

Circular RNA circ_0001105 Inhibits Progression and Metastasis of Osteosarcoma by Sponging miR-766 and Activating YTHDF2 Expression

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Background: Circular RNAs (circRNAs) play vital roles in the modulation of tumor progression. This study explored the biological functions of circ_0001105 in the progression of osteosarcoma (OS).

Methods: qRT-PCR and in situ hybridization (ISH) were performed to detect the expression status of circ_0001105 in cells and tissues. Bioinformatics analysis, dual-luciferase reporter gene assay, Western blot and qRT-PCR were performed to determine the relationships among RNAs. The CCK-8, colony formation, EdU, transwell and wound healing assays were conducted to evaluate the cell growth, invasion and migration of OS cells. Tumor xenografts were established to investigate the effects of circ_0001105 on tumor growth in vivo. Lastly, the protein expression of YTHDF2 in OS tissues was measured using immunohistochemical staining.

Results: Data showed that circ_0001105 and YTHDF2 were significantly lower, while miR-766 was higher in OS tissues compared to adjacent tissues. Low expression of circ_0001105 or YTHDF2 was associated with poor survival of OS patients as demonstrated by the Kaplan-Meier analysis. In addition, miR-766 was identified as a direct binding target of circ_0001105 and YTHDF2. Ectopic overexpression of circ_0001105 or YTHDF2 significantly suppressed OS cell viability and invasion through regulating miR-766. Last, overexpression of circ_0001105 significantly attenuated in vivo tumor growth.

Conclusion: Our findings suggest that circ_0001105 inhibits OS progression, at least partially, by regulating miR-766/YTHDF2 signaling pathway.

Keywords: circ_0001105, OS, progression, miR-766, YTHDF2

Introduction

Osteosarcoma (OS) is the most common primary malignant bone tumor posing the biggest health threats to children and young adolescents.^{1,2} More recently, the emergence of neoadjuvant combined with cooperative treatment has dramatically improved the prognosis of localized OS. However, the prognosis of patients with advanced OS metastatic or recurrent status remains poor.³ In light of this, to improve therapeutic treatment and prognostic outcome(s), a comprehensive understanding of the genetic changes associated with OS progression is vital.

Non-coding RNAs are a highly heterogeneous group of protein-coding RNAs consisting of: (i) microRNAs (miRNAs), less than 200 nt in length; (ii) long non-coding RNAs (lncRNAs), longer than 200 nt and more recently; (iii) circular RNAs (circRNAs).⁴⁻⁶ A growing body of knowledge indicates that circRNAs are differentially

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expressed in a variety of human cancers and play important roles in the regulation of gene expression by sponging miRNAs.⁷ Unlike lncRNAs and miRNAs, circRNAs are evolutionally conserved, stable and enormously abundant in cells.⁵ These features make circRNAs potential biomarkers or therapeutic targets for various human diseases, especially cancers. In addition, circRNAs have been found to be involved in the tumorigenesis and the development of neoplasms, such as esophageal squamous cell carcinoma (circRNA_100876 and circ-DLG1),^{8,9} pancreatic carcinoma (circRNA_100782),¹⁰ breast cancer (circ_0008945),¹¹ lung cancer (circ_0023404, circ-SMARCA5),^{12,13} hepatocellular carcinoma (circ_0003998),¹⁴ and OS (circ_0001721, circ-LRP6).^{15,16} However, the potential roles of circRNAs in OS are yet to be fully documented yet.

This study identified a novel dysregulated expression circRNA, circ_0001105, in osteosarcoma. So far, the expression status of circ_0001105 in OS has not been explored. Hence, we analyzed the expression of circ_0001105 and characterized its correlation with the prognosis of OS patients. Likewise, the effect of circ_0001105 on the malignant biological behavior of osteosarcoma was investigated via circ_0001105/miR-766/YTHDF2 signaling pathway ([Supplementary Figure 1](#)).

Materials and Methods

Human Specimens

This study was approved and supervised by the Ethics Committee of the First Affiliated Hospital of Zhengzhou University, Zhengzhou Central Affiliated Hospital to Zhengzhou University, and The Affiliated Cancer Hospital of Zhengzhou University. In total, 120 tissue specimens and 65 surrounding non-tumorous tissues were collected from OS patients at the above-mentioned hospitals. All the patients received primary surgical treatment and preoperative/postoperative neoadjuvant therapy. Written informed consent was obtained from all patients for research purposes and ethical approval for the study was obtained from the Ethics Committee of Zhengzhou University (protocol number: OSRE-2011-082). Clinical information and follow-up records were obtained through medical records and mobile telephones. Good or poor chemotherapy response was distinguished based on a 90% necrosis cutoff. Detailed clinicopathological parameters of the 120 patients are summarized in [Table 1](#) and [Supplementary Table 1](#).

Cell Culture

MG63, U2OS, and 143b were purchased from the American Type Culture Collection (Rockville, USA) while HOBC and HFOB were purchased from Shanghai Institute for Biological Science (Shanghai, China). Cell lines were routinely grown in suitable media as recommended and maintained at 37°C in an incubation cabinet containing 5% CO₂.

Cell Transfection and Plasmid Constructs

The miR-766 mimics, anti-miR-766 or miR-NC, shRNAs targeting circ_0001105 were synthesized by Hanbio (Shanghai, China) and transfected into MG63 and U2OS cells using the LipoFiter (Shanghai, China) according to the manufacturer's instructions. RNAi sequences used in the study are all listed in [Supplementary Table 2](#) and [3](#). The YTHDF2 human sequence was obtained from reverse transcription (RT) of total RNA.

Immunohistochemistry (IHC) and in situ Hybridization (ISH) Staining

Stained paraffin-embedded immunohistochemical tissue sections were processed as previously described.¹⁷ Primary antibodies used in the present study are listed as follows: YTHDF2 (24744-1-AP, Proteintech, Wuhan, China), Ki-67 (9449, Cell Signaling Technology, MA, USA). Three areas were chosen randomly from each section for measurement. We employed in situ hybridization to investigate the expression and intracellular location of circ_0001105 in OS tissues as previously described.^{18,19}

Western Blotting

Western blotting was processed as previously described.²⁰ Primary antibodies used in the present study are listed as follows: YTHDF2 (24744-1-AP, Proteintech), GAPDH (9449, Cell Signaling Technology).

CCK-8, Colony Formation, Cell Invasion and Migration Assay

The CCK-8, colony formation, cell invasion and migration assay were performed according to the previously described method.¹⁷

Dual-Luciferase Reporter Gene Assay

Dual-luciferase reporter gene assay was processed as previously described.²⁰ Luciferase activities were measured with a dual-luciferase reporter assay system (Promega, WI, USA).

Table I Correlation of Clinico-Pathological Features with circRNA-0001105 Expression in Osteosarcoma Cohort

Clinicopathological Features	Variables	circRNA-0001105 Expression		P-value ^a
		Low Expression (n=63)	High Expression (n=57)	
Age (years)	≤18	36 (57.1%)	31 (54.4%)	0.905
	>18	27 (42.9%)	26 (45.6%)	
Gender	Female	33 (52.4%)	38 (66.7%)	0.160
	Male	30 (47.6%)	19 (33.3%)	
Primary tumor site	Tibia/femur	22 (34.9%)	24 (42.1%)	0.019
	Elsewhere	41 (65.1%)	33 (57.9%)	
Histologic subtypes	Chondroblastic	10 (12.8%)	15 (24.7%)	0.419
	Fibroblastic	27 (43.6%)	18 (34.6%)	
	Osteoblastic	21 (30.8%)	17 (32.1%)	
	Mixed	5 (12.8%)	7 (8.6%)	
TNM stage	Stage I/II	42 (66.7%)	48 (84.2%)	0.045
	Stage III	21 (33.3%)	9 (15.8%)	
Distant metastasis	Absent	25 (39.7%)	41 (71.9%)	0.001
	Present	38 (60.3%)	16 (28.1%)	
Recurrence	No	25 (39.7%)	36 (63.2%)	0.017
	Yes	38 (60.3%)	21 (36.8%)	
Tumor size	≤7.5cm	29 (46.0%)	44 (77.2%)	0.001
	>7.5cm	34 (54.0%)	13 (22.8%)	
Chemotherapy response	Good	20 (31.7%)	27 (47.3%)	0.026
	Poor	26 (41.2%)	13 (22.8%)	
	NA	17 (26.9%)	17 (29.8%)	

Note: ^aBold values indicate statistical significance, P < 0.05.

In vivo Experiments

Animal experiments were conducted as described previously.²¹ To establish the nude mice models of OS, we injected U2OS cells with control shRNA (NC) or shRNA targeting circ_0001105 (sh-circ_0001105) into the flank of the nude mice. Tumor volume, tumor weight and luciferase signal were measured. The expression of Ki-67 was measured with IHC assay. This animal experiment was approved by the Animal Ethics Committee of the First Affiliated Hospital of Zhengzhou University. In addition, Guide for the Care and Use of Laboratory Animals (8th edition) was strictly followed in the present study.

Statistical Analysis

All data are expressed as mean ± SEM. The significant difference was determined by two-tailed Student's *t* test, ANOVA test, Chi-square test, Pearson correlation and Kaplan-Meier analysis as appropriate. P<0.05 was considered statistically significant.

Results

circ_0001105 Is Correlated with Tumor Metastasis and Survival of OS Patients

The relative expression level of circ_0001105 was measured in 30 patients with OS and surrounding non-tumorous tissues using qRT-PCR. Notably, circ_0001105 was remarkably upregulated in OS tissue (Figure 1A and B). Meanwhile, circ_0001105 level was also higher in OS cell lines than that seen in normal human osteoblast cell lines (Figure 1C). Subsequently, we determined circ_0001105 expression in OS TMA using the ISH assay, and the results revealed the elevated circ_0001105 expression level in OS (Figure 1D and E). In addition, circ_0001105 was observably highly expressed in advanced clinical TNM stage OS tumors (Figure 1F). Additionally, circ_0001105 expression was higher in metastatic tumors (Figure 1G), recurrent tumors (Figure 1H) and was associated with poor response to chemotherapy (Figure 1I) OS tissues. To further understand the significance of circ_0001105 in OS, we determined the

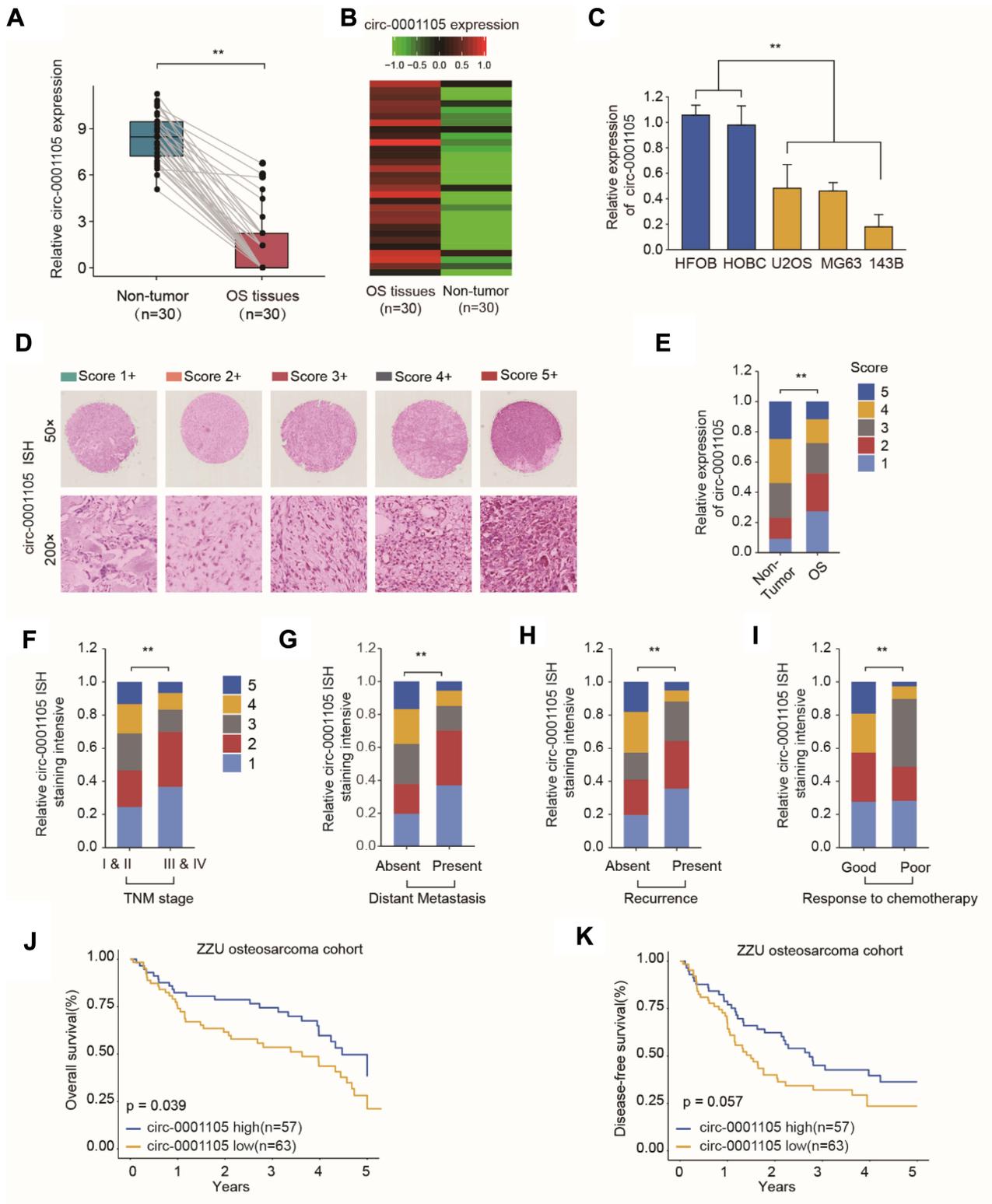


Figure 1 Circ_0001105 is down-expressed in OS tissues and cell lines. **(A, B)** Comparison of circ_0001105 expression between 30 patients' OS tissues and adjacent normal tissues. **(C)** Expression level of CIRC_0001105 in OS cell lines (MG63, U2OS, and 143B) and normal osteoblast cells HFOB and HFOB. **(D)** Representative circ-0001105 staining patterns via ISH assays. **(E)** circ_0001105 expression was significantly lower in OS tissues (n = 120) compared with that in non-tumor tissues (n = 65). The correlation of circ_0001105 expression level with TNM stage **(F)**, distant metastasis **(G)**, recurrence status **(H)** and response to chemotherapy **(I)**. Kaplan-Meier analysis revealed that down-regulated circ_0001105 was related to poorer overall survival **(J)** and disease-free survival **(K)** of OS patients. **p < 0.01. Data were shown as Mean ± SD; unpaired Student's t-test or Kaplan-Meier analysis.

potential associations between circ_0001105 expression and the patients' clinicopathological features in cohort of OS TMA cohort containing 120 OS tissues. The 120 OS patients were classified into two groups based on the circ_0001105 ISH staining intensive: (i) high-circ_0001105 group (n=57), and (ii) low-circ_0001105 group (n=63). Kaplan-Meier analysis showed that a lower circ_0001105 expression in the tumor tends to confer a significantly poor prognosis (Figure 1J and K). Moreover, univariate and multivariate Cox proportional hazards analyses confirmed that circ_0001105 level was an independent prognostic factor in patients with OS (Table 2). These findings indicated that low expression of circ_0001105 predicted favorable prognosis of patients with OS.

The Effect of circ_0001105 on the Regulation of OS Cell Proliferation and Invasion

To investigate the function of circ_0001105 in OS cells, we, respectively, carried out gain-function experiments by overexpression circ_0001105 in OS cells (Figure 2A). The ectopic overexpression of circ_0001105 significantly reduced cell proliferation in U2OS and MG63 cell lines as revealed

by the CCK-8 assays and colony formation assay (Figure 2B and C). Overexpression of circ_0001105 suppressed the DNA synthesis rate when compared with the negative control (Figure 2D). Furthermore, cell migration and transwell assays showed that overexpression of circ_0001105 significantly decreased the migration and invasion of U2OS and MG63 cells compared with the NC groups, respectively (Figure 2E and F). Overexpression of circ_0001105 also decreased the levels of the metastasis-associated protein MMP-2, MMP-7 and MMP-9 (Figure 2G). These results indicate that circ_0001105 regulates the proliferation and invasion of OS cells.

circ_0001105 Knockdown Suppresses Tumorigenesis in a Xenograft Model

Based on the in vitro results, we postulated that circ_0001105 may suppress tumor growth in animals. To test this hypothesis, U2OS cells stably overexpressing circ_0001105 or negative control cells were subcutaneously injected into the flanks of nude mice. As predicted, we observed that the mice in circ_0001105 overexpression had decreased luciferase photon flux than the negative control group (Figure 3A and B). Moreover,

Table 2 Correlation of Clinic-Pathological Features with circRNA-0001105 Expression in OS Cohort

	Univariate Analysis			Multivariate Analysis		
	HR	95% CI	P value	HR	95% CI	P value
Univariate and Multivariate Analysis of Overall Survival in OS Patients (n=120)						
Age (>18 vs ≤18)	1.167	0.906–1.463	0.188			
Gender (male vs female)	1.017	0.882–1.022	0.256			
Primary tumor site (tibia/femur vs elsewhere)	0.905	0.721–1.081	0.376			
TNM stage (III/IV vs I/II)	3.584	2.473–4.786	0.001	3.074	2.47–3.804	0.005
Distant metastasis (present vs absent)	3.081	2.989–4.326	0.005	3.245	1.914–4.752	0.016
Recurrence (present vs absent)	3.050	2.303–3.377	0.011	2.726	2.393–3.055	0.003
Tumor size (>7.5cm vs ≤7.5cm)	1.604	1.279–2.017	0.067			
Chemotherapy response (poor vs good)	2.484	1.988–2.807	0.022	1.769	1.582–2.031	0.098
circRNA-0001105 expression (low vs high)	2.876	2.290–3.540	0.012	2.665	2.137–3.068	0.018
Univariate and Multivariate Analysis of Disease-Free Survival in OS Patients (n=120)						
Age (>18 vs ≤18)	1.047	0.783–1.247	0.315			
Gender (male vs female)	0.970	0.878–1.155	0.375			
Primary tumor site (tibia/femur vs elsewhere)	0.913	0.75–1.144	0.283			
TNM stage (III/IV vs I/II)	3.256	2.479–3.651	0.023	3.325	1.968–4.579	0.011
Distant metastasis (present vs absent)	2.732	2.008–3.196	0.019	2.981	2.466–3.887	0.025
Recurrence (present vs absent)	2.219	2.058–2.717	0.016	2.295	1.895–3.556	0.035
Tumor size (>7.5cm vs ≤7.5cm)	1.580	1.385–1.895	0.089			
Chemotherapy response (poor vs good)	2.127	1.904–2.555	0.040			
circRNA-0001105 expression (low vs high)	2.595	2.127–3.097	0.016	2.565	1.985–3.458	0.020

Note: Bold values indicate statistical significance, P < 0.05.

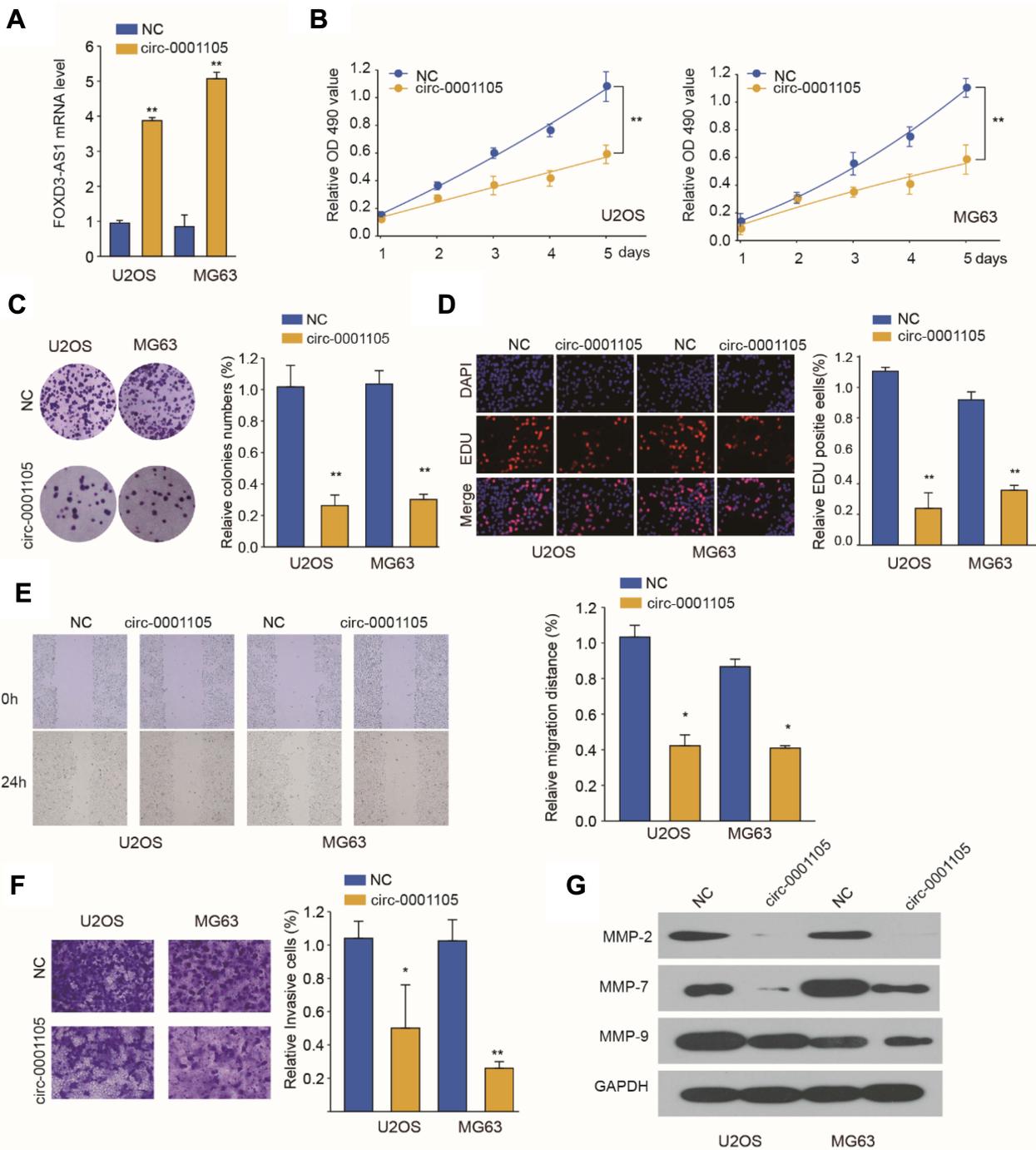


Figure 2 Ectopic overexpression of Circ_0001105 suppresses OS cell proliferation and invasion in vitro. **(A)** Circ_0001105 was stably overexpressed in U2OS and MG63 cells. **(B)** CCK-8 assays were performed to assess the viability of U2OS and MG63 cells after Circ_0001105 overexpression. **(C)** colony formation assays and **(D)** EdU staining assays were performed to determine the growth abilities of U2OS and MG63 cells. **(E)** Migration and invasion **(F)** assay of U2OS and MG63 cells after Circ_0001105 overexpression. **(G)** MMP-2, MMP-7 and MMP-9 were analyzed by Western blot. *P < 0.05, **P < 0.01; Data are shown as mean ± SD; Representative images and data are based on three independent experiments.

tumors extracted from mice injected with cells overexpressing circ_0001105 were lighter than mice in the control group (Figure 3C). Meanwhile, the rate of tumor growth was slower in the circ_0001105 overexpression group than

in the control group (Figure 3D). Furthermore, overexpression of circ_0001105 showed lower Ki-67 IHC staining intensity in the xenografted tissues relative to the control group (Figure 3E). The above results in vitro and in vivo

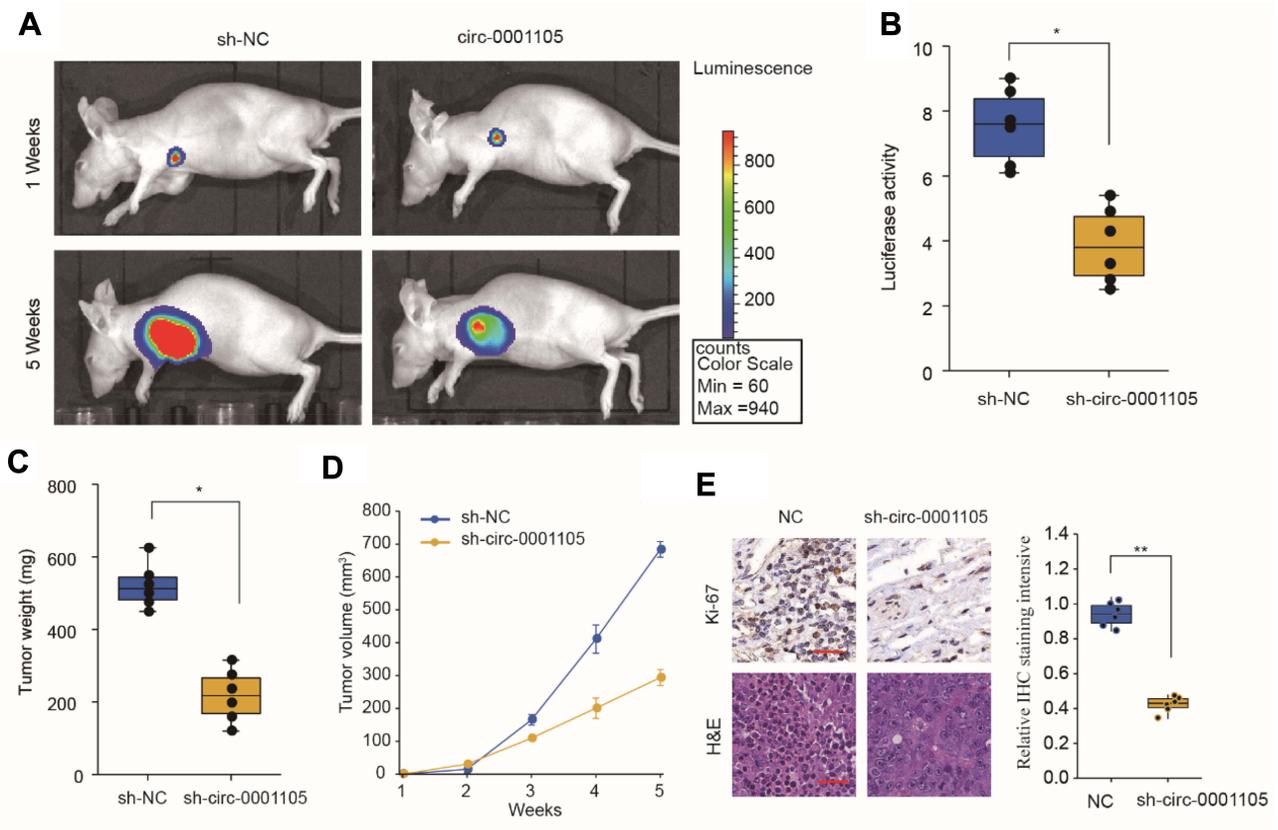


Figure 3 Ectopic overexpression of CIRC_0001105 suppresses OS cell proliferation in vivo. **(A, B)** Tumor growth was quantified by bioluminescence intensity. **(C)** Tumor weight of xenograft tumors. **(D)** Tumor growth curves of Circ_0001105 overexpression and control transfected U2OS cells in nude mice. **(E)** Xenograft tumor tissues from circ_0001105 overexpression group displayed much weaker staining of Ki-67. * $P < 0.05$, ** $P < 0.01$; Data are shown as Mean \pm SD; unpaired Student's t-test.

results further illustrate the crucial role of circ_0001105 in OS cell growth and invasion. However, the exact underlying mechanisms remain unclear.

circ_0001105 Is as a miRNA Sponge for the miR-766

Given that circRNA acts as a miRNA sponge, we explored whether circ_0001105 could bind to miRNAs. The binding sequences between circ_0001105 and miR-766 were predicted using online tools (<https://circinteractome.nia.nih.gov/>) (Figure 4A). We also found that miR-766 was evidently up-regulated in OS tissues and cell lines (Figure 4B and C). Additional, as shown in Figure 4D and E, overexpression of circ_0001105 significantly inhibited miR-766 expression, while circ_0001105-depletion dramatically increased miR-766 expression level. Moreover, the direct interaction between miR-766 and circ_0001105 was further corroborated through the luciferase reporter assay (Figure 4F). Pearson correlation analysis suggested that circ_0001105 was negatively correlated with miR-766 expression in OS tissues (Figure 4G).

To further clarify whether miR-766 was directly involved in the circ_0001105-mediated anti-proliferation effect of OS, U2OS stably overexpressing circ_0001105 were transiently transfected with miR-766 mimics. The cell viability and colony formation ability of U2OS in the miR-766 mimics group were significantly increased, while induction of circ_0001105 could abolish the role of miR-766 in promoting cell growth (Figure 5A–C). Consistently, the enhanced cell invasion following overexpression of miR-766 was reversed by co-expression of circ_0001105 (Figure 5D–E). Together, these data indicated that circ_0001105 suppresses OS progression by targeting miR-766.

YTHDF2 as the Downstream Functional Target Gene of circ_0001105/miR-766 Axis

Online tools (<http://starbase.sysu.edu.cn>) were utilized to identify the targets of miR-766, and we placed emphasis on the YTHDF2 (Figure 6A). First, dual-luciferase reporter assay

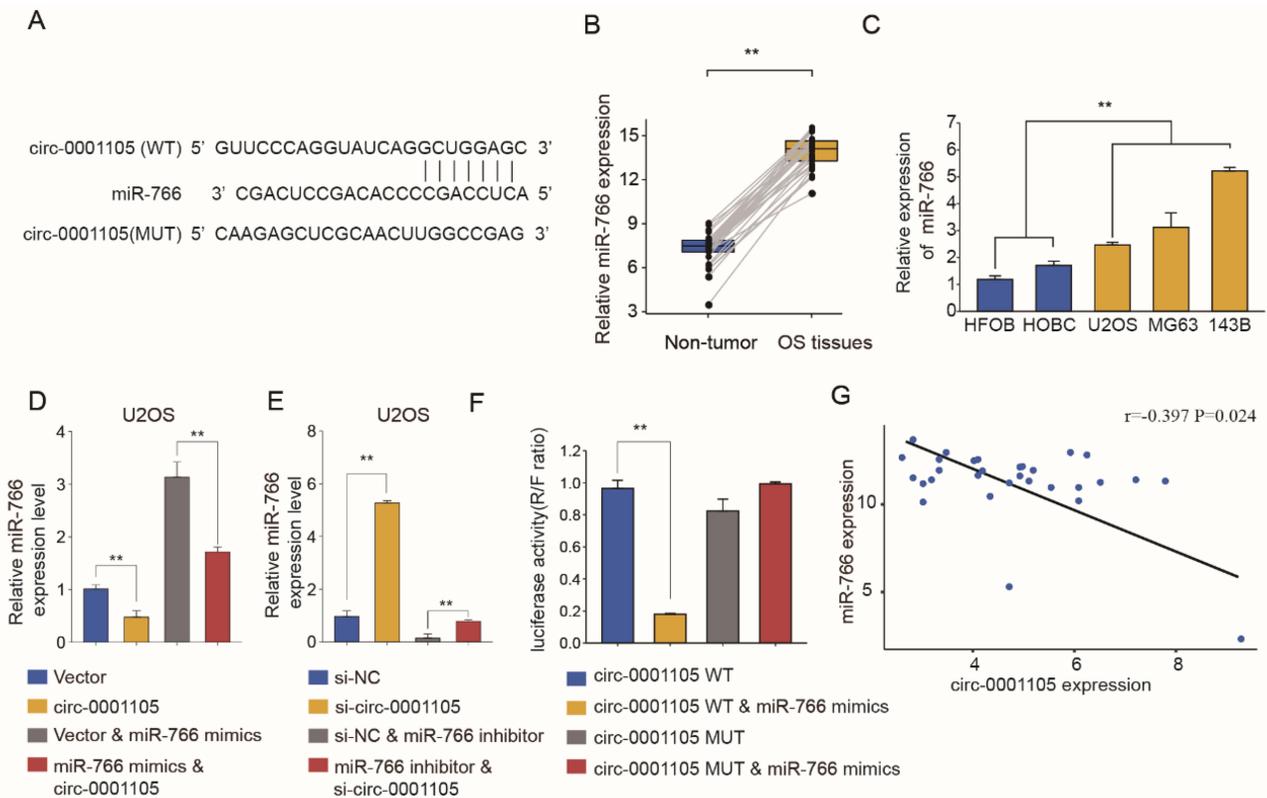


Figure 4 The interaction between Circ_0001105 and miR-766. **(A)** The putative binding sites of miR-766 on Circ_0001105 are predicted. **(B)** RT-qPCR analysis of miR-766 expression levels in OS tissues (n=30) compared with corresponding adjacent normal tissues (n=30). **(C)** MiR-766 expression levels of OS cell lines (U2OS, MG63 and 143B), compared with that in human osteoblast cells (HFOB and HOBC). qRT-PCR analysis of miR-766 expression levels in U2OS and MG63 cells after Circ_0001105 overexpression **(D)** or silencing **(E)**. **(F)** Dual-luciferase reporter assay showed the luciferase activity of the combination between miR-766 and Circ_0001105. **(G)** The correlation between miR-766 and circ_0001105 expression in OS tissues. $**p < 0.01$. Data are shown as Mean \pm SD; Representative images and data are based on three independent experiments.

was conducted to confirm the target relationship between YTHDF2 and (Figure 6B). Moreover, YTHDF2 expressions were detected using qRT-PCR after altering miR-766 expression in U2OS and SAOS cells. Interestingly, the mRNA levels of YTHDF2 were significantly decreased in miR-766 overexpression group but were downregulated in the miR-766 knockdown group (Figure 6C and D). Additionally, Pearson analysis showed a negative correlation between YTHDF2 and miR-766, but a positive correlation between YTHDF2 and circ_0001105 in OS samples (Figure 6E and F). Furthermore, Western blot assay showed that protein levels of YTHDF2 were significantly increased in the circ_0001105 overexpression group but were downregulated when transfected with miR-766 mimic. The co-transfected with circ_0001105 and miR-766 mimic reversed the inhibitory effects of miR-766 on YTHDF2 expression (Figure 6E and F).

To further figure out the role of circ_0001105/miR-766/YTHDF2 axis in tumor progress, we knockdown YTHDF2 expression in circ_0001105-overexpressed U2OS and SAOS cells. Functional experiments revealed that YTHDF2 silencing

significantly rescued OS cell proliferation invasion impaired by circ_0001105 overexpression (Figure 7A and B). Additionally, we checked the expression levels of YTHDF2 and miR-766 in tumor tissues from nude mice injected with cells overexpressing circ_0001105. We found that the levels of YTHDF2 were higher in tumor tissues of the circ_0001105 overexpression group than that in the control group, while the miR-766 level was lower (Figure 7C). Moreover, IHC analysis also indicated that there were higher protein levels of YTHDF2 in tumor tissues of the circ_0001105 overexpression group (Figure 7D). Taken together, our results demonstrated that circ_0001105 inhibitor OS cell proliferation and invasion in a miR-766/YTHDF2 axis-dependent manner.

Correlation Between YTHDF2 Expression and Clinical Characteristics of OS

Pan-cancer analysis based on the TCGA database showed that YTHDF2 expression is frequently dysregulated in various cancers (Supplementary Figure 2). To further determine the expression status and prognostic value of YTHDF2 in OS, the

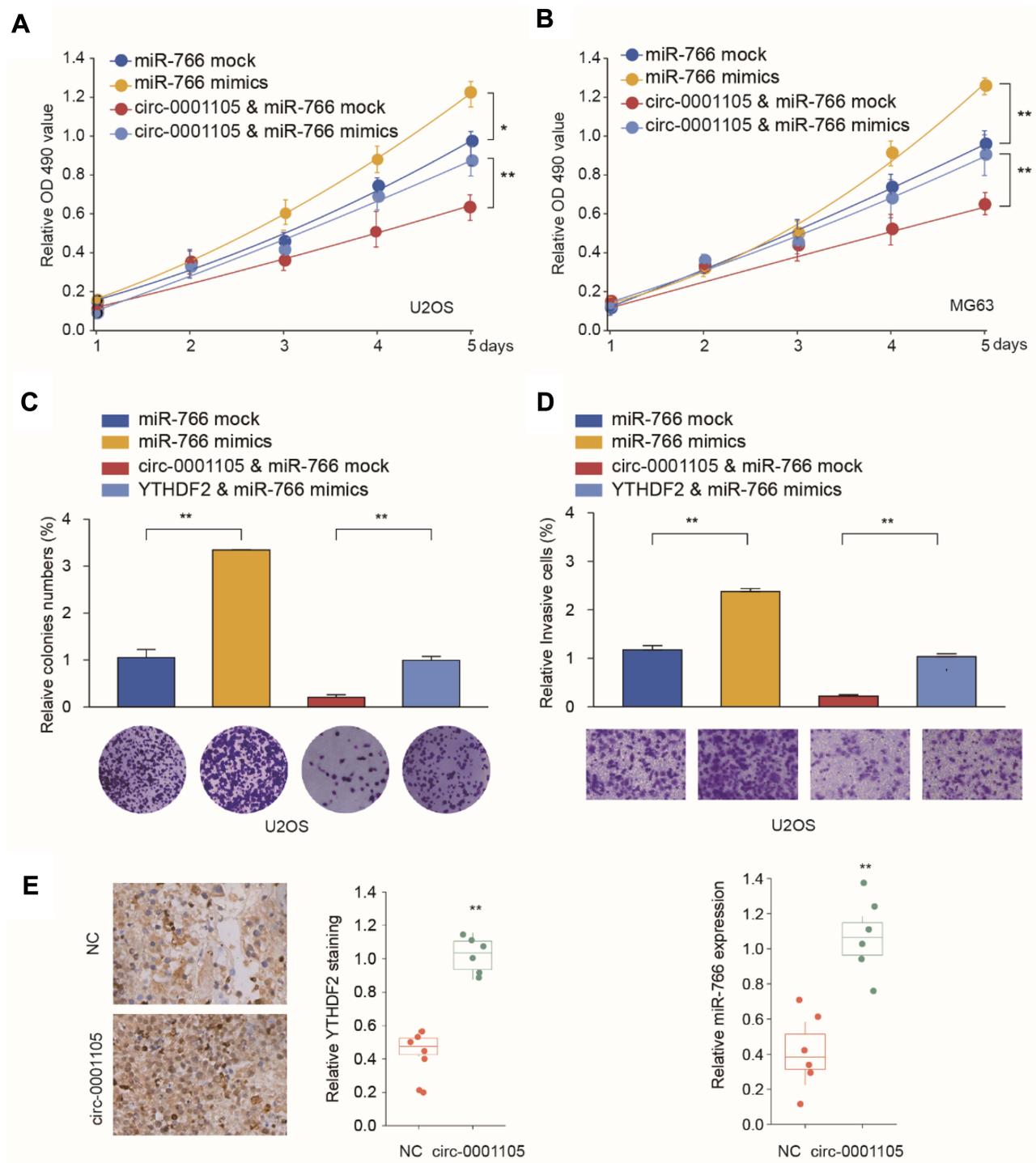


Figure 5 (D-E) Overexpression of Circ_0001105 partially reverses the promote effect of miR-766 in OS cells. U2OS and MG63 cells were transfected with miR-766 mock or miR-766 mimics, with or without Circ_0001105 overexpression vector. **(A, B)** Cell proliferation of U2OS and MG63 was analyzed at indicated time points by CCK-8 kit. Colony formation **(C)** and cell invasion ability **(D)** of U2OS were analyzed by colony formation assay and transwell assay, respectively. Xenograft tumor tissues from circ_0001105 overexpression group displayed much stronger staining of YTHDF2 **(E)**, and lower miR-766 expression **(F)**. * $p < 0.05$, ** $p < 0.01$. Data are shown as Mean \pm SD; Representative images and data are based on three independent experiments.

relationship between YTHDF2 expression level and the clinicopathological characteristics was analyzed using a TMA cohort. The results showed that YTHDF2 was significantly

downregulated in OS tissues (Figure 8A and B). In addition, low expression of YTHDF2 was significantly correlated with clinically aggressive tumor phenotypes, such as advanced

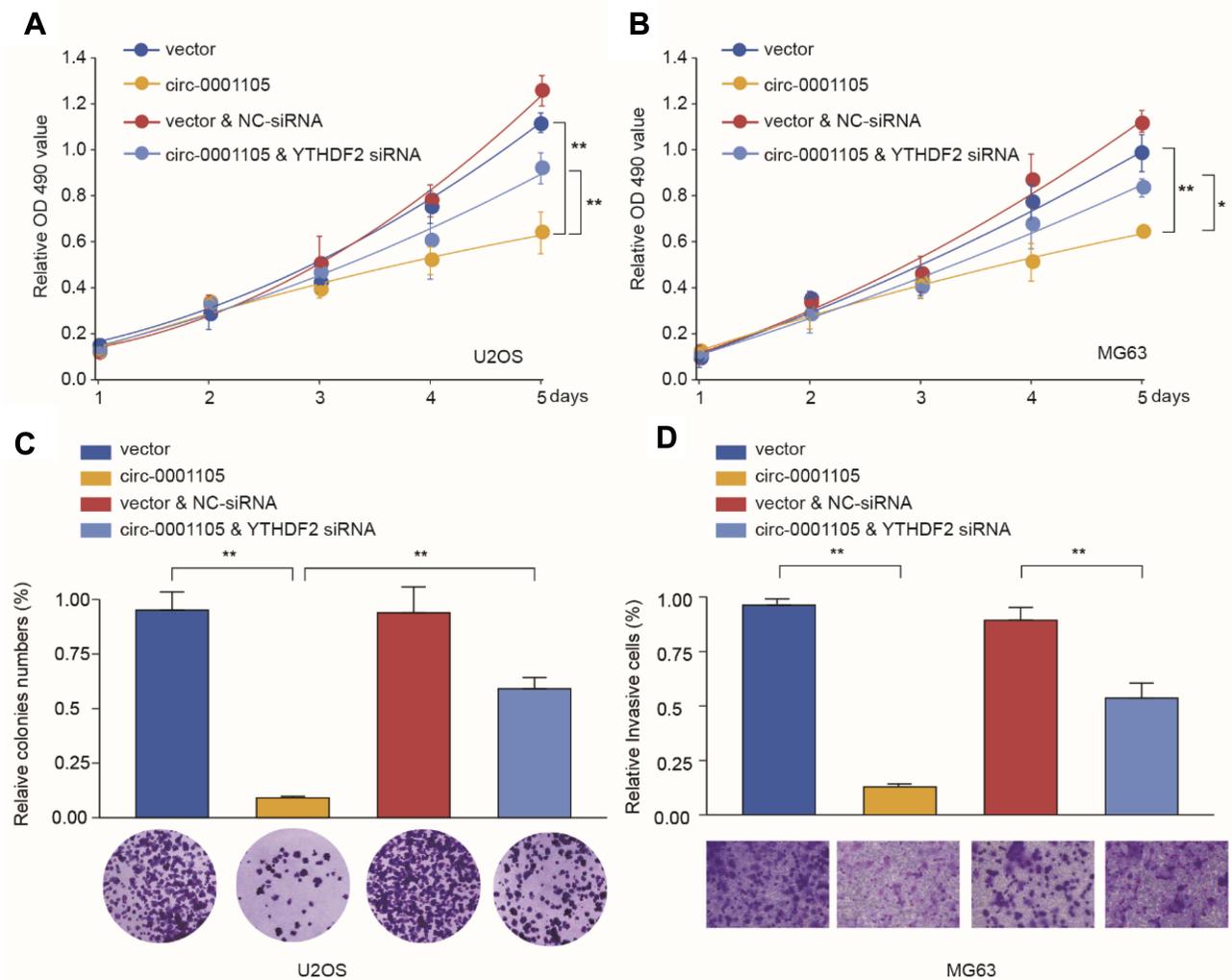


Figure 7 Knockdown of YTHDF2 partially reverses the suppressive effect of Circ_0001105 in OS cells. U2OS and MG63 cells were transfected with NC or Circ_0001105 overexpression vector, with or without siRNA targeting YTHDF2. (A–B) Cell proliferation of U2OS and MG63 was analyzed at indicated time points by CCK-8 kit. Colony formation (C) and cell invasion ability (D) of U2OS was analyzed by colony formation assay and transwell assay, respectively. * $P < 0.05$, ** $P < 0.01$; Data are shown as Mean \pm SD; Representative images and data are based on three independent experiments.

Given that circ_0001105 is downregulated in OS tissues, we tested the role of circ_0001105 in OS cells via gain-of-function in vitro and in vivo experiments. As we expected, cell proliferation and invasion potentials were strikingly impeded following circ_0001105 overexpression, indicating that circ_0001105 was a tumor suppressor in OS cells. Similarly, up-regulation of circ_0001105 significantly suppressed tumor growth in OS xenograft model. Overall, circ_0001105 might act as a tumor suppressor and contribute to the tumorigenesis and progression of OS. Thus, circ_0001105 holds huge promise as a promising therapeutic target for OS.

Previous studies have reported that circRNAs may regulate gene expression by serving as miRNA sponges.²⁷ To further explore the molecular mechanism of circ_0001105

involving in the progression of OS, we assessed whether that circ_0001105 could bind to miR-766. Jia et al found that miR-766 expression was increased in colorectal cancer and could function as an oncogene.²⁸ Similarly, Li et al demonstrated that miR-766 promoted cell proliferation of human colorectal cancer by inhibiting the expression of SOX6.²⁹ However, the expression status and biological function of miR-766 in OS are largely unknown. Similar to the results observed in colorectal cancer, we found miR-766 was significantly upregulated in OS tissues and cell lines. Functional experiments demonstrated that miR-766 mimics partly reversed the prompt-proliferation and invasion effect caused by hsa_circ_0001105 overexpression. These findings suggest that miR-766 may function as an oncogene in OS. Opposite findings were observed in lung cancer and hepatocellular

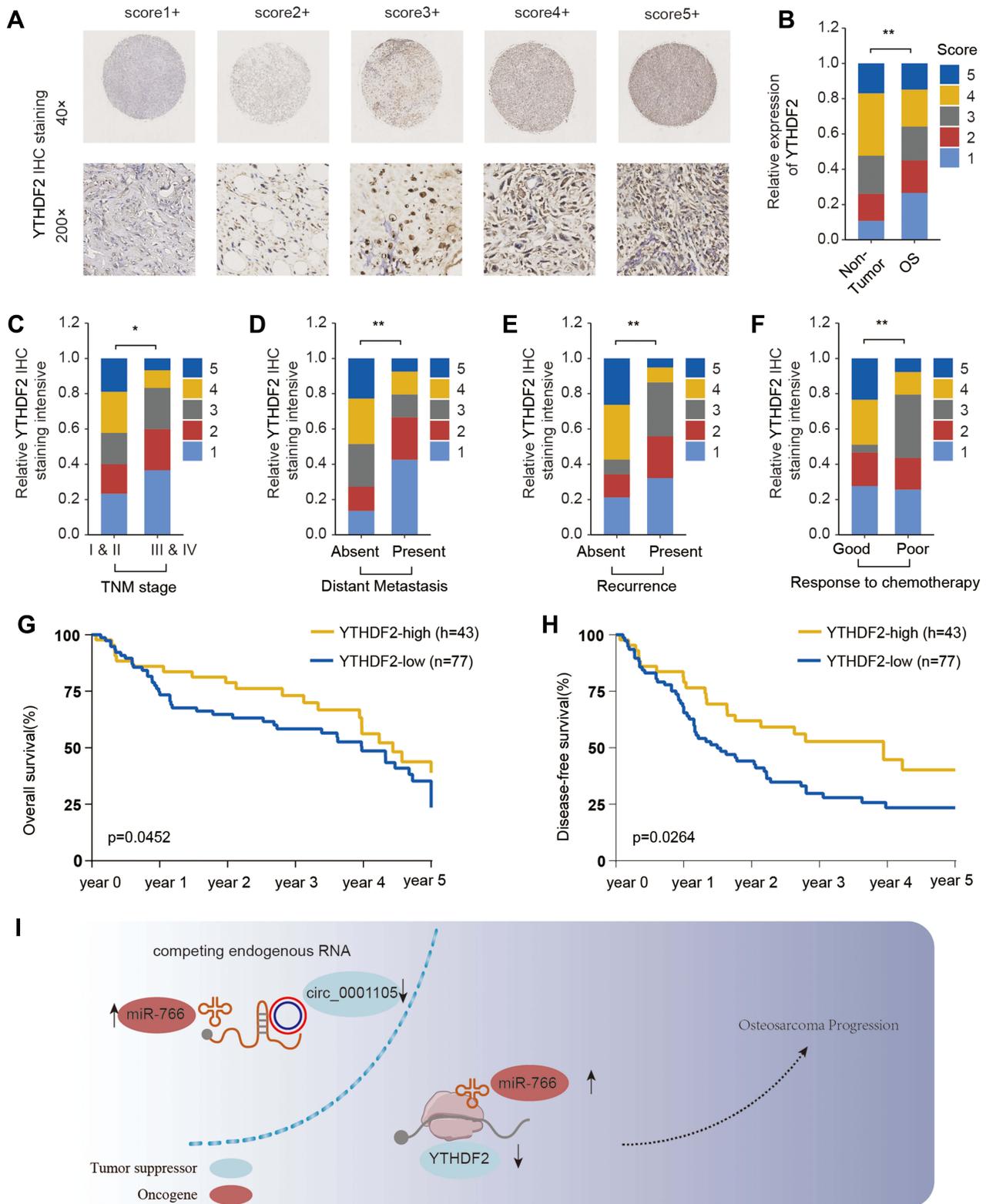


Figure 8 YTHDF2 is downregulated in OS tissues and predicts poor prognosis. **(A)** Representative YTHDF2 immunohistochemical (IHC) staining patterns with different staining scores in OS. **(B)** Representative YTHDF2 IHC staining in OS tissues and adjacent non-tumor normal tissues. The correlation of YTHDF2 expression level with TNM stage **(C)**, distant metastasis **(D)**, recurrence status **(E)** and chemotherapy response **(F)**. Kaplan-Meier analysis of overall survival **(G)** and disease-free survival **(H)** in osteosarcoma patients with high or low YTHDF2 expression (n=83). **(I)** A mechanism diagram depicting that the circ_0001105/miR-766/YTHDF2 axis affects the progression of OS. *P < 0.05, **P < 0.01.

carcinoma, which indicates the tumor-suppressor role of miR-766.^{30–32} in cancer vary with the cell type or conditions. Thus, the function role of miR-766 needs to be further investigated to draw precision conclusions.

miRNAs exert their biological functions through post-transcriptional regulating gene expression. Our findings show that YTH N6-Methyladenosine RNA Binding Protein 2 (YTHDF2) is a functional target of miR-766, while miR-766 inhibits the progression of OS by targeting YTHDF2. YTHDF2 is an m6A reader protein that promotes mRNA degradation by targeting m6A-mRNAs to the processing bodies in the cytoplasm.³³ It has been reported that YTHDF2 plays dual roles in cancer. YTHDF2 could suppress cell proliferation in acute myeloid leukemia and hepatocellular carcinoma.^{34,35} Opposite results were observed in gastric cancer, prostate cancer and lung cancer.^{36–38} Here, we report that YTHDF2 is downregulated in OS tissues and inhibited OS cell growth and invasion viability. Besides, miR-766 overexpression or hsa_circ_0001105 silencing counteract the effects of YTHDF2. Moreover, the expression level of YTHDF2 was significantly up-regulated, whereas miR-766 was distinctly decreased after hsa_circ_0001105 overexpression. Our findings suggest that hsa_circ_0001105 may act as a ceRNA (competing endogenous RNA) of miR-766 to relieve the repressive effect of miR-766 on its target YTHDF2.

Despite these results, there are few limitations and shortcomings in the current study. First, one gene may be targeted and regulated by multiple molecules; other circRNA/miRNAs might also contribute to the osteosarcoma progression through regulating YTHDF2. Secondly, YTHDF2 is an m6A regulator. Whether YTHDF2 affects the progression of osteosarcoma through an m6A modification pattern remains to be clarified.

Taken together, this study identified the decreased expression of circ_0001105 in OS tissues. A functional study revealed that circ_0001105 suppresses the progression of OS by modulating the circ_0001105/miR-766/YTHDF2 axis. Although the functional roles and molecular mechanisms of circRNAs remain largely unclear, our study underscores the role and therapeutic potential of the circ_0001105/miR-766/YTHDF2 in OS.

Disclosure

The authors report no conflicts of interest in this work.

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