 weeks) were randomly divided into three groups ($n=6$ per group). Hep3B cells (2×10^4) were subcutaneously injected into the right-back of the mice. Mice in the drug treatment group were intragastrically injected with paeonol (5 mg/kg or 80 mg/kg) every day.³⁰ Tumor volume was measured every 7 days. After 4 weeks, the mice were sacrificed by cervical dislocation. If spontaneous breathing and blink reflex were not observed within 2–3 min, the mice were considered dead. Then, the excised xenografts were weighed. The xenograft assay was authorized by the Ethical Committee of the Institution for Animal Experimentation and the Animal Welfare Committee of Luqiao Branch of Taizhou Hospital. Animal studies were performed in compliance with the ARRIVE guidelines and the Basel Declaration. All animals received humane care according to the National Institutes of Health (USA) guidelines.

Statistical Analysis

All data were validated by three independent biological replicates and calculated with the means \pm standard deviation (SD). Student's *t*-test for two groups and one-way analysis of variance (ANOVA) for at least three groups were performed to analyze the differences. All data were arranged by the GraphPad Prism 7 (GraphPad Inc., La Jolla, CA, USA). The *P*-value less than 0.05 was considered as statistically significant.

Results

Paeonol Inhibited Cell Viability and Induced Apoptosis in Hep3B and Huh-7 Cells

To investigate the potential role of paeonol in HCC, CCK-8 assay was performed to detect cell viability. After treatment with varying concentrations of paeonol (0 μ M, 50 μ M, 100 μ M, 200 μ M, 400 μ M, 800 μ M), cell viability of Hep3B and Huh-7 cells was substantially decreased in a dose-dependent manner (Figure 1A and B). Moreover, the cell apoptosis rate in Hep3B and Huh-7 cells was gradually increased with the growing concentration of paeonol (Figure 1C–E). In addition, the expression levels of proteins associated with cell proliferation or apoptosis were evaluated by Western blot assay. With the increasing concentration of paeonol, Cyclin D1, CDK4 and Bcl-2 levels were apparently blocked, whereas Bax level was obviously promoted in Hep3B and Huh-7 cells (Figure 1F and G). Taken

together, paeonol could suppress cell viability and enhance apoptosis in Hep3B and Huh-7 cells.

Paeonol Blocked Cell Migration and Invasion of Hep3B and Huh-7 Cells

To further confirm the function of paeonol in HCC, transwell assay was carried out to examine cell migration and invasion. Paeonol treatment remarkably inhibited HCC cell proliferation at 36 h (Supplement Figure 1A and B), so we performed cell migration and invasion assays at 24 h with no significant effect on cell proliferation. As shown in Figure 2A–F, the numbers of migrated and invaded cells were reduced. Besides, the expression of MMP2 and MMP9 was determined by Western blot assay. Compared with the control, the levels of MMP2 and MMP9 were inhibited in Hep3B and Huh-7 cells treated with different concentrations of paeonol (Figure 2G and H). Thus, these findings indicated that cell migration and invasion were suppressed by paeonol in HCC cells.

Paeonol down-regulated miR-21-5p level, and silencing of miR-21-5p suppressed cell viability, migration, invasion and promoted apoptosis in Hep3B and Huh-7 cells. To elucidate the relation between paeonol and miR-21-5p, the expression of miR-21-5p in Hep3B and Huh-7 cells with or without paeonol-treatment was detected by qRT-PCR. As exhibited in Figure 3A, the expression level of miR-21-5p was remarkably reduced in paeonol-treated Hep3B and Huh-7 cells compared with the control group. In addition, after transfection with miR-21-5p inhibitor, miR-21-5p expression was significantly decreased in Hep3B and Huh-7 cells (Figure 3B). Furthermore, CCK-8 assay indicated that cell viability was hindered in Hep3B and Huh-7 cells transfected with miR-21-5p inhibitor (Figure 3C). Cell apoptosis was promoted by miR-21-5p inhibitor (Figure 3D). For cell migration and invasion, the number of migrated and invaded cells in both two cell lines transfected with miR-21-5p inhibitor was lower than that in the NC control group (Figure 3E and F). As shown in Figure 3G and H, the expression of Cyclin D1, CDK4, Bcl-2, MMP2 and MMP9 was down-regulated, while Bax level was increased by miR-21-5p knockdown in Hep3B and Huh-7 cells. Collectively, these results suggested that miR-21-5p was down-regulated by paeonol, and its knockdown impeded cell proliferation, migration, invasion and promoted cell apoptosis in Hep3B and Huh-7 cells.

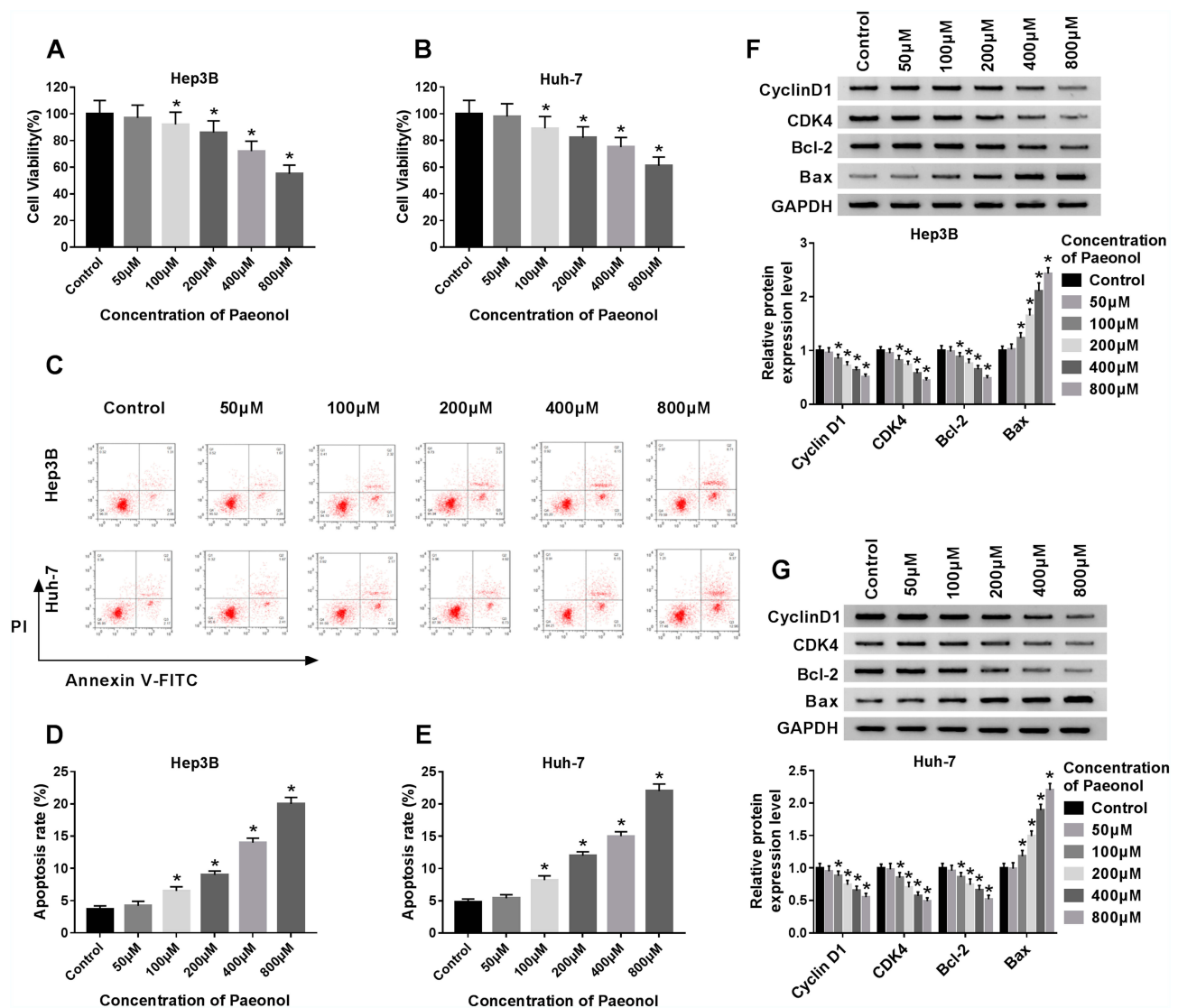


Figure 1 Paeonol inhibited cell viability and induced apoptosis in Hep3B and Huh-7 cells. (A–B) The cell viability of Hep3B and Huh-7 cells was detected by CCK-8 assay after treated with different concentrations of paeonol. (C–E) The apoptosis of Hep3B and Huh-7 cells was measured by flow cytometry assay after treated with varying concentrations of paeonol. (F–G) The expression levels of Cyclin D1, CDK4, Bcl-2 and Bax were determined by Western blot assay in paeonol-treated Hep3B and Huh-7 cells. * $P < 0.05$.

KLF6 Was a Target of miR-21-5p

To clarify the underlying molecular mechanism of miR-21-5p in the progression of Hep3B and Huh-7 cells, TargetScan website tool was used to predict the target of miR-21-5p. The binding sites of miR-21-5p on KLF6 3'-UTR and its mutant are exhibited in Figure 4A. Dual-luciferase reporter assay was conducted to confirm the relationship between miR-21-5p and KLF6. After transfection with miR-21-5p mimic and wt-KLF6, the relative luciferase activity was drastically suppressed in wt-KLF6 group, while there was no significant change of luciferase activity in Hep3B and Huh-7 cells co-transfected with miR-21-5p mimic and mut-KLF6 (Figure 4B and C).

The efficiency of miR-21-5p transfection was also verified (Figure 4D). In addition, RNA pull-down assay revealed that bio-KLF6 significantly enriched miR-21-5p compared to bio-NC control in Hep3B and Huh-7 cells (Figure 4E). Moreover, Western blot analysis implied that miR-21-5p knockdown obviously elevated the expression of KLF6 in comparison with NC control in Hep3B and Huh-7 cells (Figure 4F and G). In a word, miR-21-5p targeted KLF6 and down-regulated the expression of KLF6.

Knockdown of KLF6 relieved the effect of silenced miR-21-5p on cell viability, migration, invasion and apoptosis in Hep3B and Huh-7 cells. The knockdown efficiency of KLF6

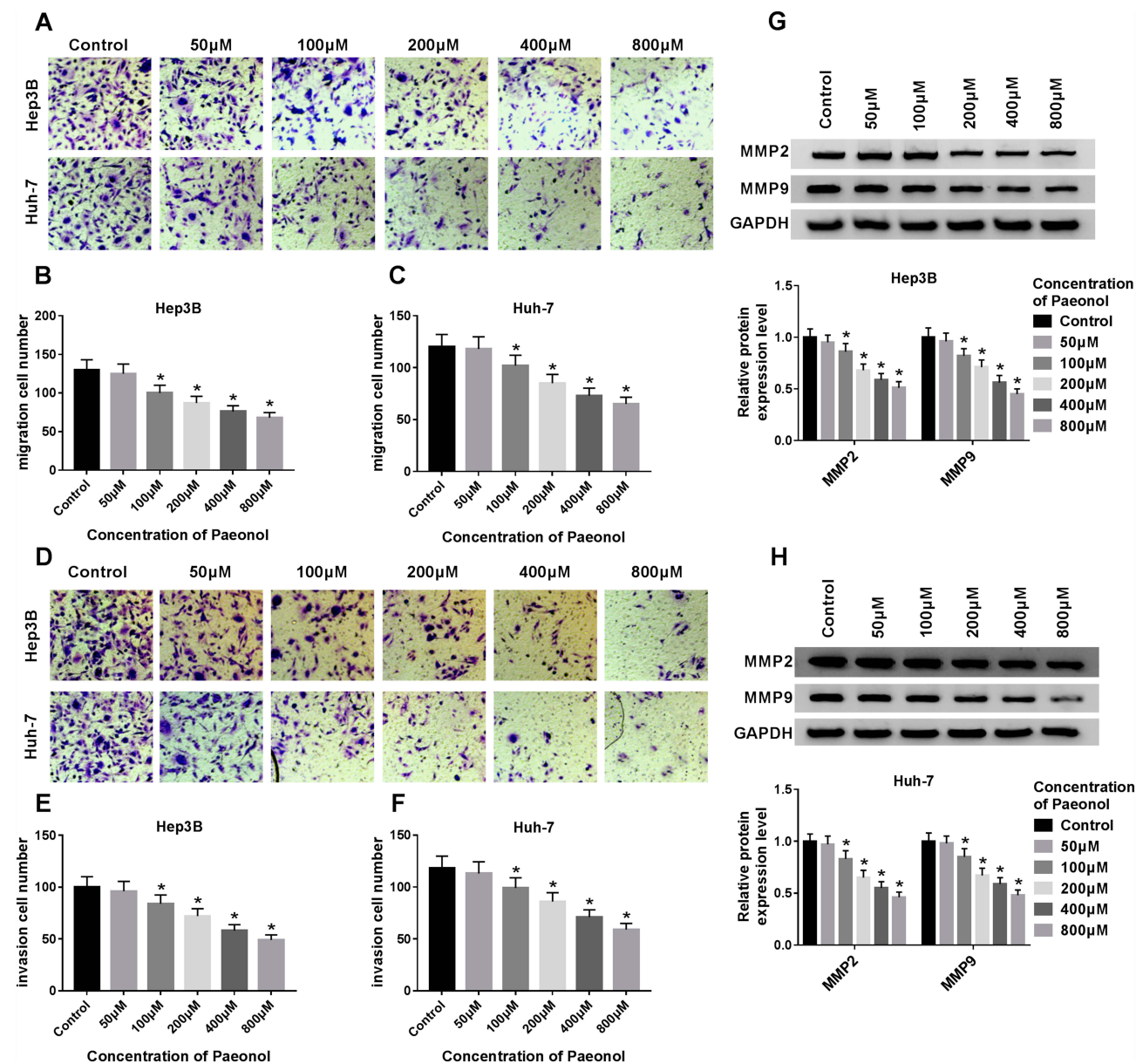


Figure 2 Paeonol suppressed cell migration and invasion in Hep3B and Huh-7 cells. (A–F) Transwell assay was conducted to assess cell migration and invasion in Hep3B and Huh-7 cells treated with various concentrations of paeonol for 24 h. (G and H) Western blot assay was performed to measure the expression of MMP2 and MMP9 in Hep3B and Huh-7 cells after treated with different concentrations of paeonol. * $P < 0.05$.

was detected using Western blot assay. The results suggested that KLF6 expression was evidently reduced in Hep3B and Huh-7 cells transfected with si-KLF6 (Figure 5A). Then, Hep3B and Huh-7 cells were introduced with miR-21-5p inhibitor + si-NC or miR-21-5p inhibitor + si-KLF6 to further investigate the interaction between miR-21-5p and KLF6. After transfection with miR-21-5p inhibitor, the expression of KLF6 was markedly enhanced compared with the NC control group, whereas a distinct reduction of KLF6 expression was detected when cells were co-transfected with miR-21-5p inhibitor and si-KLF6

(Figure 5B and C). As shown in Figure 5D–G, inhibition of miR-21-5p reduced cell viability, migration and invasion and elevated cell apoptosis, while the effects were restored after reintroduction with si-KLF6. Furthermore, Western blot analysis suggested that miR-21-5p inhibitor led to a decrease in the levels of Cyclin D1, CDK4, Bcl-2, MMP2 and MMP9, and an increase in Bax expression, which was recovered by KLF6 depletion (Figure 5H and I). These data implied that the down-regulation of KLF6 could reverse the inhibitory effect of miR-21-5p inhibition on HCC progression.

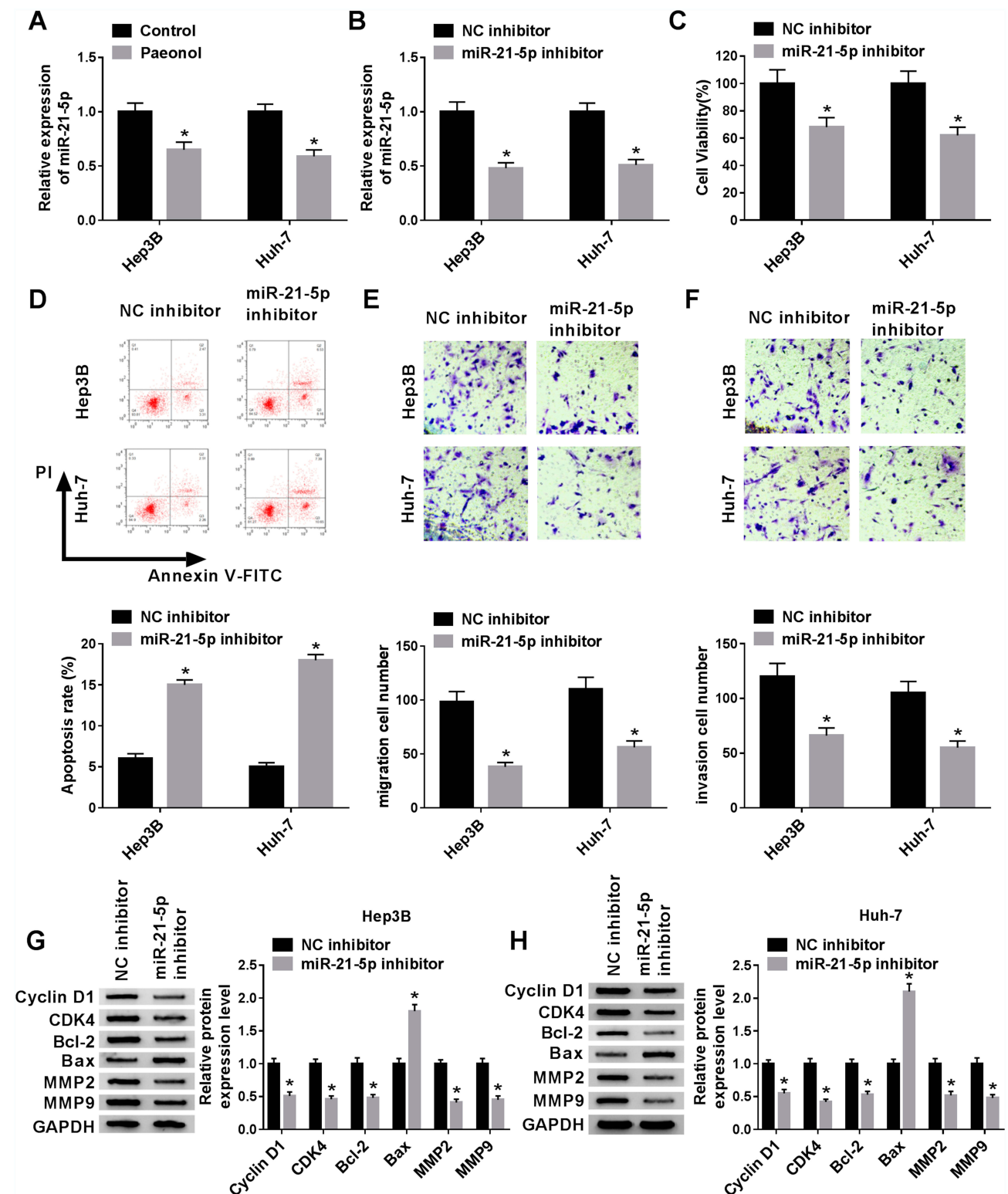


Figure 3 Paeonol down-regulated miR-21-5p level, and silencing of miR-21-5p inhibited cell viability, migration, invasion and promoted apoptosis in Hep3B and Huh-7 cells. (A) The expression of miR-21-5p was measured in Hep3B and Huh-7 cells treated with or without paeonol by qRT-PCR. (B) MiR-21-5p level was detected in Hep3B and Huh-7 cells transfected with NC inhibitor or miR-21-5p inhibitor by qRT-PCR. (C and D) Cell viability and apoptosis of Hep3B and Huh-7 cells were detected by CCK-8 or flow cytometry after transfected with NC inhibitor or miR-21-5p. (E and F) Transwell assay was conducted to assess the cell migration and invasion in transfected Hep3B and Huh-7 cells. (G and H) The expression of Cyclin D1, CDK4, Bcl-2, Bax, MMP2 and MMP9 was measured by Western blot assay in Hep3B and Huh-7 cells transfected with NC inhibitor or miR-21-5p inhibitor. * $P < 0.05$.

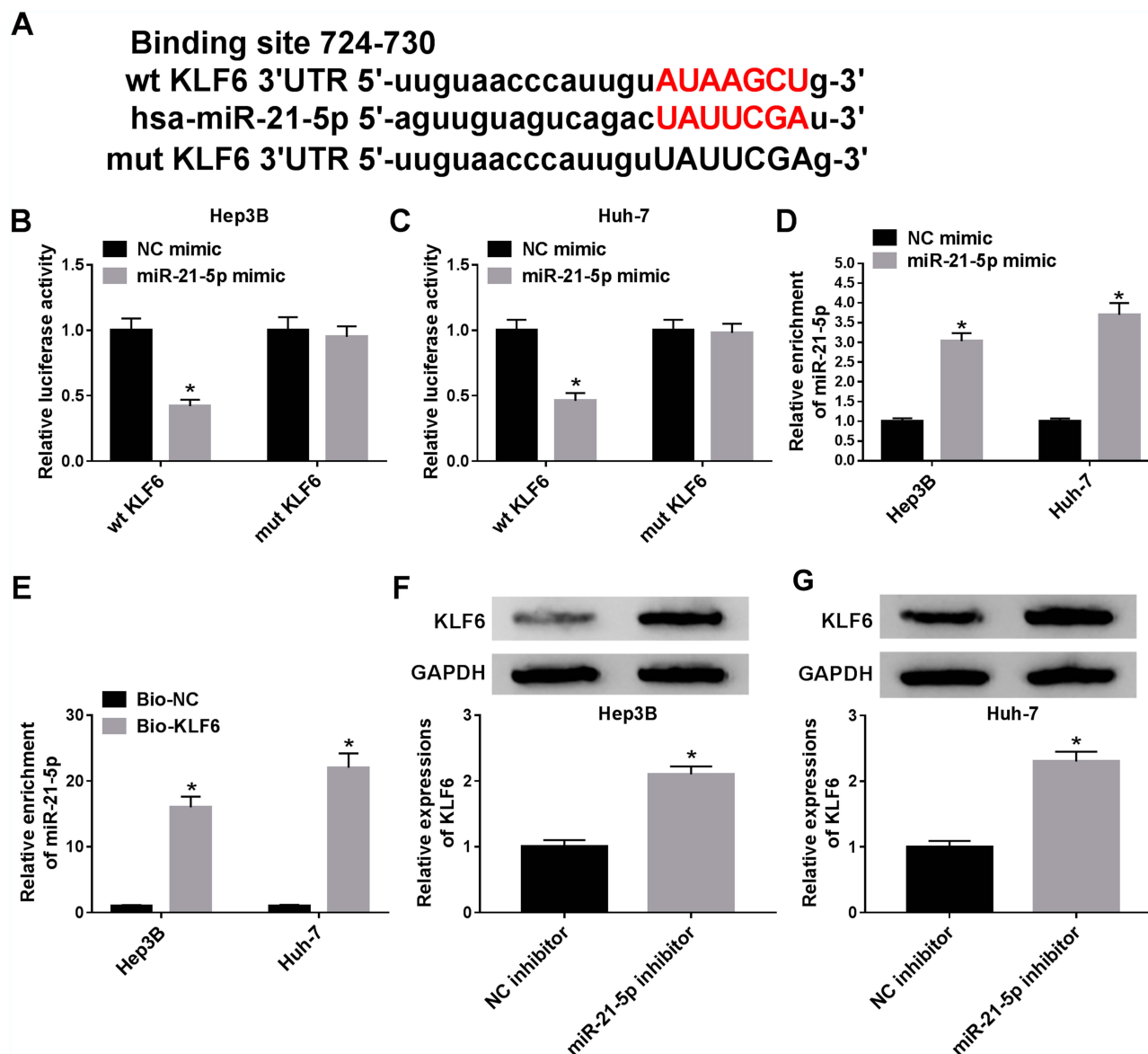


Figure 4 MiR-21-5p targeted KLF6 and down-regulated KLF6 expression. (A) The binding sites of miR-21-5p on KLF6 3'-UTR and its mutant were shown. (B and C) Luciferase activity in Hep3B and Huh-7 cells transfected with NC mimic or miR-21-5p mimic was detected. (D) The level of miR-21-5p was detected by qRT-PCR in Hep3B and Huh-7 cells transfected with miR-NC and miR-21-5p mimic. (E) RNA pull-down was conducted to measure the enrichment of miR-21-5p in Hep3B and Huh-7 cells transfected with Bio-NC or Bio-KLF6. (F and G) The level of KLF6 was detected by Western blot assay in transfected Hep3B and Huh-7 cells. * $P < 0.05$.

Paeonol Reversed the Suppression of miR-21-5p on KLF6 Expression

To explore the function of miR-21-5p/KLF6/paeonol axis, miR-21-5p were transfected into Hep3B and Huh-7 cells treated with or without paeonol, and the expression of KLF6 was measured by Western blot assay (Figure 6A and B). After transfection with miR-21-5p mimic, the level of KLF6 was conspicuously inhibited in Hep3B and Huh-7 cells, whereas paeonol treatment weakened the suppression on KLF6 expression triggered by miR-21-5p up-regulation. In total, paeonol declined

the miR-21-5p-mediated inhibitory effect on KLF6 level in Hep3B and Huh-7 cells.

Paeonol Treatment Impeded the Cell Cycle at the G1 to S Transition

Paeonol treatment inhibited the expression of the cell cycle regulatory proteins (Cyclin D1 and CDK4), so we speculated that paeonol could regulate the cell cycle. Flow cytometry was conducted to detect the cell cycle, and the data indicated that paeonol treatment prominently facilitated cell cycle arrest compared to the control group (Figure 7A and B).

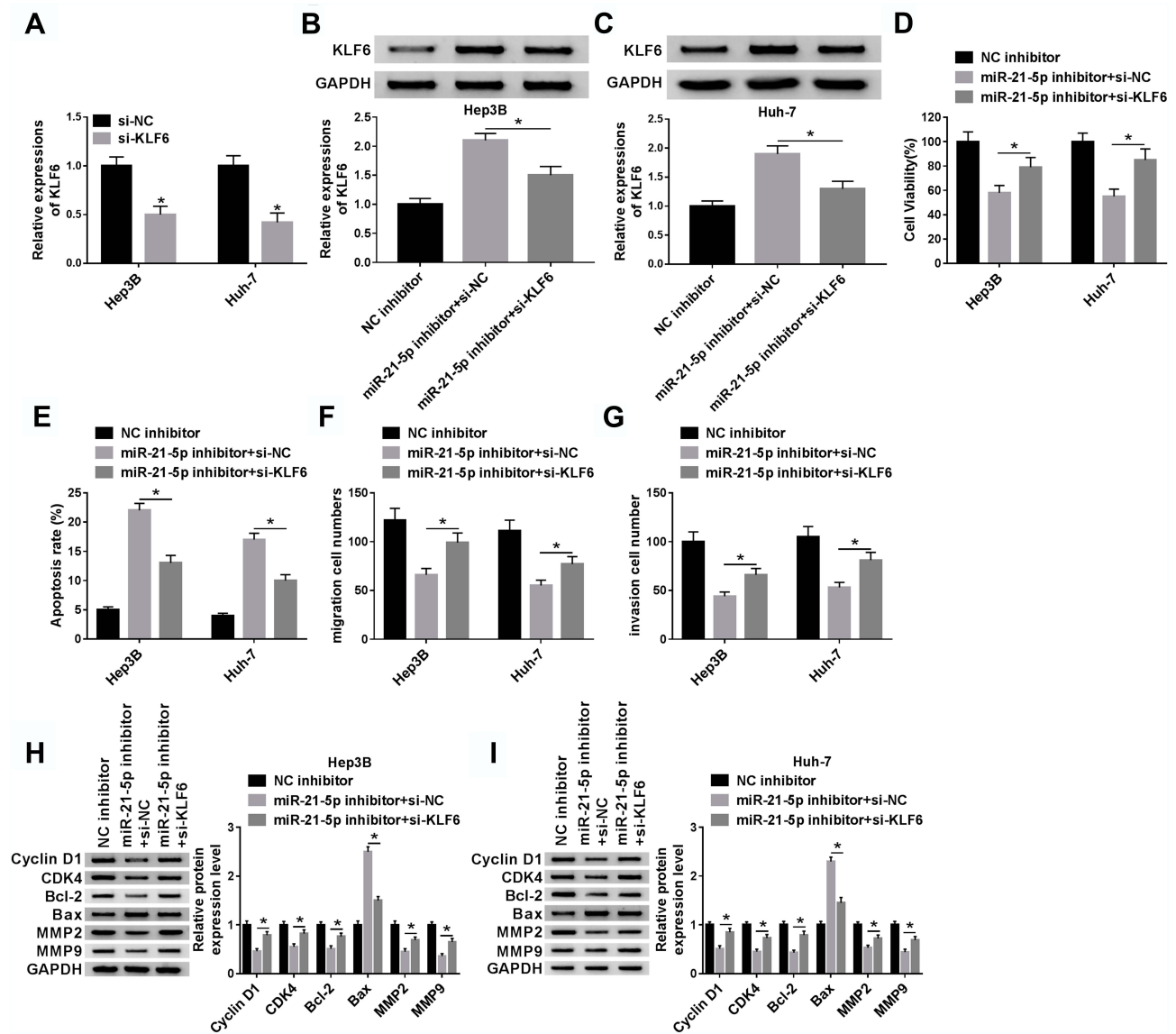


Figure 5 Knockdown of KLF6 relieved the effect of silenced miR-21-5p on cell viability, migration, invasion and apoptosis in Hep3B and Huh-7 cells. **(A)** The knockdown efficiency of KLF6 was determined by Western blot in Hep3B and Huh-7 cells introduced with si-NC or si-KLF6. Hep3B and Huh-7 cells were transfected with NC inhibitor, miR-21-5p + si-NC or miR-21-5p + si-KLF6. **(B and C)** Western blot was carried out to detect the KLF6 expression in transfected Hep3B and Huh-7 cells. **(D and E)** Cell viability and apoptosis were assessed by CCK-8 or flow cytometry, respectively. **(F and G)** Cell migration and invasion were examined by transwell assay in transfected Hep3B and Huh-7 cells. **(H and I)** The levels of Cyclin D1, CDK4, Bcl-2, Bax, MMP2 and MMP9 were determined by Western blot in transfected Hep3B and Huh-7 cells. * $P < 0.05$.

Administration of Paeonol Blocked Tumor Growth in Xenograft Mice

To investigate the role of paeonol in tumorigenesis in vivo, xenograft mice models were established. The results suggested that tumor volume and weight of xenografts treated with 800 μ M paeonol were significantly reduced (Figure 8A and B). In addition, paeonol treatment inhibited the expression of miR-21-5p and promoted the expression of KLF6, confirming the results of in vitro experiments (Figure 8C and D). These data certified that paeonol hindered tumor growth in vivo.

Discussion

HCC is one of the common cancers and the third leading cause of death in the world.³¹ Recent studies revealed that paeonol had been applied in many cancers, including HCC.³² Thus, it is necessary to elucidate the molecular mechanism of paeonol for the treatment of HCC.

In this study, we revealed that paeonol restrained cell viability, migration and invasion, and enhanced cell apoptosis in HCC cells and high concentration of paeonol significantly decreased miR-21-5p expression in HCC

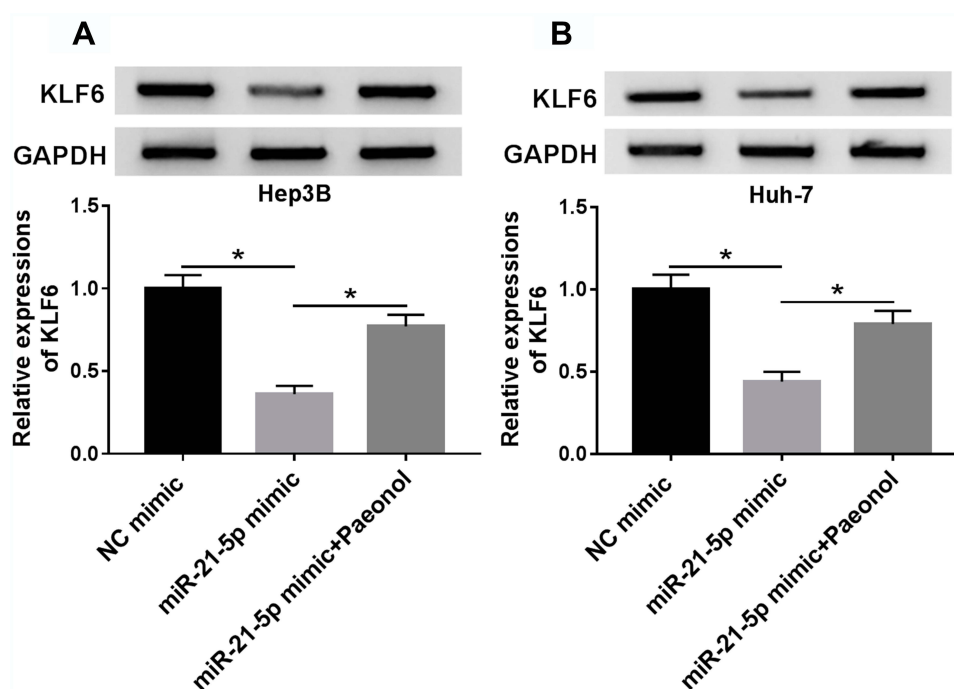


Figure 6 Paeonol attenuated miR-21-5p-mediated inhibitory effect on KLF6 level in Hep3B and Huh-7 cells. **(A and B)** The expression level of KLF6 was assessed by Western blot assay in Hep3B and Huh-7 cells transfected with NC mimic, miR-21-5p mimic or miR-21-5p + paeonol. * $P < 0.05$.

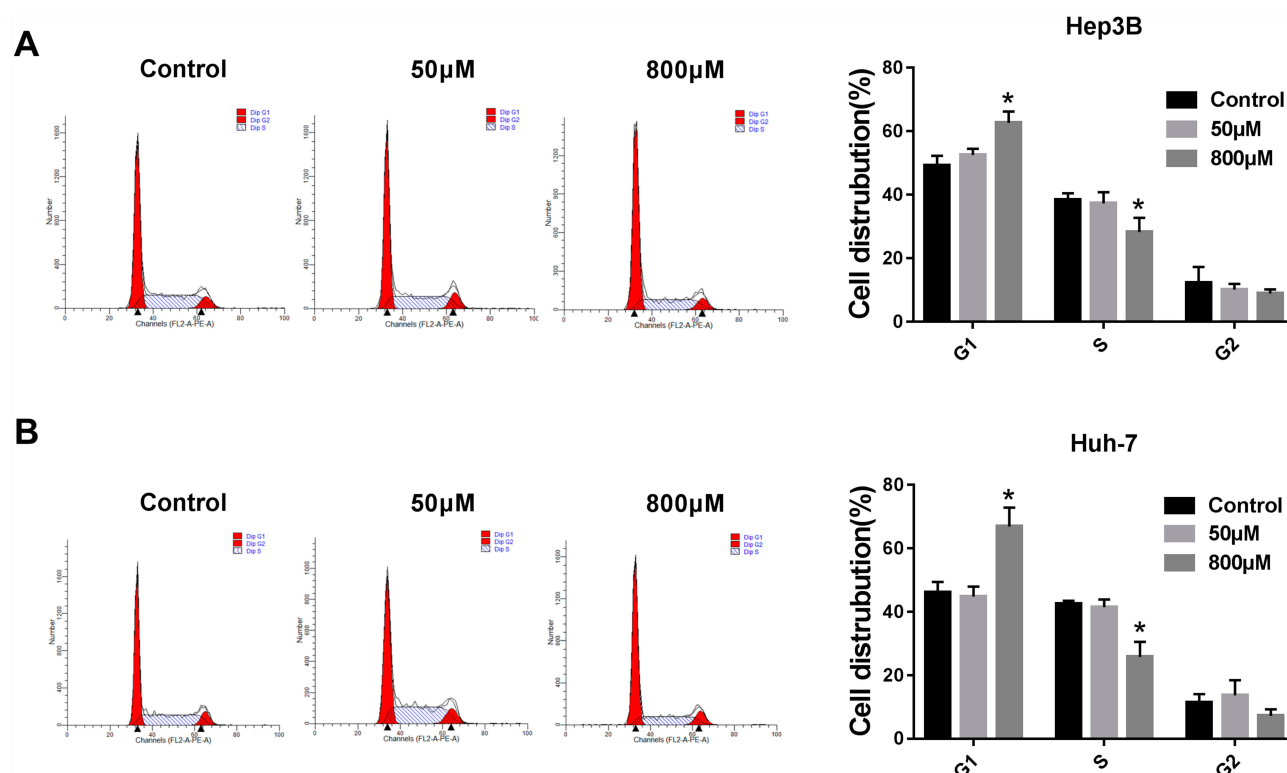


Figure 7 Paeonol treatment impeded the cell cycle at the G1 to S transition. After incubation with paeonol (50 μM or 800 μM), cell cycle of Hep3B and Huh-7 cells was monitored by flow cytometry. **(A and B)** The statistics of cell number in G1 and S stages were shown. * $P < 0.05$.

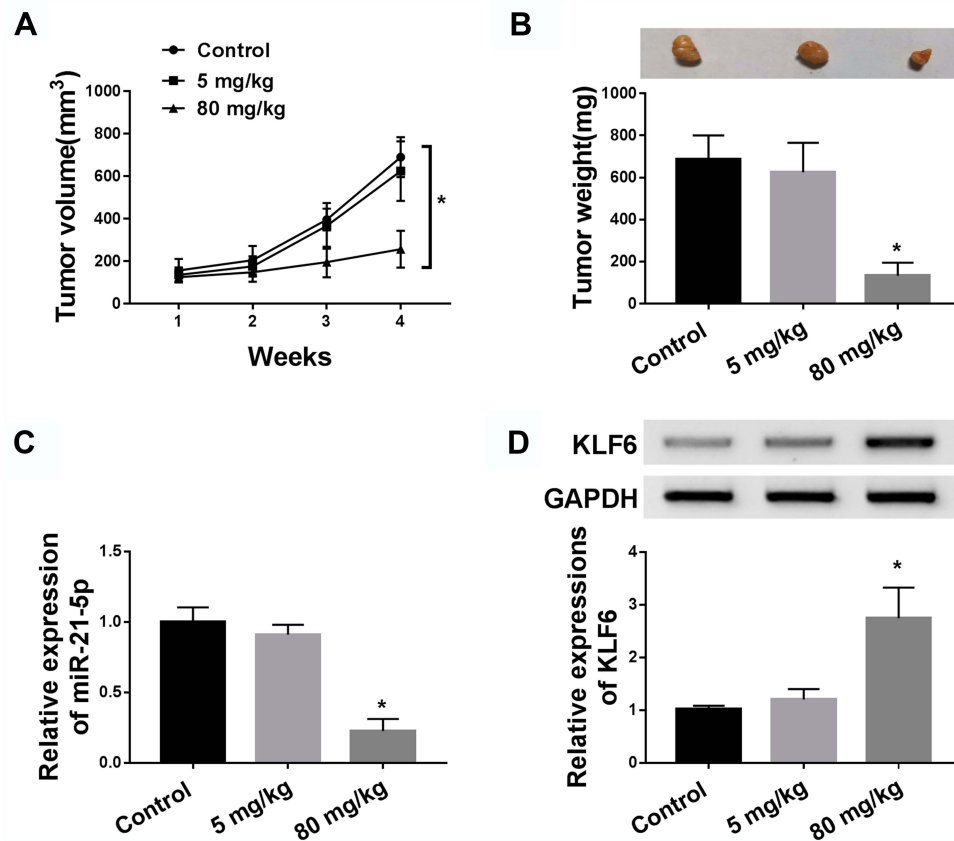


Figure 8 Administration of paeonol blocked tumor growth in xenograft mice. BALB/c mice were subcutaneously injected with Hep3B cells and treated with paeonol (50 μ M or 800 μ M). **(A)** Tumor volume was measured every 7 d. **(B)** Tumor weight was examined after mice were sacrificed. **(C)** The expression of miR-21-5p was determined by qRT-PCR. **(D)** The protein expression of KLF6 was detected by Western blot assay. * $P < 0.05$.

cells. Meanwhile, miR-21-5p targeted KLF6 and repressed the expression of KLF6. Also, KLF6 interference relieved the effect of miR-21-5p knockdown on cell viability, apoptosis, migration and invasion in HCC cells. Moreover, the level of KLF6 was apparently inhibited by miR-21-5p overexpression, which was abated by paeonol introduction. These data implied that paeonol modulated HCC progression via regulating miR-21-5p/KLF6 axis.

Paeonol has attracted the attention of researchers all over the world due to its anti-inflammatory, antioxidant and anti-thrombotic effects. The previous study indicated that the proliferation of typhoid bacillus could be inhibited by paeonol.³³ Also, through studying the HCC xenograft model, paeonol was identified to reduce oxidative injury and facilitate immunity function in vivo.³² Consistent with the above studies, cell viability, migration and invasion of Hep3B and Huh-7 cells were significantly inhibited, and cell apoptosis was apparently promoted by paeonol in a dose-dependent manner. These results indicated that paeonol might play a crucial role in the progression of HCC.

The regulatory effects of miR-21-5p on tumor development have been extensively identified in many cancers. For instance, miR-21-5p enhanced cell proliferation, migration and invasion in NSCLC via targeting SMAD7.³⁴ Also, miR-21-5p was highly expressed in the recurrence group and might be a potential predictor of gastric cancer recurrence.³⁵ Furthermore, increasing studies determined that miR-21-5p was involved in the progression of HCC. Pu et al reported that miR-21-5p was markedly elevated in the serum of HCC patients.³⁶ Bioinformatics analysis revealed that miR-21-5p might be a novel tumor marker for the diagnosis of HCC.²² However, the network of miR-21-5p in HCC was still unknown. In the current study, we assessed the miR-21-5p expression level in paeonol-treated Hep3B and Huh-7 cells. The level of miR-21-5p was inhibited by paeonol, and its knockdown promoted cell viability, migration and invasion, which was consistent with the previous study.²⁶ Also, miR-21-5p was predicted to target KLF6 mRNA and blocked the expression of KLF6. Hence, we hypothesized that paeonol regulated HCC progression via miR-21-5p/KLF6 axis.

KLF6 is a vital transcription factor and regulates numerous physiology processes, such as cell proliferation, differentiation and adipogenesis. Previous studies showed that KLF6 expression was decreased in HCC,²⁹ and its down-regulation could facilitate tumor activity by targeting the promoter of the oncogene PTTG1.³⁷ To further clarify the function of KLF6 in HCC, we co-transfected si-KLF6 and miR-21-5p into Hep3B and Huh-7 cells. The results indicated that the down-regulation of KLF6 reversed the effects of miR-21-5p silencing on cell viability, apoptosis, migration and invasion. Moreover, paeonol enhanced KLF6 level by restoring the miR-21-5p-mediated effect on KLF6 in Hep3B and Huh-7 cells.

Taken together, this study concluded that paeonol promoted cell viability, migration, invasion and blocked apoptosis by regulating miR-21-5p/KLF6 axis in Hep3B and Huh-7 cells, providing a potential treatment strategy for HCC and new evidence to understand paeonol/miR21-5p/KLF6 axis in HCC. In addition, paeonol hindered tumor growth in vivo.

Disclosure

The authors declare that they have no conflict of interests.

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