Emergence of Terbinafine Resistant *Trichophyton mentagrophytes* in Iran, Harboring Mutations in the Squalene Epoxidase (SQLE) Gene

**Introduction:** *Trichophyton mentagrophytes* and *T. interdigitale* are important causative agents of superficial mycoses, demonstrating emergent antifungal drug resistance. We studied the antifungal susceptibility profiles in Iranian isolates of these two species.

**Methods:** A total of 96 *T. interdigitale* and 45 *T. mentagrophytes* isolates were subjected to molecular typing by ribosomal ITS region. Antifungal susceptibility profiles for terbinafine, griseofulvin, clotrimazole, efinaconazole, luliconazole, amorolfine and ciclopirox were obtained by CLSI broth microdilution method. The squalene epoxidase (SQLE) gene was subjected to sequencing for mutations, if any, in isolates exhibiting elevated MICs for terbinafine.

**Results:** Luliconazole and efinaconazole showed the lowest MIC values against *T. mentagrophytes* and *T. interdigitale* isolates. There were five isolates with terbinafine MICs \( \geq 32 \) µg/mL in our sample. They belonged to *T. mentagrophytes* type VIII and harbored two alternative SQLE gene sequence variants, leading to Phe397Leu and Ala448Thr or Leu393Ser and Ala448Thr substitutions in the enzyme. All terbinafine resistant strains could be inhibited by luliconazole and efinaconazole.

**Conclusion:** This study documented a step in the global spread of resistance mechanisms in *T. mentagrophytes*. However, treatment alternatives for resistant isolates were available.

**Keywords:** *Trichophyton mentagrophytes*, *SQLE*, terbinafine, antifungal drug resistance, Iran

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**Introduction**

Dermatophytosis or tinea is known as the most common superficial mycosis in dermatological practice. It involves skin, nails and hair. Dermatophytosis is caused by a group of fungi, called dermatophytes. The dermatophytes are cosmopolitan and encompass more than 50 species from the genera *Trichophyton*, *Microsporum*, *Epidermophyton*, *Arthroderma*, *Nannizzia*, *Lophophyton*, and *Paraphyton*. Most skin infections by dermatophytes, especially by *Trichophyton* spp., are successfully treated by using terbinafine (TRB), an allylamine compound which is the first-line oral medication for the treatment of such infections. The drug blocks the formation of ergosterol, the major component of fungal membrane, by inhibiting squalene epoxidase enzyme (SQLE) and subsequently inhibits the fungal growth. However, an increasing number of difficultly treated cases is being documented. This phenomenon can be connected in part with relapses because of poor adherence to antifungal treatment regime. However, the most important part is emergence of...
recalcitrant dermatophytosis due to verified in vivo/vitro resistance to TRB. In the first decade of the 21st century, terbinafine resistance in dermatophytes was found to be rare and primarily limited to T. rubrum isolates.

Nonetheless, recent reports from India and some other Asian and European countries indicate that clinical/microbial TRB resistance now involves T. interdigitale and T. mentagrophytes. Since these fungi are essentially conspecific, they can be treated together as the T. mentagrophytes/T. interdigitale species group (TMTISG). In most mentioned reports, clinical therapeutic failures have been correlated with nonsynonymous point mutations in the SQLE gene. But, there has been no report on TRB resistance and molecular mechanisms underlying reduced susceptibility to this antifungal agent in Iranian TMTISG isolates.

Our study aimed to assess antifungal susceptibility of TMTISG isolates. We described five T. mentagrophytes strains, harboring some known point mutations in SQLE gene, which probably were responsible for high terbinafine MICs and proposed treatment alternatives for TRB resistant cases.

Materials and Methods

Clinical Isolates

A total of 141 clinical TMTISG isolates from different provinces of Iran were included in the study. These isolates were a part of a recent investigation on the epidemiological aspects of infections due to the TMTISG in Iran over a two-year study during 2016–2018. The study was approved by the Ethic Committee of Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran (approval ID: IR.AJUMS.REC.1398.851).

Molecular Identification of the Isolates

Identification of all isolates was performed by sequencing of the internal transcribed spacer region of ribosomal DNA (ITS). Briefly, the isolates were sub-cultured on Sabouraud dextrose agar (SDA; BD Diagnostics, USA) and incubated at 28°C for one week. Genomic DNA was isolated from the mycelium of each strain by mechanical homogenization in lysis buffer, consisting of 200 mM Tris- HCl [pH 8.0], 0.5% sodium dodecyl sulfate, 250 mM NaCl, and 25 mM EDTA with the use of SpeedMill device (Analytik Jena, Germany), followed by purification with phenol-chloroform-isoamyl alcohol (25:24:1) and precipitation with ethanol. ITS region was amplified in each isolate by V9G (5′-TTACGTCCTGCCCTTTTGA-3′) and LS266 (5′-GCATTCCACAACAATCGACTC-3′) primers. The amplification program could be schematically represented in the following way: 6 min 94°C; 35 × [30 s 94°C, 30 s 58°C, 1 min 72°C]; 10 min 72°C. The amplified products were sequenced with the use of mentioned primers and the BigDye Terminator Kit version 3.1 (Applied Biosystems, USA), in an ABI Prism 3130XL Genetic Analyzer (Applied Biosystems, USA). The obtained raw sequences were imported to MEGA software ver. 7.0 and quality-checked. Each isolate was identified down to the species level by BLAST search of respective ITS sequence against annotated sequences, deposited in the Westerdijk Fungal Biodiversity Institute (CBS) database (http://www.cbs.knaw.nl).

Antifungal Susceptibility Testing

In vitro AFST against TMTISG isolates was done by the Clinical and Laboratory Standards Institute broth microdilution method, according to CLSI M38-A2 document. The following drugs were tested: terbinafine (TRB; Combi-Blocks, USA), itraconazole (ITC; Sigma-Aldrich, USA), griseofulvin (GRE; Wako Pure Chemical, Japan), clotrimazole (CLT; Sigma-Aldrich, USA), efinaconazole (EFN; Nihon Nohyaku, Japan), luliconazole (LUZ; Funakoshi, Japan), amorolfin hydrochloride (AHC; LKT Laboratories, USA) and ciclopirox olamine (CPO; LKT Laboratories, USA). The final concentration of each antifungal agent was as follows: 0.001–0.5 µg/mL for TRB, 0.016–8 µg/mL for ITC, 0.015–8 µg/mL for GRE, 0.0625–32 µg/mL for CLT, 0.001 to 0.5 µg/mL for EFN, 0.00006–0.031 µg/mL for LUZ, 0.03–16 µg/mL for AHC and 0.004–2 µg/mL for CPO. To promote conidiation, we cultured the strains for a week at 28°C on Petri dishes containing modified Sabouraud Glucose agar, diluted 10-fold (peptone 0.2%, KH2PO4 0.1%, MgSO4·7H2O 0.1%, glucose 0.1%, agar 1.5%). The sterile saline containing 0.05% (w/v) Tween 80 was added to the surface of each mature colony and inoculum suspension was prepared by rubbing with a sterile scraper. Conidial suspension was harvested by a sterile syringe and transferred to a sterile filter (0.4 µm) to eliminate the hyphal masses. After gentle shaking conidial suspension was diluted to achieve 65–70% light transmission at wavelength of 530 nm. To obtain final density of 1 × 10^3 to 3 × 10^3 CFU/mL, the conidial suspension was diluted in RPMI 1640 medium, with the ratio of 1:50. For each isolate, the dermatophyte-free and antifungal-free controls were used and microplates were incubated at 30°C. To check the performance accuracy of AFST, the CLSI
reference strains of Candida parapsilosis ATCC 22019 and
C. kruase ATCC 6258 were used as control for every new
batch of tested isolates. The minimal inhibitory concentra-
tions (MICs) for all used antifungals were defined as the
lowest concentration that led to complete inhibition of observ-
able growth after 96 hrs.

Sequencing of Squalene Epoxidase (SQLE) Gene and Analysis

In order to determine whether a mutation in the SQLE
gene could be potentially involved in elevated TRB
MICs, the partial SQLE gene in five resistant isolates, and
also in five susceptible isolates for comparison, was
amplified and sequenced with the TrSQLE-F1 (5’-
ATGGTTGTAGAGGCTCCTCCC-3’) and TrSQLE-R1 (5’-
CTAGCTTTGAAGTTCGGCAAA-3’) primer pair.11

A PCR machine was run according to a program, modified
from Singh et al: 5 min 95°C; 34× [30 s 95°C, 30 s 60°C,
3 min 72°C]; 10 min 72°C.5 The predicted amino acid
sequences of SQLE in all tested T. mentagrophytes isolates
were compared with the reference sequence for
T. mentagrophytes and T. interdigitale (GenBank accession
number KU242352).

ITS Typing

Given that some recent reports correlated in vitro/in vivo
TRB resistance in the TMTISG with a distinct ITS geno-
type (Type VIII), the nucleotide sequences for ribosomal
ITS1-5.8S-ITS2 region in all isolates were used for typing
by a script (https://github.com/Ivan-Pchelin/genotyping-by-
sequencing). All point mutations and indel events strictly
within the borders of ITS region were considered
significant.21

Results

By ITS sequencing, 96 isolates were identified as
T. interdigitale and 45 isolates as T. mentagrophytes. The
T. interdigitale isolates originated from tinea pedis (n = 71),
tinea unguium (n = 20) and tinea corporis (n = 5) infections,
while the majority of T. mentagrophytes isolates were from
tinea corporis (n = 43), followed by tinea capitis (n = 1) and
nail infection (n = 1).

Table 1 summarizes MIC ranges, geometric means
(GMs) of MICs, and the MIC50/MIC90 ratios of 8 antifungal
drugs used against 141 TMTISG isolates. GM MIC values of
LUZ and EFN against T. interdigitale isolates were 0.0016
and 0.0057 µg/mL while these values for T. mentagrophytes
isolates were 0.0024 and 0.009 µg/mL, respectively. Whereas
GRE had the highest GM MIC value (1.1 µg/mL) for
T. interdigitale isolates, CLT showed the highest GM MIC
value (3.25 µg/mL) against T. mentagrophytes isolates. In
view of susceptibility to TRB, all T. interdigitale isolates
were susceptible to this agent (MICs range = 0.003–0.25
µg/mL and MIC90 = 0.0125 µg/mL) while the MICs of
TRB for T. mentagrophytes isolates were in the range
0.007–32 µg/mL. We found 5 T. mentagrophytes isolates
with TRB MICs ≥32 µg/mL (resistant). From a total of 45
T. mentagrophytes isolates, 28 isolates harbored ITS type
VIII and all five TRB resistant strains were from this geno-
type (5/28; 18%).

The partial SQLE sequence was successfully amplified
in all 5 sensitive and 5 resistant strains. The obtained
sequences were deposited in GenBank under accession
numbers MN893286 and MN901902–MN901905 for sus-
ceptible and MN901906–MN901910 for resistant strains.
All T. mentagrophytes resistant isolates harbored missense
mutations in SQLE, corresponding to amino acid substitu-
tion Ala448Thr in combination with Leu393Ser (isolate

Table 1 MIC Values (µg/mL) of 8 Antifungal Agents Against 141 Trichophyton interdigitale and T. mentagrophytes Isolates

<table>
<thead>
<tr>
<th>Species</th>
<th>Antifungal MICs</th>
<th>TRB</th>
<th>ITC</th>
<th>EFN</th>
<th>LUZ</th>
<th>CLT</th>
<th>AHC</th>
<th>CPO</th>
<th>GRE</th>
</tr>
</thead>
<tbody>
<tr>
<td>T. interdigitale (n=96)</td>
<td>MIC&lt;sub&gt;GM&lt;/sub&gt;</td>
<td>0.038</td>
<td>0.36</td>
<td>0.0057</td>
<td>0.0016</td>
<td>1.43</td>
<td>0.92</td>
<td>0.45</td>
<td>1.1</td>
</tr>
<tr>
<td></td>
<td>MIC&lt;sub&gt;50&lt;/sub&gt;</td>
<td>0.062</td>
<td>0.5</td>
<td>0.007</td>
<td>0.003</td>
<td>1</td>
<td>1</td>
<td>0.5</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>MIC&lt;sub&gt;90&lt;/sub&gt;</td>
<td>0.125</td>
<td>1</td>
<td>0.015</td>
<td>0.001</td>
<td>4</td>
<td>2</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>MIC range</td>
<td>0.003–0.25</td>
<td>0.062–8</td>
<td>0.001–0.125</td>
<td>0.0004–0.015</td>
<td>0.25–16</td>
<td>0.125–4</td>
<td>0.062–1</td>
<td>0.062–4</td>
</tr>
<tr>
<td>T. mentagrophytes</td>
<td>MIC&lt;sub&gt;GM&lt;/sub&gt;</td>
<td>0.12</td>
<td>0.32</td>
<td>0.009</td>
<td>0.0024</td>
<td>3.25</td>
<td>0.92</td>
<td>0.56</td>
<td>1.8</td>
</tr>
<tr>
<td>(n=45)</td>
<td>MIC&lt;sub&gt;50&lt;/sub&gt;</td>
<td>0.625</td>
<td>0.25</td>
<td>0.0075</td>
<td>0.003</td>
<td>4</td>
<td>1</td>
<td>0.5</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>MIC&lt;sub&gt;90&lt;/sub&gt;</td>
<td>&gt;32</td>
<td>0.5</td>
<td>0.031</td>
<td>0.001</td>
<td>16</td>
<td>2</td>
<td>1</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>MIC range</td>
<td>0.007–32</td>
<td>0.062–2</td>
<td>0.001–0.125</td>
<td>0.0004–0.015</td>
<td>0.5–32</td>
<td>0.125–4</td>
<td>0.062–1</td>
<td>0.25–4</td>
</tr>
</tbody>
</table>

Abbreviations: TRB, terbinafine; ITC, itraconazole; EFN, efinaconazole; LUZ, luliconazole; CLT, clotrimazole; AHC, amorolfine hydrochloride; CPO, ciclopirox olamine; GRE, griseofulvin.
R1) or Phe397Leu (isolates R2 to R5). On the other hand, two susceptible isolates (MICs = 0.125–0.25 µg/mL) revealed wild type SQLE sequence, while the three other isolates (MICs = 0.0075–0.25 µg/mL) showed a single Ala448Thr substitution (Table 2).

**Discussion**

The antifungal susceptibility profile of Iranian TMTISG isolates, described in our results (Table 1), was comparable to that ones from other Iranian and foreign studies. The MIC values of EFN against TMTISG isolates from Japan, Canada, US and Iran in three studies all in all ranged from 0.001 to 0.03 µg/mL. They did not differ significantly from the MIC values for EFN in the current study (MICs = 0.001–0.125 µg/mL). Similarly, LUZ inhibitory concentrations against our TMTISG isolates ranged from 0.0004 to 0.015 µg/mL and were comparable with 0.0001–0.004 µg/mL for this imidazole against TMTISG strains in two previous reports from US and Iran. In two other investigations, LUZ has also had the most pronounced in vitro effect against TMTISG isolates from India and Iran, when compared with other antifungals, though the MIC values were higher (0.016–0.25 µg/mL) than in the current study. In the current report, similar to other countries varied in the therapeutic potencies of LUZ and EFN against TMTISG isolates (Table 1). The GM MIC value for CPO against T. interdigitale (0.45 µg/mL) in the present study, was similar to those found by Rudramurthy et al (0.25 µg/mL) and Magagnin et al (0.6 µg/mL). In the present study, the MIC90 for AHC against TMTISG isolates was 2 µg/mL, which was not compatible with those reported for T. interdigitale (0.02 µg/mL) and T. mentagrophytes (0.125 µg/mL) in India and US. In agreement with the reports by Singh et al and Baghi et al, TMTISG strains showed increased susceptibility to CLT and GRE.

To the extent of our knowledge, TRB resistance in TMTISG isolates has already been reported from at least eight Asian and European countries, including India, Switzerland, Japan, Finland, Denmark, Bahrain, Russia, and Germany. The TRB MIC values in resistant TMTISG isolates from Iran and other countries varied in the range ≥1–≥32 µg/mL. In the current report, similar to some other studies, all TRB resistant isolates belonged to T. mentagrophytes species and ITS type VIII (5/28; 18%). Despite this, careful analysis of the literature revealed resistant Type II strains (Table 3). Hence, more data are needed to clarify whether in the TMTISG there is an association between genotypes and a potential to develop antifungal drug resistance. Among amino acid substitutions in SQLE, leading to TRB resistance, the most commonly encountered are Phe397Leu and Leu393Phe (Tables 2 and 3). The substitution Leu393Ser or other less common substitutions were also correlated with a high MIC value (≥32 µg/mL) of TRB. In some cases, in vitro resistance could not be explained by the presence of any mutation in SQLE (wild type). Then, mechanisms other than Phe397Leu and Leu393Phe substitution should still be considered as alternatives for TRB treatment failure. For example, Santos et al, recently showed that TRB resistance in T. rubrum can be mediated by multiplication of salicylate 1-monooxygenase (salA) gene.

**Table 2** MIC Values (µg/mL) of Used Antifungal Agents Against 10 Terbinafine Resistant/Susceptible *T. mentagrophytes* Isolates and Corresponding GenBank Accession with Consequential Amino Acid Substitutions in SQLE Gene

<table>
<thead>
<tr>
<th>Amino Acid Substitution</th>
<th>Resistant</th>
<th>Sensitive</th>
</tr>
</thead>
<tbody>
<tr>
<td>SQLE accession number</td>
<td>MN901906–MN901910</td>
<td>MN901906–MN901905, MN9328</td>
</tr>
<tr>
<td>Phe397Leu/Ala448Thr</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>Leu393Ser/Ala448Thr</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Ala448Thr</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>Wild type</td>
<td>0</td>
<td>2</td>
</tr>
<tr>
<td>MIC Values (µg/mL)</td>
<td>≥32</td>
<td>0.125–0.25</td>
</tr>
<tr>
<td>TRB</td>
<td>0.125–2</td>
<td>0.25–2</td>
</tr>
<tr>
<td>ITC</td>
<td>0.002–0.008</td>
<td>0.008–0.125</td>
</tr>
<tr>
<td>EFN</td>
<td>0.008–0.004</td>
<td>0.008–0.015</td>
</tr>
<tr>
<td>LUZ</td>
<td>2–8</td>
<td>2–16</td>
</tr>
<tr>
<td>CLT</td>
<td>1–4</td>
<td>1–4</td>
</tr>
<tr>
<td>AHC</td>
<td>0.5–2</td>
<td>0.25–4</td>
</tr>
<tr>
<td>CPO</td>
<td>0.5–1</td>
<td>0.5–1</td>
</tr>
<tr>
<td>GRE</td>
<td>1–4</td>
<td>1–4</td>
</tr>
</tbody>
</table>

**Conclusion**

Overall, in both *T. mentagrophytes* and *T. interdigitale*, the potencies of LUZ and EFN against TMTISG isolates were apparently greater than those of other agents. Here, for the first time in Iran, we described TRB resistance in TMTISG isolates with its molecular mechanisms. The emergence of high level of in vitro TRB resistance with proven mutation in
Table 3 Number of Terbinafine (TRB) Resistant Isolates Showing Mutations in the Squalene Epoxidase (SQLE) Gene in Regards to the ITS Genotypes and TRB MIC Values in Different Studies

<table>
<thead>
<tr>
<th>Row</th>
<th>Species</th>
<th>ITS Genotype</th>
<th>Country of Origin</th>
<th>MIC Rates of Resistant Isolates (µg/mL)</th>
<th>Number of Resistant Isolates</th>
<th>Amino Acid Substitution Found in Resistance Isolates</th>
<th>Study</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td><em>T. interdigitale</em></td>
<td>ND</td>
<td>Switzerland</td>
<td>≥1.2</td>
<td>1</td>
<td>Phe397Leu</td>
<td>Yamada et al., 2017</td>
</tr>
<tr>
<td>2</td>
<td><em>T. interdigitale</em></td>
<td>II</td>
<td>Japan</td>
<td>1</td>
<td>2</td>
<td>Wild type</td>
<td>Hiruma et al., 2018</td>
</tr>
<tr>
<td>3</td>
<td><em>T. mentagrophytes</em></td>
<td>VIII</td>
<td>India</td>
<td>1–≤32</td>
<td>20</td>
<td>Phe397Leu Leu393Phe</td>
<td>Singh et al., 2018</td>
</tr>
<tr>
<td>4</td>
<td><em>T. interdigitale</em></td>
<td>ND</td>
<td>India</td>
<td>4–≥16</td>
<td>4</td>
<td>Phe397Leu Wild type</td>
<td>Rudramurthy et al., 2018</td>
</tr>
<tr>
<td>5</td>
<td><em>T. interdigitale</em></td>
<td>ND</td>
<td>India</td>
<td>4–≥32</td>
<td>13</td>
<td>Phe397Leu Leu393Phe Wild type</td>
<td>Khurana et al., 2018</td>
</tr>
<tr>
<td>6</td>
<td><em>T. interdigitale</em></td>
<td>II</td>
<td>Denmark</td>
<td>4–≤8</td>
<td>2</td>
<td>Phe397Leu Leu393Phe Wild type</td>
<td>Saunte et al., 2019</td>
</tr>
<tr>
<td>7</td>
<td><em>T. mentagrophytes</em></td>
<td>VIII</td>
<td>Finland</td>
<td>4–8</td>
<td>4</td>
<td>Phe397Leu</td>
<td>Järv et al., 2019 TIMM</td>
</tr>
<tr>
<td>8</td>
<td><em>T. mentagrophytes</em></td>
<td>ND</td>
<td>Switzerland</td>
<td>≥1</td>
<td>1</td>
<td>Gin408Leu</td>
<td>Hsieh et al., 2019</td>
</tr>
<tr>
<td>9</td>
<td><em>T. mentagrophytes</em></td>
<td>VIII</td>
<td>Bahrain</td>
<td>–&lt;sup&gt;2&lt;/sup&gt;</td>
<td>1</td>
<td>Phe397Leu</td>
<td>Swi et al., 2019</td>
</tr>
<tr>
<td>10</td>
<td><em>T. mentagrophytes</em></td>
<td>VIII</td>
<td>India</td>
<td>–&lt;sup&gt;2&lt;/sup&gt;</td>
<td>1</td>
<td>Phe397Leu</td>
<td>Burmester et al., 2019</td>
</tr>
<tr>
<td>11</td>
<td><em>T. mentagrophytes</em></td>
<td>VIII</td>
<td>Iran</td>
<td>≥32</td>
<td>5</td>
<td>Phe397Leu/Ala448Thr Leu393Ser/Ala448Thr</td>
<td>Current study</td>
</tr>
</tbody>
</table>

Notes: <sup>1</sup>No ITS sequence accession numbers were mentioned. <sup>2</sup>The TRB resistant isolates were reported as *T. interdigitale* but the true identity of isolates based on ITS-sequencing is *T. mentagrophytes*. <sup>3</sup>Resistance was not determined by the broth microdilution method, but by the agar diffusion method using Sabouraud's dextrose agar containing 0.2 µg/mL terbinafine.

Abbreviation: ND, not determined.

SQLE gene in Iranian *T. mentagrophytes* isolates is unpromising and warrants the genotyping of isolates primarily resistant to TRB.

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Disclosure

The authors declare that they have no competing interests in this work.

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