

Lnc HAGLR Promotes Colon Cancer Progression Through Sponging miR-185-5p and Activating CDK4 and CDK6 in vitro and in vivo

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Background/Aim: LncRNA plays a key role in tumor progression. HAGLR functions as an oncogene in many cancers. However, the molecular mechanism of HAGLR in colon cancer is still unclear.

Methods: qRT-PCR was used to measure the expression of HAGLR, miR-185-5p in colon cancer. The expression of CDK4 and CDK6 was detected by Western blot. CCK-8 assay, EdU staining, transwell and Annexin V-FITC/PI assay were used to analyze the effect of HAGLR and miR-185-5p on cell proliferation, invasion, migration and apoptosis. Bioinformatic analysis and luciferase were used to analyze the target genes of HAGLR and miR-185-5p. Nude mice were used to detect mouse tumor changes.

Results: Compared with normal colon cancer tissues and cells, the expression of HAGLR was increased in colon cancer tissues and cells. In addition, the expression of HAGLR down-regulation inhibited the growth, migration, and invasion of colon cancer cells. MiR-185-5p was reduced in colon cancer, and CDK4 and CDK6 acted as target genes of miR-185-5p to regulate the progress of colon cancer. And CDK4 and CDK6 were predicted as downstream targets of miR-185-5p. Finally, it was demonstrated that HAGLR regulated tumor progression in vivo.

Conclusion: Lnc HAGLR promoted the development of colon cancer by miR-185-5p/CDK4/CDK6 axis, and lnc HAGLR might be potential target for colon cancer.

Keywords: lnc HAGLR, miR-185-5p, CDK4/CDK6, colon cancer, proliferation, apoptosis

Introduction

Colon cancer is a frequently-occurring disease in the clinic.^{1,2} New cases of colon cancer have increased. It is worth noting that the incidence of colon cancer varies not only globally, but also its development trends in different countries and regions.³ The pathogenesis of colon cancer is not fully understood, and it is currently considered to be a combination of environmental factors and genetic factors.⁴ The main factors affecting the incidence of colon cancer include environmental factors, intestinal homeostasis, dietary structure, tobacco and alcohol habits, and physical exercise.^{5,6} At present, the principles for the treatment of colon cancer tumors are mainly comprehensive treatment based on surgical resection, chemotherapy, radiation therapy, and targeted therapy.^{7,8} From the perspective of the effect of treatment, individualized differences between patients with colon cancer tumors are obvious. The above treatments have obvious deficiencies and poor overall efficacy, which can only partially relieve clinical symptoms and have a poor prognosis.^{9,10}

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Therefore, finding new markers for early diagnosis of colon cancer is a hot topic of current research.

In the human genome sequence, during the transcription process, most products are non-coding transcripts. The length of long-chain non-coding RNA (lnc RNA) is generally greater than 200 nt.^{11,12} The regulation of lncRNA in cancer has been sought by many researchers.^{13,14} lncRNA is closely related to various tumor behaviors of malignant tumors, such as tumor autophagy, tumor resistance and tumor immunity.^{15,16} At present, it has been found that lncRNAs are dysregulated in colon cancer, and they can play a regulatory role in the tumor formation process, thereby affecting the occurrence, metastasis and prognosis of colon cancer.^{17,18} lncRNA HAGLR is a recently discovered lncRNA, which has been found abnormally expressed in many malignant tumors.¹⁹ For example, the study found that HAGLR in NSCLC was increased.²⁰ But the mechanism of lncRNA HAGLR in colon cancer have not been studied.

The regulatory relationship of lncRNA-miRNAs has attracted the attention, and the interactive regulation of lncRNAs and miRNAs is the current research focus.²¹ MicroRNAs (miRNAs) are a single species derived from the organism's own genome.²² miRNAs can play a carcinogenic role and promote the development of tumor cells. It can act as a proto-oncogene to inhibit tumor growth.^{23,24} In solid cancer patients, the expression of miRNAs in tumor tissue are dysregulated, which are related to tumorigenesis, invasion, metastasis, judgment of prognosis, and drug resistance.^{25,26} miRNAs have received more attention in the occurrence of colon cancer.^{27,28} miR-185-5p has been confirmed to be abnormally expressed in many tumor tissues.²⁹ But its function in colon cancer is currently unknown. Cyclin-dependent kinases (CDKs) are the core part of the cell cycle regulation mechanism, and considered as important biomarkers for treatment of malignant tumors.³⁰ CDK4 and CDK6 have been poorly studied in colon cancer.^{31,32} Therefore, it was speculated that lncRNA HAGLR can control the progression of colon cancer by CDK4 and CDK6 through miR-185-5p. The purpose of this study was to explore the mechanism of lncRNA HAGLR regulation of colon cancer.

Materials and Methods

Tissue Sample

The study was approved by the ethics committee of the First Affiliated Hospital of Jinzhou Medical University.

A total of 25 colon cancer tissues and adjacent normal tissues were obtained from patients undergoing colon cancer surgery in our hospital. All patients have not received any radiation therapy or chemotherapy. Each patient signed an informed consent form.

Cell Culture

HCT-116, HCT-8, HT-29, DLD-1, SW620, and FHC cells were obtained from ATCC (Manassas, Virginia). The cells were cultured in RPMI 1640 medium containing 10% FBS (GIBCO, Carlsbad, CA) at 37 ° C in a cell incubator containing 5% CO₂.

Cell Transfection

Lipofectamine 2000 reagent (Invitrogen) was used to transfect miR-185-5p mimic, inhibitor or its negative control into cells. A cDNA of human HAGLR was amplified from the mRNA of colon cancer cells. The sequence was designed using shRNA-luciferase (shLuc) as a negative control. The target product was subcloned into pcDNA3.1 (Invitrogen, Carlsbad, CA) and the lentiviral packaging vector pMD2.G was used to verify transfection by qRT-PCR.

Cell Proliferation Assay

SW480 cells were seeded in a 96-well plate at a density of 5,000 cells per well. 100 μ L CCK-8 solution (Liji, Shanghai, China) was added. And absorbance was measured using a microplate reader (Potenov, Beijing, China) at 450nm.

Apoptosis Analysis

SW480 cells were plated into 6-well plates at a density of 5×10^5 cells/well, and the cells were collected and counted. The cells were centrifuged and 195 μ L of Annexin V-FITC binding solution was added. After mixed with 5 μ L Annexin V-FITC and 10 μ L propidium iodide staining solution, the cells were incubated for 10–20 min, and then placed in an ice bath.

Transwell Assay

Transwell assays was used to detect cell invasion and metastasis. The upper basement membrane of the transwell chamber was pre-coated with 20 μ g Matrigel and cultured overnight in a 24-well plate. After 12 hours of culture, it was washed for 3 times the PBS, fixed with 90% formaldehyde, and then stained in the crystal violet solution for 15 min. Then the photograph was taken under an inverted

microscope. In the cell migration experiment, the upper chamber of the transwell chamber was free of matrigel coating, and the rest of the operation was the same as the invasion experiment.

EdU Analysis

After the SW480 cells were treated, the 10 μ mol/L EdU was added to the culture medium and incubated in the incubator for 24 h. Then, 1 mL 0.5% triaxone and 0.5 mL of reaction solution were added, 0.5 mL of Hoechst 33342 solution was added for 30 min, and the observation and counting were performed under inverted fluorescence microscope.

Nude Mouse Tumor Xenograft Model

Male athymic BALB/c nude mice were purchased from the National Laboratory Animal Center (Beijing, China). Each mouse was injected subcutaneously with HCT-116 cells to establish a mouse xenograft model. A total of 5×10^6 shRNAHAGLR-HCT-116 and NC-HCT-116 cells were suspended in 100 μ L phosphate buffer and injected subcutaneously into nude mice. Five weeks later, mice were euthanized. The study has been approved by the Ethics Committee of Beijing Friendship Hospital and is based on the the National Institutes of Health's Guide to the Care and Use of Laboratory Animals.

RNA Pull-Down Assay

RNA immunoprecipitation assay was performed using EZMagna RIP kit (Millipore). Cells were lysed using complete RIP lysis buffer. Cell extracts were incubated with magnetic beads conjugated with anti AGO2 or anti IgG antibodies for 6 h. Then the RNA was purified by RT-qPCR analysis.

RNA Extraction and qRT-PCR

Total RNA in tissues and cells was extracted by TRIzol reagent (APPLYGEN, Beijing, China). And qRT-PCR was performed using ViiATM 7 real-time PCR system (Life Technologies, Grand Island, NY). 2- $\Delta\Delta$ Ct Method was used to calculate the relative gene expression level. qRT-PCR methods were performed by the reference.³³ β -actin was used as the internal control of HAGLR. U6 was used as the internal control of miR-185-5p. The primer sequence was as follows:

HAGLR

forward: 5'-GGCTCTTCCCTAATGTGTGG-3',

reverse: 5'-CAGGTCCAGCATGAAACAGA-3';

β -actin

forward 5'-CGCTCTCTGGTCCTCCTGTTC-3',

reverse: 5'-ATCCGTTGGCTCCGACCTTCAC-3';

miR-185-5p

forward: 5'-TGCGGTGGAGAGAAAGGCAG-3',

reverse: 5'-CTGCCTTTCTCTCCTCCGCA-3';

U6

forward: 5'-TGCGGGTGCTCGCTTCGGCAGC-3',

reverse: 5'-TGCGGGTGCTCGCTTCGGCAGC-3';

Determination of Luciferase Activity

Wild type (WT) or mutant (MUT) HAGLR binding miR-185-5p was subcloned into the pGL3 Basic vector. MiR-185-5p (RiboBio, Guangzhou, China) with 10 μ g pLUC-WT-HAGLR or pLUC-MUT- HAGLR was co-transfected into HCT-116 cells. Wild-type (WT) or mutant (MUT) CDK4 and CDK6 combined with miR-185-5p were subcloned into pGL3 base vector (Promega). MiR-185-5p (RiboBio (Guangzhou, China) was co-transfected with 10 μ g pLUC-WT-CDK4 and pLUC-WT-CDK6 or pLUC-MUT-CDK4 and pLUC-MUT-CDK6. After 48 h transfection, a dual luciferase assay system (Promega Corporation, Fitchburg, WI, USA) was used for luciferase activity.

Immunohistochemistry

Colon tissue were incubated with a primary antibody (Abcam) against Ki-67. Envision™ ABC kit (Gene-Tech Co.,Ltd., Shanghai, China) was used for immunoassay. Leica DM4000B/M microscope (Leica Microsystems, Inc., Buffalo Grove, IL) was used for analysis.

Western Blot

The transfected cells were collected, total protein was extracted, and the protein concentration was quantified using the BCA Protein Assay Kit. It was incubated with anti-CDK4 (1: 1000, Xinyu, Shanghai, China) and anti-CDK6 (1: 1000, Xinyu, Shanghai, China) and anti-GAPDH antibody (1: 1000, Xinyu, Shanghai, China) at 4 °C. Then it was incubated with anti-rabbit secondary antibody (1: 1000, Cell Signaling Technology, Boston, MA, USA) for 1 h.³⁴

Statistical Method

The monitoring data were analyzed by SPSS19.0 statistical software. The results of data analysis were represented as mean \pm standard deviation (mean \pm SD). Multigroup data analysis was founded on one-way ANOVA. LSD test was

leveraged for subsequent analysis. $P < 0.05$ indicated the difference was significant.

Results

HAGLR Was Upregulated and miR-185-5p Was Downregulated in Colon Cancer Tissues and Cell Lines

The results were shown in [Figure 1A](#). Compared with adjacent normal tissues, the expression levels HAGLR in colon cancer tissues were significantly increased ($P < 0.05$), while the expression levels of miR-185-5p were significantly reduced ($P < 0.05$) ([Figure 1B](#)). There was a significant negative correlation between the expression levels of HAGLR and miR-185-5p in colon cancer tissues ([Figure 1C](#)). Compared with the normal colonic epithelial cell line FHC, the expression level of HAGLR in the colon cancer cell line (DLD-1, SW620, HCT-116, LOVO, SW480) was significantly increased ($P < 0.05$, [Figure 1D](#)), and the expression level of miR-185-5p was also

significantly reduced ($P < 0.05$, [Figure 1E](#)). Because of the highest expression level of HCT-116 in SW480 cells, SW480 cells were selected for follow-up test.

HAGLR Expression Regulated Proliferation and Apoptosis in CC Cells

As shown in [Figure 2A](#), in the SW480 cells, the expression level of HAGLR in the shHAGLR1 group and the shHAGLR2 group was significantly reduced, while the expression level of HAGLR in the pc HAGLR1 group was significantly increased, indicating that the transfection was successful ($P < 0.05$). As shown in [Figure 2B](#) and [C](#), compared with the sh-NC group, the cell viability in the shHAGLR1 group and the shHAGLR2 group was significantly reduced, and the positive rate of EdU staining was significantly reduced ($P < 0.05$). As shown in [Figure 2D](#), Tanswell results showed that compared with sh-NC group, the cell migration and invasion of the shHAGLR1 and the shHAGLR2 group were significantly reduced ($P < 0.05$).

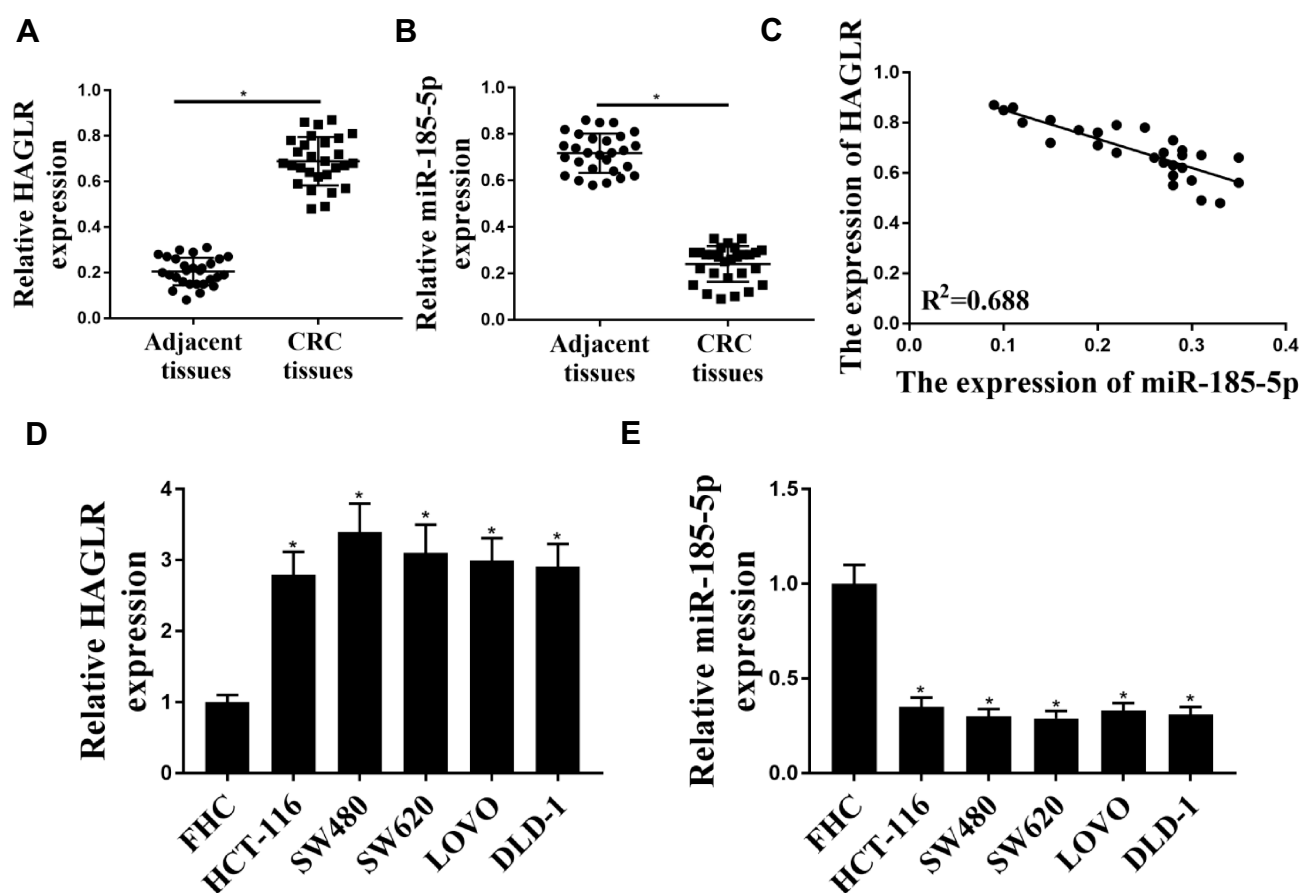


Figure 1 Expression levels of HAGLR and miR-185-5p in colon cancer tissues and cell lines. (**A** and **B**) HAGLR and normal colon tissue ($n = 28$) and colon cancer tissue ($n = 28$) miR-185-5p expression level. (**C**) Relationship between HAGLR and miR-185-5p in colon cancer tissues. (**D** and **E**) Expression level of HAGLR and miR-185-5p in colon cancer cell lines and normal cells (FHC). * $P < 0.05$, $n = 3$.

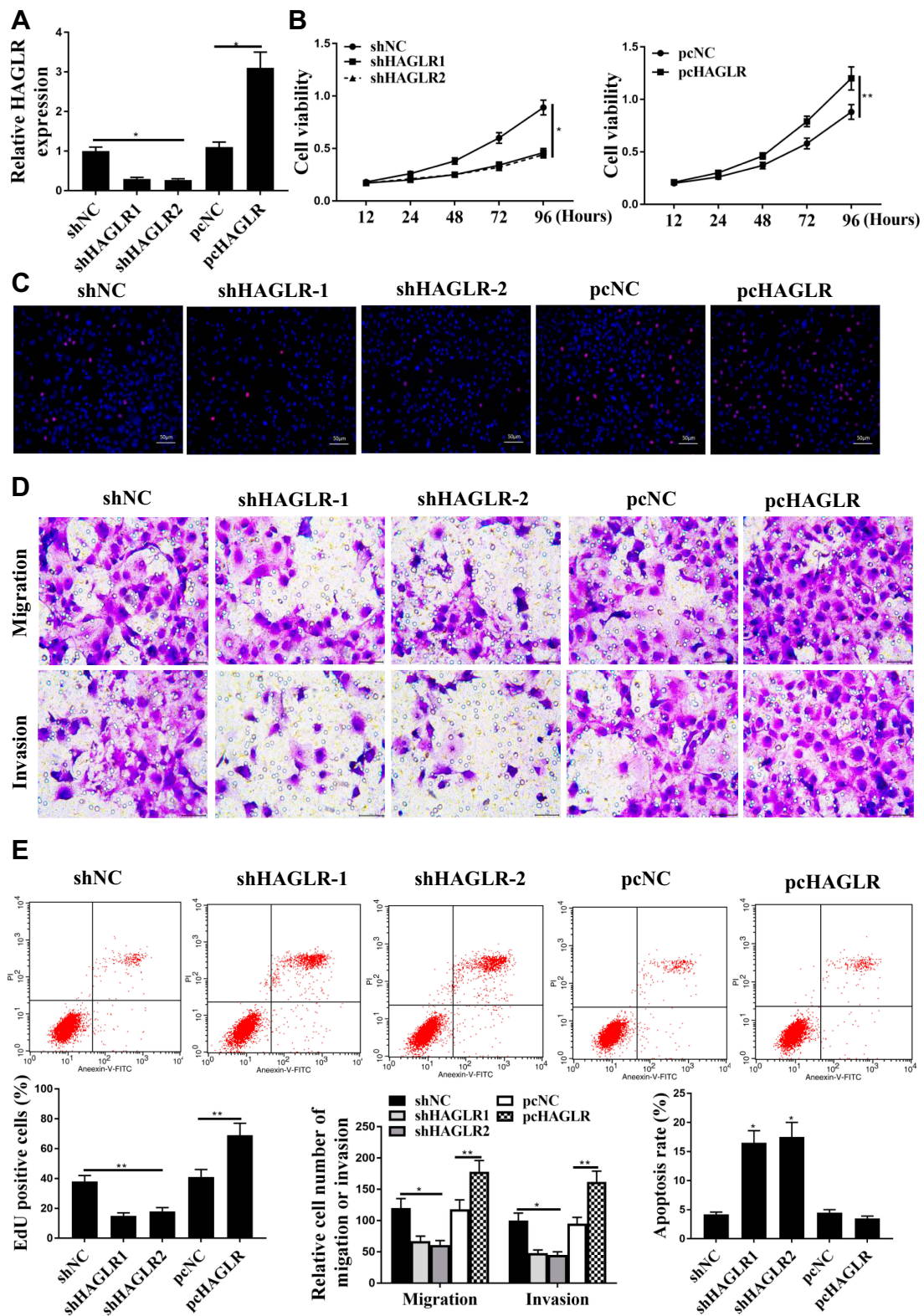


Figure 2 HAGLR regulated colon cancer progression in vitro. **(A)** HAGLR expression level in SW480 cells. **(B)** CCK-8 assay was used to detect for cell viability. **(C)** EdU staining was used to detect for cell proliferation. **(D)** Transwell assay was used to detect for cell migration and cell invasion. **(E)** Apoptosis was determined by flow cytometry. * $P < 0.05$, ** $P < 0.01$, $n = 3$. pcHAGLR: high expression of HAGLR.

As shown in [Figure 2E](#), the shHAGLR1 group and the shHAGLR2 group could significantly induce apoptosis ($P < 0.05$). The pcHAGLR2 result was the opposite of the shHAGLR1 result.

miR-185-5p Overexpression Inhibited Proliferation and Induced Apoptosis in CC Cells

As shown in [Figure 3A](#), compared with miR-NC group, the expression level of miR-185-5p in miR-185-5p mimic group was significantly increased in SW480 cells, indicating successful transfection ($P < 0.05$). The cell viability was significantly reduced in the miR-185-5p mimic group, and the positive rate of EdU staining was significantly reduced ($P < 0.05$, [Figure 3B](#) and [C](#)). Tanswell results showed that miR-185-5p mimic was significantly reduced cell migration and invasion ($P < 0.05$, [Figure 3D](#)). MiR-185-5p mimic was significantly induced apoptosis ($P < 0.05$, [Figure 3E](#)).

HAGLR Acted as a Sponge for miR-185-5p in CC Cells

We used the Starbase 2.0 online prediction tool and miR-185-5p was identified as a potential target for HAGLR ([Figure 4A](#)). The WT-HAGLR or mutant (Mut) -HAGLR luciferase reporter plasmid was used for luciferin enzyme reporter assay. The results showed that the luciferase activity of pGL3-REPOR-HAGLR-WT was reduced by miR-185-5p mimics ($P < 0.05$), but the luciferase activity of pGL3-REPOR-HAGLR-Mut did not change significantly ([Figure 4B](#)). In addition, the expression level of HAGLR using biotinylated miR-185-5p (miR-185-5p-bio) probes were significantly increased compared with control (NC-bio) or miR-185-5p probes ([Figure 4C](#)) ($P < 0.05$). As shown in [Figure 4D-G](#), compared with the shNC group, the cell viability, EdU staining positive rate, and cell invasion and migration rate of the shHAGLR1 group were significantly reduced ($P < 0.05$). The apoptosis rate was significantly increased ($P < 0.05$). And shHAGLR1 co-transfected with miR-185-5p inhibitor group could reverse the effects of shHAGLR1 on cell viability, invasion, migration and apoptosis ($P < 0.05$). The results demonstrated that HAGLR exerted biological effects on colon cancer cells through miR-185-5p.

CDK6 and CDK4 Were the Target of miR-185-5p

We used the Starbase 2.0 online prediction tool and CDK4 and CDK6 were identified as potential targets for miR-185-5p ([Figure 5A](#)). To verify the prediction results, WT-CDK4, WT-CDK6 or mutant (Mut) -CDK4 and (Mut)-CDK6 luciferase reporter plasmid were used for luciferase reporter gene assay. The luciferase activity of pGL3-REPOR-CDK4-WT and pGL3-REPOR-CDK6-WT was reduced by miR-185-5p mimic ($P < 0.05$). However, the luciferase activity of pGL3-REPOR-CDK4-Mut and pGL3-REPOR-CDK6-Mut did not change significantly ([Figure 5B](#)). Compared with the control group, miR-185-5p mimic could significantly reduce the protein expression level of CDK4 and CDK6 ($P < 0.05$), miR-185-5p inhibitor significantly increase the protein expression levels of CDK4 and CDK6 ($P < 0.05$, [Figure 5C](#)). As shown in [Figure 5D](#), shHAGLR1 and shHAGLR2 were significantly reduced the protein expression levels of CDK4 and CDK6 ($P < 0.05$), while pcHAGLR was significantly increased the protein expression levels of CDK4 and CDK6 ($P < 0.05$).

Downregulation of HAGLR Repressed Colon Cancer Progression in vivo

In vivo experiments were used to analyze whether HAGLR knockdown inhibited colon cancer progression. The results were shown in [Figure 6A-C](#). Compared with the sh-NC group, shHAGLR1 was inhibited tumor growth, while co-transfection of shHAGLR1 with miR-185-5p inhibitor were significantly reversed the growth of shHAGLR1 on tumors. Ki-67 staining results showed that the number of positive cells in shHAGLR1 group was decreased, while co-transfection of shHAGLR1 with miR-185-5p inhibitor significantly reversed the effect of shHAGLR1 on the number of positive cells ([Figure 6D](#)). In addition, as shown in [Figure 6E](#), the protein expression levels of CDK4 and CDK6 in the shHAGLR1 group were significantly reduced ($P < 0.05$), and co-transfection of shHAGLR1 with miR-185-5p inhibitor significantly reversed the effect of shHAGLR1 on the protein expression levels of CDK4 and CDK6 ($P < 0.05$, [Figure 6E](#)).

Discussion

Colon cancer is a common malignant tumor that originates in the colon and is the third most common gastrointestinal tumor.³⁵ And 40–50 years old is the high incidence population.³⁶ According to the pathological types, colon

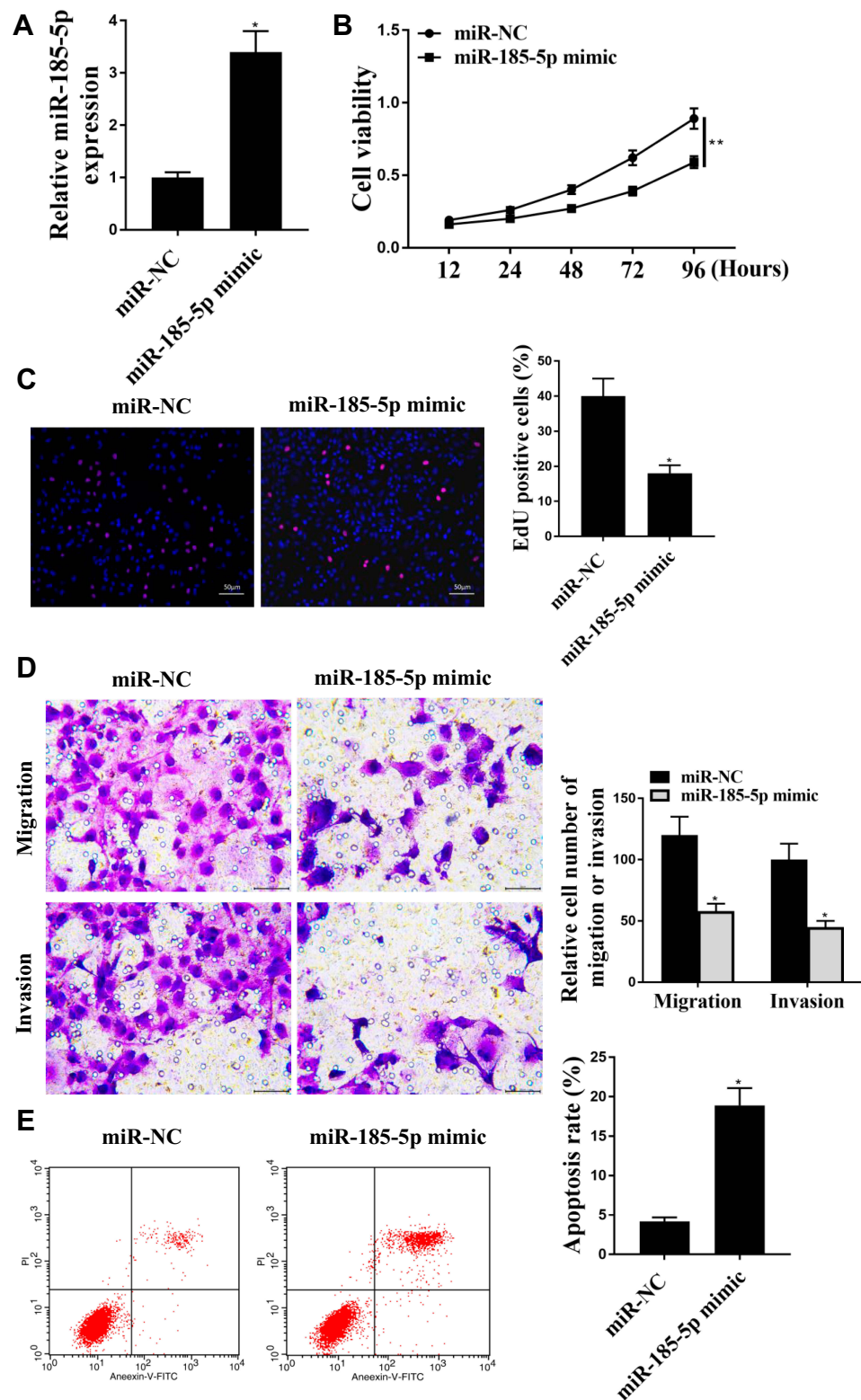


Figure 3 MiR-185-5p regulated colon cancer progression in vitro. **(A)** miR-185-5p expression level in SW480 cells was used to detect by q-PCR. **(B)** Cell viability was used to detect for determined by CCK-8. **(C)** EdU staining was used to detect for cell proliferation. **(D)** Transwell was used to detect by cell migration and cell invasion. **(E)** Flow cytometry was used to detect by apoptosis. * $P < 0.05$, ** $p < 0.01$, $n = 3$.

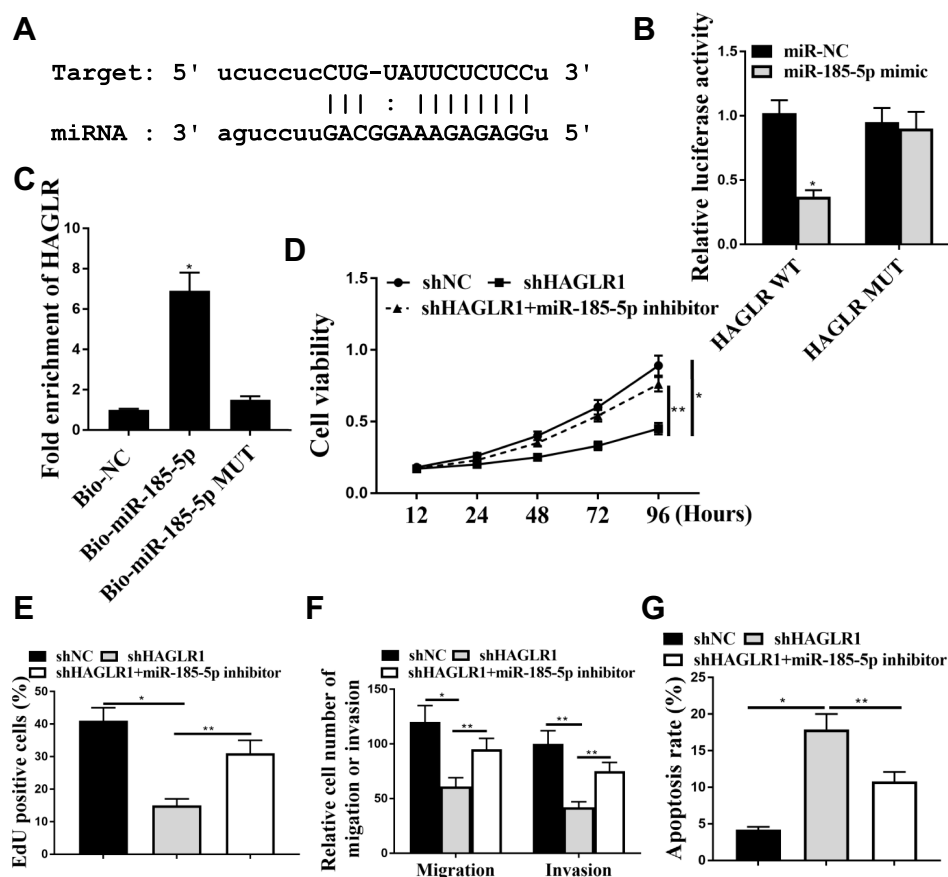


Figure 4 HAGLR acted as a sponge for miR-185-5p in CC cells. **(A)** Putative binding site between HAGLR and miR-185-5p. **(B)** Relative luciferase activity. **(C)** RNA pull-down assay. **(D)** Cell viability was used to detect by CCK-8. **(E)** EdU staining was used to detect for cell proliferation. **(F)** Transwell was used to detect by cell migration and cell invasion. **(G)** Flow cytometry measured apoptosis. * $P < 0.001$, ** $p < 0.05$, $n = 3$.

cancer can be divided into three major categories, adenocarcinoma, mucinous adenocarcinoma and undifferentiated carcinoma.³⁷ However, invasive colon cancer is prone to metastasis in the early stage and easily relapses after treatment, which seriously affects the diagnosis and treatment and prognosis of colon cancer patients.³⁸ The occurrence of colon cancer are the result of the interaction of multiple pathogenic factors, high-fat diet, insufficient food fiber and intestinal flora disturbance are the main environmental factors involved in colon cancer. Familial gonadal tumor polyposis and familial hereditary non-polyposis are typical hereditary factors that lead to colon cancer, and gene mutations are also important pathogenic factors that have been studied.³⁹ The abnormal gene expression play a key role in the occurrence of colon cancer.⁴⁰

Long-chain non-coding ribonucleic acid (lncRNA) is a type of RNA consisting of about 200–600 nucleotides. They generally do not encode proteins, but regulate the expression level of genes through RNA in the form of

transcriptional regulation, epigenetic regulation and post transcriptional regulation.⁴¹ Studies have shown that lncRNA play a key role in many regulatory processes.⁴² LncRNAs were found to be abnormally expressed in colon cancer.⁴³ The effect of lncRNA RP11-462c24.1 on the clinicopathological characteristics of colon cancer, it has been found to be reduced in colorectal cancer tissues, and was significantly related to distant metastasis of tumors.⁴⁴ In addition, it has been found that up-regulation of MALAT1 expression can promote the protein level of HIF-1 α and endothelial cells, which affects the development of colon cancer.⁴⁵ Studies have found that lncRNA HAGLR is abnormally expressed in many cancers. For example, The expression level of lncRNA HAGLR is increased in NSCLC.¹⁹ This study found that the expression level of lncRNA HAGLR was significantly increased in CRC. ShHAGLR could inhibited cell proliferation, invasion and migration, and induced apoptosis. shHAGLR could effectively inhibit the weight and volume

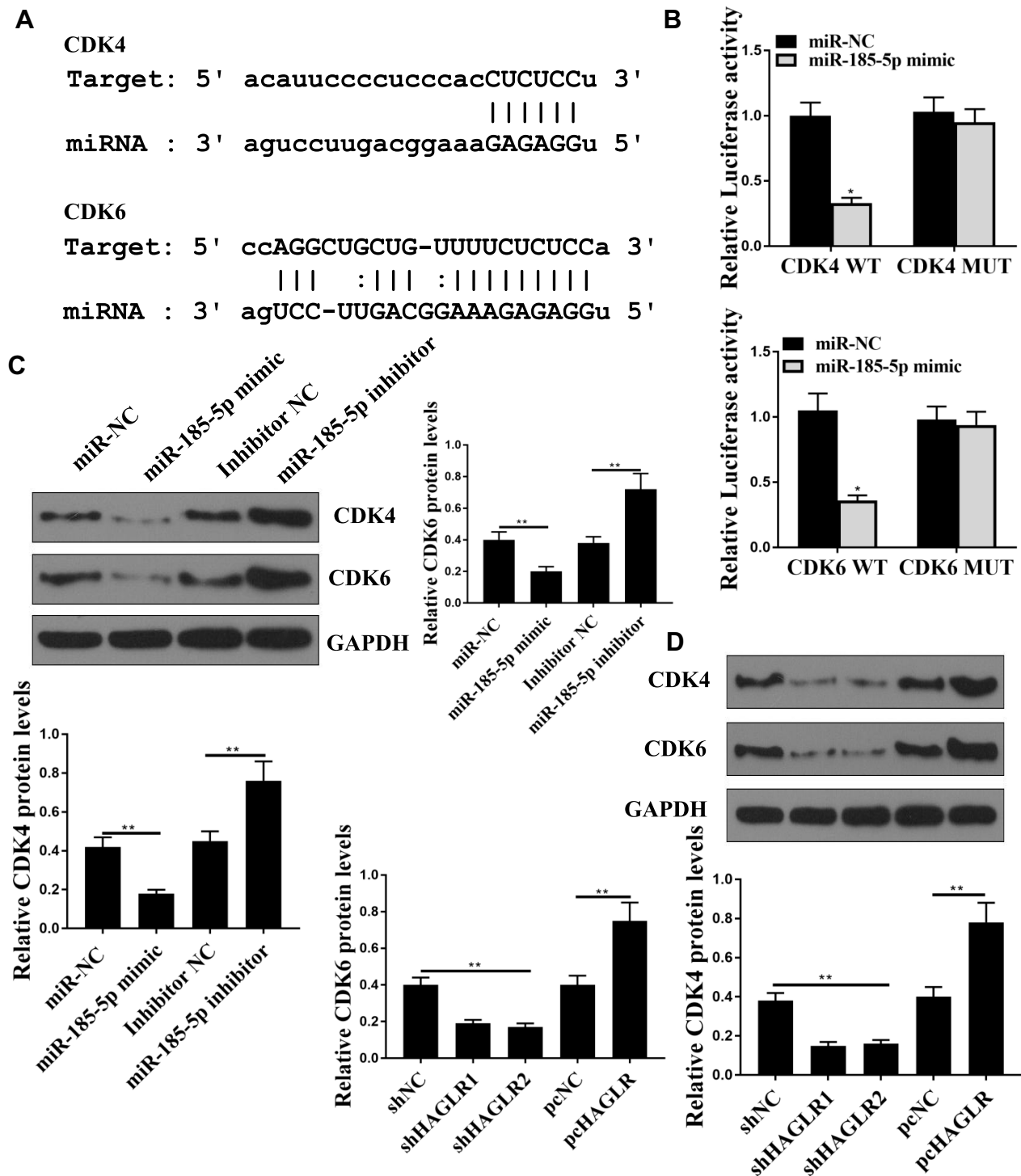


Figure 5 CDK4 and CDK6 were direct targets of miR-185-5p. (A) Starbase predicted the binding site between miR-185-5p and CDK4 and CDK6. (B) Relative luciferase activity. (C, D) The protein expression levels CDK4 and CDK6 in SW480 cells was used to detect by q-PCR. * $P < 0.05$, ** $p < 0.01$, $n = 3$.

of tumors in vivo. Therefore, HAGLR can control the development of colon cancer.

LncRNA regulates miRNA, thereby regulating protein translation and cell activity.⁴⁶ At present, researchers have

identified human miRNA, and miRNAs regulate the expression levels of various genes in the human body.²³ The relationship between miRNAs and colon cancer has been obtained more attention. MiRNAs are expected to become targets for

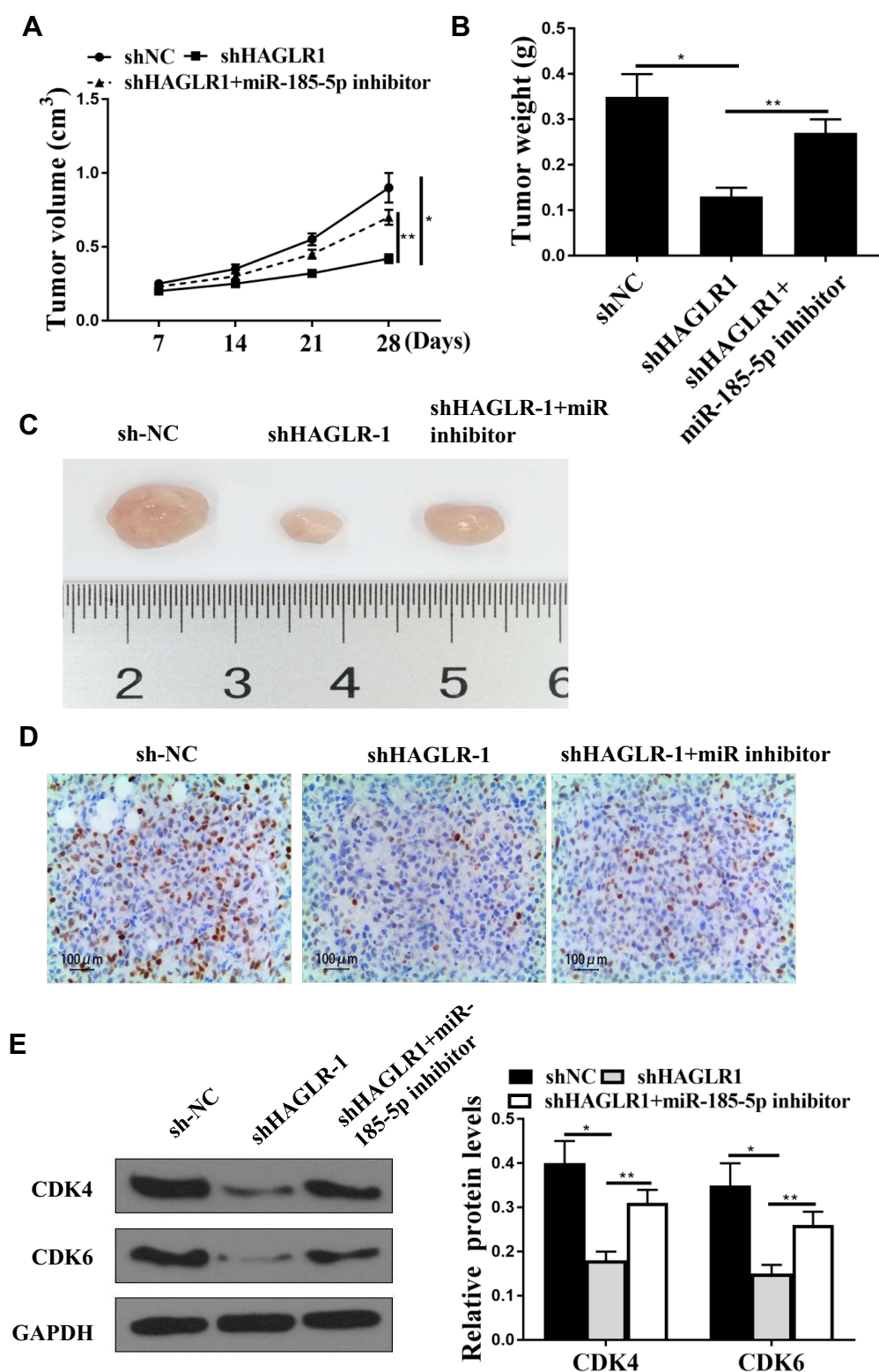


Figure 6 Downregulation of HAGLR1 inhibited the progression of colon cancer in vivo. (A) Measurement of tumor volume in nude mice weekly. (B) Comparison of tumor weight in nude mice. (C) Representative images of three groups of subcutaneous tumors. (D) Ki-67 staining. (E) Protein expression levels of CDK4 and CDK6 was used to detect by Western blot. * $P < 0.05$, ** $p < 0.01$, $n = 5$.

colon cancer treatment.⁴⁷ For example, it has been found that the expression of miRNA-21 is abnormally increased in serum of colon cancer patients, and its expression level is related to TNM stage, chemotherapy effect and prognosis of patients.⁴⁸ Screening results of colon cancer-related miRNA from a large sample showed that in clinical colon cancer samples and their corresponding cancer 21 miRNAs with up-regulated expression are found in tissues.⁴⁹ MiR-185-5p is a miRNA with tumor growth inhibitory effect found. For example, studies have found that miR-185-5p suppresses prostate cancer proliferation.²⁹ MiR-185-5p was screened as a target gene for HAGLR. MiR-185-5p was negatively correlated with HAGLR. The expression level of miR-185-5p was significantly reduced in colon cancer. And miR-185-5p mimic could inhibit cell proliferation, invasion and migration, and induce apoptosis. Co-transfection of shHAGLR with miR-185-5p inhibitor could reverse the effects of shHAGLR on cell proliferation, apoptosis and tumor weight and volume. These results suggested that HAGLR might promote colon cancer growth by regulating miR-185-5p.

miRNAs participate in tumor formation and progression by regulating genes during tumor formation.⁵⁰ Cyclin-dependent kinases (CDKs) belong to the serine/threonine kinase family. The activities of CDK4 and CDK6 are considered to be closely related to the pathogenesis of cancer.⁵¹ CDK4 and CDK6 could promote tumorigenesis and development when over-activated.⁵² In this study, it was found that CDK4 and CDK6 were target gene of miR-185-5p. In addition, miR-185-5p mimic was reduced the expression levels of CDK4 and CDK6, miR-185-5p inhibitor was increased the expression levels of CDK4 and CDK6, while shHAGLR was reduced the expression levels of CDK4 and CDK6, pcHAGLR was increased the expression levels of CDK4 and CDK6. These data indicated that Lnc HAGLR promoted the development of colon cancer by targeting the miR-185-5p/CDK4/CDK6 axis.

There are several limitations in this research. Our study initially explained the mechanism of HAGLR/miR/CDK6 axis, while the deeper control mechanism has not been fully explained clearly. In addition, since our research focuses on CDK4 and CDK6, and the entire cell cycle regulation process involves multiple proteins, there may be more confounding factors in the real in vivo environment. Therefore, further experiments are necessary to provided more profound evidence in the future.

Conclusion

Lnc HAGLR promoted colon cancer progression by regulating the miR-185-5p/CDK4/CDK6 axis, suggesting that

Lnc HAGLR might be a potential oncogene for colon cancer.

Disclosure

The authors declare that they have no competing interests.

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