

SPOCD1 accelerates ovarian cancer progression and inhibits cell apoptosis via the PI3K/AKT pathway

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Background: Ovarian cancer (OC) is the most common type of gynecological malignant tumors with poor prognosis. The spen paralogue and orthologue C-terminal domain containing 1 (SPOCD1) is a newly identified molecule that has been indicated to discriminate progressive in human solid tumors. However, the role of SPOCD1 in OC remains unknown.

Methods: The expression of SPOCD1 in OC and non-cancerous tissue was detected by Realtime polymerase chain reaction and immunohistochemical staining. The expression of SPOCD1 in OC cells (SKOV3 and CAOV3) was also detected by immunohistochemical staining. The effect of SPOCD1 on cell proliferation was analyzed by Cell Counting Kit 8 and colony formation assay, and cell migration was analyzed by transwell assay. Apoptosis was analyzed by flow cytometry. The protein expression of SPOCD1, PTEN, PI3K, p-AKT, and mTOR in OC cells was measured by Western blot.

Results: SPOCD1 expression was significantly upregulated in OC tissues compared with non-cancerous tissues ($P < 0.01$), and was positively correlated to FIGO stage and tumor grade of OC. Also, SPOCD1 was significantly expressed in nucleus and cytoplasm of SKOV3 and CAOV3 cells. Kaplan–Meier analysis indicated that patients with high SPOCD1 expression had shorter overall survival (HR = 1.512, 95%CI: 1.321–2.793, $P = 0.031$) and progression-free survival (HR = 1.875, 95%CI: 1.435–3.157, $P = 0.028$). SPOCD1 was upregulated in OC SKOV3 and CAOV3 cells. Further investigation revealed that downregulation of SPOCD1 inhibited the SKOV3 and CAOV3 cells proliferation and migration. In addition, the deficit of SPOCD1 increased the apoptosis in SKOV3 and CAOV3 cells. PI3K/AKT pathway was inhibited by knockdown of SPOCD1 in SKOV3 and CAOV3 cells.

Conclusions: Our data suggest that SPOCD1 may act as a carcinogenesis factor by activating the PI3K/AKT pathway to restrained cell apoptosis in OC.

Keywords: ovarian cancer, SPOCD1, proliferation, AKT

Introduction

Ovarian cancer (OC) is the leading cause of cancer-associated death among gynecological malignancies in worldwide. An estimated 521,000 new cases and 225,000 deaths occurred in 2015 in China, and 58% of new cases occurred in developing countries.^{1,2} OC is difficult to early detection and diagnosis due to deep anatomical location, lacking clinical symptoms, and effective index.³ Approximately 70–75% of OC patients have undergone abdominal cavity or distant metastasis at first diagnosis, with five years survival rate only about 30%.^{4,5} The clinical prognosis

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of OC is difficult to predict.^{6,7} Therefore, it is necessary to identify new prognostic biomarkers and therapeutic targets to improve treatment strategies.

The spen paralogue and orthologue C-terminal domain containing 1 (SPOCD1) is a recently identified novel gene that encodes a protein belonging to the transcription factor S-II (TFIIS) family.⁸ SPOCD1 was initially found to interact with testis protein phosphatase 1 which is a major eukaryotic serine/threonine-specific phosphatase regulating cellular signaling.⁹ However, the exact role of SPOCD1 in biological processes remains unknown. Recent studies have shown that SPOCD1 can predict progression and prognosis in T1G3 bladder cancer.¹⁰ SPOCD1 was overexpressed in gastric tumors and SPOCD1 knocking out reduced gastric cancer cell proliferation, invasion, and migration activities in vitro and in vivo.^{11,12} Furthermore, SPOCD1 promoted cell proliferation and inhibited cell apoptosis through the regulation of vascular endothelial growth factor A (VEGF-A) in osteosarcoma.¹³ The PI3K/Akt pathway is frequently changed in OC in an array comparative genomic hybridization studies.¹⁴ Meanwhile, the expression levels of both PIK3CA and phosphorylated Akt were analyzed in OC patients and found to be associated with poor survival.¹⁵

SPOCD1 expression has not been studied in OC. It is still unclear whether SPOCD1 could serve as a potential biomarker or tumor promoter in OC. Thus, the aim of this study was to characterize the expression and localization of SPOCD1. In addition, correlation analysis was performed to investigate the association between SPOCD1 expression and clinicopathological features and prognosis in OC patients. The mechanisms of SPOCD1 on carcinogenesis in OC cells were explored. Our study might explore new insights into the treatment of OC in clinical practice.

Materials and methods

Complete datasets of 93 OC and 32 non-cancerous tissue samples were obtained from patients undergoing first curative treatment resection in the Department of Obstetrics and Gynecology of the First Hospital of Lanzhou University (Lanzhou, China) between January 2009 and December 2012. None of the enrolled patients received chemotherapy or radiotherapy prior to surgery. Diagnosis and differentiation of OC were defined according to World Health Organization criteria. The characteristics of 93 OC are shown in Table 1. The study was approved by the Ethics Committee of the first Hospital of Lanzhou University, and the specimens were collected after informed consent from each patient.

Table 1 Correlations between SPOCD1 expression and clinicopathological parameters in ovarian cancer patients

Parameter	Group	Total number of patients	High	Low	P-Value
Age (years)	<55	42	18 (42.8%)	24 (57.2%)	0.231
	≥55	51	30 (58.8%)	21 (41.2%)	
FIGO stage	I+II	30	11 (36.7%)	19 (63.3%)	0.021*
	III+IV	63	40 (63.5%)	23 (36.5%)	
Tumor size (cm)	<10	64	35 (54.7%)	29 (45.3%)	0.328
	≥10	29	14 (48.3%)	15 (51.7%)	
Tumor grade	G1	36	14 (38.9%)	22 (61.1%)	0.012*
	G2+G3	57	34 (59.6%)	23 (40.4%)	
Tumor type	Serous	45	21 (46.7%)	24 (53.3%)	1.233
	Mucinous	38	18 (47.4%)	20 (52.6%)	
	Endometrioid	10	6 (60.0%)	4 (40.0%)	

Note: *P<0.05.

The main clinical and pathological features of these patients were also collected. All the patients got follow-up for five years.

Immunohistochemistry

The paraffin sections of tissues were dewaxed as routine. Endogenous peroxide activity was blocked by methanol containing 0.3% hydrogen peroxide for 30 mins. After exposure to 10% non-immunized goat serum in PBS for 10 mins, sections were treated at four Cover night with primary rabbit antibodies to SPOCD1 (1:200, Abcam, USA). The sections were then incubated with biotinylated anti-rabbit immunoglobulin serum for 30 mins, followed by incubation with peroxidase-labeled streptavidin for 20 mins at room temperature. Reaction products were visualized by diaminobenzidine tetrahydrochloride as chromogen. Finally, the sections were counter stained with hematoxylin. The percentage was scored as follows: 0 (0%), 1 (0–10%), 2 (10–50%), and 3 (>50%). The intensity was scored as follows: 0

(negative staining), 1 (weak staining), 2 (moderate staining), and 3 (intense staining). The final scores of SPOCD1 expression, ranging from 0 to 9, were calculated by multiplying the percentage score by the intensity score. Patients with a final score of <4 were classified as low expression group and vice versa.

Realtime PCR

Total RNA was extracted by the conventional Trizol method. First-strand cDNA was synthesized with a Prime Script Reverse Transcriptase Kit (GenePharma, Shanghai) according to the manufacturer's instructions. The specific primers were as follows: GAPDH forward 5'-TCCACCA CCCTGTTGTGTA-3', and reverse 5'-ACCACAGTCCA TGCCATCAC-3'; SPOCD1: Forward: 5'-CCCCATGGAG TGAAGCTTGT and Reverse: 5'-GCACCATGGGCCTTT TCTTC-3'.

Western blot

Total protein was extracted and quantified according to the manufacturer's instructions. Western blot assay was performed as described previously. A polyclonal rabbit anti-human antibody against SPOCD1 (1:1,000; Abcam, USA) and p-AKT (1:1,000; Abcam, USA) and the goat anti-rabbit secondary antibody (1:2,000; Invitrogen) were used. GAPDH (1:1,000; Abcam, USA) was used as a loading control. The ECL substrate was used to detect their expression. The band intensities were determined using the Bio-Rad imaging system (Hercules, USA).

Cell culture, treatment, and reagents

Human OC cell lines SKOV3 and CAOV3 were purchased from ATCC and routinely cultured. Cell transfection involved use of lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) with SPOCD1 shRNA according to the manufacturer's protocol. The primer sequence for SPOCD1 shRNA was designed by Shanghai GenePharma Co., Ltd. The efficiency in knocking down the target protein was determined by Western blot analysis after 48 hrs. SKOV3 and CAOV3 were treated by IGF-I after 24 hrs.

Cell growth assays

SKOV3 and CAOV3 cells were seeded in a 96-well plates at a density of 2,000 cells per well and maintained in DMEM containing 10% FBS at 37°C. Cell proliferation was detected by Cell Counting Kit 8 assay at various time points according to the manufacturer's instructions (Dojindo, Japan). The experiments were repeated three times.

Colony formation

Cells were plated in six-well culture plates at 500 cells/well. After incubation for 14 days at 37°C, cells were washed twice with PBS and stained with 0.5% crystal violet solution. The number of colonies containing <50 cells was counted under a microscope. Colonies containing 50 cells or more were counted as colonies. The experiments were repeated three times.

Cell invasion assay

A total of 5×10^4 cells were suspended in 500 μ L DMEM without serum and added to the upper chamber, while 750 μ L DMEM containing 10% FBS added in the lower chamber. After 48 hrs incubation, the cells on the upper layer were gently scratched with a cotton stick. The cells that invaded into the chamber membrane were fixed in 100% methanol for 30 mins, then stained with 0.5% crystal violet. At least three random microscopic fields (magnification, 40 \times) were analyzed for each insert. The experiments were repeated three times.

Flow cytometry assay

Apoptosis was detected and assayed by an Annexin-V/FITC Kit (BD Biosciences, San Jose, CA, USA) according to the manufacturer's instructions and analyzed by flow cytometry after a series of processes as described previously.

Statistical analysis

Data are expressed as mean \pm SD and were analyzed by use of GraphPad Prism 5. Two groups were compared by Student's *t*-test, and multiple groups by One-way ANOVA, with further two group comparison by Multiple Comparison Test Tukey's test. When the variance of the two groups was not homogeneous, nonparametric tests were used. Correlations were examined by Spearman correlation analysis. The cumulative progression-free survival (PFS) and overall survival (OS) rates were calculated using the Kaplan-Meier, and differences were compared using a log-rank test. $P < 0.05$ was considered statistically significant.

Results

SPOCD1 high expressed in OC tissues

We determined mRNA expression levels of SPOCD1 in 32 ovarian tumors and 32 non-tumor tissues. Realtime PCR analysis revealed that SPOCD1 mRNA levels were

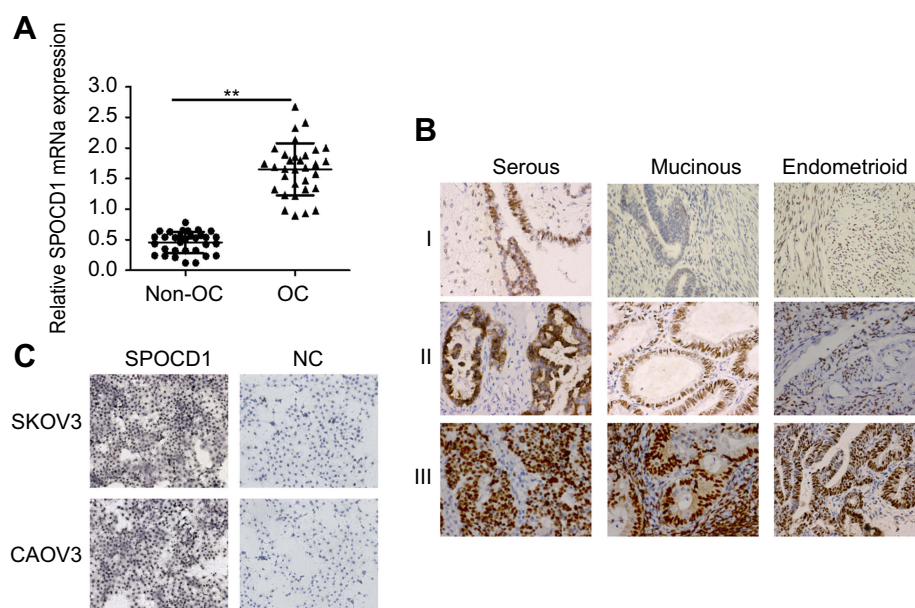


Figure 1 Expression of SPOCD1 in ovarian cancer tissues and non-cancerous tissues. **(A)** Realtime PCR analysis of SPOCD1 expression levels in ovarian cancer tissues and non-cancerous tissues (** $P < 0.01$). **(B)** IHC staining of SPOCD1 expression in ovarian cancer tissues (including serous, Mucinous and Endometrioid) (I: low expression; II: moderate expression; III: high expression). $\times 400$. **(C)** IHC staining of SPOCD1 expression in ovarian cancer SKOV3 and CAOV3 cells. $\times 200$.

Abbreviations: NC, negative control; OC, ovarian cancer.

significantly increased in OC tissues compared with non-cancerous tissues ($P < 0.01$, Figure 1A). IHC in 93 samples analysis showed positive expression of SPOCD1 in the nucleus and cytoplasm of OC tissues (Figure 1B). Also, SPOCD1 was significantly expressed in nucleus and cytoplasm of SKOV3 and CAOV3 cells (Figure 1C).

Relationships between SPOCD1 expression and clinicopathological features

To evaluate the significance of SPOCD1 expression in OC, we examined the associations between SPOCD1 expression and several clinicopathological features in 93 patients OC. As shown in Table 1, high SPOCD1 expression showed no association with patient age ($P = 0.231$), histological subtype ($P = 1.233$), or tumor size ($P = 0.328$). However, high SPOCD1 expression was positively correlated with FIGO stage ($P = 0.021$), tumor grade ($P = 0.012$). Taken together, these results supported that SPOCD1 promoted the progression and development of OC.

Prognostic value of SPOCD1 in OC

We also performed a survival analysis. Kaplan–Meier plots of all patients with clinical follow-up whose tumors had either stained positively or negatively for SPOCD1 were carried out. Kaplan–Meier survival curve analysis

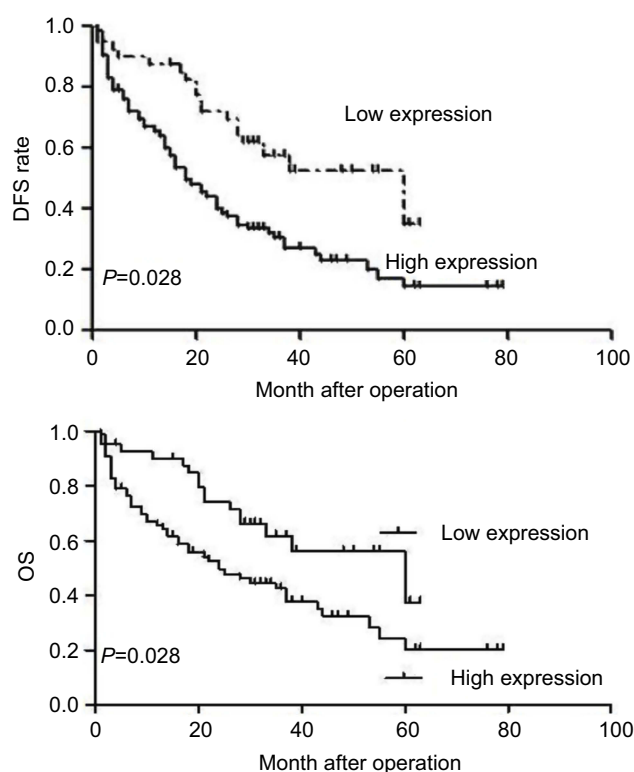


Figure 2 Kaplan–Meier survival curves for patients with ovarian cancer according to SPOCD1 expression level. Patients with high SPOCD1 expression levels exhibited a poor prognosis with respect to OS ($P = 0.031$) and PFS ($P = 0.028$).

was performed to predict OS and PFS in 93 OC patients. The results showed that high expression of SPOCD1 was

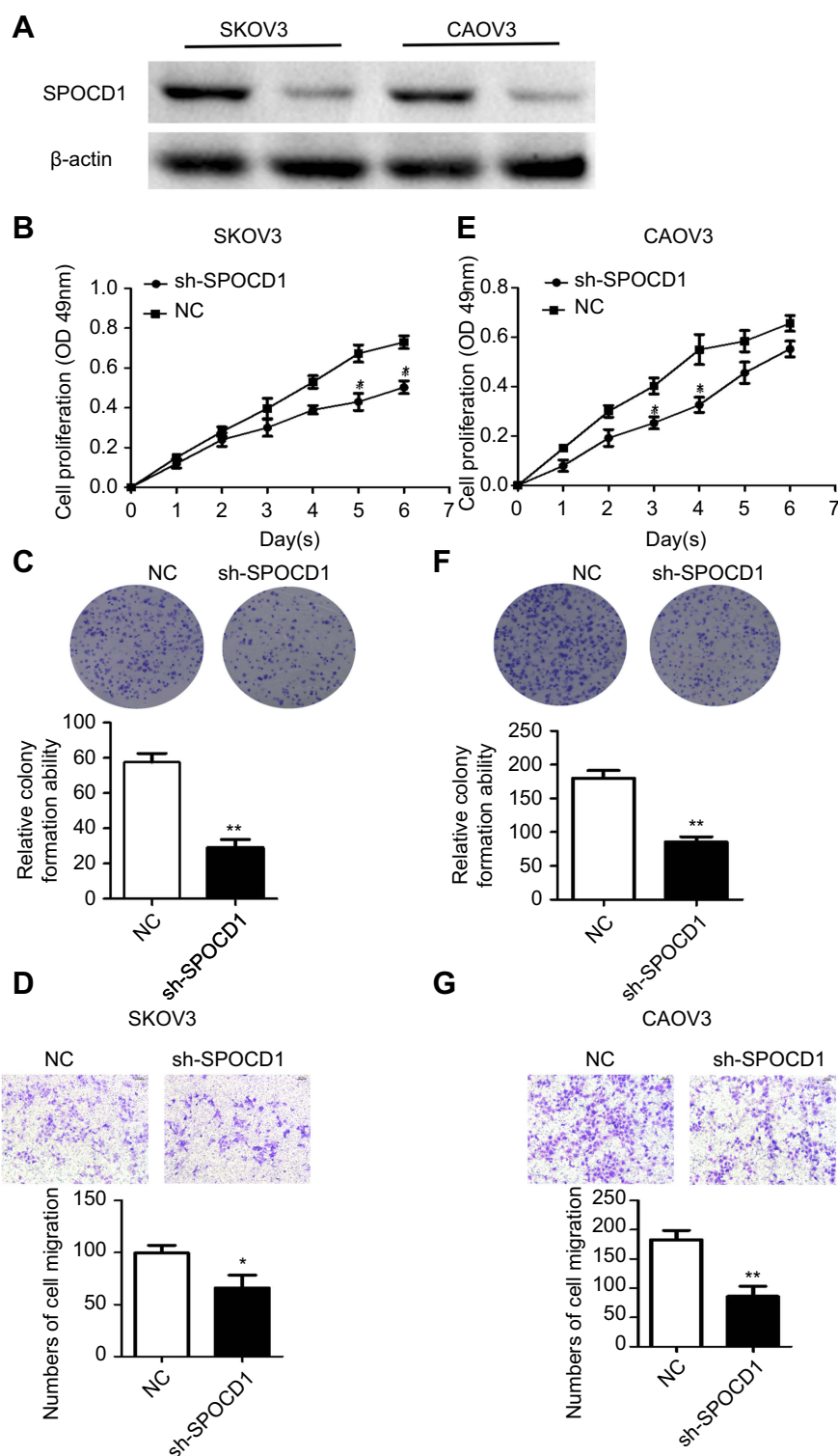


Figure 3 Effects of sh-SPOCD1 in SKOV3 and CAOV3 cells proliferation and metastasis. **(A)** Effect of knockdown SPOCD1 in SKOV3 and CAOV3 cells. **(B, E)** Effect of sh-SPOCD1 on the cell proliferation rate in SKOV3 and CAOV3 cells expressing empty vector or sh-SPOCD1 and determined by CCK8 assay at the indicated time point, for $P < 0.05$ (Vector vs sh-SPOCD1). **(C, F)** Effect of sh-SPOCD1 on colony formation. SKOV3 and CAOV3 cells with stable expression of vector or sh-SPOCD1 were seeded into six-well plates at 900 cells per well and cultured for 14 days, followed by crystal violet staining and colony counting. **(D, G)** Effect of sh-SPOCD1 on transwell assay. Data were analyzed as mean \pm SD and for $P < 0.05$ (Vector vs sh-SPOCD1).

Abbreviation: NC, normal control.

negatively correlated with OS (HR=1.512, 95% CI:1.321–2.793, $P=0.031$) and PFS (HR =1.875, 95% CI:1.435–3.157, $P=0.028$) ($P<0.05$, Figure 2).

Knockdown of SPOCD1 with shRNA inhibits cell proliferation and metastasis in SKOV3 and CAOV3 cells

The effect of SPOCD1 knockdown by shRNA was detected by Western blot in SKOV3 and CAOV3 cells (Figure 3A). We found that knocking down endogenous SPOCD1 expression decreased cell proliferation compared to the NC groups in SKOV3 cells ($P<0.05$) (Figure 3B). In addition, SPOCD1 shRNA-transfected cells starkly decreased the colony formation ability compared with NC cells (Figure 3C) ($P<0.05$). shRNA-mediated suppression of SPOCD1 also suppressed cell migration compared to the NC groups ($P<0.05$) (Figure 3D). The role of sh-SPOCD1 in CAOV3 cells was similar to that in SKOV3 cells. Suppression of SPOCD1 inhibited CAOV3 cells proliferation and migration (Figure 3E–G).

The deficit of SPOCD1 increases the apoptosis in SKOV3 and CAOV3 cells

In order to further figure out whether the ovarian SKOV3 and CAOV3 cells' proliferation were caused by increased cell division or apoptosis. We detected the

cell cycle and apoptosis, respectively, by PI and Annexin-V-FITC. The result showed that the deficit of SPOCD1 increased the apoptosis in SKOV3 and CAOV3 cells (Figure 4), but the cell cycle was no changes between NC and sh-SPOCD1.

Knockdown of SPOCD1 with shRNA inhibits PI3K/AKT pathway

In order to figure out the mechanism of SPOCD1 resulted in SKOV3 and CAOV3 cells apoptosis, the PI3K/AKT pathway was examined. As shown in Figure 5, when SPOCD1 was knocked down in SKOV3 cells, excluding PTEN, the protein levels of p-PI3k, p-AKT, and mTOR were notably downregulated. Interestingly, the protein level of p-PI3K, p-AKT, and mTOR was unchanged by adding PI3K/AKT pathway activator (IGF-1) in SPOCD1 shRNA-transfected SKOV3 cells. The protein expression of PTEN was unchanged in SPOCD1 shRNA-transfected and negative control SKOV3 cells (Figure 5A). The result of sh-SPOCD1 in CAOV3 cells was similar to that in SKOV3 cells. Knockdown of SPOCD1 down-regulated the protein levels of p-PI3K, p-AKT, and mTOR in CAOV3 cells, which unchanged by adding PI3K/AKT pathway activator (IGF-1) (Figure 5B). These data revealed that SPOCD1 regulated OC cell proliferation by restrained cell apoptosis through activating PI3K/AKT pathway.

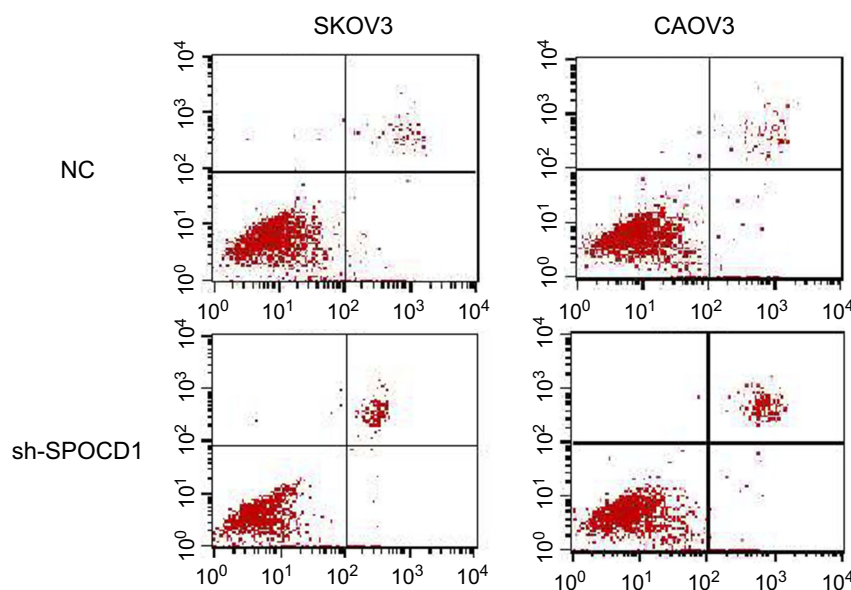


Figure 4 The deficit of SPOCD1 increases the apoptosis in SKOV3 and CAOV3 cells SKOV3 and CAOV3 cells expressing empty vector or sh-SPOCD1 were stained with Annexin-V-FITC (20 mg/mL). Apoptosis was analyzed by flow cytometry.

Discussion

Our results show that SPOCD1 was upregulated in OC compared with non-cancerous tissue. In addition, the association between SPOCD1 expression and the clinicopathological features of OC patients with were explored. Interestingly, patients with overexpression SPOCD1 were associated with advanced FIGO stage and low tumor grade. Also, high SPOCD1 expression had notably decreased OS and PFS compared with those with low SPOCD1 expression. The results suggested that SPOCD1 expression plays a crucial role in OC progression. A mechanistic study showed that the antitumor effects of SPOCD1 knocking down in OC cells. Downregulation of SPOCD1 caused significant decreases in cell proliferation and migration. Further study proposes that deficit of SPOCD1 increased the

apoptosis and inhibited PI3K/AKT pathway in OC cells. The study explores a novel mechanism that the PI3K/AKT pathway are the responsible factors for the antitumor effects of sh-SPOCD1.

To our knowledge, the research about tumorigenesis role of SPOCD1 is little in OC. Recent study shows that SPOCD1 high expressed in bladder cancer and gastric cancer.^{10,12} SPOCD1 promotes the proliferation and metastasis of glioma cells by PTX3. SPOCD1 promotes cell proliferation and inhibits cell apoptosis in osteosarcoma via VEGF-A.¹³ Our study showed that SPOCD1 was significantly upregulated in OC samples and SKOV3 and CAOV3 cells. Importantly, statistical analysis revealed that a high level of SPOCD1 correlated with advanced clinicopathological characteristics and

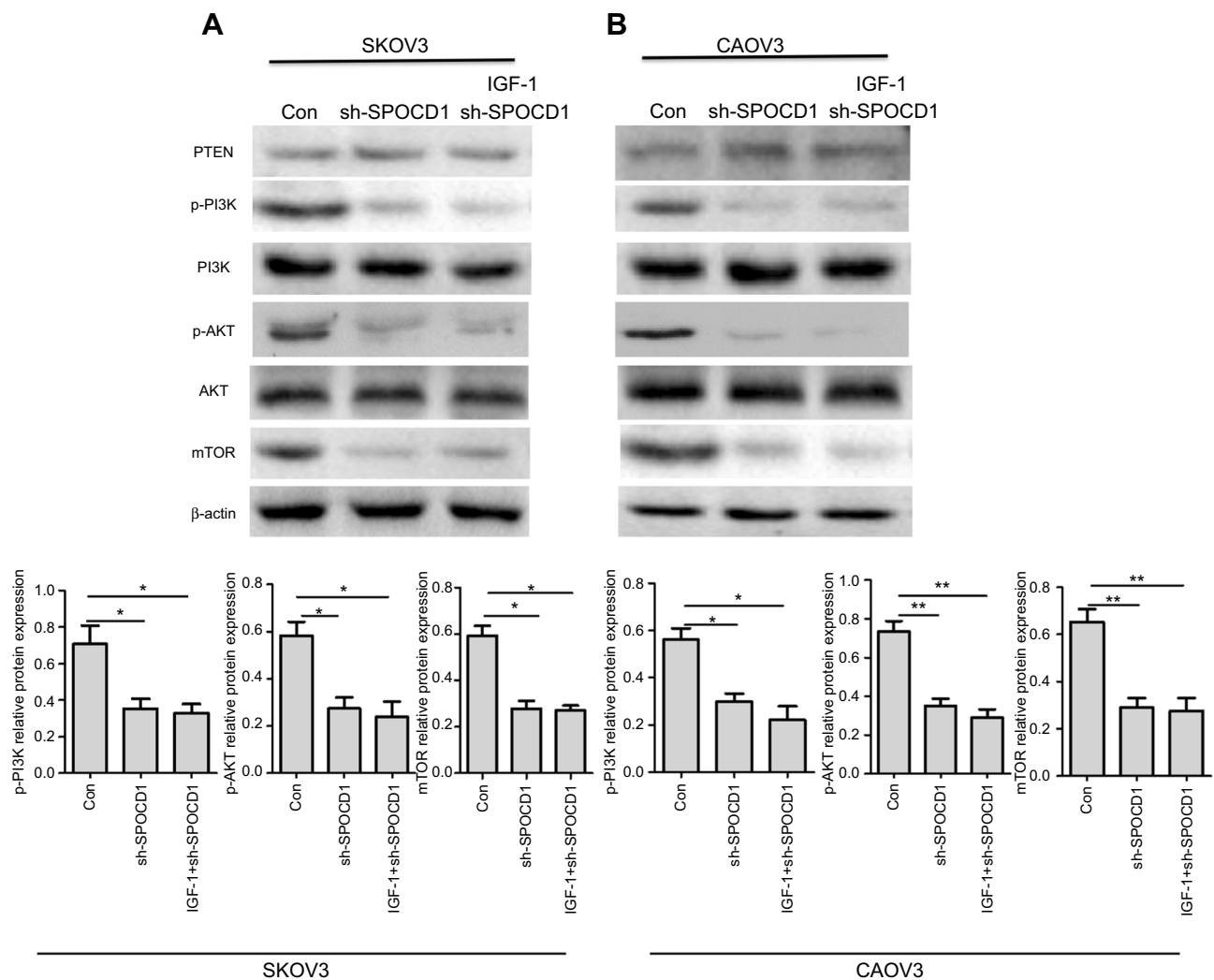


Figure 5 SPOCD1 deficit inhibited PI3K/AKT pathway in ovarian cancer cells. **(A)** In SKOV3 cells, SPOCD1 was knocked, and IGF-I was used as the PI3K/AKT pathway activator. **(B)** In CAOV3 cells, SPOCD1 was knocked, and IGF-I was used as the PI3K/AKT pathway activator. Relative protein level of PTEN, p-PI3K, PI3K, p-AKT, AKT, and β-actin was measured using Image J. Data were presented as mean ± SD from at least three independent experiments. ** $P < 0.01$ compared with control group.

poor prognosis. Moreover, reducing SPOCD1 significantly promoted proliferation and migration of ovarian cells. Further study showed that deficit of SPOCD1 increased the apoptosis. It demonstrated that SPOCD1 promoted proliferation via inhibiting cell the increasing cell apoptosis. These data support the suggestion that SPOCD1 could be a promising independent prognostic factor for OC patients and favors an oncogenic role for SPOCD1 in OC.

The PI3K/AKT pathway has demonstrated to participate in the regulation of proliferation, apoptosis, and survival of many cancer cells.^{16,17} The PI3K/AKT signal AKT activator stimulated proliferation in various carcinoma cells.¹⁸ In this study, our results demonstrated that SPOCD1 contributed to induced acceleration of proliferation and migration through inhibiting cell apoptosis and activated the PI3K/AKT pathway in OC cells. This study identifies SPOCD1 as an oncogene in OC through the PI3K/AKT pathway. The mechanism underlying how SPOCD1 activates the PI3K/AKT pathway needs further investigation. Further experiments in cell and animal models of OC are needed to understand biological processes regulated by SPOCD1 in OC progression.

In conclusion, SPOCD1 has a crucial role in promoting tumorigenesis and growth of OC. The specific mechanisms have been primarily expounded and proved in this paper. Results from this study have the direct potential to develop SPOCD1 as an important biomarker for prognosis of OC and a therapeutic target for OC, especially for those with SPOCD1 overexpression.

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Disclosure

The authors report no conflicts of interest in this work.

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