

# Micro-CT imaging of breast tumors in rodents using a liposomal, nanoparticle contrast agent

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**Abstract:** A long circulating liposomal, nanoscale blood pool agent encapsulating traditional iodinated contrast agent (65 mg I/mL) was used for micro-computed tomography (CT) imaging of rats implanted with R3230AC mammary carcinoma. Three-dimensional vascular architecture of tumors was imaged at 100-micron isotropic resolution. The image data showed good qualitative correlation with pathologic findings. The approach holds promise for studying tumor angiogenesis and for evaluating anti-angiogenesis therapies.

**Keywords:** molecular imaging, liposome, angiogenesis, breast cancer, small animal imaging, micro-CT

## Introduction

Angiogenesis plays an important role in breast tumor growth and in the process of metastasis.<sup>1</sup> However, imaging angiogenesis using X-ray-based techniques has proved extremely difficult due to the small amount of contrast enhancement and the limited size of the tumor vessels. To surmount these difficulties, several conditions need to be met: a high concentration of contrast agent to provide adequate imaging signal, a stable contrast enhancement during the imaging procedure, and an imaging system with high spatial, contrast, and temporal resolution.

In the past few years, liposomes have been studied as excellent vehicles for drug delivery.<sup>2–7</sup> They have also been used as contrast agents in computed tomography (CT) and magnetic resonance (MR) imaging,<sup>8–10</sup> and as a delivery mechanism of contrast molecules to tumor regions.<sup>11</sup> Some members of this group have previously developed and evaluated a long circulating liposomal-based, nanoparticle contrast agent for blood pool imaging.<sup>10</sup> The agent has longer *in vivo* residence time than traditional contrast agents. The hypothesis of this study was that this new contrast agent would have utility in breast cancer imaging, providing long-lasting opacification of tumor microvasculature in micro-CT images of rats with implanted mammary tumors. The purpose of the study was to perform a proof-of-principle imaging experiment with this contrast agent by obtaining high-resolution images of mammary tumors in the rat model and qualitatively compare the images to histological specimens.

## Materials and methods

### Contrast agent formation

A lipid mixture (200 mM) consisting of 1,2-dipalmitoyl-sn-glycero-3-phosphocholine (DPPC), cholesterol, and 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-N-[methoxy(polyethylene glycol)-2000] (DSPE-MPEG2000) in the molar ratio 55:40:5

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was dissolved in ethanol at 70 °C. The ethanol solution was then hydrated for two hours with Iopamidol solution (Isovue® 370, Bracco Diagnostics, Princeton, NJ, USA). Liposomes were extruded in a 10-mL Lipex Thermoline extruder (Northern Lipids, Vancouver, British Columbia, Canada) with five passes and seven passes, respectively, through a 200- and a 100-nm Nuclepore membrane (Waterman, Newton, MA). Un-encapsulated iodine was then removed with a MicroKros® filtration module (Spectrum Labs, Rancho Dominguez, CA, USA) with a 400-kDa cut-off. Further detail on this synthesis process can be found in an earlier publication.<sup>10</sup>

The size distribution of the liposomes in the final formulation was determined by dynamic light scattering using a ZetaPlus Analyzer (Brookhaven Instruments, Inc., Chapel House, UK). The resultant liposomes were found to be 113 nm in size with a polydispersity index of 0.14. The iodine concentration in the liposomal formulations was  $65 \pm 5$  mg/ml, assessed based on the absorption of ultraviolet light at 242 nm measured with an ultraviolet-visible light spectrophotometer.

### **In vivo studies**

All animal procedures were performed using protocols approved by the Duke University Institutional Animal Care and Use Committee. Eight female Fischer 344 rats (National Cancer Institute, Frederick Cancer Center, Frederick, MD, USA) with a weight of 126.6–145.5 g (140 g average) were used for the study. Each rat was implanted with the R3230AC Rat Mammary Adenocarcinoma.<sup>12</sup> For tumor implantation, small pieces of tumor tissue (1–2 mm<sup>3</sup>) were obtained from tumor-bearing donor rats and transplanted subcutaneously in the right flank of each recipient animal. The animals were imaged approximately three weeks after implantation when the tumors reached 1–1.5 cm in diameter. They were imaged twice; at one and four hours after contrast injection.

### **Animal procedures**

The rats were anesthetized using an intra-peritoneal 50 mg/kg injection of sodium pentobarbital and 2 mg/kg butorphanol. Following endotracheal intubation, anesthesia was maintained with 2%–3% isoflurane, delivered by a custom-made ventilator.<sup>13</sup> Animals were mechanically ventilated at 90 breaths/min and at a tidal volume of 1.0 ml.

The iodinated liposomes were delivered via injection into a microcatheter inserted into the tail vein at a volume dose of approximately 0.02 ml/kg of animal weight. This was chosen as a standard volume for infusion as it results in

adequate opacification of the blood vessels and tumors, and was well tolerated by the animals.

Animals were placed vertically in a cradle. Flexible tubes carrying anesthesia gas and wires conveying physiologic telemetric signals were suspended from above to allow free rotation of the animal throughout the 190° rotation (Figure 1). All physiological telemetry data were displayed on a computer (LabVIEW, National Instruments, Austin, TX, USA). The monitored physiologic parameters were airway pressure tracing, measured with a solid-state pressure transducer on the breathing valve, and ECG, measured by electrodes taped to the footpads. Body temperature was maintained at 36.5 °C with heat lamps, an oral thermistor, and feedback controller (Digi-Sense®, Cole-Parmer, Chicago, IL, USA).

Following the study, the animals were euthanized with an overdose of nembutal. Tumors were excised, sectioned into 5 to 10 micron slices, and stained using hematoxylin and eosin (H&E) staining. Histological analysis was performed on the stained slices.

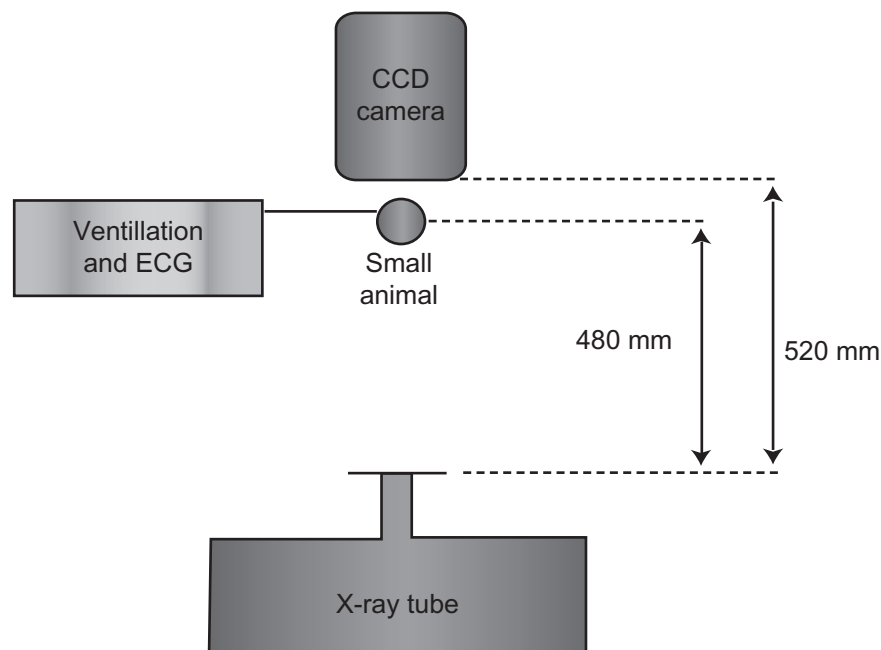
### **Micro-CT system**

The micro-CT system used, described previously<sup>14</sup> consisted of a high-flux, rotating anode X-ray tube (Philips SRO 09 50, Cleveland, OH, USA) with a dual 0.3/1.0-mm focal spot. The tube was capable of 10 msec exposures to minimize cardiac motion blur. The tube was coupled to a cooled charge-coupled device (CCD) camera with a Gd<sub>2</sub>O<sub>2</sub>S phosphor on a fiber-optic coupler with 3:1 de-magnification ratio (X-ray Image-Star, Photonics Science, East Sussex, UK). The camera had a 106 mm<sup>2</sup> active field-of-view with a pixel size of 51 μm. The tube and detector were mounted in the horizontal plane on aluminum frame components (80/20 Inc., Columbia City, IN, USA) (see Figure 1).

The animal was centered vertically at 40 mm from the detector face. The distance between the animal and X-ray source was 480 mm. This configuration essentially eliminated scatter, and focal spot blur by matching the penumbra to the Nyquist limiting frequency of the detector.<sup>14,15</sup>

### **Image acquisition and reconstruction**

Animals were imaged using 40 kVp, 400 mA, and 12.5 msec exposure time. This low-kVp radiographic technique was chosen to enhance soft-tissue contrast within the animal. Each image set consisted of 380 projections regularly spaced over a circular orbit of 190°. Each dataset acquisition per animal lasted for 8–10 minutes and led to an average absorbed dose of 27 cGy. All projections were acquired with ventilatory synchronization at end-expiration. Projection images were



**Figure 1** Placement of the animal in the micro-CT gantry.

**Abbreviations:** CCD, charge-coupled device; CT, computed tomography; ECG, electrocardiograph.

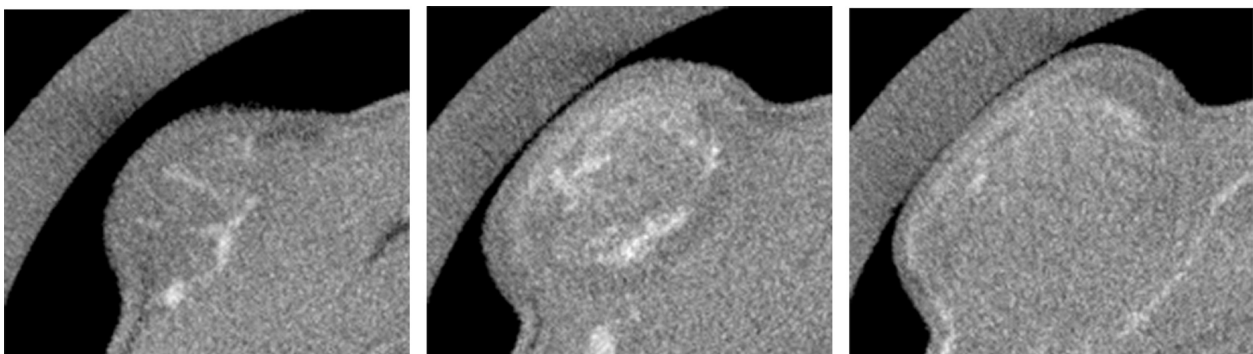
used to reconstruct tomograms with a Feldkamp algorithm using Parker weighting (Cobra EXXIM, EXXIM Computing Corp., Livermore, CA). The reconstructed datasets were 3D image arrays  $5.12 \text{ cm}^3$  in size with  $(0.1 \text{ mm})^3$  voxels.

Contrast enhancement was measured in regions of interest (ROIs) in the tumor as well as in the muscle. To tumor ROI was confined to within a vascular enhancement region within each tumor. The muscle region was approximately  $5 \times 5 \text{ mm}$  chosen on the contra-lateral flank of the animal. Tumor volumes and characteristics were obtained by hand segmentation using ImageJ software (<http://rsb.info.nih.gov/ij/>). Qualitative comparison was made of microvessel distribution, and regions of necrosis between CT images and corresponding histological sections.

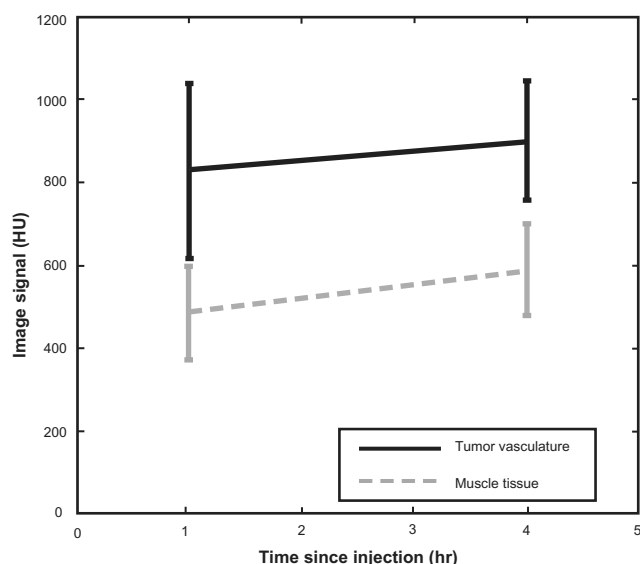
## Results

The liposomal contrast agent produced an average CT number of  $861 \pm 180 \text{ HU}$  (mean  $\pm$  standard deviation) in tumor vasculature and  $535 \pm 113 \text{ HU}$  in muscle, for an average contrast difference of  $326 \pm 151 \text{ HU}$ . Within the tumor, vessels as small as 200–300 microns were readily visible. The average tumor volume determined by hand segmentation was  $0.52 \pm 0.12 \text{ cm}^3$ . Enhancement, presumably of the microvasculature, of the tumor was clearly apparent on all images of the tumor (Figure 2).

Enhancement persisted for several hours post-injection (Figure 3). Even at four hours post-injection, the tumor signal remained high, especially in comparison to the signal in normal musculature. Figure 4 shows the



**Figure 2** Three tomographic images of an R3230AC tumor in a rat. Enhancement is clearly visible.



**Figure 3** Image value (in HU) at two time points for the tumor vasculature and normal tissue. Error bars reflect the standard deviation of the values within the ROI.

**Abbreviations:** HU, Hounsfield units; ROI, region of interest.

tumor enhancement at 1.2 and 4.2 hours after injection. The findings show the continued presence of the nanoparticle contrast agent in the blood pool at four hours post-injection.

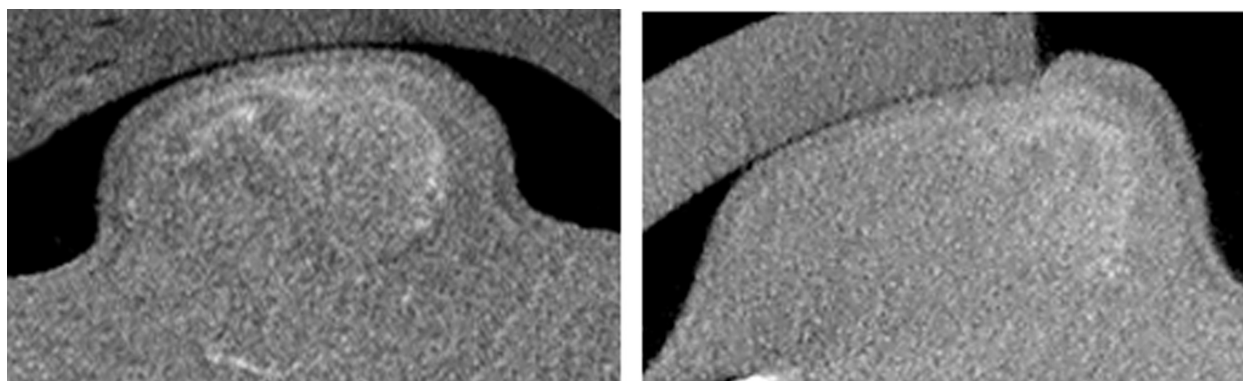
## Discussion and conclusions

Comparison between the micro-CT images and corresponding histological sections allows for interpretation of the imaging findings (Figure 5). Blood vessels are present on histological sections in the same areas where contrast enhancement is noted on the CT images. Moreover, evaluation of two typical regions of the tumor, a central slice through the core of the mass, and a peripheral slice along the capsular margin yield additional insight. A region of

central necrosis (pink-staining on histological section) is readily evident on the central section and corresponds to region of subtle hypodensity on the corresponding CT image (Figures 5a–c). This finding is in keeping with the known behaviour of liposomes which circulate within the bloodpool without significant extravasation. Therefore, enhancement would only be expected within the viable (purple-staining portions) of the mass. On the peripheral section, blood vessel proliferation is demonstrated along the margins of the tumor on histological section and correspond to peripheral enhancement of the tumor.

The findings demonstrate the utility of the nanoparticle contrast agent for imaging angiogenesis in breast cancer. The agent enabled an attenuation difference between the tumor vasculature and the muscle of  $326 \pm 151$  HU, allowing visualization of the 3D vascular architecture. This level of attenuation difference also enables estimations of tumor volumes and vascular percentages. As vascular enhancement was observed at four hours post-injection, its residence time in the blood pool was at least four hours. This was significantly longer enhancement, on the order of hours, compared to traditional, non-encapsulated contrast agents. In addition, the liposomal envelope shields the body contrast agent, resulting in elimination via the liver and spleen rather than excretion via the kidneys.

Several other liposomal-based agents have been utilized. Liposomes encapsulating both iodine-based and gadolinium-based molecules have been developed for multimodality imaging using CT and MRI.<sup>16,17</sup> These agents were prepared with a diluted starting iodine solution to incorporate the gadolinium-chelates, thus resulting in low iodine concentration within each liposome. Prior studies have not focused on breast cancer cell lines as a precursor for human applications.



**Figure 4** Persistent enhancement of the tumor of a rodent several hours after contrast injection of a rodent with liposomal contrast. These two images were acquired 1.2 hours (left) and 4.2 hours (right) after contrast injection. The images reflect positioning differences between two acquisitions.



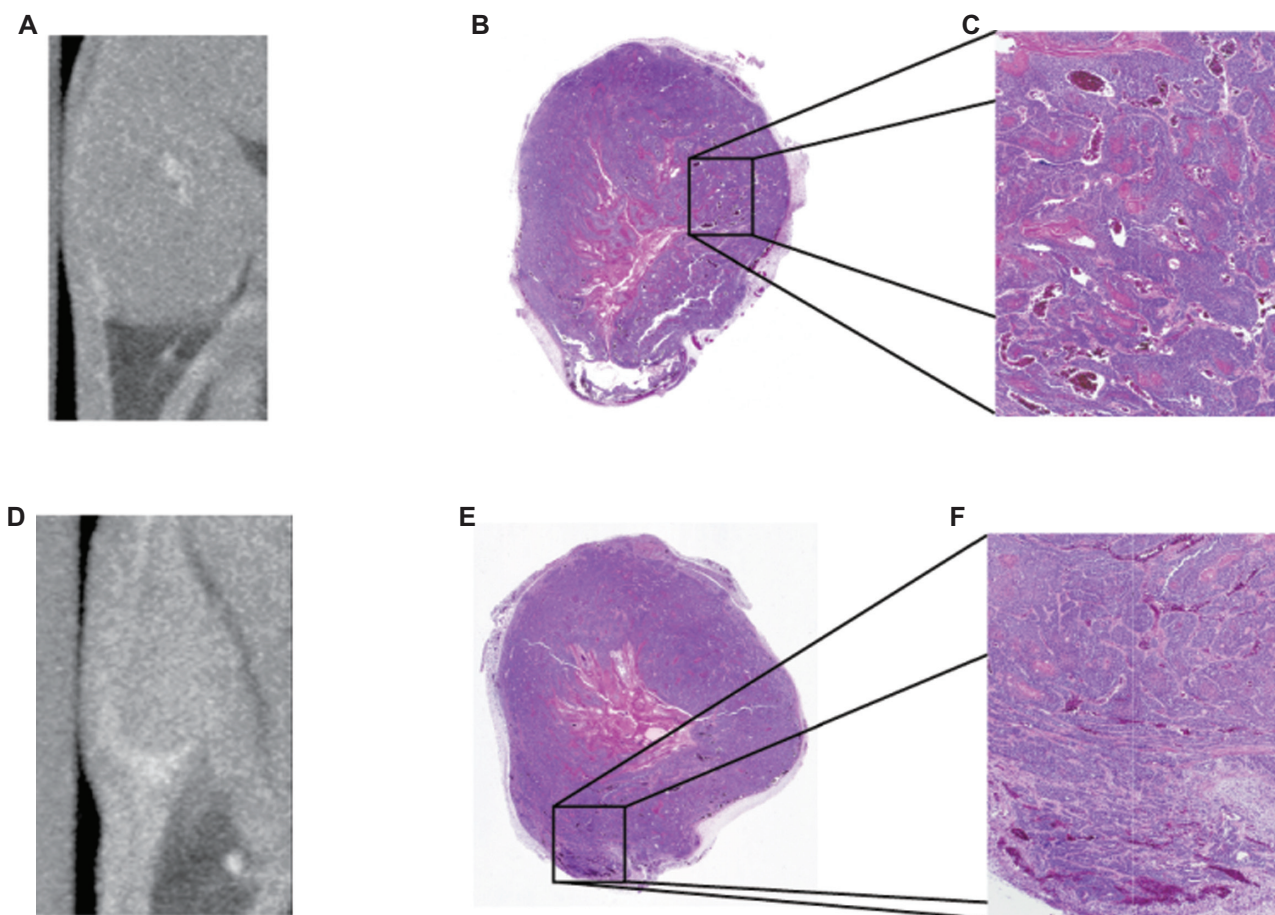
Liposomes containing iodine conjugated to the lipids have also been developed.<sup>18</sup> The conjugation of iodine to lipid results in low iodine:liposome ratio compared to our formulation, wherein the iodinated molecules are encapsulated within the core interior. Although not used in this study, liposomes with an even higher concentration of iodine (eg, 100 mg/ml) have been successfully used.<sup>10</sup> Increases in iodine concentration in combination with a diffusion-optimized size for the liposomes would allow for even better visualization of tumors in small animals and may open the door to delayed micro-CT imaging studies to examine tumor leakage via contrast agent extravasation.

While these findings are qualitative, due to the pilot nature of the study, they suggest that a liposomal, nanoparticle contrast agent is a promising tool for micro-CT

evaluation of tumor vascular architecture in small animals. As such, this approach may be useful for the study of tumor angiogenesis including the evaluation of antiangiogenic therapies in small animals with potential extensions and applications to human imaging. The findings pave the way for a future more extensive study with a larger cohort of animals to provide statistically justified quantitative results.

## Acknowledgments

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**Figure 5** Micro-CT images and micrographs of histological sections of the same R3230AC tumor implanted in a rat. For a central section of the tumor, this figure shows the CT image **A**) the corresponding histological section micrograph (low-power) **B**) and a high-power micrograph of the region contained within the box **C**). Focal enhancement demonstrated on the micro-CT image corresponds to a cluster of blood vessels in corresponding region of the histological section. For a peripheral region of the tumor, this figure shows the CT image **D**) the histological section (low-power micrograph) **E**) and a high-power micrograph of the region contained within the box **F**). In this slice, enhancement of the rim of the tumor is demonstrated. The corresponding region of the histological section demonstrates significant blood vessel growth. While the CT and pathology findings cannot be correlated quantitatively, they show similar qualitative indications.

**Abbreviation:** CT, computed tomography.

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