Phosphoglycerate Mutase 1: Its Glycolytic and Non-Glycolytic Roles in Tumor Malignant Behaviors and Potential Therapeutic Significance

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Abstract: Phosphoglycerate mutase 1 (PGAM1) is an important enzyme that catalyzes the reversible conversion of 3-phosphoglycerate and 2-phosphoglycerate during the process of glycolysis. Increasing evidence suggests that PGAM1 is widely overexpressed in various cancer tissues and plays a significant role in promoting cancer progression and metastasis. Although PGAM1 is a potential target in cancer therapy, the specific mechanisms of action remain unknown. This review introduces the basic structure and functions of PGAM1 and its family members and summarizes recent advances in the role of PGAM1 and various inhibitors of cancer cell proliferation and metastasis from a glycolytic and non-glycolytic perspective. Recent studies have highlighted a correlation between PGAM1 and clinical features and prognosis of cancer as well as the development of target drugs for PGAM1. The integrated information in this review will help better understand the specific roles of PGAM1 in cancer progression. Furthermore, the information highlights the non-glycolytic functions of PGAM1 in tumor metastasis, providing an innovative basis and direction for clinical drug research.

Keywords: PGAM1, glycolysis, non-glycolytic, proliferation, metastasis, cancer therapy

Introduction

Even in aerobic environments, most cancer cells rely mainly on glycolysis to generate energy, unlike normal cells, which mainly rely on mitochondrial oxidative phosphorylation to generate energy. This phenomenon was discovered by Warburg in 1924 and was named the “Warburg effect.” Glycolysis is not an effective process for generating adenosine triphosphate (ATP) and the preference of cancer cells for this type of metabolic pattern has aroused intense interest and has been thought to be a hallmark of cancer therapy in past decades. Following the discovery of the Warburg effect, many glycolytic proteins were subsequently found to be involved in cancer progression, including lactate dehydrogenase A (LDHA), phosphoglycerate dehydrogenase (PHGDH), hexokinase 2 (HK2), and glucose transporter 1 (GLUT1). Among these proteins, phosphoglycerate mutase 1 (PGAM1), a key enzyme in the glycolytic pathway that catalyzes the reversible conversion of 3-phosphoglycerate (3-PG) into 2-phosphoglycerate (2-PG), has also received increasing attention. PGAM1 is over-expressed in colorectal cancer, hepatocellular carcinoma (HCC), non-small cell lung cancer (NSCLC), pancreatic ductal adenocarcinoma (PDAC), oral squamous cell carcinoma (OSCC), prostate cancer (PCA), urothelial carcinoma (UBC), and...
and breast cancer. Furthermore, it plays an important role in tumor proliferation and tumor metastasis in some of these cancer types. The expression of PGAM1 was higher in tumor tissues than in adjacent normal tissues. Altogether, these findings indicate that PGAM1 could be a potential target for cancer therapy. Until recently, several factors of PGAM1 biology were still unknown such as how it affected tumor proliferation and metastasis through the regulation of glycolysis, whether its non-glycolytic effect participated in the malignant behavior of cancer and whether it is a clinically relevant therapeutic target or biomarker for cancer. In this review, we summarized the current knowledge of the role of PGAM1 and its inhibitors in the regulation of tumor malignant behaviors, as well as current developments on target drugs for PGAM1. Such information will provide novel concepts for future investigation of PGAM1 as a potential target for cancer therapy.

Basic Structure and Function of PGAM1 and Its Family Members

PGAM1 belongs to the phosphoglycerate mutase family, which can be subdivided into monophosphoglycerate mutases (mPGAM) and bisphosphoglycerate mutases (BPGAM). The interconversion of 3-PG and 2-PG is mainly catalyzed by mPGAM, whereas the conversion of 1,3-bisphosphoglycerate (BP) to 2,3-BPG in the presence of 3-PG is catalyzed by BPGAM. Additionally, mPGAM can be further subdivided into two distinct categories, cofactor-dependent (dPGM) and cofactor-independent (iPGM). Previous studies provided evidence indicating that dPGM and BPGAM have kinetic and structural similarities and are thought to be paralog structures. For example, dPGM participates in three catalytic reactions: the reversible conversion of 3-PG to 2-PG, the phosphatase reaction transforming 2,3-BPG to PG, and the synthase reaction producing 2,3-BPG from 1,3-BPG, which is similar to BPGAM. In adult mammals, dPGM has two different subunits, BB-PGAM and MM-PGAM. In humans, BB-PGAM, another form of PGAM1, was originally isolated from the brain but has recently been found in the liver, breast and other tissues. MM-PGAM (also known as PGAM2) is a muscle-specific form mainly expressed in mature cardiac tissues and skeletal muscles.

In humans, the cytogenetic location of PGAM1 is 10q24.1, with its cDNA encoding a 254 amino acid protein. PGAM1 is a homodimer with a molecular weight (MW) of 28,804 Da (Figure 1A). The phosphorylated HIS11 residues in the active domain are donors and acceptors of phosphate groups, with 2,3-BPG acting as an intermediate (Figure 1B). PGAM1 is primarily found in the cytoplasm, but has also been found on the cell membrane.

The primary role of PGAM1 is to catalyze the reversible conversion of 3-PG to 2-PG, a critical step in glycolysis (Figure 2). According to a study by Liu et al., PGAM1 is a downstream target of the PI3K/Akt/mTOR/HIF-1α pathway, which regulates cellular metabolism (Figure 2). Schrade et al found that altered expression of PGAM1 is associated with GATA4, which mostly modulates tight and adherens junction formation and extracellular matrix reorganization in mouse Sertoli cells (SCs).

Glycolytic Role of PGAM1 in Cancer Proliferation

Glycolysis is an oxygen-independent metabolic pathway that converts glucose to ATP and combines ten enzyme-catalyzed reactions. The glycolytic pathway is the first step in glucose metabolism in all living cells, with multiple enzymes involved in the precise regulation of the pathway for the maintenance of homeostasis. Most normal cells generate energy through glycolysis under oxygen deficient conditions. However, the Warburg effect highlights that cancer cells mainly produce energy via glycolysis, even in an aerobic environment. Therefore, to provide sufficient ATP and carbon for the necessary building blocks of the cellular processes such as nucleotides, amino acids, lipids and NADPH, cancer cells require a higher glucose intake than normal cells to meet the energy requirements for rapid proliferation. Subsequently, this overactive glycolysis may play a role in promoting tumor cell proliferation.

The alternative recombinant metabolic pattern of cancer cells was considered to provide new opportunities for cancer treatment, which lead researchers to investigate the roles of metabolic enzymes during the development of cancer. Therefore, the relationship between the metabolic changes brought by PGAM1 and cancer are gradually being explored. Hitosugi et al. found that PGMI-004A, a small molecule inhibitor of PGAM1, was able to decrease the glycolytic function of PGAM1. Subsequently, a significant decrease in the pentose phosphate pathway (PPP) flux and biosynthesis as well as an attenuated cell proliferation and tumor growth were observed in the breast cancer cell line MDA-MB-231, the lung cancer cell line H1299, the acute myeloid leukemia cell line Molm14, and in the head and neck cancer cell line 212LN. In this in-depth study, several new findings were...
discovered. First, knocking down PGAM1 led to a significant decrease in the glycolytic rate, lactate production, lipogenesis, and RNA biosynthesis and, correspondingly, cell proliferation in H1299 cells. Second, the role of PGAM1 in promoting tumor proliferation has also been shown to be modulated by intracellular levels of 3-PG and conversely by 2-PG. Third, Y26 phosphorylation of PGAM1 was found to represent a common, short-term molecular mechanism that contributed to the upregulation of PGAM1 activity and promotion of cancer cell proliferation and tumor growth. This mechanism differs from the previously described chronic mechanism in which the upregulation of PGAM1 was thought to be caused by loss of TP53. Fourth, the crystal structure of the mechanism of Y26 phosphorylation has been revealed, showing that activation of PGAM1 is enhanced by the release of inhibitory E19 that typically blocks the active site, thereby stabilizing cofactor 2,3-BPG binding and H11 phosphorylation. In addition, Engel et al. also indicated that PGAM1 activity can be inhibited by exogenous polypeptides, resulting in a decrease in the glycolytic rate and cell growth arrest in a breast cancer cell line. Although there are still many unknown factors, there is a correlation between PGAM1 and cancer proliferation. Moreover, PGAM1 is thought to affect cancer cell proliferation through the
regulation of glycolysis in the cell. In addition to the proliferation of tumor cells, tumor metastasis is also an important factor affecting the prognosis of cancer patients. DM et al. reported PGAM1 was overexpressed in the cytoplasm of capillary/artery endothelial cells, suggesting a potential correlation between PGAM1 and tumor invasion and metastasis. However, the relationship between the metabolic role of PGAM1 and tumor metastasis has been infrequently reported. It is difficult to confirm whether the mechanisms by which PGAM1 affects tumor metastasis are also achieved through glycolytic regulation.

**Non-Glycolytic Role of PGAM1 in Tumor Invasion and Metastasis**

Many studies have highlighted the metabolic role of PGAM1 in promoting cancer cell proliferation. However, it remains unclear whether PGAM1 can promote cancer malignant behaviors through a non-metabolic pathway. Previously, metabolites such as adenosine monophosphate (AMP), an allosteric activator for AMP-activated protein kinase which senses intracellular energy levels (ATP/AMP ratio), have been suggested to function as signaling molecules. Glutamine, which activates leucine uptake, leads to mTOR activation. The non-glycolytic role of PGAM1 has been recently uncovered. Hitosugi et al. found that targeting PGAM1 did not significantly influence intracellular ATP levels and showed that the decrease in ATP production caused by the attenuated glycolysis in PGAM1 knockdown cells was compensated by rescue treatment with methyl-2-PG. However, methyl-2-PG treatment only partially rescued the attenuated cell proliferation in the PGAM1 knockdown cells or cells treated with PGMI-004A, indicating that PGAM1 might contribute to cell proliferation in a 2-PG-dependent and -independent manner. The latter has been associated with the non-glycolytic function of PGAM1. The promoting role of PGAM1 on tumor metastasis has also been unveiled, but rarely related to the glycolytic functions of PGAM1. Recently, Zhang et al. confirmed that PGAM1 can promote tumor metastasis through a non-metabolic function. In this study, PGAM1 was found to directly interact with α-smooth muscle actin 2 (ACTA2) independent of its metabolic activity. To exclude the impact of the glycolytic pathway, numbers of glycolytic enzymes, such as HK2, PKM2, LDHA, and PDK1, were individually depleted in MDA-MB-231 cells. Following depletion of these enzymes, knocking down the expression of PGAM1 still reduced cancer cell motility. The PGAM1 metabolic inhibitor PGMI-004A also failed to affect cell migration in HEK 293 cells regardless of the effects of decreased PGAM1 enzymatic activity in cancer cell proliferation. This metabolism-independent role of PGAM1 in tumor invasion and metastasis has been verified through its association with ACTA2.

As well as interacting with non-glycolytic proteins, the promoting role of PGAM1 in tumor invasion and
metastasis was also found to correlate with other non-glycolytic pathways. Zhang et al.\(^\text{17}\) showed that reduced expression of PGAM1 in HN12 and Cal27 cells lead to a significant decrease in cell migration and in the expression levels of corresponding regulatory pathway molecules, such as focal adhesion kinase, the proto-oncogene c-SRC, and paxillin. Liu et al.\(^\text{36}\) also found that PGAM1 can promote migration and invasion of pancreatic cancer cells and may promote epithelial-to-mesenchymal transition (EMT) in pancreatic cancer cells by regulation of the Wnt/\(\beta\)-catenin pathway. Although the non-glycolytic function of PGAM1 was rarely described, it provided a better explanation of the mechanism by which PGAM1 modulates tumor progression, especially invasion and metastasis and led to an important new pathway for anti-cancer therapy.

### Potential Clinical Value of PGAM1 as a Target for Cancer Therapy

The role of PGAM1 in cancer progression is receiving increasing attention. Recent clinical data showed a correlation between PGAM1 and the clinical features and prognosis of cancer, suggesting that PGAM1 can be a novel potential therapeutic target. Zhang et al.\(^\text{17}\) reported that PGAM1 expression was correlated with age, lymphatic metastasis, and tumor recurrence and was closely associated with poorer overall survival (OS) and disease-free survival (DFS). PGAM1 was also suggested to be an independent risk factor for OS and DFS. It also correlated with a poor differentiation status and was identified as a potential therapeutic target for urothelial cancer by Peng et al.\(^\text{19}\) who conducted a two-dimensional electrophoresis proteomic analysis of clinical tissues. Li et al.\(^\text{49}\) found that PGAM1 was highly expressed in clear cell renal cell carcinoma and that its expression was significantly associated with age, tumor size, and TNM stage. Ren et al.\(^\text{14}\) analyzed the expression of PGAM1 in 54 paired HCC samples and 21 normal liver tissues and suggested PGAM1 as a potential diagnostic biomarker, as well as an attractive therapeutic target for HCC. Finally, Liu et al.\(^\text{36}\) found that the overexpression of PGAM1 correlated with poor prognosis in PDAC patients after analyzing 54 PDAC clinical tissues. Taken together, these clinical data have emphasized the clinical research value of PGAM1 and suggest that PGAM1 is a potential therapeutic target for the treatment of cancer.

### Summary of PGAM1 Inhibitors and Research Directions to Explore PGAM1-Targeted Drugs

Since PGAM1 was suggested as a potential therapeutic target for multiple cancer types, several PGAM1 inhibitors have been developed for cancer therapy.\(^\text{21,22,28,50–53}\) A summary of these inhibitors and their related functions are listed in Table 1. PGAM1 inhibitors are divided into pharmacological inhibitors and genetic inhibitors. The pharmacological inhibitors are small molecular compounds, with six types of small molecules reported to inhibit PGAM1, and which are mainly associated with metabolism and cancer cell proliferation. MJE3 was the first cell-permeable, small-molecule compound inhibitor of PGAM1. It reacted specifically with lysine-100 (K100) in the PGAM1 active site and hydrolyzed in situ to produce acid products that decreased breast cancer cell proliferation.\(^\text{21,22}\) The anthraquinone derivative 3, also named PGMI-004A, is another small-molecule inhibitor of PGAM1 that inhibits the conversion of 3-PG to 2-PG in cancer cells, leading to significant inhibition of the glycolytic pathway, PPP flux and biosynthesis, subsequently decreasing cancer cell proliferation and tumor growth.\(^\text{28}\) However, this inhibitor has been reported to be ineffective for tumor invasion or metastasis.\(^\text{23}\) Epigallocatechin-3-gallate (EGCG), a natural product derived from green tea, was also identified as a PGAM1 inhibitor. EGCG was reported to inhibit PGAM1 enzymatic activity by directly impairing glycolysis and PPP flux, regardless of 3-PG competition, and further, it was shown to inhibit cancer cell proliferation by modulating the intracellular level of 2-PG.\(^\text{22}\) However, because of its multiple targets, its specificity to PGAM1 is poor.\(^\text{28,54}\) Wang et al.\(^\text{53}\) used scaffold hopping and a sulfonamide reversal strategy based on the lead compound PGMI-004A to discover a series of xanthone derivatives (12a–12s) as novel PGAM1 inhibitors. These xanthone derivatives showed stronger efficacy and better specificity than PGMI-004A in the inhibition of PGAM1, as well as an increased anti-proliferative effect in the H1299 cell line. Huang et al.\(^\text{51}\) revealed that F22, K100, and R116 of PGAM1 residues were critical for the binding of inhibitors and that compound 9i, an anthraquinone inhibitor, significantly decreased lung cancer cell proliferation in different cell lines, which is a promising inhibitor for PGAM1. Moreover, in the recent research of Wen CL et al.\(^\text{55}\) an allosteric inhibitor of PGAM1 named KH3 has been explored that dramatically inhibited the proliferation of PDAC cell lines by hampering the canonical cancer metabolic pathways. In summary, inhibitors targeting PGAM1 have been developed.
rapidly. However, most of the PGAM1 inhibitors were glycolysis-targeted with minimal to no effect on the invasion and metastasis of cancer cells (Table 1).

Genetic inhibitors, unlike pharmacological inhibitors, interfere with RNA levels and appear to have increased inhibitory effects on cancer. Genetic inhibitors such as PGAM1-siRNA or shRNA proved to not only inhibit cancer cell proliferation, but also invasion and metastasis.\(^ 17,18,23,36\) Liu et al showed that following PGAM1 inhibition in PDAC cell lines, the decrease in PDAC cell invasion occurred earlier than proliferation.\(^ 16,36\) This points to the presence of an active site in PGAM1 that regulates its non-glycolytic functions and has a greater correlation with cancer metastasis. Therefore, in the future, the development of PGAM1-targeted drugs should also consider the non-glycolytic pathway. Surprisingly, in the latest study by Huang et al.\(^ 56\) reported the first allosteric PGAM1 inhibitor HKB99, which suppresses NSCLC tumor growth through ROS-dependent activation of JNK/c-Jun and metastasis by abrogating the interaction between PGAM1 and ACTA2. This discovery provides a new understanding of the function of PGAM1’s undiscovered domain. PGAM1-targeted drugs that integrate these two functions would more likely produce a more substantial effect in tumor therapy (Figure 2).

**Conclusion**

Increasing evidence has indicated the vital biological roles of PGAM1 in tumor progression. On one hand, PGAM1 is thought to be involved in the glycolytic pathway to regulate tumor cells’ metabolic pattern and promote cancer cell proliferation. On the other hand, PGAM1 can promote cancer cell invasion and metastasis through a specific non-glycolytic function. Future studies should focus on the molecular pathways modulated by PGAM1 to induce cancer cell motility during invasion and metastasis and development of drugs that can target the non-glycolytic functions of PGAM1, as its metabolic changes are mainly associated with cancer cell proliferation. The latter requires a deeper understanding of the non-glycolytic functions of PGAM1. Finally, the correlation between PGAM1 and cancer prognosis has been gaining attention and further research will identify whether PGAM1 can be used as a biomarker for early cancer detection. Although

### Table 1 Effects of Different Inhibitors of PGAM1 on Proliferation and Metastasis of Various Cancer

<table>
<thead>
<tr>
<th>Involved Organ</th>
<th>PGAM1 Inhibitor</th>
<th>Inhibit Tumor Proliferation</th>
<th>Inhibit Tumor Metastasis</th>
<th>Signal Pathway</th>
<th>References</th>
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<tr>
<td>Breast cancer</td>
<td>1) siRNA</td>
<td>+</td>
<td>+</td>
<td>ACTA2</td>
<td>[23, 28]</td>
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<tr>
<td></td>
<td>2) PGMI-004A</td>
<td>+</td>
<td>—</td>
<td>/</td>
<td>[28]</td>
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<tr>
<td></td>
<td>3) MjE3</td>
<td>+</td>
<td>/</td>
<td>/</td>
<td>[21, 22]</td>
</tr>
<tr>
<td></td>
<td>4) Xanthone derivatives</td>
<td>+</td>
<td>/</td>
<td>/</td>
<td>[53]</td>
</tr>
<tr>
<td>Glioma</td>
<td>siRNA</td>
<td>+</td>
<td>+</td>
<td>/</td>
<td>[20]</td>
</tr>
<tr>
<td>HCC</td>
<td>shRNA</td>
<td>+</td>
<td>/</td>
<td>/</td>
<td>[14]</td>
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<tr>
<td>Leukemia</td>
<td>PGMI-004A</td>
<td>+</td>
<td>+</td>
<td>/</td>
<td>[28]</td>
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<tr>
<td>NSCLC</td>
<td>1) shRNA</td>
<td>+</td>
<td>/</td>
<td>RTK/PI3K/AKT/mTOR</td>
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<td></td>
<td>2) Compound 9i</td>
<td>+</td>
<td>/</td>
<td>/</td>
<td>[51]</td>
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<td>+</td>
<td>/</td>
<td>/</td>
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<tr>
<td></td>
<td>4) EGCG</td>
<td>+</td>
<td>/</td>
<td>/</td>
<td>[52]</td>
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<td></td>
<td>5) PGMI-004A</td>
<td>+</td>
<td>+</td>
<td>/</td>
<td>[28]</td>
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<tr>
<td></td>
<td>6) HKB99</td>
<td>+</td>
<td>/</td>
<td>ROS;JNK/c-JUN; ACTA2</td>
<td>[56]</td>
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<tr>
<td>OSCC</td>
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<td>+</td>
<td>Paxillin/FAK/SRC</td>
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<tr>
<td>PCa</td>
<td>siRNA</td>
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<td>+</td>
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<td>PDAC</td>
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<td>+</td>
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<tr>
<td></td>
<td>3) KH3</td>
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<td>UBC</td>
<td>shRNA</td>
<td>+</td>
<td>/</td>
<td>/</td>
<td>[19]</td>
</tr>
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</table>

**Notes:** “+” means “positive result,” “—” means “negative result,” “/” means “not research.”

**Abbreviations:** HCC, Hepatocellular carcinoma; NSCLC, Non-small cell lung cancer; OSCC, Oral squamous cell carcinoma; PCa, Prostate cancer; PDAC, Pancreatic ductal adenocarcinoma; UBC, Urothelial bladder cancer.
there are many unsolved questions around the roles of PGAM1 in tumor malignant behaviors, increasing evidence suggests that it has become an emerging and promising target for cancer therapy and worth further investigation in the future.

**Ethical Approval**

This article does not include any experiments involving humans or animals.

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**Author Contributions**

Dr. Li was involved in the conception and design, the first draft of the article, final approval of the article, and agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of the work are appropriately investigated and resolved. Dr. Liu was involved in the conception and design, study supervision, initial drafting of the article, critical revision of the article for important intellectual content, final approval of the article, and agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of the work are appropriately investigated and resolved. All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; gave final approval of the version to be published; and agree to be accountable for all aspects of the work”.

**Disclosure**

The authors report no conflicts of interest in this work.

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