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ORIGINAL RESEARCH

# Antifungal Activity of ZnO Nanoparticles and Nystatin and Downregulation of SAPI-3 Genes Expression in Fluconazole-Resistant *Candida albicans* Isolates from Vulvovaginal Candidiasis

This article was published in the following Dove Press journal: Infection and Drug Resistance

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**Purpose:** Antifungal resistance and virulence properties of *Candida albicans* (*C. albicans*) are growing health problems worldwide. The present study aims to investigate the effect of Zinc Oxide (ZnO) nanoparticles and Nystatin on SAP1-3 genes expression in *C. albicans* isolates of females with Vulvovaginal Candidiasis (VVC) isolated from Sayad Shirazi Obstetrics and Gynecology Hospital in Northeastern Iran during 2017–2018.

**Patients and Methods:** In this descriptive-analytic study, vaginal samples were collected from 280 VVC women. 196 (70%) of *C. albicans* isolates were identified by phenotypic and ITS genotypic methods. Susceptibility to Fluconazole *C. albicans* isolates was determined by the disk diffusion method. Detection of ERG11 gene was done by RT-PCR technique.

**Results:** It was revealed that PCR amplified the ERG11 gene in all of the Fluconazole-resistant isolates. Real-time PCR was used to survey the effects of  $3\pm1.7\mu$ g/mL concentrations of ZnO nanoparticles and Nystatin on expression of SAP1-3 genes before and after treatment. 186 (95%) susceptible *C. albicans* and 10 (5%) Fluconazole-resistant *C. albicans* isolates from VVC were exposed to sub-minimum inhibitory concentrations (Sub-MIC) of ZnO-np (range=0.02–12 µg/mL). Sub-MIC concentration was used for each strain, which reduced the expression of SAP1-3 genes to 1.8 MIC in the vaginal swabs. The observed reduction in gene expression was significant for both ZnO nanoparticles and Nystatin (P=0.01 and P=0.07, respectively).

**Conclusion:** ZnO as antifungal agent can well reduce the growth and gene expression of SAP1-3 in the pathogenesis of VVC.

Keywords: ZnO, gene expression, pathogenesis, vulvovaginal candidiasis

#### Introduction

Vulvovaginal candidiasis (VVC) is one of the most common gynecologic infections in women that are seen daily in clinics.<sup>1</sup> In 85–95% of patients, vaginal fungal infections are caused by *Candida albicans* (*C. albicans*), and in the rest, *Candida glabrata* and *Candida tropicalis* are the main causes of the disease.<sup>2</sup>

Increasing the resistance of *C. albicans* to antifungal agents, mainly to azoles, is a serious health problem and prevents VVC treatment. Resistance of candida species against antifungal agents can infect approximately 5% of women at least four times a year.<sup>3–5</sup> The causes of the increasing incidence of VVC could be low-dosage azole maintenance regimens and the use of antifungal agents such as Fluconazole without a prescription.<sup>6–8</sup>

Infection and Drug Resistance 2020:13 385-394

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© 2020 Hosseini et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms. work you hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission foro Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, is see aparagraphs 4.2 and 5 of our Terms (http://www.dovepress.com/terms.php). Predisposing factors for VVC include reproductive age, use of contraceptive steroid medications and bacterial antibiotics, and being diabetic, pregnant, or immunocompromised. Increased Fluconazole-resistance is nowadays one of the most important problems in the long-term treatment of recurrent VVC. Resistance to Fluconazole may occur through various mechanisms including alphademethylase in sterol biosynthesis that enhances the expression of ERG11 gene.

Overexpression of ER11 gene results in the production of a large amount of Lanosterol 14 $\alpha$ -demethylase; this helps to continuous synthesis of ergosterol and maintaining the cell wall integrity that can cause the candida be resistant to Fluconazole.<sup>6–9</sup> This type of resistance has been associated with widespread and continuous usage of Fluconazole as prophylaxis.<sup>6</sup> Also, mutations in the ERG11 gene exist and contribute to resistance in about 65% of Fluconazole-resistant candida species.<sup>6,10</sup> Among the Fluconazole-resistant isolates, mutations in the ERG11 gene among the candida species are responsible for significant differences in their susceptibility to Fluconazole.<sup>11</sup>

Secreted Aspartyl Proteinases (SAP) are considered as a potential factor that may affect the virulence of this yeast. Extracellular proteolytic activity of candida species is caused by SAP. As the best member of the hydrolytic enzyme group, SAP play an important role in adhesion, colonization, penetration and tissue attack by destroying cell membranes and host surface molecules.<sup>12</sup> SAP are coded by 10 families of the SAP genes; all are effective in *C. albicans* pathogenesis.<sup>13,14</sup> According to previous studies, SAP1-3 genes are involved in mucosal infections. SAP1 and SAP3 play important roles in the adhesion of *C. albicans*. SAP2 plays an important role in *C. albicans* colonization at vaginal mucosal surface and development of *C. albicans* pathogenesis.<sup>15</sup> Therefore, inhibition of SAP has a protective effect on the host.<sup>16</sup>

Nystatin is one of the drug combinations used to treat recurrent VVC. It belongs to the polyenes family, which changes the membrane permeability by binding to the sterols in the fungus cell membrane and causes the essential contents of the intracellular fungi to be removed. In this study, Nystatin was used for treatment of VVC infections. Then, Nanomaterials were used to inhibit the growth of VVC isolates.

The main principle of nanotechnology is the application of new compounds with nanoscale structures. Booster nanocomposites of ZnO have been studied in the last decade, remarkably. The reasons for using ZnO are its high energy bandgap, chemical and thermal stability, high oxidation energy (60mev), and good antimicrobial properties.

ZnO has been used for long years such that the Romans called it "cadmium" and used it to produce rice alloys. Metallic ions like silver and copper have traditionally been used to treat diseases.<sup>17</sup> ZnO improves durability and resistance to mildew; it has also been used in the pharmaceutical and cosmetic industries in powder and ointment forms.<sup>18</sup>

ZnO increases the permeability of the cell wall and cytoplasmic membrane through binding to lipids and proteins and the destruction of protein's 3-D structure.<sup>17</sup>

We conducted Fluconazole susceptibility testing on the clinical C. albicans isolates obtained from some young VVC women. The resistant strains and susceptible dose-dependent species were subjected to polymerase chain reaction using primers that identify ERG11 gene expression. We investigated the secretion of SAP and their activity in VVC strains. There is a clear link between the ability to produce SAP and the development of disease in Fluconazole-resistant strains.<sup>19,20</sup> In addition, we investigated the antifungal effect of ZnO nanoparticles and Nystatin on inhibiting the growth of VVC C. albicans isolates. Subsequently, the analysis of the expression of SAP1-3 genes in C. albicans isolates was performed using real-time PCR. Additionally, changes in the expression of SAP1-3 genes were determined by Real-time PCR after the addition of sub-inhibitory concentrations of ZnO-np and Nystatin onto VVC C. albicans isolates.

# **Materials and Methods**

# Sampling, Culture, and Identification of Yeasts

This descriptive cross-sectional study was carried from August 2017 to July 2018 in Sayad Shirazi teaching hospital of Golestan University of Medical Sciences. Approval for the study was granted by the Ethics Committee of Golestan University of Medical Sciences.

Inclusion criteria were vaginal swabs from the VVC women willing to participate in the study. The drug susceptibility test of Nystatin and ZnO nanoparticles in VVC *C. albicans* isolates was done by previous informed consent of the participants.

All procedures performed in studies involving human participants were in accordance with the rules on patient sampling and fully complied with the Ethics Committee (Code no.IR.Gums.rec.1397.008) and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. Written informed consent was obtained from all participants, and the questionnaire was sent to the journal as an Additional File.

A semi-structured questionnaire was administered after explaining the details of the study and obtaining their consent. The vaginal samples were obtained from 280 patients (aged 18–46 years) suffering from burning, itching, malodorous and cheesy vaginal secretions, with a confirmed diagnosis by Dacron swabs. The vaginal swab specimens were collected by the attending physician following standard procedures.<sup>3</sup>

The yeasts were first identified according to their morphological characteristics using several phenotypic techniques such as production of germ tube in human serum, culture on CHROMagar Candida (Chromagar, France), and Cornmeal agar (Sharlo, USA), containing Tween 80.<sup>10</sup> After identification of primary morphology, DNA extraction was performed using the Phenol-chloroform-isoamyl alcohol method. DNA was amplified using the ITS1 (5'- TCC GTA GGT GAA CCT GCG G-3') and ITS4 (5'- TCC TCC GCT TAT TGA TAT GC-3') primers, which were used for PCR amplification of the ITS1-5.8S-ITS2 rDNA region. Both VVC isolates and standard strain of *C. albicans* produced bands with an approximate length of 535 bp (Figure 1). Digestion of the ITS region of *C. albicans* using HpaII enzyme produced two bands of 297 and 338 bp.<sup>21</sup>

# Susceptibility Testing of Isolates to Fluconazole by Disk

The standard disk diffusion assay (Clinical and Laboratory Institute (CLSI) M44-A2) was used. In summary,  $1 \times 10^{6}$  CFU/mL yeasts were prepared for each suspension in a Buffered

Phosphate Solution (PBS) and distributed on Mull-Hinton (MH) agar with 2% glucose and  $0.5\mu$ g/mL Methylene blue. Fluconazole (Rosco, France) was used as a positive control. Fluconazole disks ( $25\mu$ g) were placed on the agar, and PBS (as negative control) was added to the blank disk. Then the M-H agar plates were incubated for 24 hrs at 37°C and the diameter of the non-growth zone was measured.<sup>22</sup> *C. albicans* isolates were reported as susceptible S (zone diameter  $\geq$ 19 mm), susceptible dose-dependent SDD (15 to 18 mm), and resistant R ( $\leq$ 14 mm). 10 (5%) isolates showed resistance to Fluconazole while 18 (9.13%) were susceptible

# Reverse Transcription-PCR (RT-PCR) Analysis of the ERGII Gene

to dose-dependent and 168 (85.71%) were susceptible.

RNA was extracted from the *C. albicans* isolates using TRIzol<sup>TM</sup> reagent (Catalog#15596026, Invitrogen, Thermo Fisher Scientific, Waltham, MA, USA) according to the manufacturer's protocol. Before the synthesis of cDNA, all samples of RNA were treated with DNase1 (Thermo Fisher Scientific Co., EN0521, USA). Synthesis kit of cDNA (Fermentase Co., K1622, USA) was utilized for the synthesis of cDNA. Primer pair with the Accession Number: XM\_711668.2, 5'- CTTGGTTTGGCAACCCCATGAGT -3' (forward) and 5'- ACATTGGCAACCCCATGAGT -3' (reverse) was designed to amplify an expected PCR product of 397 bp, which includes the complete ERG11 gene sequence. The PCR was performed in a Thermocycler (Eppendorf, Germany). The system was programmed to 2 min of denaturation at 92°C followed by 30 cycles, each consisting of 1 min of



Figure I Identification of VVC C. albicans isolates.

denaturation at 92°C, 1 min of annealing at 43°C, and 1 min of extension at 72°C. At the final cycle, an additional 10 min of incubation at 72°C was performed for a complete extension. For each PCR run, a negative control was also included containing the reaction buffer, dNTPs, Taq polymerase without the target DNA. Reference strain *C. albicans* ATCC10231 was included in each run. The ERG11 gene was detected by electrophoresis on a 2% agarose gel performed at 70 V for 2.5 hrs. The gel was stained with ethidium bromide (0.5 µg/mL) and photographed under UV light with a Photo documentation system (GenoSens 1500). A DNA ladder of 100 bp (Promega, USA) was used as molecular weight marker.<sup>23</sup>

# Antifungal Drug Susceptibility Testing of ZnO and Nystatin on VVC *C. albicans* Isolates

ZnO was synthesized by the sole gel method<sup>24</sup> was prepared in 1mL distilled water, and Nystatin (1477003, Sigma Aldrich) was prepared in 1 mL DMSO 10%. The inhibitory effects of ZnO and Nystatin were tested on the surface of 96-well microplates using the CLSI M27-A3 method. Absorption was measured after incubation for 48 hrs at 30°C. MIC (Minimum Inhibitory Concentration) was determined by comparing it with the control group without the drug. For ZnO and Nystatin, MIC was determined by a significant reduction in growth after 48 hrs of incubation. MIC was considered to be the lowest concentration in which growth was reduced. To ensure non-growth, a negative control was considered for each isolate.<sup>23</sup>

# Effect of ZnO Nanoparticles and Nystatin on the Growth of VVC *C. albicans* Isolates Using the MTT

The effect of ZnO nanoparticles and Nystatin on the growth of *C. albicans* isolates was assessed according to the CLSI-M27-A3 guidelines.<sup>19</sup> Briefly, *C. albicans* isolates from the overnight-grown cultures in YPD broth were harvested by centrifugation and adjusted to a final concentration of  $1 \times 10^6$  cell/mL in RPMI-1640 medium. 100 µL of the cell suspension was added to each well of the 96-well microplates (Thermo Scientific Nunc Microwell). ZnO-np dissolved in water was added to the wells to achieve a two-fold series of concentrations of ZnO, ranging from 0.02 µg/mL to 256 µg/mL. The wells containing the same volume of water were set as negative controls, while the wells treated with 8 µg/mL Nystatin were served as blank controls. After incubation at 37°C for 24 hrs, the inhibitory effect of ZnO was assessed by visual

inspection. Then, 10  $\mu$ L of sterile MTT solution (5mg/mL in PBS) was added to each well of the microplates, followed by another incubation at 37°C in dark for 4 hrs. The supernatant in each well was discarded, and then 100 mL of DMSO water-soluble supernatant was added to each well.

An Eliza Reader was used to capture the optical absorption of the 96-well microplates (Nank, Germany) (OD) at 570 nm. The vitality of the fungal cells in each well was calculated as a percentage of surviving cells = $(1-X/C)\times100$ 

X:OD570 treatment, C: OD570 control. All assays were performed in triplicate.<sup>25</sup>

# Analysis of SAPI-3 Genes Expression Level in VVC *C. albicans* Isolates

Analysis of the SAP1-3 genes expression in VVC *C. albicans* isolates was performed in the presence and absence of sub-MIC concentrations of ZnO-np (2–18.1µg/mL) and Nystatin (0.031–128µg/mL). RNA was extracted from the *C. albicans* strains using TRIzol<sup>TM</sup> reagent (Catalog #15596026, Invitrogen, Thermo Fisher Scientific, Waltham, MA) according to the manufacturer's protocol. Before cDNA synthesis, all samples of RNA were treated with DNase1 (Thermo Fisher Scientific Co., EN0521). Synthesis kit of cDNA (Fermentase Co., K1622, USA) was utilized for the synthesis of cDNA.

# **Real-Time PCR**

The Primers designed by the PRIMER3 web-based software (<u>http://frodo.wi.mit.edu/primer3</u>) were used for realtime PCR analysis.

The primers were checked for specificity through the BLAST search available on the NCBI website (<u>http://blast.ncbi.nlm.nih.gov/Blast.cgi</u>). A primer set for the hemochromatosis gene was designed to be used as an internal control (Table 1). Amplification and detection of SAP1-3 genes expression in the presence of sub-MIC concentrations of ZnO and Nystatin was performed using the Real-time PCR with the Rotor-Gene SYBR Green PCR kit (Amplicon) with specific primers listed in Table 1 for SAP1 [GeneBank: XM\_712960.1], SAP2 [GeneBank: XM\_705969.1], SAP3 [GeneBank: XM\_717232.1].<sup>13</sup> The final volume of each reaction was 25  $\mu$ L, consisting of 12.5  $\mu$ L of the SYBR Green Master Mix, 1  $\mu$ L of forward primer (1  $\mu$ M), 1  $\mu$ L of RNase-free water.

Thermal cycling was performed in the ABI system 7300 (Applied Biosystems, Foster City, CA) scheduled to 95°C for 10 min (enzyme activation), 40 cycles of 95°C

Sequence (5´–3´)	Primer	Product Size (bp)	Accession Number
GTTGGTTTTGGTGGTGCTTC	SAPIF	161	XM-
GCAGCATTGGGAGAGTTGAG	SAPIR		712960.1
TGTGGATTCAGGTACCACCA	SAP2F	192	XM-
GCAAATTCGGAAGCTGGA	SAP2R		705969.1
TGGTCAAGGACAAGATCCAA	SAP3F	231	XM-
CCAATCCCTAAAATCCCTTG	SAP3R		718117.1
CCAGCTTTCTACGTTTCC	ATCIF	209	XM-
CTGTAACCACGTTCAGAC	ATCIR		717232.1

 Table I The Specific Primer Sequences and PCR Product Length

for 10 second (denaturing), 58°C for 15 second (annealing), and 72°C for 30 second (extension). Negative (water) and positive (*C. albicans* ATCC10231) controls were included in each run. The SAP1-3 genes expression was normalized to the ACT1 housekeeping genes, and evaluated using the  $2^{-\Delta\Delta Ct}$  method.

Data were presented as transcriptional mRNA (preferred units) relative to ACT1 or equal difference in expression relative to the preculture cell. Each experiment was performed in three replications, and the reactions (25 mL) were also performed in three replications.

## Statistical Analysis

To analyze the obtained data, the significance level was considered at P<0.05. The statistical software of Rest (ver. 2009) was used to test the ratio of target and reference molecules. For this purpose, the expression of the target gene was normalized to the gene expression of ACT1 as a reference gene. Data were entered into the SPSS software (ver. 16) and the results were analyzed by the one-way ANOVA.  $P \le 0.05$  was regarded as statistically significant.

## Results

## Identification of VVC C. albicans Isolates

Overall, 196 out of the 280 direct smeared samples were culture positive with five different yeast strains, 53.6% of which were *C. albicans*. All of these samples were detected by phenotypic and genotypic methods (Figure 1).

# Susceptibility Testing of Fluconazole Against VVC C. albicans Isolates by Disk

C. albicans strains were reported as susceptible S (zone diameter  $\geq 19$  mm), susceptible dose-dependent SDD (15 to 18 mm) and resistant R (≤14 mm). In this study, a disk diffusion test was used to determine the susceptibility of the samples to Fluconazole. Clinical specimens were divided into three groups based on the zone of inhibition of growth: less than 14 mm, 10 resistant isolates; 14-21 mm, 18 intermediate isolates; and more than 21 mm, 168 sensitive isolates. The ERG11 gene was tested in the Fluconazole-susceptible, intermediate and Fluconazole-resistant strains. 10 Fluconazoleresistant isolates had ERG11 gene expression; 18 intermediate and 168 Fluconazole-sensitive isolates had no ERG11 gene expression. Therefore, 196 isolates were divided into two groups of 10 Fluconazole-resistant and 186 Fluconazolesusceptible isolates based on their level of ERG11 gene expression. Zones of inhibition showing the interpretive categories in accordance with the disk diffusion technique are presented in Figure 2A and B. ERG11 gene was amplified and detected in five resistant C. albicans isolates (Figure 3).



Figure 2 Susceptibility pattern of C. albicans isolates to Fluconazole (25 µg) showing different sizes of inhibition zone: (A) 14 mm (R), and (B) 22mm (S).

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Figure 3 Agarose gel electrophoresis of the amplicon lane M: DNA marker, lane I, positive control; (+) lane 6, negative control; (-) and lanes 2–4 visible amplification of ERGII gene with a band size of 397 bp for Fluconazole-resistant *C. albicans* isolates.

# Antifungal Drug Susceptibility Testing of ZnO and Nystatin Against VVC *C. albicans* Isolates

To investigate the antifungal effects of ZnO nanoparticles and Nystatin, *C. albicans* isolates were exposed to various concentrations of these drugs. It was observed that all isolates were sensible to ZnO and Nystatin, and had inhibitory activity in low concentrations (Table 2). The lowest MIC value of ZnO nanoparticles and Nystatin was 2 and  $0.6\mu$ g/mL, respectively. The MIC measurement revealed that both ZnO nanoparticles and Nystatin might be good inhibitors of candida isolates (Table 2).

**Table 2** In vitro Susceptibility Testing of VVC C. albicans Isolates

 Against ZnO Nanoparticles and Nystatin

Strains	Antifungal Agents	MIC Range (µg/mL)	MIC 50%	MIC 90%
C. albicans (n=10)	ZnO nanoparticles	0.02–269	5	11.3
	Nystatin	0.6–32	3	8

Abbreviations: MIC50, Minimum Inhibitory Concentration 50%; MIC90, Minimum Inhibitory Concentration 90%.

Determining Is the Effect of the Sub-MIC Concentration of ZnO-np and Nystatin on VVC *C. albicans* Isolates by MTT Test We performed antifungal tests according to the CLSI-M27 -A3 guideline. As shown in Figure 4, the sub-MIC concentration of ZnO nanoparticles and Nystatin against VVC *C. albicans* isolates was 11.1 and 4  $\mu$ g/mL, respectively, indicating that more than 50% of the *C. albicans* isolates were viable at the sub-MIC concentrations of ZnO nanoparticles and Nystatin (11.1 and 4  $\mu$ g/mL, respectively).

# SAPI-3 Genes Expression in VVC C. albicans Isolates

Sensitive *C. albicans* strains showed lower expression of the target genes, while Fluconazole-resistant strains showed a significant increase in the expression of SAP1-3 genes. SAP1-3 genes expressions level was higher in 10 isolates (Fluconazole-resistant) comparing to that of 18 SDD and 168 sensitive isolates. The mean of target gene expression level SAP1-3 in the resistant isolates was significantly higher than in the sensitive strains (P < 0.001) (Table 3).

# Downregulation of SAPI-3 Genes Expression in VVC *C. albicans* Isolates Affected by Different Sub-MIC Concentrations of ZnO Nanoparticles in Comparison to Nystatin

In this model, the target gene expression was normalized by ACT1 expression, which is a reference gene. The normalized value was then applied to compare differential gene expression in different samples.

The candida strains were treated with different sub-MIC concentrations of ZnO nanoparticles and Nystatin. The gene expression of SAP1 was decreased about 2.44 times after treatment with ZnO nanoparticles comparing to the control group (untreated) (P<0.02).

It was also decreased around 4.16 times after treatment with Nystatin (P<0.01) (Figure 5). The gene expression of SAP2 in the strains indicated a decrease of 3.12 and 5.55 times after treatment with Nystatin and ZnO nanoparticles, respectively (P<0.03) (Figure 5). Also, the gene expression of SAP3 was remarkably decreased about 2.6 and 4.8 times after treatment with ZnO nanoparticles and Nystatin, respectively (P<0.05) (Figure 5).



Figure 4 The antifungal activity of ZnO nanoparticles and Nystatin on VVC C. *albicans* isolates: (A, B) MIC values of ZnO nanoparticles and Nystatin against C. *albicans* were determined by microdilution method. The isolates were incubated with various concentrations of ZnO nanoparticles and Nystatin for 24 hrs before conducting the MTTassay.

#### Discussion

Among the isolates studied, nearly 53.6% (196/280) were from VVC patients and 15% (43/280) were from RVVC patients. In some reports from Iran, the incidence of VVC was 5-10%,<sup>12</sup> 33.3%<sup>13</sup> and 47.3%.<sup>5</sup> We found that the frequency of VVC in our study is higher than the rates found in Nigerian [14.0%], Indian [20.47%], and Turkish [16%] studies.<sup>1,23,25</sup> In the present study, the species distribution of C. albicans isolates was similar to the study of Shi et al and Yano et al.<sup>11,12</sup> There are limited data regarding the antifungal susceptibility of yeast species causing VVC. Our study indicated that the ratio of resistance to Fluconazole C. albicans isolates from VVC was 5% (10/ 196), which is significantly higher than the rate in Southern China,<sup>11</sup> indicating differences in the genotype of the circulating isolates or in the frequency of the use of Fluconazole in different regions of China.

These changes can be due to the social and economic characteristics of sample individuals, as well as the knowledge of patients about personal health and self-treatment. The Fluconazole resistance rate in the RVVC group was significantly higher than in the VVC group; this indicates that more increased fluconazole use of Fluconazole can generate Fluconazole resistance in candida species due to overexpression of ERG11 gene.<sup>6</sup>

In this study, PCR amplified ERG11 gene expression in ten resistant isolates. Constant exposure of candida species to Fluconazole has frequently been associated with resistance. This study analyzed SAP1-3 genes expression in *C. albicans* isolates during vulvovaginal infection.

It was revealed that the average level of SAP in *C. albicans* was significantly more in the Fluconazoleresistant isolates than in the sensitive *C. albicans* strains (Figure 5). In addition, the SAP1-3 genes expression was increased in the Fluconazole-resistant *C. albicans* strains comparing to the sensitive *C. albicans* strains. This indicates a clear link between the expression of SAP1-3 genes exposure to antifungal agents and their drug resistance. The results of the present study imply that the importance of SAPs for pathogenicity during vulvovaginal infections.

Table 3 SAPI-3 Genes E	xpression Level	in Vulvovaginal	C. albicans	Isolates
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Evaluated Genes	Strain	SAPI-Gene	SAP2-Gene	SAP3-Gene
	Number	Expression (%)	Expression (%)	Expression (%)
Number of Fluconazole-resistant strains	10	10 (100%)	9 (90%)	6 (60%)
Number of SDD	18	18 (100%)	15 (83%)	12 (67%)
Number of susceptible strains	168	148 (88%)	148 (88%)	118 (70%)



Relative expression SAP1-3 *C. albicans* strains treated sub-MIC Nystatin and ZnO-nanoparticles

Figure 5 Comparison of the expression of SAP1-3 of VVC C. albicans isolates with  $3\pm1.7\mu$ g/mL concentrations of ZnO-nanoparticles and Nystatin compared to the calibrated gene in the control group (Value of the nonadjacent samples was zero). The ACT1 gene was used as an internal control (P<0.05). The numbers were obtained as mean  $\pm$  standard deviation (SD) after three replicates.

However, the expression levels of SAP1 and SAP3 were higher in the Fluconazole-resistant *C. albicans* isolates than in the sensitive *C. albicans* isolates. We also found that the expression level of SAP2 was significantly greater in the Fluconazole-resistant strains than in the sensitive *C. albicans* isolates. This indicates the importance of SAP2 in the development of pathogenesis *C. albicans* (P <0.05) (Figure 5). These results are in agreement with the results of Hosseini et al, who showed that the total surface area of SAP1-3 mRNA was higher in Fluconazole-resistant *C. albicans* <sup>14</sup>

Hence, it seems necessary to find new compounds to treat drug resistance to antifungals. Nystatin is a polyene antifungal that is frequently used as a topical agent in the treatment of VVC.<sup>1</sup> We previously reported<sup>22</sup> the antifungal effects of ZnO nanoparticles on the growth inhibition of standard strains of urinary catheter *C. albicans* in comparison with Fluconazole drug.<sup>19</sup> This is the first study to investigate the effect of ZnO on SAP1-3 genes expression in VVC *C. albicans* isolates. Previous investigations have identified the effects of ZnO-np and sodium dodecyl sulfate on the growth inhibition of standard strains of *C. albicans* in comparison with Fluconazole drug.<sup>25</sup> The MIC results of ZnO nanoparticles on Fluconazole-resistant VVC *C. albicans* isolates showed that the nanoparticles have intensive antifungal property.

The results of drug susceptibility testing of *C. albicans* isolates in the presence of ZnO nanoparticles and Nystatin revealed that the highest sensitivity of isolates to ZnO was MIC90 $\geq$ 0.5µg/mL compared to the non-treatment control samples (Table 1). These results agree with the finding of Kanathila et al, who showed an antifungal effect of Magnesium Oxide, ZnO nanoparticles, Silicon Oxide and Copper Oxide nanoparticles against the growth of *C. albicans*.<sup>7,17,26</sup> Also, Sardella et al reported that ZnO-np could inhibit the growth of Penicillium expansum by automated turbidometric analysis.<sup>7,22</sup> Therefore, ZnO seems to be an active ingredient for the control, prevention, and treatment of Candidiasis. In several studies, Nystatin was used for treating VVC patients with Fluconazole-resistant *C. albicans* isolates.

A study conducted by Hosseini et al<sup>13</sup> found that *C. albicans* resistant to azoles showed a higher SAP activity than the susceptible isolates. Similarly, in this study, Fluconazole-resistant strains were shown to increase the expression of SAP.<sup>17,27</sup> We used sub-MIC ( $3\pm1.7\mu$ g/mL) concentrations of ZnO nanoparticles and Nystatin to inhibit the expression of SAP1-3 genes. The results revealed that both ZnO and Nystatin decreased the SAP1-3 genes expression in all isolates that is in accordance with the findings of Cho et al.<sup>10,28–30</sup> This finding shows that ZnO nanoparticles  $5\pm1.3\mu$ g/mL and Nystatin  $3\pm1.3\mu$ g/mL

significantly reduced the SAP1-3 genes expression in Fluconazole-resistant VVC *C. albicans* isolates (P<0.05) (Figure 5).

ZnO has high permeability to the cell wall and cytoplasmic membrane through binding to lipids and proteins and denaturing through the destruction of the 3-D structure of proteins.<sup>26</sup> In total, the present research findings confirmed that low concentrations of ZnO nanoparticles and Nystatin reduced the expression of SAP1-3 genes in Fluconazole-resistant isolates.

## Conclusion

This is the first work done in Sayad Shirazi Obstetrics and Gynecology Hospital in Northeastern Iran on the expression of different virulence markers and azole resistance genes in clinical isolates of C. albicans. We identified the importance of SAPs1-3 genes role in the pathogenesis of Fluconazole-resistant VVC isolates; hence, we decided to use ZnO-np and Nystatin to inhibit the activity of these genes. Therefore, for the first time, Nystatin and ZnO nanoparticles (that we synthesized ourselves) were used to inhibit the activity of Aspartyl Proteinase Secreted in Fluconazole-resistant C. albicans VVC isolates. ZnO has a high cell wall and cytoplasmic permeability to the membrane by binding to lipids and proteins, and by destroying the 3-D structure of proteins.<sup>17</sup> The highest expression of SAP1-3 genes was observed in VVC Fluconazole-resistant C. albicans isolates with ERG11 gene.<sup>13</sup>

Each of these genes can enhance the resistance of the organism to antifungal drugs by different molecular mechanisms.<sup>31,32</sup> These results indicate that ZnO-np can be more effective in inhibiting the growth and activity of fungi by different mechanisms compared to Nystatin as an antifungal agent. According to the current research results and numerous tests of nanoparticles, these ingredients appear to be suitable for eliminating the infection in combination with new pharmaceutical formulations along with antifungal chemical drugs conjugated with medications. Therefore, there is a need for further studies on natural target cells, such as human vaginal epithelial cells as well as in vivo studies on mouse models to clarify whether ZnO nanoparticles could be applicable as an alternative antifungal.

# **Ethics Statement**

Inclusion criteria were vaginal swabs from women with vulvovaginal candidiasis who tended to participate in the study.

All procedures performed in studies involving human participants were in accordance with the rules on patient sampling and fully complied with the Ethics Committee of Gorgan University of Medical Sciences (Code no. IR. Gums.rec.1397.008) and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards. Written informed consent was obtained from all participants, and the questionnaire was sent to the journal as an additional file.

# **Data Sharing Statement**

All data are presented in the manuscript or as additional files.

# Acknowledgments

The current study was extracted from a research protocol approved by Golestan University of Medical Sciences, Gorgan, Iran. The study was approved by the university's Ethics Committee (Code no. IR. Gums. rec. 1397.008).

# **Author Contributions**

HSS conceived the study, participated in its design and analysis of the results and revised the manuscript. JHR, ST, AA and MZ participated in the design and coordination of the study, contacted the drug regulatory authorities, analyzed the results and drafted the manuscript. All authors gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

# Disclosure

The authors declare that they have no conflicts of interest.

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