LncRNA DSCAM-AS1 Promotes Colon Cancer Cells Proliferation and Migration via Regulating the miR-204/SOX4 Axis

This article was published in the following Dove Press journal:
Cancer Management and Research

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Introduction: Long non-coding RNA (lncRNA) DSCAM-AS1 was reported to be aberrantly expressed and play pivotal roles in various human cancers. The aim of the present study was to investigate the expression and roles of DSCAM-AS1 in colon cancer (CC).

Methods: Quantitative real-time PCR (qRT-PCR) was used to detect the expression of DSCAM-AS1, miR-204 and the mRNA level of SOX4. Cell proliferation and cell cycle were analyzed by MTT assay and flow cytometry, respectively. Transwell assay was used for migration capacity detection. Luciferase activity assay was conducted to verify the direct binding of DSCAM-AS1 and miR-204 or miR-204 and SOX4. The protein expression of SOX4 was determined by Western blot. Kaplan–Meier curves were calculated and the Log rank test was performed for the survival data analysis.

Results: DSCAM-AS1 was significantly upregulated in CC and high expression of DSCAM-AS1 was associated with poor prognosis in patients with colon cancer. Knockdown of DSCAM-AS1 significantly suppressed CC cells proliferation and migration. In addition, DSCAM-AS1 acted as a molecular sponge for miR-204 and SOX4 was identified as a direct target of miR-204 in CC. Moreover, the rescue assay revealed that miR-204 inhibition partly abolished the effects of DSCAM-AS1 knockdown on CC cells proliferation, migration and SOX4 expression.

Discussion: The present study demonstrated that DSCAM-AS1 acted as an oncogenic lncRNA in CC progression by regulating miR-204/SOX4 axis and DSCAM-AS1 may serve as a novel therapeutic target in the treatment of colon cancer.

Keywords: DSCAM-AS1, miR-204/SOX4, colon cancer, proliferation, migration

Introduction

Colon cancer (CC), derived from digestive tract, is the third most common malignant tumor and one of the leading causes of cancer-related deaths among men and women.¹ What is worse, the incidence and mortality rate of CC is still increasing each year. It was estimated that there will be about 2 million new cases and nearly 1 million deaths of CC occur one year by 2030.² At present, surgical resection is the most important choice for colon cancer treatment, supplemented by radiotherapy, chemotherapy, and immunotherapy.^{3,4} Although a lot of advances in the early diagnosis and treatment methods of CC have been achieved during the past few decades, the prognosis of CC patients is still not satisfactory.⁵ Therefore, it is necessary to identify some novel molecule involved in the initiation and development of colon cancer, so as to develop some new therapeutic methods for CC patients.

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Long non-coding RNAs (lncRNAs) are a subgroup of poorly conserved endogenous RNAs containing more than 200 nucleotides in length and have little protein coding ability.⁶ lncRNAs always play their roles by regulating the RNA and/or the protein content of a cell on the transcriptional and the post-transcriptional levels.⁷ Accumulating evidence showed that lncRNAs may act as tumor suppressors or oncogenes by participating in multiple processes of cancer development, such as cell proliferation, apoptosis, migration and invasion.^{8,9} lncRNA Down Syndrome Cell Adhesion Molecule anti-sense (DSCAM-AS1), located on 21q22.2, is belonged to the immunoglobulin superfamily of cell adhesion molecules.¹⁰ Recently, emerging literatures reported that DSCAM-AS1 play crucial roles in various human cancers. For example, Liao et al showed that DSCAM-AS1 was upregulated in non-small cell lung cancer (NSCLC). Its overexpression could enhance NSCLC cell migration and invasion via upregulating BCL11A.¹¹ Huang et al reported that DSCAM-AS1 was significantly upregulated in melanoma. Knockdown of DSCAM-AS1 inhibited proliferation, colony formation, migration and invasion of melanoma cells.¹² Ji and colleagues demonstrated that DSCAM-AS1 promoted the progression of hepatocellular carcinoma (HCC) via sponging miR-338-3p and increased DSCAM-AS1 was associated with poor prognosis in HCC.¹³ However, the expression level and functions of DSCAM-AS1 in colon cancer are still not fully understood.

MicroRNAs (miRNAs) are a class of single-stranded small molecule RNAs that regulate the expression of post-transcriptional genes via binding to its 3'untranslated region (3'-UTR).¹⁴ miR-204 was reported to act as a tumor suppressor in multiple human cancers such as glioblastoma, hepatocellular cancer and bladder cancer.¹⁵⁻¹⁷ Moreover, it was demonstrated that SOX4 was a target gene of miR-204 and the miR-204/SOX4 axis played important roles in cancers development. For example, Hu et al showed that miR-204 inhibited cell metastasis and EMT in lung adenocarcinoma through targeting SOX4.¹⁸ Yin et al demonstrated that miR-204 negatively regulates SOX4 and inhibited proliferation, migration and invasion of T-cell acute lymphoblastic leukemia cell lines.¹⁹ Wu et al reported that miR-204 inhibited cell proliferation, migration and invasion in human renal cell carcinoma cells by downregulating SOX4.²⁰ In colon cancer, previous studies showed that SOX4 was overexpressed and associate with a relative poor

prognosis.^{21,22} However, whether lncRNA DSCAM-AS1 play its roles in cancer progression via regulating the miR-204/SOX4 axis is unknown.

In the current study, we aimed to investigate the expression pattern and roles of DSCAM-AS1 in colon cancer. Firstly, we detected the expression levels of DSCAM-AS1 in colon cancer tissues and cell lines in comparison with normal tissues and cell line. Then, we determined the biological functions of DSCAM-AS1 in colon cancer cells by in vitro assays. Finally, we demonstrated that DSCAM-AS1 exerted its effects in colon cancer via regulating the miR-204/SOX4 axis. These results suggest that lncRNA DSCAM-AS1 may serve as a novel biomarker or therapeutic target in colon cancer.

Materials and Methods

Clinical Tissue Samples

In this study, the colon cancer tissues and paired adjacent normal tissues were collected from 37 primary colon cancer patients who underwent surgical treatment at Department of General Surgery, Chinese People's Liberation Army General Hospital. The clinicopathological characteristics of CC patients were summarized in Table 1. Patients received no chemotherapy or radiotherapy prior surgery. After surgical resection, tissue samples were all frozen in liquid nitrogen immediately and stored at -80°C until use. All the patients enrolled were followed up for at least 60 months and the survival time was recorded. This study was approved by the Ethics Committee of Chinese People's Liberation Army General Hospital and each patient has signed a written informed consent for all the procedures.

Cell Culture and Transfection

The human colon cancer cell lines (HT29, HCT8, SW480, and LOVO) and the normal human colon epithelial NCM460 cell line were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were all cultured in Dulbecco's Modified Eagle medium (DMEM, Hyclone) containing 10% FBS (Gibco), 100 U/mL penicillin and 100 mg/mL streptomycin (Corning). All cell lines were cultured in an incubator at 37°C with 5% CO₂.

Cell transfections were all performed using Lipofectamine[®] 2000 reagent (Invitrogen) according to the manufacturer's protocol. Briefly, after seeded in 6-well plates and grown to a cell density of 30%, the

Table 1 Correlation Between DSCAM-AS1 Expression and Clinicopathological Characteristics in Colon Cancer Patients

Characteristics	Number	DSCAM-AS1 Expression		P value
		Low (18)	High (19)	
Age (years)				0.508
≤60	14	8	6	
>60	23	10	13	
Sex				0.743
Male	22	10	12	
Female	15	8	7	
Tumor size (cm)				0.325
≤5	21	12	9	
>5	16	6	10	
Stage				0.045*
I/II	22	14	8	
III/IV	15	4	11	
Differentiation				0.495
Well	24	13	11	
Poor	13	5	8	
Lymph nodes metastasis				0.038*
Negative	24	15	9	
Positive	13	3	10	
Distant metastasis				0.232
Negative	29	16	13	
Positive	8	2	6	

Note: *P<0.05.

cells were transfected and cultured at 37°C for further 48 hours. Then, the cells were harvested for qRT-PCR and other experiments. The siRNA targeting lncRNA DSCAM-AS1 (si-DSCAM-AS1) and the negative control (si-NC) were purchased from GenePharma Co. (Shanghai, China). The miRNA mimics, inhibitors and their negative controls were all provided by RiboBio Co. (Guangzhou, China).

RNA Extraction and qRT-PCR

Total RNA was extracted from clinical tissue samples or cultured cells using TRIzol reagent (Thermo Fisher Scientific, Inc.) according to the manufacturer's protocol. RNA was reverse transcribed into cDNA using the SuperScript III Reverse Transcriptase kit (Thermo Fisher Scientific, Inc.) and qRT-PCR was performed using the SYBR Green PCR Master mix (Thermo Fisher Scientific, Inc.) according to manufacturer's protocol. β -actin was used as an internal control for DSCAM-AS1 and SOX4

mRNA levels. U6 was used as the internal control for miR-204. The primers were shown as follow: lncRNA DSCAM-AS1 forward, 5'-GTGACACAGCAAGACTCCCT-3' and reverse, 5'-GATCCGTCGTCCATCTCTGT-3'; β -actin forward, 5'-GGCCCAGAATGCAGTTCGCCTT-3' and reverse, 5'-AATGGCACCCCTGCTCACGCA-3'; miR-204 forward, 5'-GACGCTTTCCTTTGTCATCCT-3' and reverse, 5'-GTGCAGGGTCCGAGGTATTC-3'; U6 forward, 5'-ATTGGAACGATACAGAGAAGATT-3' and reverse, 5'-GGAACGCTTCACGAATTG-3'; SOX4 forward, 5'-CTTGACATGATTAGCTGGCATGATT-3' and reverse, 5'-CCTGTGCAATATGCCGTGTAGA-3'. The relative expression levels were normalized with the internal controls and calculated according to the $2^{-\Delta\Delta CT}$ method.

MTT Assays

Cell proliferation was determined by using (3-(4,5-dimethyl-thiazol-2-yl) 2,5-diphenyl tetrazolium bromide) (MTT) assay. In brief, the transfected cells were seeded into 96-well plates at a density of 5×10^3 cells/well and cultured at 37°C. Following incubation for 24, 48, 72 or 96 hours, 100 μ L full culture medium containing 0.5 mg/mL MTT (Sigma-Aldrich, St. Louis, MO, USA) was added into each well and cells were cultured for additional 4 hours. Subsequently, the medium was removed and 150 μ L DMSO (Sigma-Aldrich) was added to dissolve the formazan crystals. The absorbance was measured at 490 nm using a microplate reader (Molecular Devices, USA).

Cell Cycle Analysis

Cell cycle distribution was detected by propidium iodide (PI) staining. In short, the transfected cells were collected and fixed with 70% ethanol overnight at 4°C. Then, the cells were washed twice and stained with PI (Sigma-Aldrich) following the manufacturer's protocol. Cell cycle distribution was determined by using a BD FACS Calibur Flow Cytometer (BD Biosciences, CA, USA).

Transwell Migration Assay

Transwell Chambers (8.0 μ m pore, BD Biosciences, CA, USA) were used to measure cell migration capacity. In brief, 2×10^5 cells were isolated, resuspended in 200 μ L serum-free medium and seeded into the upper chambers. The lower chambers were supplemented with 600 μ L complete medium with 10% FBS. After incubated at 37°C for 24 h, cells on the upper side of the membrane were wiped

off by a cotton swab and the migrated cells were fixed with 4% paraformaldehyde for 20 min. Thereafter, the migrated cells were stained with 0.1% crystal violet for 5 min at room temperature and washed with PBS 3 times. The images were photographed using a microscope (Olympus, Tokyo, Japan), and the migrated cells were counted in three random fields.

Luciferase Reporter Assay

Bioinformatics analysis indicated that miR-204 was identified as a candidate target of DSCAM-AS1 by Starbase and SOX4 was identified a direct target of miR-204 by TargetScan. For luciferase reporter assay, the wild type or mutant 3'-UTR of DSCAM-AS1 or SOX4 containing the miR-204 binding site was cloned into the firefly luciferase-expressing pmirGLO vector (Promega, Madison, WI, USA) to generate the wild type or mutant 3'-UTR reporters, named DSCAM-AS1 WT, SOX4 WT, DSCAM-AS1 Mut or SOX4 Mut respectively. The wild type (WT) or mutant (Mut) luciferase vectors were co-transfected with miR-NC or miR-204 mimics into colon cancer cells using Lipofectamine® 2000 according to the manufacturer's protocol. After 48 hours, the luciferase activity was assessed by using a Dual Luciferase Reporter Assay System (Promega, Madison, WI, USA) and normalized to Renilla luciferase activity.

Western Blot

Total protein was extracted from cultured cells using radioimmunoprecipitation assay (RIPA) and then quantified by bicinchoninic acid (BCA) method (Pierce, Rockford, IL, USA). Equal amount of protein sample was separated using 9–11% SDS-PAGE and then transferred to nitrocellulose membranes (Millipore, Billerica, MA, USA). Membranes were blocked with 5% non-fat milk in PBS for 1 hour at room temperature and incubated with primary antibody (both from Abcam, Cambridge, UK) at 4 °C overnight. After washed three times, membranes were incubated with horseradish peroxidase-labeled secondary antibody (Abcam, Cambridge, UK) for 1 h. Subsequently, the signals were detected using an enhanced chemiluminescent (ECL) Western blot analysis kit (Pierce, Rockford, IL).

Statistical Analysis

Data were expressed as mean $\pm$ SD and each experiment was repeated at least three times. All statistical analyses were performed by using GraphPad Prism 5.0 software

(San Diego, CA). The comparisons between groups were compared by using the *t*-test or one-way analysis of variance. The correlation between lncRNA DSCAM-AS1 expression and clinicopathological characteristics of CC patients was analyzed by the Chi-square test. Kaplan–Meier curves were calculated and the Log rank test was performed for the survival data analysis. The potential linear relationship of DSCAM-AS1 and miR-204 expression level was determined by Spearman correlation analysis. The difference was significant when $P < 0.05$.

Results

DSCAM-AS1 Was Overexpressed and Associated with Poor Prognosis in CC

The relative expression level of lncRNA DSCAM-AS1 was detected in CC tissues and cell lines by qRT-PCR. Data revealed that DSCAM-AS1 was significantly overexpressed in CC tissues in comparison with matched adjacent normal tissues (Figure 1A). Meanwhile, we observed that DSCAM-AS1 expression was remarkably higher in CC tissues with advanced clinical stage (III–IV phase) than that with early clinical stage (I–II phase) (Figure 1B). Moreover, our results of qRT-PCR showed that the expression of DSCAM-AS1 was significantly increased in CC cell lines compared with normal human colon epithelial cell line NCM460 (Figure 1C). To evaluate the clinical significance of DSCAM-AS1 in CC, patients were divided into two groups according to the median expression: low DSCAM-AS1 expression and high DSCAM-AS1 expression group. The correlation between DSCAM-AS1 expression and the clinicopathological characteristics of CC patients was evaluated. Data revealed that the expression of DSCAM-AS1 was closely correlated with clinical tumor stage and lymph node metastasis in CC patients (Table 1). In addition, Kaplan–Meier analysis showed that patients with high DSCAM-AS1 expression have a relative poor overall survival (Figure 1D). These results suggest that DSCAM-AS1 may play important roles in the development of CC.

DSCAM-AS1 Knockdown Suppressed CC Cells Proliferation and Migration

To explore the biological effects of DSCAM-AS1 on CC cells proliferation and migration, the expression of DSCAM-AS1 was effectively knocked down in CC cells HCT8 and SW480 by DSCAM-AS1 siRNAs transfection (Figure 2A). Then, functional assays were conducted on

Figure 1 DSCAM-AS1 was overexpressed and associated with poor prognosis in CC. (A) The expression level of lncRNA DSCAM-AS1 determined in 37 CC and matched adjacent normal tissues using qRT-PCR. (B) The relative expression of DSCAM-AS1 in CC tissues with early (I-II) or advanced (III-IV) clinical stage. (C) The relative DSCAM-AS1 expression in 4 CC cell lines and the normal colon epithelial cell line NCM460. (D) Kaplan-Meier survival analysis of patients with high or low DSCAM-AS1 expression. Data were mean $\pm$ SD. * $P < 0.05$.

transfected cells. Results of MTT assay showed that knockdown of DSCAM-AS1 significantly suppressed HCT8 and SW480 cells proliferation (Figure 2B). Flow cytometry was used to detect cell cycle distribution of CC cells after DSCAM-AS1 knockdown. Data showed that DSCAM-AS1 knockdown remarkably inhibited cell cycle transition in HCT8 and SW480 cells (Figure 2C). In addition, the migration capacity of transfected CC cells was determined by transwell migration assay, and results revealed that knockdown of DSCAM-AS1 significantly reduced CC cells migration capacity (Figure 2D). These results suggest that DSCAM-AS1 has a significant effect on the progression of colon cancer.

DSCAM-AS1 Acted as a Molecular Sponge for miR-204 in CC Cells

Accumulating studies reported that lncRNAs always exert their functions via acting as competing endogenous RNAs

(ceRNA) for some microRNAs in cancer development.^{23,24} We supposed that DSCAM-AS1 may also play its roles by sponging some microRNA in CC. Bioinformatic analysis by Starbase2.0 (<http://starbase.sysu.edu.cn/>) revealed that miR-204 contains the potential binding site for DSCAM-AS1 (Figure 3A). Then, the luciferase reporter assay was conducted in HCT8 and SW480 cells to confirm that. Data showed that miR-204 significantly suppressed the luciferase activity of the wild type DSCAM-AS1 reporters (DSCAM-AS1 WT) but not of the mutant reporters (DSCAM-AS1 Mut) in CC cells (Figure 3B). Meanwhile, we found that DSCAM-AS1 expression was obviously reduced following miR-204 overexpression and knockdown of DSCAM-AS1 significantly upregulated miR-204 expression in CC cells (Figure 3C, D). Moreover, the relative expression levels of miR-204 in CC tissues and cell lines were also detected. The results showed that miR-204 was significantly down-regulated in CC tissues compared with adjacent normal

Figure 2 DSCAM-AS1 knockdown suppressed CC cells proliferation and migration. **(A)** DSCAM-AS1 siRNA was transfected into CC cell lines HCT8 and SW480. The expression of DSCAM-AS1 was detected by qRT-PCR following transfection. **(B)** CC cells proliferation was determined by MTT assay after DSCAM-AS1 knockdown. **(C)** Cell cycle distribution of HCT8 and SW480 cells following DSCAM-AS1 knockdown. **(D)** The migration capacity of CC cells after effectively knockdown of DSCAM-AS1. Data were mean $\pm$ SD. * $P < 0.05$.

tissues (Figure 3E). CC cell lines also showed a relative lower expression level of miR-204 than that of the normal colon epithelial cell line NCM460 (Figure 3F). In addition, the relationship between DSCAM-AS1 and miR-204 expression level in CC tissue samples was also evaluated. We found that DSCAM-AS1 expression was negatively correlated with miR-204 level in CC (Figure 3G). These results indicated that DSCAM-AS1 may act as a molecular sponge for miR-204 in CC cells.

SOX4 Was a Direct Target of miR-204 in CC

Our results indicated that miR-204 was downregulated in CC tissues and cell lines. Here, we explored the downstream target of miR-204 in CC. We used TargetScan tool to predict the candidate target of miR-204 and found that SOX4 was a direct target of miR-204 (Figure 4A). To confirm that, a luciferase activity assay was conducted in CC cells. Results of luciferase activity assay showed the

luciferase activity was significantly reduced following cotransfection of miR-204 mimics and wild-type SOX4 vectors (SOX4 WT), but no change following cotransfection of miR-204 mimics and mutant SOX4 vectors (SOX4 Mut) (Figure 4B). Then, the mRNA and protein expression of SOX4 in CC cells were determined following miR-204 overexpression. Results of qRT-PCR showed that the mRNA level of SOX4 was significantly reduced after overexpression of miR-204 in HCT8 and SW480 cells (Figure 4C). Data from Western blot revealed that the protein expression of SOX4 was also dramatically decreased after miR-204 overexpression in CC cells (Figure 4D). These results suggest that SOX4 was a direct target of miR-204 in CC.

DSCAM-AS1 Exerted Its Effects on CC Cells via miR-204/SOX4 Axis

The results above suggest that DSCAM-AS1 acted as a molecular sponge for miR-204 and SOX4 was a direct

Figure 3 DSCAM-AS1 acted as a molecular sponge for miR-204 in CC cells. **(A)** miR-204 contains the potential binding site for DSCAM-AS1 predicted by Starbase2.0. **(B)** The luciferase activity was detected in CC cells after co-transfected with wild type (WT) or mutant (Mut) DSCAM-AS1 reporter and miR-204 mimics or miR-NC. **(C)** The relative DSCAM-AS1 expression levels in CC cells following miR-204 mimics transfection. **(D)** The relative miR-204 expression levels in CC cells after DSCAM-AS1 knockdown. **(E)** The relative expression of miR-204 in colon cancer tissues and adjacent normal tissues detected by qRT-PCR. **(F)** miR-204 expression in 4 CC cell lines and the normal colon epithelial cell line NCM460 determined by qRT-PCR. **(G)** The linear relationship between DSCAM-AS1 and miR-204 expression levels in CC tissue samples. Data were presented as mean $\pm$ SD. * $P < 0.05$.

target of miR-204 in CC. Previous study reported that SOX4 was overexpressed and associate with a relative poor prognosis in colon cancer.^{21,22} We supposed that DSCAM-AS1 may perform its effects on CC cells via miR-204/SOX4 axis. To verify it, we transfected miR-204 inhibitors to downregulate its expression in CC cells following DSCAM-AS1 knockdown (Figure 5A). MTT assay indicated that inhibiting of miR-204 partially reversed the suppressive effects of DSCAM-AS1 knockdown on CC cells proliferation (Figure 5B). The results of migration assay also indicated that the effects of DSCAM-AS1 knockdown on CC cells migration capacity was partly reversed (Figure 5C). Furthermore, we detected the protein expression of SOX4 in the treated cells. Data of Western blot showed that the protein expression of SOX4 was significantly reduced after knockdown of DSCAM-AS1, while this effect was partly restored when

co-transfected with miR-204 inhibitors (Figure 5D). These results suggest that DSCAM-AS1 exerted its effects on CC cells via regulating miR-204/SOX4 axis.

Discussion

An increasing number of studies have reported that the dysregulation of lncRNAs are involved in the tumorigenesis and development of multiple human cancers including colon cancer. For examples, it was reported that lncRNA UCA1 was upregulated in colon cancer and could modulate progression of colon cancer through regulating the miR-28-5p/HOXB3 axis.²⁵ LncRNA FENDRR was demonstrated to function as a tumor-suppressor gene by repressing SOX4 and may serve as a potential therapeutic target in colon cancer.²² Besides, lncRNA HOTAIR was showed to be overexpressed in colon cancer and promote malignant progression of colon cancer by regulating

Figure 4 SOX4 was a direct target of miR-204 in CC. (A) SOX4 was a candidate target of miR-204 predicted by TargetScan. (B) Luciferase reporter assay was conducted in HCT8 and SW480 cells after transfected with wild-type (WT) or mutant (Mut) SOX4 vectors and miR-204 mimics or miR-NC. (C) The relative mRNA levels of SOX4 in CC cells HCT8 and SW480 after miR-204 overexpression. (D) The protein expression of SOX4 in CC cells HCT8 and SW480 after miR-204 overexpression. Data were presented as mean $\pm$ SD. *P < 0.05.

miR-34a.²⁶ However, the clinical significance and biological effects of lncRNA DSCAM-AS1 in colon cancer have not full investigated.

In this study, we are aimed to investigate the expression pattern and functions of DSCAM-AS1 in colon cancer. Firstly, we used qRT-PCR to detect the expression level of DSCAM-AS1 in colon cancer. We found that DSCAM-AS1 was significantly upregulated in CC tissue samples and cell lines when compared with normal control. We also observed that the expression of DSCAM-AS1 was closely correlated with clinical tumor stage and lymph node metastasis in CC patients. Kaplan–Meier analysis showed that patients with high expression of DSCAM-AS1 have a significantly shorter overall survival time. Then, we transfected siRNAs to downregulate the expression of DSCAM-AS1 in CC cells and performed some

functional assays. Our results showed that knockdown of DSCAM-AS1 significantly suppressed CC cells proliferation and cell cycle transition. Moreover, the transwell migration assay indicated that the migration capacity of CC cells was significantly reduced following DSCAM-AS1 knockdown. These results suggest that DSCAM-AS1 was upregulated and acted as an oncogene in colon cancer.

During the past few years, accumulating evidence suggest that lncRNAs can regulate the progression of human cancers by acting as competing sponges for some miRNAs. Here, it was predicted that miR-204 contains the potential binding site for DSCAM-AS1 by Starbase2.0. We further demonstrated that DSCAM-AS1 acted as a molecular sponge for miR-204 in CC cells and SOX4 was identified as a direct target of miR-204 in CC.

Figure 5 DSCAM-AS1 exerted its effects on CC cells via miR-204/SOX4 axis. (A) The expression of miR-204 was inhibited in CC cells after miR-204 inhibitors transfection. (B) miR-204 inhibition partially reversed the suppressive effects of DSCAM-AS1 knockdown on CC cells proliferation. (C) The suppressive effects of DSCAM-AS1 knockdown on CC cells migration capacity was partly reversed by miR-204 inhibition. (D) The protein expression of SOX4 in HCT8 and SW480 cells after DSCAM-AS1 knockdown and/or miR-204 inhibition. Data were mean $\pm$ SD. * $P < 0.05$.

These results revealed that DSCAM-AS1 may play its roles in CC by regulating miR-204/SOX4 axis. To verify this, a rescue assay was performed on CC cells. We observed that miR-204 inhibition partially reversed the suppressive effects of DSCAM-AS1 knockdown on CC cells proliferation and migration. Results of Western blot showed that the protein levels of SOX4 were reduced after DSCAM-AS1 knockdown and these effects were partly abolished by miR-204 inhibition. These results suggest that DSCAM-AS1 may play its roles via regulating miR-204/SOX4 axis in colon cancer.

Taken together, the present study demonstrated that DSCAM-AS1 was upregulated in CC and associated with a poor prognosis. DSCAM-AS1 acted as an oncogenic lncRNA in CC progression by regulating miR-204/SOX4 axis. These results suggest that DSCAM-AS1 may serve as a novel therapeutic target in the treatment of colon cancer.

Conclusions

The present study demonstrated that lncRNA DSCAM-AS1 was upregulated and associated with a poor prognosis in CC. DSCAM-AS1 knockdown suppressed CC cells

proliferation and migration by regulating miR-204/SOX4 axis. DSCAM-AS1 may serve as a novel therapeutic target in the treatment of colon cancer.

Author Contributions

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; gave final approval of the version to be published; and agree to be accountable for all aspects of the work.

Disclosure

The authors declared no conflicts of interest.

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