ARPP-19 Mediates Herceptin Resistance via Regulation of CD44 in Gastric Cancer

Xiang Gao 1,*, Changwen Lu 1,*, Changyu Chen 2, Kang Sun 1, Qixin Liang 1, Jianfeng Shuai 1, Xiaoming Wang 1, Yuxing Xu 1*

1Department of General Surgery, The First Affiliated Hospital of Anhui University of Chinese Medicine, Hefei, People’s Republic of China; 2Department of Gastrointestinal Surgery, The First Affiliated Hospital of Nanchang University, Nanchang, People’s Republic of China

*These authors contributed equally to this work

Purpose: As the first-line drug for treatment of HER2-positive metastatic gastric cancer (GC), Herceptin exhibits significant therapeutic efficacy. However, acquired resistance of Herceptin limits the therapeutic benefit of gastric cancer patients, in which the molecular mechanisms remain to be further determined.

Methods: Quantitative real-time polymerase chain reaction was performed to detect the mRNA levels of ARPP-19 and CD44 in GC cells. Protein levels were determined using Western blot and IHC staining. MTT and soft agar colony formation assays were used to measure cell proliferation. Xenograft model was established to verify the functional role of ARPP-19 in Herceptin resistance in vivo. Sphere formation assay was conducted to determine cell stemness.

Results: We observed ARPP-19 was up-regulated in Herceptin resistance gastric cancer cells NCI-N87-HR and MKN45-HR. The forced expression of ARPP-19 promoted, whereas the silencing of ARPP-19 impaired Herceptin resistance of HER2-positive gastric cancer cells both in vitro and in vivo. Moreover, ARPP-19 significantly enhanced the sphere formation capacity and CD44 expression, CD44 was also a positive factor of Herceptin resistance in HER2-positive gastric cancer cells. In addition, high level of ARPP-19 was positively associated with Herceptin resistance and poor survival rate of gastric cancer patients.

Conclusion: We have demonstrated that ARPP-19 promoted Herceptin resistance of gastric cancer via up-regulation of CD44, our study suggested that ARPP-19 could be a potential diagnostic and therapeutic candidate for HER2-positive gastric cancer.

Keywords: Herceptin resistance, HER2, gastric cancer, ARPP-19, CD44

Introduction

Gastric cancer (GC) is one of the most commonly diagnosed malignancies and contributes to the third leading cancer-specific mortality worldwide. Although notable progress in GC diagnosis and treatment has been achieved, the prognosis of patients with recurrent or metastatic GC remains much unsatisfactory due to the high cancer heterogeneity. Human epidermal growth factor receptor 2 (HER2), whose expression dysregulated in as many as 7–34% of GC, is a 185 kDa transmembrane tyrosine kinase receptor. Herceptin (the only monoclonal humanized antibody targeting at HER2 approved by the FDA) has been demonstrated to significantly improve the clinical outcome of patients with HER2-positive gastric cancer in the ToGA trial. However, most patients developed Herceptin resistance after their one-year initial response.

Increasing studies have reported that the complicated molecular mechanisms of Herceptin resistance in GC, with the involvement of some crucial molecules and signal pathway. Shi et al reported the HER4-YAP1 axis promotes Herceptin...
resistance in HER2-positive gastric cancer by inducing epithelial and mesenchymal transition.\textsuperscript{8} The sensitivity of gastric cancer to Herceptin is regulated by the \textit{mir-223/FBXW7} pathway.\textsuperscript{9} Tang et al demonstrated that \textit{NES1/KLK10} promotes Herceptin resistance via activation of \textit{PI3K/AKT} signaling pathway in gastric cancer.\textsuperscript{10} Nevertheless, the diverse and deep molecular mechanisms of Herceptin resistance in GC are still urgent to be characterized to provide diagnosis and treatment strategies for patients suffering from inevitable resistance.

\textit{ARPP-19} (cAMP-regulated phosphoprotein 19) is a member of the alpha-endsulfine (\textit{ENSA}) family and characterized as a substrate for protein kinase A in mammalian brain.\textsuperscript{11,12} \textit{ARPP-19} is ubiquitously expressed and its related proteins have also been identified in \textit{Caenorhabditis elegans}, \textit{Schistosoma mansoni}, \textit{Drosophila melanogaster} and yeast genomes.\textsuperscript{12,13} In the neuronal system, \textit{ARPP-19} links the nerve growth factor signaling to the post-transcriptional regulation of neuronal gene expression to promote the growth of synapses and synaptic plasticity.\textsuperscript{14} \textit{ARPP-19} has also been reported to play a pivotal role in the pathogenesis of neuronal system diseases, such as Down syndrome and Alzheimer’s disease.\textsuperscript{15} Furthermore, \textit{ARPP-19} has been demonstrated to act as an oncogene in tumorigenesis and progression. For example, it has been reported that \textit{ARPP-19} promotes proliferation and metastasis of human glioma;\textsuperscript{16} \textit{ARPP-19} has also been reported to be targeted by \textit{microRNA-320a} and mediate tamoxifen resistance of breast cancer cells; in addition, increased \textit{ARPP-19} expression is associated with hepatocellular carcinoma.\textsuperscript{17} However, the functions and underlying mechanisms of \textit{ARPP-19} in Herceptin resistance of \textit{HER2}-positive gastric cancer cells have been rarely explored.

In the present study, \textit{ARPP-19} was observed to be remarkably up-regulated in GC cells and tissues with Herceptin resistance. We also demonstrated that \textit{ARPP-19} enhanced Herceptin resistance of \textit{HER2}-positive GC cells both in vitro and in vivo. Besides, \textit{CD44} was positively regulated by \textit{ARPP-19} and silence of \textit{CD44} resensitized Herceptin resistant GC cells to Herceptin. Furthermore, elevated \textit{ARPP-19} predicted poorer overall survival rate of GC patients. Therefore, these results help throw light upon the complicated mechanisms of Herceptin resistance and \textit{ARPP-19} may be used as a new prognosis bio-marker or therapeutic target in GC patients with Herceptin resistance.

Materials and Methods
Clinical Gastric Cancer Specimens
Fifty paraffin-embedded \textit{HER2}-positive (diagnosed as \textit{HER2++} or \textit{HER2+++} by immunohistochemistry in the department of pathology) gastric cancer tissues were collected at Department of General Surgery, the First Affiliated Hospital of Anhui Medical University between 2005 and 2015. The clinicopathological characteristics of these patients without any other diseases or special therapies were recorded according to World Health Organization (WHO) classification system. The follow-up time is from the diagnosis to mortality or the date last known alive. Moreover, all of these 50 patients were received Herceptin-based therapy and their prognosis was collected. Tissues from patients with good prognosis after Herceptin treatment were designated to be Herceptin sensitive, and tissues from patients with poor prognosis or with tumor recurrence shortly were designated to be Herceptin resistant. Informed consent was signed by each patient enrolled. The study protocol about the clinical samples has been reviewed and approved by the ethics committee of the First Affiliated Hospital of Anhui Medical University in accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki).

Cell Lines and Cell Culture
Two human \textit{HER2}-positive GC cell lines (NCI-N87 and MKN45) were purchased from ATCC. All cell lines were cultured in RPMI 1640 (Hyclone) supplemented with 10% fetal bovine serum (FBS, Gibco-BRL; Invitrogen) and incubated at 37°C in a humidified chamber containing 5% CO\textsubscript{2}.

Herceptin resistant GC cell lines (NCI-N87-HR and MKN45-HR) were established as described previously.\textsuperscript{18} Briefly, the parental NCI-N87 and MKN45 cells were treated with increasing doses of Herceptin gradually for 6 months until Herceptin resistance was stably acquired. NCI-N87-HR and MKN45-HR cells were normally maintained with 10 μg/mL Herceptin.

RNA Extraction and qRT-PCR Analysis
Total RNA from cells was isolated by using Trizol (Invitrogen, Carlsbad, CA) and converted to cDNA using RevertAid First Strand cDNA Synthesis Kit (Thermo Scientific Bio) following the manufacturer’s instructions. qRT-PCR analysis was performed by SYBR green Master MIX (Applied Biosystem) as
recommended.\textsuperscript{19,20} GAPDH was used as an internal control to normalized the mRNA expression. Sequences of all the primers used are listed in Table 1.

**Plasmid Constructs and Transfection**

Human ARPP-19 coding sequence transcript (Gene ID: 10776) was cloned and inserted into mammalian expression vector pIRESneo3 (Invitrogen). Lipofectamine 2000 (QIAGEN) was used for plasmid transfection as described previously.\textsuperscript{10,16}

**RNA Oligonucleotides and Transfection**

siRNAs against ARPP-19 and CD44 were designed and synthesized by GenePharma (Shanghai, China). The sequences of siRNAs used in this study are listed in Table 1. siRNAs transfection was performed by using Lipofectamine 2000 (QIAGEN) according to the manufacturer’s instructions as recommended.\textsuperscript{21,22}

**Protein Extraction and Western Blot**

Cells were lysed in modified RIPA buffer and total protein was extracted according to the manufacturer’s instructions. Western blot was conducted as previously described.\textsuperscript{17,23} The primary antibodies against ARPP-19 (11678-1-AP) and CD44 (3570) were purchased from Proteintech and Cell Signaling Technology, respectively. Anti-β-actin antibody (sc-47778, Santa Cruz Biotechnology, USA) was used as a protein loading control.

**Immunohistochemical Staining**

The UltraSensitive-SP kit (Maxin-Bio, Fuzhou, China) was used to conduct immunohistochemistry (IHC) analysis. All steps were performed strictly according to the kit’s instructions. Protein levels of ARPP-19 in the paraffin-embedded HER2-positive gastric cancer tissues were detected by using the rabbit polyclonal antibody against ARPP-19 (1:200, 11678-1-AP, Proteintech, Wuhan, China), and rabbit polyclonal antibody against CD44 (1:200, 15675-1-AP, Proteintech, Wuhan, China) was used to detect CD44. Sections with ≥15% stained cells were considered to be ARPP-19/CD44 high expression, and sections with <15% stained cells were considered to be ARPP-19/CD44 low expression.

**MTT Assays**

Logarithmically growing cells (5.0x10^5 cells/well) were seeded in the 96-well plates and treated with the indicated concentrations of Herceptin for 6 days. MTT reagent (Promega) was added into each well and the absorbance at 570 nm was measured after 2 hours of incubation.

**Soft Agar Colony Formation Assay**

Soft agar colony formation assay was performed as described earlier.\textsuperscript{24,25} In brief, 48h after the indicated transfection, cells were digested and resuspended with 0.3% soft agar in RPMI-1640 containing 10% FBS. Three hundred cells were seeded in the six-well plate per well, and coated with 0.6% solidified agar in RPMI-1640 containing 10% FBS. Media were refreshed every 3 days. About two weeks later, cells were fixed with methanol for 10 min at −20°C and stained with 0.005% crystal violet (Sigma-Aldrich Co., St Louis, MO, USA) for 10 min at room temperature. The colonies were counted for statistical analysis.

**Sphere Formation Assays**

Cells (10^5) suspended in the serum-free DMEM medium containing 0.4% BSA (Sigma), 2% B27 (BD Pharmingen), 5 μg/mL insulin (Sigma), 20 ng/mL basic fibroblast growth factor (bFGF, Invitrogen) and 20 ng/mL epidermal growth factor (EGF, Invitrogen) were plated in the six-well ultra-low attachment plates (Corning). Two weeks later, images of the spheres were taken in three randomly chosen fields with the microscope (Nikon) and NIS-Element F3.0 program (Nikon) and the numbers of spheres in each field were counted for statistical analysis.

**Xenograft and Ki-67 Staining**

For animal experiments, all procedures were reviewed and approved by the Animal Study Committee of the First Affiliated Hospital of Anhui Medical University in Hefei, Anhui Province, China, and in compliance with the
National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals and Animal Welfare Act. MKN45 Vec and MKN45 ARPP-19 cells (5x10^6 per 125μL per site) were injected subcutaneously into the right flanks of 5 week-old nude female mice (12 mice each cells). Mice injected with MKN45 Vec or MKN45 ARPP-19 cells were randomized to receive Herceptin (10 mg/kg) or vehicle as control (6 mice each group) by tail vein injection once a week after the palpable tumors were formed, respectively. Tumor growth was recorded by measuring the length (L) and width (W) and tumor volume was calculated using the formula: Volume (mm3) = L x W^2 x \Pi/6^6.

Mice were sacrificed and tumors were resected after 39 days. Ki-67 staining was performed to assess the tumor cell viability of the harvested tumor tissues and the percentage of Ki-67 positive cell population was calculated.

Statistical Analysis
All the experiments were repeated at least three times. Student’s t-test was used to analyze the difference between the two groups; chi-squared test was used to analyze expression of ARPP-19 in HER2-positive gastric cancer tissues; Kaplan–Meier’s analysis was used for prognostic analyses. All statistical analyses were performed using GraphPad Prism (San Diego, CA) and P<0.05 was considered as statistically significant.

Results
The Expression of ARPP-19 Was Elevated in Human HER2-Positive Gastric Cancer Cells with Herceptin Resistance
To evaluate the expression pattern of ARPP-19 in human HER2-positive gastric cancer cells with Herceptin resistance, we first established two human HER2-positive Herceptin resistant GC cell lines: NCI-N87-HR and MKN45-HR. We found both NCI-N87-HR and MKN45-HR cells exhibited remarkable resistance to Herceptin, compared to their parental NCI-N87 and MKN45 cells (Figure 1A). Subsequently, the expression levels of ARPP-19 in NCI-N87-HR and MKN45-HR cells and their parental cells were determined by qRT-PCR and Western blot. Both the mRNA (Figure 1B) and protein (Figure 1C) levels of ARPP-19 were obviously up-regulated in NCI-N87-HR and MKN45-HR cells compared with NCI-N87 and MKN45 cells, respectively. Thereby, ARPP-19 was over-expressed in GC cells with acquired Herceptin resistance.

Silence of ARPP-19 Sensitized Acquired Herceptin Resistant Gastric Cancer Cells to Herceptin
We further explored the detailed role of ARPP-19 in Herceptin resistance of GC cells. The specific siRNAs against ARPP-19 were transfected into NCI-N87-HR and MKN45-HR cells, siARPP-19 significantly silenced the protein levels of ARPP-19 compared with siNC in NCI-N87-HR and MKN45-HR cells (Figure 2A). Cell viability of NCI-N87-HR and MKN45-HR transfected with siNC showed no significant difference when treated with Herceptin compared with vehicle control. However, depleted ARPP-19 expression led to significantly reduced cell viability with the treatment of Herceptin compared with vehicle control (Figure 2B). The fold change of vehicle-treated group compared with Herceptin treated group was 0.98 (P>0.05) in NCI-N87-HR-siNC cells, 1.38 (P<0.05) in NCI-N87-HR-siARPP-19 cells, 1.04 (P>0.05) in MKN45-HR-siNC cells, and 1.47 (P<0.05) in MKN45-HR-siARPP-19 cells (Figure 2B). Besides, as showed in Figure 2C, siRNA-mediated depletion of ARPP-19 decreased cell colony formation in soft agar on exposure to Herceptin in NCI-N87-HR and MKN45-HR cells, with the fold change of vehicle/Herceptin: NCI-N87-HR-siNC 1.01 (P>0.05), NCI-N87-HR-siARPP-19 1.41 (P<0.05), MKN45-HR-siNC 1.05 (P>0.05) and MKN45-HR-siARPP-19 1.52 (P<0.05). Taken together, these results suggested that silence of ARPP-19 significantly re-sensitized Herceptin resistant cells NCI-N87-HR and MKN45-HR to Herceptin.

ARPP-19 Promoted Resistance to Herceptin of Gastric Cancer Cells Both in vitro and in vivo
Further on, NCI-N87 and MKN45 cells were stably transfected with ARPP-19 overexpression plasmid or the empty vector, the overexpression efficacy was confirmed by Western blot (Figure 3A). Results in Figure 3B and C showed cell viability and cell colony formative capacity in soft agar of NCI-N87 and MKN45 cells increased after transfection with ARPP-19 overexpression plasmid compared with Vector control. Treatment with Herceptin dramatically reduced cell viability and cell colony formative capacity in soft agar of NCI-N87
Vec and MKN45 Vec cells, while NCI-N87 and MKN45 cells with overexpression of ARPP-19 exhibited less significant inhibitory effects on cell viability and cell colony formative capacity in soft agar with treatment of Herceptin (Fold change of vehicle/Herceptin for cell viability: NCI-N87 Vec 2.48 (P<0.05), NCI-N87 ARPP-19 1.81 (P<0.05), MKN45 Vec 2.83 (P<0.05) and MKN45 ARPP-19 1.73 (P<0.05). Fold change of vehicle/Herceptin for cell colony formative capacity in soft agar: NCI-N87 Vec 2.16 (P<0.05), NCI-N87 ARPP-19 1.77 (P<0.05), MKN45 Vec 2.32 (P<0.05) and MKN45 ARPP-19 1.76 (P<0.05)). Therefore, forced expression of ARPP-19 decreased the sensitivity of NCI-N87 and MKN45 cells to Herceptin in vitro.

To determine the role of ARPP-19 in Herceptin sensitivity of gastric cancer in vivo. MKN45 ARPP-19 and MKN45 Vec cells were injected subcutaneously into the right flanks of 5-week-old female BALB/c nude mice. Mice injected with these two group of cells were, respectively, randomized to receive either Herceptin or vehicle as control when the palpable tumors were formed. Tumor sizes were measured every 5 days and tumor growth curves were analyzed at the 40th day. Tumors derived from MKN45 ARPP-19 cells grew much faster, resulting in mean tumor volume and

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**Figure 1** Expression of ARPP-19 was elevated in HER2-positive gastric cancer with acquired Herceptin resistance. (A) MTT assay was performed to detect cell viability of Herceptin resistant cells NCI-N87-HR and MKN45-HR or their parental cells NCI-N87 and MKN45 after the treatment with the indicated concentrations of Herceptin for 6 days. (B) qRT-PCR analysis of ARPP-19 mRNA level in NCI-N87-HR and MKN45-HR cells or their corresponding parental cells. (C) Western blot was conducted to examine the protein level of ARPP-19 in NCI-N87-HR and MKN45-HR cells or their corresponding parental cells. *P<0.05.
weight approximately two-fold of those of tumors derived from MKN45 Vec cells. Treatment with Herceptin decreased both MKN45 ARPP-19 cells-derived and MKN45 Vec cells-derived tumor volume and weight, but MKN45 ARPP-19 cells-derived tumors showed dramatically less decrease on exposure to Herceptin with a growth curve similar with MKN45 Vec cells without the treatment of Herceptin (Figure 4A and B). At day 40, mice were sacrificed and the resected tumors were made into paraffin sections for Ki-67 staining analysis. In consistence with tumor volume and weight, Ki-67 positive cell population in MKN45 ARPP-19 cells-derived tumors was significantly higher than the tumors formed by MKN45 Vec cells. Herceptin reduced Ki-67 positivity in both MKN45 ARPP-19 cells-derived...
and MKN45 Vec cells-derived tumors, whereas tumors formed by MKN45 ARPP-19 cells with the treatment of Herceptin maintained comparative Ki-67 positivity with tumors derived from MKN45 Vec cells in the absence of Herceptin treatment (Figure 4C). Collectively, forced expression of ARPP-19 reduced sensitivity of gastric cancer cells to Herceptin in vivo.

**CD44 Was Regulated by ARPP-19 and Overexpressed in Herceptin Resistant Gastric Cancer Cells**

As reported previously, cancer stem cell (CSC) played a pivotal role in the acquired Herceptin resistance of gastric cancer, and CD44 was an important factor contributed to cancer stem cell-like properties in gastric cancer.
We examined the expression level of \( \text{CD44} \) in Herceptin resistance gastric cancer cells NCI-N87-HR and MKN45-HR with the silence of \( \text{ARPP-19} \), the mRNA (Figure 5A and B) and protein (Figure 5C) levels of \( \text{CD44} \) obviously decreased in NCI-N87-HR and MKN45-HR cells transfected with siARPP-19 compared with siNC (another two cancer-stem-cell-related genes \( \text{SOX2} \) and \( \text{c-MYC} \) were also determined, but they did not change significantly (data not shown)). Besides, the mRNA (Figure 5D and E) and protein (Figure 5F) levels of \( \text{CD44} \) were significantly up-regulated in NCI-N87-HR and MKN45-HR cells compared to their parental cells.

**CD44-Mediated Herceptin Resistance of Gastric Cancer Cells**

To assess the role of \( \text{CD44} \) played in acquired Herceptin resistant gastric cancer cells. We depleted the endogenous \( \text{CD44} \) in NCI-N87-HR and MKN45-HR cells by transfection of specific siRNAs against \( \text{CD44} \), and the knockdown efficacy of \( \text{CD44} \) was verified by Western blot (Figure 6A). Cell viability and cell colony formative capacity in soft agar of NCI-N87-HR and MKN45-HR cells transfected with si\( \text{CD44} \) or siNC and treated with Herceptin or vehicle were determined by MTT assay and soft agar colony formation assay, respectively. Similarly, we found cell viability and soft agar colony formation of NCI-N87-HR and MKN45-HR transfected with siNC showed no significant difference when treated with Herceptin compared with vehicle control. However, depleted \( \text{CD44} \) expression led to significantly reduced cell viability and soft agar colony formation with the treatment of Herceptin compared with vehicle control (Figure 6B and C). Collectively, depletion of \( \text{CD44} \) re-sensitized the Herceptin resistant gastric cancer cells to Herceptin.

**ARPP-19 Promoted Cancer-Stem-Cell-Like Properties in Herceptin Resistant Gastric Cancer Cells**

As \( \text{CD44} \) was an important factor in gastric cancer stem cells and \( \text{ARPP-19} \) positively regulated the expression of \( \text{CD44} \) in Herceptin resistant gastric cancer cells, \( \text{ARPP-19} \) might perform an important role in cancer-stem-cell-like properties of Herceptin resistant gastric cancer cells.

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**Figure 4** Forced expression of \( \text{ARPP-19} \) decreased Herceptin sensitivity of MKN45 cells in xenograft model. (A) Tumor growth curves of MKN45 ARPP-19- or MKN45 Vec-derived tumors of mice with the treatment of Herceptin (10 mg/kg) or vehicle. (B) Tumor weights of each group were measured after mice were sacrificed and tumors were resected at day 40. (C) Ki-67 staining of tumor sections from mice of each group. * \( P < 0.05 \).
Figure 5 CD44 was regulated by ARPP-19 in NCI-N87-HR and MKN45-HR cells. (A and B) mRNA level of CD44 in NCI-N87-HR and MKN45-HR cells transfected with siRNAs against ARPP-19 (siARPP-19) or negative control siRNA (siNC) was determined by qRT-PCR. (C) Protein level of CD44 in NCI-N87-HR and MKN45-HR cells transfected with siRNAs against ARPP-19 (siARPP-19) or negative control siRNA (siNC) was determined by Western blot. (D and E) mRNA level of CD44 in NCI-N87-HR and MKN45-HR cells or their parental NCI-N87 and MKN45 cells was determined by qRT-PCR. (F) Protein level of CD44 in NCI-N87-HR and MKN45-HR cells or their parental NCI-N87 and MKN45 cells was determined by Western blot. *P<0.05.
Concordantly, an enhanced sphere formation capacity was observed in NCI-N87-HR cells as compared to its parental NCI-N87 cells (Figure 7A). Forced expression of \textit{ARPP-19} promoted sphere formation of NCI-N87 cells (Figure 7B), whereas knockdown of \textit{ARPP-19} inhibited sphere formation of NCI-N87-HR cells (Figure 7C). The expression of \textit{ARPP-19} was elevated in NCI-N87-derived sphere as compared to its parental NCI-N87 cells, at both mRNA and protein levels (Figure 7D and E). Taken together, these results suggested \textit{ARPP-19} regulated stemness in Herceptin resistant gastric cancer cells and this might be mediated by \textit{CD44}.

**Figure 6** Depletion of \textit{CD44} reduced Herceptin resistance of NCI-N87-HR and MKN45-HR cells. (A) Protein level of \textit{CD44} in NCI-N87-HR and MKN45-HR cells transfected with siRNAs against \textit{CD44} (siCD44) or negative control siRNA (siNC) was determined by Western blot. (B) NCI-N87-HR and MKN45-HR cells transfected with siRNAs against \textit{CD44} (siCD44) or negative control siRNA (siNC) were treated with Herceptin (10 µg/mL) or vehicle for 6 days; cell viability was determined by MTT assay. (C) NCI-N87-HR and MKN45-HR cells transfected with siRNAs against \textit{CD44} (siCD44) or negative control siRNA (siNC) were treated with Herceptin (10 µg/mL) or vehicle for 6 days; cell colony formation in soft agar was examined. *P<0.05.
ARPP-19 was associated with Herceptin resistance and poor survival of HER2-positive gastric cancer patients

In further, 50 paraffin-embedded HER2-positive gastric cancer tissues were collected and protein levels of ARPP-19 and CD44 in these HER2-positive gastric cancer tissues were examined by IHC staining. ARPP-19 and CD44 were both up-regulated in gastric cancer tissues with Herceptin resistance as compared with Herceptin-sensitive gastric cancer tissues (Figure 8A). Elevated ARPP-19 was significantly positively associated with Herceptin resistance of gastric cancer.
patients (Figure 8B). Besides, patients with high ARPP-19 levels showed lower overall survival rate ($P=0.024$) compared with patients with low ARPP-19 levels (Figure 8C). Moreover, Figure 8D shows that ARPP-19 and CD44 were correlated in these 50 HER2-positive gastric cancer tissues. Taken together, elevated expression of ARPP-19 in HER2-positive gastric cancer predicted poor prognosis.

**Discussion**

In the current study, we comprehensively investigated the expression pattern, function and underlying mechanisms of ARPP-19 in Herceptin resistance of human HER2-positive gastric cancer. Expression levels of ARPP-19 were elevated in HER2-positive gastric cancer cells and tissues with acquired Herceptin resistance, and prognosis study showed that patients with a higher expression level
of ARPP-19 had a shorter overall survival rate. Functionally, silence of ARPP-19 decreased, while the forced expression of ARPP-19 promoted the Herceptin resistance of HER2-positive gastric cancer cells in vitro. Furthermore, overexpression of ARPP-19 increased the Herceptin resistance of MKN45 cells in xenograft model in vivo. Mechanically, ARPP-19 promoted the cancer stem properties by up-regulation of CD44, a famous cancer stem cell marker in gastric cancer. In addition, depletion of CD44 re-sensitized the Herceptin resistant gastric cancer cells to Herceptin, which indicated CD44 was also a positive regulator of Herceptin resistance in HER2-positive gastric cancer. Thereby, this study expanded our understanding of ARPP-19 in HER2-positive gastric cancer with Herceptin resistance.

Recent studies reported that the expression of ARPP-19 was deregulated in hepatocellular carcinoma. Besides, ARPP-19 has also been reported to contribute to tumorigenesis and progression of glioma and acute myeloid leukemia. Moreover, it was also reported that ARPP-19, which was negatively regulated by miR-320a, promoted tamoxifen resistance of breast cancer cells. However, the functions and underlying mechanisms of ARPP-19 in gastric cancer and Herceptin resistance were poorly understood.

Tumors were heterogeneous and a small proportion of cells with stem cell features in tumor tissues was called cancer stem cell (CSCs), which was considered to contribute to tumor initiation. CSCs were characterized with self-renewal, resistance to treatment and high metastasis potential, all of which make them the source of cancer relapse and treatment failure. Especially, CSCs played crucial roles in cancers with acquired Herceptin resistance. We herein demonstrated that CD44 was regulated by ARPP-19 and mediated Herceptin resistance of human HER2-positive gastric cancer cells. Plenty of literatures demonstrated that CD44 played oncogenic roles in various human cancers, gastric cancer included. Furthermore, CD44 was a famous mediator to regulate cancer stem cell-like properties. As reported previously, lsohaptogenin (ISO) inhibits stem cell-like properties and invasion of bladder cancer cell by attenuating CD44 expression. EGCG inhibits CSC-like properties through targeting miR-485/CD44 axis in A549-cisplatin-resistant cells. LncRNA DGCR5 contributes to CSC-like properties via modulating miR-330-5p/CD44 in NSCLC. In this study, we demonstrated ARPP-19 up-regulated the expression of CD44 and promoted sphere formation, which indicated the enhancement of cancer stem cell-like properties in gastric cancer cells. Therefore, we concluded ARPP-19 promoted Herceptin resistance of HER2-positive gastric cancer through regulation of CD44-mediated cancer stem cell-like properties. However, it still needs further study to clarify the specific molecular mechanisms on regulation of CD44 by ARPP-19.

In conclusion, we herein verified the functional roles and underlying molecular mechanisms of ARPP-19 in Herceptin resistance of human HER2-positive gastric cancer cells, and revealed the expression patterns and prognostic correlation of ARPP-19 in HER2-positive gastric cancer patients. ARPP-19 might thereby be a potential novel therapeutic target and diagnostic biomarker in human HER2-positive gastric cancer.

Disclosure
The authors declare that they have no conflicts of interest.

References


