

ASH2L-Promoted *HOXC8* Gene Expression Plays a Role in Mixed Lineage Leukemia-Rearranged Acute Leukemia

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Background: Mixed lineage leukemia (MLL) fusion protein alone exhibits poor histone lysine methyltransferase (HKMT) activity in catalyzing histone H3 Lys4 trimethylation (H3K4me3) in MLL-rearranged acute leukemia.

Methods: To explore the HKMT effect of another regulatory protein within the complex of proteins associated with Set 1 (COMPASS), we analyzed the H3K4me3 modification of the *HOXC8* promoter under the action of ASH2L regulation. Small interfering RNA of ASH2L, chromatin immunoprecipitation, real-time-PCR (RT-PCR), and Western blotting were used to detect the expression of specific regions of the *HOXC8* promoter, *RBBP5*, *WDR5*, *MLL*, and *BRTF* in two MLL-rearranged acute leukemia cell lines (RS4:11 and THP-1 cells).

Results: The gene and protein expression levels of *HOXC8* were significantly downregulated upon treatment with ASH2L-siRNA (as analyzed by targeting specific regions of the *HOXC8* promoter located 0 and 3 kb (–3.0 kb) upstream of the transcriptional start site in RSH:11 cells; and –3.0 and –2.0 kb upstream of the transcriptional start site, and +1.4 kb downstream of the transcriptional start site in THP-1 cells). The expression levels of the *BRTF*, *RBBP5*, *WDR5*, and *MLL* genes were significantly downregulated from the different transcriptional start sites of the *HOXC8* promoter in the RSH:11 cell line ($P < 0.05$). Furthermore, the *BPTF* and *RBBP5* genes were downregulated from the *HOXC8* promoter in the THP-1 cell line ($P < 0.05$).

Conclusion: Based on these results, we suggest a new concept of histone modification of the ASH2L protein in MLL-rearranged acute leukemia, which cannot carry out methyltransferase activity independently. The protein–protein interactions of ASH2L with other COMPASS members, such as *MLL*, *WDR5*, *RBBP5*, and chromatin remodeling factor *BRTF*, appear to be essential for its role in the activation of *HOXC8* gene transcription.

Keywords: *MLL*-rearranged acute leukemia, *HOXC8* gene, *ASH2L*, COMPASS complex, H3K4 trimethylation

Introduction

The mixed lineage leukemia (*MLL*) gene is located in q23 of the long arm of chromosome 11 (11q23), the rearrangement of which is a common chromosomal abnormality in adult and childhood acute lymphoblastic leukemia (ALL) and acute myeloid leukemia (AML). Patients that harbor this gene rearrangement exhibit unique clinical and biological characteristics. The WHO Classification of Tumors of Hematopoietic and Lymphoid Tissues (2008) individually lists “mixed phenotype leukemia (v; 11q23) with *MLL* rearrangement” as one of the important subtypes of acute leukemia. This type of leukemia is not usually sensitive to

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conventional chemotherapy and is associated with a low complete remission rate and a short survival period.^{1–3}

The *MLL* gene, which encodes an important methyltransferase, is involved in *MLL* gene rearrangement during the development of leukemia through the regulation of downstream target genes.

The proteins ASH2L, WDR5, RBBP5, and Dpy30 together constitute the *MLL1* complex (COMPASS), which exhibits strong enzymatic activity.⁴ As an important regulatory protein, ASH2L is likely involved in transcriptional activation of the *HOXC8* gene in *MLL*-rearranged acute leukemia.⁵ To explore the histone lysine methyltransferase (HKMT) effect of another regulatory protein within the complex of proteins associated with Set 1 (COMPASS), we examined the expression of ASH2L and the stability of the *HOXC8* gene and analyzed H3K4H3K4 trimethylation (H3K4-me3) modification of the *HOXC8* promoter under ASH2L regulation in *MLL*-rearranged leukemia cells.

Materials and Methods

Cell Lines

Human RS4:11 and THP-1 cell lines were purchased from the Shanghai Cell Bank, of the Chinese Academy of Sciences. The cells were placed in a 5% CO₂ incubator at 37°C and the cell density was controlled at 0.5–1×10⁶/mL in RPMI 1640 medium with 10% premium fetal bovine serum.

Fluorescence in-situ Hybridization (FISH)

An *MLL* two-color fluorescent probe (Vysis Inc., USA) was used in the study. The hybridization signals of red and green fluorescence of cells were detected by excitation of the DAPI/TR filter (Texas Red) and observation under a Leica DRMA2 fluorescence microscope, according to a previous method.⁶ At least 300 cells were analyzed for each sample. Nuclei with clear boundaries and intact structures that were non-overlapping were counted to calculate the percentage of positive cells. The *MLL* two-color break-point separation probe diagnostic criteria were that cells with *MLL* rearrangement showed red-green fusion signals (i.e., a red signal and a green signal), whereas normal cells showed two red-green fusion fluorescence signals.

Quantitative Real-Time PCR (qRT-PCR) Detection of the *HOXC8* Gene

For all experiments, three independent samples were detected for the RS4:11 and THP-1 cell lines. The high-purity total RNA rapid extraction kit (General, USA) was used to extract

total RNA from the cells. The RNA was then reverse-transcribed into cDNA with the RevertAid First Strand cDNA Synthesis Kit (Fermentas, USA). Quantitative RT-PCR was performed using an SYBR Green RT-PCR kit (Bio-Rad, Hercules, CA, USA) with a quantitative PCR instrument (Bio-Rad). The primers were synthesized by Sangon Biotech (Shanghai, China). The primer sequences for amplification of *HOXC8* and the internal reference gene GAPDH are listed in [Supplemental Table 1](#). Relative quantitative analysis was performed for *HOXC8* according to the PCR amplification curve, and standard curves for the negative and positive controls were also determined for each experiment.

Small Interfering RNA (siRNA) Transfection

The FAM fluorescently labeled siRNA-ASH2L fragments were chemically synthesized and transfected into cells using Lipofectamine 2000 reagent according to the manufacturer's instructions. Three sets of primers were used for the ASH2L-siRNA group that specifically interfere with the target gene ASH2L ([Supplemental Table 2](#)). Cells were harvested 24 hrs after transfection with siRNA, lysed by the addition of 1 mL of Trizol, and then total RNA was extracted by centrifugation and reverse transcribed into cDNA for qPCR. qPCR was performed in triplicate using SuperReal PreMix Plus (SYBR Green; TIANGEN Biotech) on a CFX connect RT-PCR System (Bio-Rad). The expression levels of *HOXC8* and ASH2L cDNAs were normalized to β -actin cDNA by the comparative cycle threshold (Ct) method. The primer sequences for *HOXC8* and ASH2L are listed in [Supplemental Table 3](#).

Western Blot Analysis After ASH2L siRNA Treatment

Immunoblot signals were detected with Pierce ECL Western blotting substrate (Thermo Fisher). Cells were lysed in RIPA buffer containing protease inhibitors (Beyotime). The proteins were analyzed by SDS-PAGE and transferred to PVDF membrane. The membrane was exposed to X-ray film after incubation with the indicated ASH2L antibody (Abcam; ab181117) and GAPDH was used as a control.

Chromatin Immunoprecipitation (ChIP)

Before or after transfection for 48 h, cells were fixed in formalin and ChIP was performed against anti-H3K4me3, anti-ASH2L, and IgG antibodies (Abcam). ChIP experiments were performed using a SimpleChIP[®] Plus Sonication

Chromatin IP Kit (Cell Signaling Technology) according to the manufacturer's instructions. Associated DNA isolated via ChIP was then analyzed by quantitative real-time RT-PCR using IQ SYBR Green Supermix (Bio-Rad, cat no:170-8882AP) and primers to the HOXC8 promoter region, MLL, BPTF, RBBP5, and WDR5, which are listed in [Supplemental Table 4a](#) and [4b](#). The PCR products were electrophoresed and sent to Biosune Inc. (Shanghai) for sequencing using the primers listed in [Supplemental Table 4c](#).

Statistical Methods

For FISH, 200 interphase cells were analyzed for each sample. According to the criterion of the number of positive cells $> (x \pm 3s)$, the positive criterion for MLL-rearrangement was determined to be $>8.9\%$. To analyze gene expression, after PCR, the cycle threshold (Ct value) was recorded. Using GAPDH expression as the internal reference, the expression level of the gene of interest was measured according to the equation $2^{-\Delta CT}$, where $\Delta CT = \text{mean value of CT of the gene to be measured} - \text{mean value of CT of the internal reference gene}$. The results were automatically calculated by the fluorescence quantitative PCR software Bio-Rad CFX Manager and were reported in the generated PDF files and the EXCEL raw datasheet. The measured data are represented by $x \pm s$, and the data between the two groups were analyzed by a paired *t*-test: ** $P < 0.01$, * $P < 0.05$.

Results

Detection of Both Cell Types by FISH

In total, 300 interphase cells of the RS4:11 and THP-1 cell types were analyzed. More than 90% of the cells showed

a positive signal (one red, one green, and one fusion signal), which confirmed two types of MLL rearrangements in 11q23 ([Figure 1](#)).

HOXC8 Gene Expression in the Two Cell Lines

HOXC8 gene expression was analyzed in both cell lines by the qPCR method, and the mRNA level in THP-1 cells was found to be 2.7704 (1.0232–6.343), compared with 4.0487 (3.8011–4.2248) in RS4:11 cells. The Ct values of HOXC8 in both cell lines were no more than three times that of the control, indicating a relatively high level of gene expression, and thus the expression of HOXC8 in RS4:11 cells was more stable.

Change in Expression of the *ASH2L* and *HOXC8* Genes Following *ASH2L* Interference as Determined by Quantitative PCR

After 24 hrs incubation, the green fluorescent proteins carried by the plasmid were observed under a fluorescence microscope to calculate the transfection rate. No green fluorescence was observed in the untransfected group, indicating that the *ASH2L* gene had been successfully transfected. The results showed that the transfection efficiency of the RS4:11 and THP-1 cells was $>90\%$ by inverted fluorescence microscope and $>85\%$ by flow cytometry ([Figure 2](#)).

Three independent tests confirmed that the expression levels of the *ASH2L* and *HOXC8* genes were downregulated after transfection with *ASH2L* siRNA. The expression of *ASH2L* in sample 2 of THP-1 cells and sample 2 of

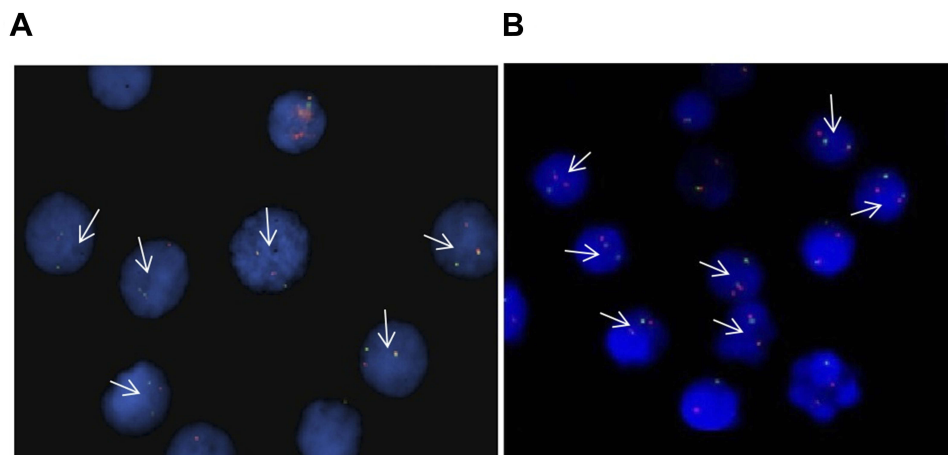


Figure 1 FISH detection of the two cell lines of MLL-rearranged acute leukemia. Arrow: positive signal cells (one red, one green, and one fusion signal), (A) THP-1 cells, (B) RS4:11 cells.

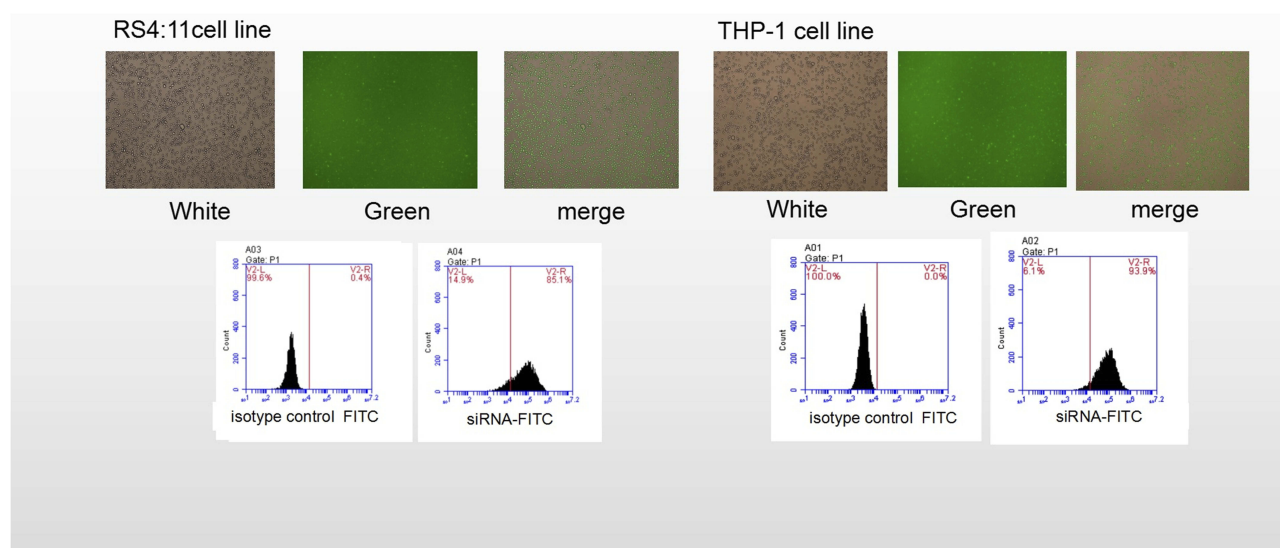


Figure 2 Detection of siRNA transfection efficiency. Transfection of the RS4:11 cells and the THP-1 cells with the plasmid resulted in an intracellular distribution of the green fluorescence. Upper: The green fluorescent proteins were observed under a fluorescence microscope to calculate the transfection rate (left: 95%, in RS4:11 cells; right: 91% of THP-1 cells). Lower figure: Detection of the transfer efficiency was observed by flow cytometry (left: 85% of RS4:11 cells, right: 89% of THP-1 cells).

RS4:11 cells was significantly downregulated ($P < 0.01$); the expression of *HOXC8* in sample 2 of THP-1 cells and samples 1, 2, and 3 of RS4:11 cells was also significantly downregulated ($P < 0.01$), indicating that *ASH2L* siRNA successfully interfered with the expression of *ASH2L* in THP-1 and RS4:11 cells. This result also indicated that the expression of *HOXC8* was downregulated following *ASH2L* interference in RS4:11 and THP-1 cells.

qPCR Detection of *ASH2L* and *HOXC8* Gene Expression After siRNA Interference

Three independent qPCR assays showed that the expression of both the *ASH2L* and *HOXC8* genes was downregulated following siRNA interference. The expression of *ASH2L* was significantly downregulated in sample No. 2 of THP-1 cells and sample No. 2 of RS4:11 cells ($P < 0.01$); *HOXC8* was significantly downregulated ($P < 0.01$) in sample No. 2 of THP-1 cells and sample No. 1, 2, and 3 of RS4:11 cells (Supplemental Figure 1). The second pair of primers showed the highest efficiency among the three pairs of primers tested so were used in subsequent studies.

Change in *ASH2L* Protein Expression Following siRNA Interference

Three independent Western blot experiments showed that *ASH2L* protein expression was downregulated following siRNA interference. The protein expression levels of *ASH2L*

in sample No. 1 and 2 of THP-1 cells and sample No. 2 and 3 of RS4:11 cells were significantly lower than that of the negative control group ($P < 0.05$), indicating that siRNA successfully interfered with the expression of the target *ASH2L* protein in THP-1 and RS4:11 cells (Figure 3).

ChIP-qRT-PCR and Sequencing Results Before siRNA Interference

In both RS4:11 and THP-1 cells, the *ASH2L* protein showed the strongest binding to locus -2 kb of the promoter region of *HOXC8*, although it also showed a degree of enrichment and binding in other regions. In RS4:11 cells, the modification of H3K4me3 was positively correlated with the binding of *ASH2L*. In THP-1 cells, the modification of H3K4-me3 at locus 1.4 kb of the first intron was elevated abnormally (Supplemental Figure 2). The sequencing results showed that the PCR products correlated with distinct sequences in the *HOXC8* promoter (Supplemental Figure 3).

ChIP-qRT-PCR Results After siRNA Interference

After siRNA interference of *ASH2L* expression, DNA from RS4:11 and THP-1 cells was fixed with formaldehyde and broken into fragments of 200–500 bp by ultrasonic fragmentation. DNA was then extracted by ChIP using a specific negative control antibody (anti-RbIgG) and anti-H3K4-me3. Subsequently, the -2, -3, and 1.4 kb loci of the promoter region of *HOXC8* were detected by qRT-PCR. The results

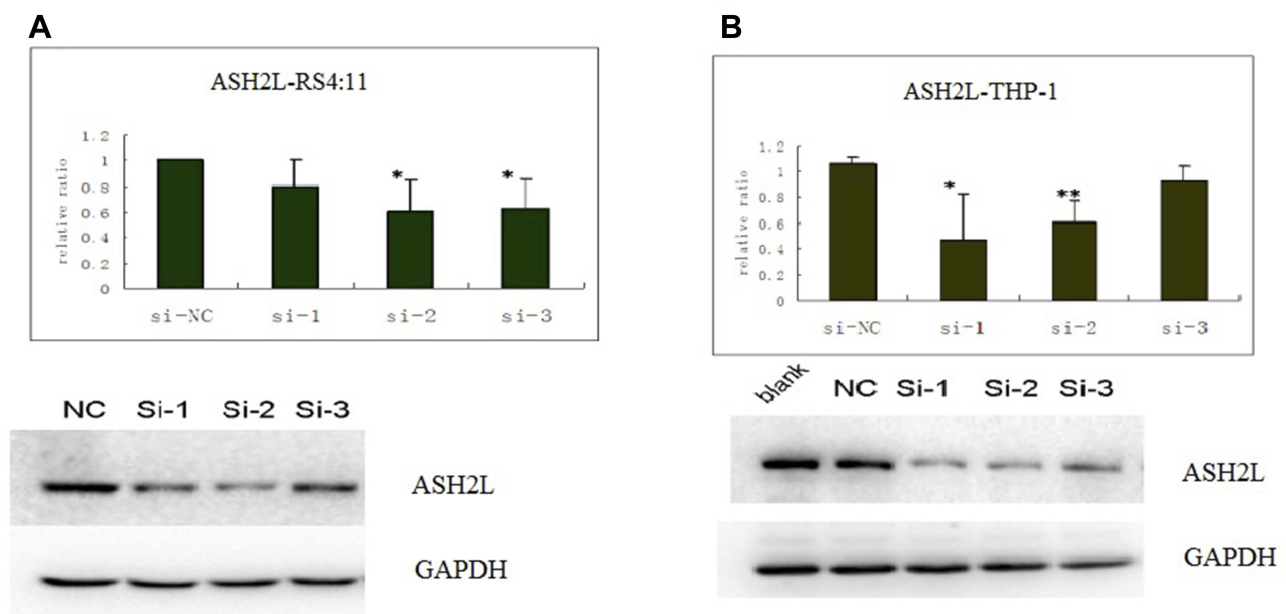


Figure 3 ASH2L levels were analyzed by Western blotting in RS4:11 and THP-1 cell lines after siRNA interference. The protein level of ASH2L in sample No. 1 and 2 of THP-1 cells and sample No. 2 and 3 of RS4:11 cells were lower than that of the negative control group. (A) Expression in RS4:11 cells; (B) expression in THP-1 cells. Upper: column chart; Lower: electrophoresis map, ASH2L (Si-1: 741 bp, Si-1: 1412 bp, Si-1: 1664 bp). Western Blot Statistics (* $P < 0.05$, ** $P < 0.01$).

showed that compared with the negative control, modification at locus -3 kb of *HOXC8* and the transcriptional start region of H3K4-me3 were significantly reduced. In THP-1 cells, modification of loci -2 and -3 kb of the *HOXC8* promoter region and H3K4-me3 of locus 1.4 kb of the first intron were significantly reduced ($P < 0.05$) ([Supplemental Figure 4](#)).

Activity of MLL, BPTF, RBBP5, and WDR5 Was Reduced in the *HOXC8* Promoter Region After Treatment with siRNA-ASH2L

After siRNA-ASH2L treatment of RS4:11 cells, decreased activity at the promoter region of *HOXC8* was detected for: BPTF at loci -1.4, +2, and +3 kb ($P < 0.05$); RBBP5 at loci -1.4, +2, +3, and 0 kb ($P < 0.05$); WDR5 at loci -1.4 and +2 kb ($P < 0.05$); and MLL at loci -1.4 and +2 kb ($P > 0.05$).

After siRNA-ASH2L treatment of THP-1 cells, decreased activity at the promoter region of *HOXC8* was detected for: BPTF at loci -1.4 and +3 kb ($P < 0.05$); and RBBP5 at loci -1.4 and +3 kb ($P < 0.05$); however, the activities of WDR5 and MLL did not change at the four loci of the *HOXC8* promoter ($P > 0.05$) ([Supplemental Figure 5](#)).

Discussion

HOX genes are widespread among organisms and can be divided into four clusters phylogenetically: A, B, C, and

D. Previous studies have shown that *HOX* expression is closely related to the development of tumors. In normal tissues, the expression of *HOX* (which is only usually expressed in embryonic tissues) may lead to the development of tumors.⁷ The *HOXC8* cluster is located on chromosome 12, and its subgroup *HOXC8* is a type of evolutionarily highly conserved developmental regulatory gene that regulates the proliferation, apoptosis, migration, and differentiation of cells by regulating many genes at the transcriptional level. There is growing evidence that *HOXC8* plays an important role in the development and progression of human malignancies, but the related mechanism is unclear.^{8–10}

The MLL family of proteins is a class of methyltransferases specific for H3K4 whose methyltransferase activity depends on a conserved SET domain at the C-terminus. In MLL-rearranged acute leukemia, MLL fusion protein cannot directly activate downstream target genes by modifying the promoter region. Therefore, it is highly likely that there are other regulatory proteins in the promoter region of the target gene, or the regulatory protein plays a synergistic regulatory role by binding to the *MLL* gene. MLL family protein is different from other methyltransferases with a SET domain because its function requires a number of intrinsic auxiliary proteins, including WDR5, RBBP5, ASH2L and DPY30 to form a functional complex (COMPASS) to complete the methylation modification process.^{4,10} How this complex

performs methylation modification remains in dispute. In recent years, research has focused on the role of ASH2L because ASH2L is an important and complex regulatory protein, and it has binding sites that can easily bind to other proteins. Moreover, ASH2L can bind to the promoter region of *HOXC8*, initiate the movement of nucleosomes and thereby activate the target gene.^{11–15} Based on these characteristics, we considered that ASH2L is likely to be involved in the transcriptional activation of the *HOXC8* gene in MLL-rearranged acute leukemia.

To explore whether there are interactions between ASH2L and *HOXC8* in MLL-rearranged acute leukemia, this study for the first time used siRNA transfection techniques to specifically interfere with the expression of ASH2L in RS4:11 and THP-1 MLL-rearranged acute leukemia cell lines. The results showed that the transfection efficiency was greater than 90%. We further studied the gene expression levels of *ASH2L* and *HOXC8* after transfection, and the results showed significantly downregulated expression of both ASH2L and *HOXC8*.

To investigate whether ASH2L causes a decrease of H3K4-me3 in the *HOXC8* promoter region, we next used ChIP-qRT-PCR to study the acute leukemia cells with two MLL genes rearranged after siRNA-ASH2L treatment. The H3K4-me3 levels at loci 0 kb (transcription start site in the promoter region of *HOXC8*), -2 and -3 kb (both upstream of the start site), and 1.4 kb (downstream of the start site) were analyzed. The results showed that, in RS4:11 cells after ASH2L interference, modification of H3K4-me3 at locus -3 kb upstream of the start site of the promoter region of *HOXC8* was remarkably reduced. In THP-1 cells, modification of H3K4-me3 at loci -3 and -2 kb in the promoter region and locus 1.4 kb in the first intron of *HOXC8* was significantly reduced ($P < 0.05$). These results confirmed our original hypothesis that ASH2L plays an important role in the regulation of histone methylation of acute leukemia with MLL rearrangement. ASH2L may directly or indirectly affect the expression of the target gene *HOXC8* via interaction with MLL protein, increasing the H3K4-me3 level in the promoter region of *HOXC8* and thereby activating *HOXC8* expression. Chromatin remodeling is a recently described concept and the mechanism involved in chromatin remodeling is complex. The nucleosome slide along the chromatin, changing its structure from a tight to a loosened structure. The loosened structure may facilitate the binding of various transcription factors to the regulatory motif, thereby activating the gene.^{16–18} Chromatin remodeling is not an independent mechanism, but is regulated by multiple

proteins, and is closely related to histone modifications. we thus further tested the binding between MLL, WDR5, and RBBP5 in the promoter region of *HOXC8* and maximum nucleosome remodeling factor subunit PHD finger transcription factor (bromodomain and PHD finger transcription factor, BPTF) after siRNA-ASH2L interference. The results showed that after siRNA-ASH2L treatment of RS4:11 cells, decreased levels of BPTF and RBBP5 were detected at loci -1.4, +2, +3, and 0 kb in the promoter region of *HOXC8* ($P < 0.05$); and MLL and WDR5 showed decreased activity at loci -1.4 and +2 kb in the promoter region of *HOXC8* ($P < 0.05$). However, after siRNA-ASH2L treatment of THP-1 cells, BPTF and RBBP5 showed decreased activity at loci -1 and +3 kb in the promoter region of *HOXC8* ($P < 0.05$); while no significant changes in WDR5 and MLL levels were observed for any of the four loci of *HOXC8* ($P > 0.05$). This is possibly because the cell viability of THP-1 cells was relatively low compared with RS4:11 cells, thus leading to less reliable results. However, taken together, our results show that decreased expression of H3K4-me3 in the promoter region of *HOXC8* was not directly due to the silencing of ASH2L.

In summary, our results provide insight into the association between histone methylation of the *HOXC8* gene and MLL-rearranged acute leukemia. ASH2L may activate *HOXC8* expression via interaction with MLL, WDR5, and RBBP5 of the MLL methylase complex, as well as BPTF protein, which modifies histone methylation and remodeling of the chromatin structure. Identification of ASH2L as a component of the MLL complex required for specific regulation of H3K4-me3 provides insight into the molecular mechanism of histone modification. Future genetic and biochemical studies on ASH2L and histone modification will be instrumental in defining the biological role of H3K4 methylation during development and differentiation.

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Author Contributions

Yu-Jie Wu and Li-xia Li designed the research; Lu Liu, Si-shu Zhao, and Qiu Hai-rong performed the research; Hui Wang analyzed the data; Yu-Jie Wu and Li-xia Li wrote the paper. All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to

be published, and agree to be accountable for all aspects of the work. Yu-Jie Wu and Li-xia are co-first authors.

Disclosure

The authors report no conflicts of interest relating to this work.

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