

Nano-Hydroxyapatite Coating Promotes Porous Calcium Phosphate Ceramic-Induced Osteogenesis Via BMP/Smad Signaling Pathway

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Background: The hierarchical porous structure and surface topography of calcium phosphate (CaP) bioceramics have a crucial impact on their osteoinductivity.

Purpose: To fabricate a biomimetic bone graft with an interconnected porous structure analogous to that of trabecular bone and a bioactive nanostructured surface with excellent osteoinductive potential.

Materials and methods: A biphasic CaP (BCP) substrate with highly porous structure was fabricated by an improved sponge replication method. Surface modification was performed by uniformly depositing a hydroxyapatite (HA) nanoparticle layer to create nHA-coated BCP scaffolds. The effects of these scaffolds on osteogenic differentiation of murine bone marrow-derived stem cells (BMSCs) were investigated in vitro, and their osteoinductivity was further assessed in vivo.

Results: The BCP and nHA-coated BCP scaffolds had similar trabecular bone-like architectures but different surface structures, with mean grain sizes of ~55 nm and ~1 µm, respectively. Compared with the BCP substrate, the nHA-coated BCP scaffolds favored cell adhesion and promoted osteogenic differentiation of BMSCs, as evidenced by upregulated expression of osteogenic genes, enhanced alkaline phosphatase activity, and increased osteocalcin production. This could be attributed to activation of the BMP/Smad signaling pathway, as significantly higher expression levels of BMPRI, Smad1, Smad4, and Smad5 were observed in the nHA-coated BCP group. The nHA-coated BCP scaffold not only maintained scaffold integrity but also induced ectopic bone formation when implanted into rabbit dorsal muscle in vivo for 90 days, whereas the BCP substrate underwent marked biodegradation that led to severe inflammation with no sign of osteogenesis.

Conclusion: The present study demonstrates the potential of this biomimetic bone graft with a trabecular framework and nanotopography for use in orthopedic applications.

Keywords: nano-hydroxyapatite, calcium phosphate ceramics, porous scaffolds, MSCs, osteoblastic differentiation, osteoinduction

Introduction

Calcium phosphate (CaP) bioceramics are of considerable interest in the orthopedic clinic for use as bone graft substitutes.^{1,2} It is well known that the physicochemical properties of CaP scaffolds influence their functionalities. Our previous studies have demonstrated that an interconnected porous structure is a prerequisite for the osteoinductivity and osteoconductivity of CaP bioceramics; it is generally believed that macropores (>100 µm) facilitate the in-growth of newly

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formed bone and blood vessels, whereas micropores (0.1–100 μm) are favorable for cell proliferation and growth.^{3–5} Thus, many techniques have been developed to fabricate porous scaffolds, including 3D printing, gas foaming, and freeze casting.^{3,6,7} The sponge replication method has attracted considerable attention as it can be used to readily make bone substitutes with highly porous structures, uniform pore geometry, and fine interporous connections that resemble the hierarchical structure of trabecular bone. These can be used to provide transportation highways and induce the growth of newly formed bone deep inside the host body.^{3,8,9}

Moreover, inspired by its compositional and structural similarity to the inorganic constituent of native bone, nano-sized hydroxyapatite (nHA) has become a research hotspot in the field of bone regeneration. Extensive studies have demonstrated that nHA has excellent bioaffinity and the ability to promote bone integration.^{10–12} For instance, Cai et al¹³ found that nHA could better upregulate gene expression of type I collagen and promote osteoblastic differentiation of bone marrow-derived mesenchymal stem cells (BMSCs) compared with a micro-sized structure. Recently, a unique phenomenon was discovered in which nHA particles showed suppressive and apoptotic effects on osteosarcoma cells but improved the viability and proliferation of BMSCs; this was believed to be closely related to particle size.^{14–17} These findings suggest that nHA not only induces osteogenesis to repair hard tissue injury but also promotes selective apoptosis of tumor cells. Thus, nHA has emerged as an ideal candidate for biomedical applications.^{18,19} However, it is difficult to endow CaP ceramics with both an interconnected porous structure and a bioactive nanostructured surface, as grains grow rapidly and pores collapse easily during the sintering process.¹ Therefore, multiple approaches have been developed to incorporate nHA particles with artificial bone grafts, including the deposition of nHA coating onto a scaffold surface, which provides a simple and effective means to functionalize an implant and improve its hydrophilicity and biocompatibility.^{20–22}

The aim of this study was to fabricate a biomimetic bone graft with both an interconnected porous structure analogous to that of trabecular bone and a bioactive nanostructured surface, via a combination of spongy replication and dip-coating technologies, and to investigate its osteoinductive potential *in vitro* and *in vivo*.

Materials And Methods

Fabrication Of BCP Scaffolds With Trabecular Framework

Biphasic CaP (BCP) powders consisting of about 80% tricalcium phosphate (TCP) and 20% HA were supplied by the Engineering Research Center in Biomaterials of Sichuan University, China. To prepare open porous bodies, an improved sponge replication method was adopted, following a procedure similar to that described previously.²³ In brief, cylindrical polyurethane (PU) foams with high porosity (40 pores per inch) were impregnated into a series of BCP slurries with decreased viscosities, prepared by thoroughly dispersing gradually reduced amounts of BCP powders in a liquid containing polyvinyl alcohol (as a binder) and 25% ethyl alcohol (as a dispersant). After several repetitions of coating/centrifuging/drying, a uniform slurry coating layer was formed on the spongy struts. Then, the green bodies were cut into discs ($\Phi 14 \times 3.5$ mm), which were sintered at 1100°C for 2 hrs to fabricate porous BCP scaffolds (ie, the BCP substrate).

Preparation Of BCP Scaffolds With Nanostructured Surface

An nHA surface layer was coated onto the porous BCP discs. In brief, nHA particles were synthesized by combining wet chemical synthesis and hydrothermal treatment.²⁴ A uniform nHA suspension was obtained by dispersing nHA particles in ultrapure water (upH_2O) at a concentration of 20 mg/mL and treating with ultrasonic vibrations for 2 hrs. Pre-made porous BCP discs were soaked in the nHA slurry, kept under vacuum at 10^{-3} Pa for 0.5 hrs, washed with upH_2O , and then dried at 60°C. The above process was repeated three times to ensure that the surface of the porous ceramic bodies was fully covered by nHA particles. Finally, heating at 650°C for 4 hrs was used to fabricate BCP scaffolds with nanostructured surfaces (ie, nHA-coated BCP). Prior to *in vitro* and *in vivo* studies, the ceramic discs were sterilized by dry heat at 250°C for 2 hrs.

Scaffold Characterization

The surface morphology and micro-nano structure of nHA-coated and uncoated discs (BCP substrate and nHA-coated BCP) were observed by stereo microscopy (Leica, M205FA, Germany) and scanning electron microscopy (SEM, JSE-5900LV, Japan). To quantitatively measure scaffold structure, 10 randomly selected areas were

selected from each optical or SEM image ($n=3$) and analyzed using the Image J 1.44p software. Energy dispersive spectrometry (EDS) was performed to characterize the chemical composition of the scaffolds.

To investigate changes in the micro-nano surface topography of the porous ceramic scaffolds after the coating process and to eliminate interference from the 3D geometry, BCP ceramic discs with the same grain sizes but a smooth surface were obtained by uniaxially compacting the powder slurry and calcining it into tablets with surface structure that resembled that of the BCP substrates. Subsequently, nano-sized HA particles were deposited onto these tablets, following the process described above, to mimic the surface structure of porous nHA-coated BCP scaffolds. Finally, their surface area and roughness were measured by atomic force microscopy (AFM, Asylum Research, MFP-3D, Oxford Instruments, UK) in the trapping mode. The average surface roughness of each sample was quantified based on six randomly selected fields from three images.

Cell Seeding And Cell Morphology

Murine BMSCs were cultured and seeded as previously described.²³ In brief, BMSCs were cultured in α -minimum essential medium (α -MEM, Gibco, USA) containing 10% v/v fetal bovine serum (Gibco, USA), 100 U/mL penicillin, and 100 μ g/mL streptomycin (Gibco, USA). After 90% confluence was reached, BMSCs were detached with 0.25% trypsin solution, evenly dispersed in culture medium, and seeded onto porous ceramic discs (5×10 cells/disc). Culture was performed in a humidified incubator at 5% CO_2 and 37°C. At day 3, the samples were harvested, fixed in 2.5% glutaraldehyde at 4°C, and then subjected to gradient dehydration using ethanol with volume fractions of 30%, 50%, 70%, 80%, 90%, and 100% (twice). After sputter coating with gold, the samples were examined by SEM.

Real-Time Quantitative Reverse Transcription PCR

BMSCs were seeded onto porous ceramic discs (1×10^5 cells/disc). Cells cultured on acid-cleaned glass coverslips were used as a control. Real-time quantitative reverse transcription PCR (qRT-PCR) analysis was used to assess the expression of specific osteogenic genes and cell signaling-related genes associated with the BMP/Smad signaling pathway, including bone morphogenetic protein-2 (BMP-2), runt-related

transcription factor (Runx-2/Cbfa-1), osterix (OSX), alkaline phosphatase (ALP), bone sialoprotein (BSP), osteopontin (OPN), BMP receptor type IA (BMPRI), and mothers against decapentaplegic homolog 1 (Smad1) Smad4, and Smad5. Primers are listed in Table 1. In brief, at 1, 4, and 7 days, total RNA was extracted using an RNeasy Mini Kit RNA (Qiagen, Germany) from BMSCs seeded on BCP and nHA-coated BCP scaffolds ($n=3$) and then reversed into cDNA using an iScript™ cDNA Synthesis kit (Bio-Rad, USA). Gene expression was detected using a CFX96 real-time thermal cycler system (Bio-Rad, USA) with SsoFast™ EvaGreen® Supermix (Bio-Rad, USA). Relative expression of each target gene was calculated by the $\Delta\Delta\text{Ct}$ -value method, using GAPDH as an internal reference to normalize the test results.

ALP Activity And OCN Secretion

To further evaluate material-stimulated osteoblastic differentiation, both osteogenic marker proteins, ie, ALP and

Table 1 Primer Sequences Used For Real-Time Quantitative Reverse Transcriptase Polymerase Chain Reaction (qRT-PCR)

Gene	5' to 3'	Primers
BMP-2	Sense Anti-sense	GATCTGTACCGCAGGCACTCA AAGCTTCCTGTATCTGTTCCCG
Runx-2	Sense Anti-sense	AGCCTCTTCAGCGCAGTGAC CTGGTGCTCGGATCCCAAA
OSX	Sense Anti-sense	ATGGCGTCCTCTCTGCTTG TGAAAGGTCAGCGTATGGCTT
ALP	Sense Anti-sense	ATCTTTGGTCTGGCTCCCATG TTTCCCGTTACCGTCCAC
BSP	Sense Anti-sense	GTTTCAGCAGACAACCCACC CTCCCCGTTCTCGTTGTCATA
OPN	Sense Anti-sense	CCCTCGATGTCATCCCTGTT CCCTTCCGTTGTTGTCCTG
BMPRI	Sense Anti-sense	CCTGTTGTTATAGGTCCGTTCTT AGCTGGAGAAGATGATCATAGCA
Smad1	Sense Anti-sense	GGATGAGCTTCGTGAAGGGTTGG GCAAGAGACGGAAGCCACAGG
Smad4	Sense Anti-sense	GTGACTGTGGATGGCTATGTGG GCAACCTCGCTCTCTCAATCG
Smad5	Sense Anti-sense	ATCCCACTGTCTGTAAG TTTATCCAGCCACTGAAGAGG
GAPDH	Sense Anti-sense	TGGTGAAGGTCGGTGTGAAC CCATGTAGTTGAGGTCAATGAAGG

osteocalcin (OCN), were quantitatively measured. After co-culturing BMSCs with BCP and nHA-coated BCP scaffolds for 7 and 14 days, cells were harvested, lysed using RIPA Lysis Buffer (Boster, Wuhan, China), and centrifuged at 8000 rpm for 10 mins to collect the supernatant. Then, according to the manufacturer's specification, ALP and OCN contents were determined using a commercialized ALP assay kit (SensoLyte® pNPP ALP assay kit, CA, USA) and murine osteocalcin ELISA kit (Cloud-clone Corp, USA), respectively. The relative amount of ALP and OCN per unit protein was normalized to the total protein content, which was measured with a Pierce™ BCA Protein Assay Kit (Thermo Scientific, USA).

In Vivo Animal Studies

The animal experimental procedure was approved by the Animal Care and Use Committee of Sichuan University, China, following the *Guide for the Care and Use of Laboratory Animals* published by the Chinese National Academy of Sciences. Three healthy male New Zealand white rabbits (2–2.5 kg) purchased from Chengdu Dashuo Experimental Animal Co., Ltd. were subjected to intramuscular implantation surgery to evaluate the osteoinductivity of the two porous scaffolds. Briefly, each rabbit was anesthetized by pentobarbital sodium with a dose of 40 mg per kg body weight. Then, three longitudinal blunt incisions (2 cm in length) spaced about 3 cm apart were made within the dorsal muscle on each side of the bilateral spine and filled with porous scaffolds (one per incision). Finally, incisions were sutured layer by layer. All rabbits were able to function normally post-surgery. Three months after implantation, samples (n=9) were harvested and fixed in 4% phosphate-buffered paraformaldehyde solution for 5 days before further analysis.

Histological And Immunofluorescent Analyses

The fixed samples were decalcified with 10% EDTA (pH 7.4), dehydrated in ascending concentrations of ethanol from 70% to 100%, and then embedded in paraffin. Each specimen was transversely cut into thin sections (5 µm in thickness) parallel to the direction of the disc plane using a microtome (Leica, Germany). Finally, some tissue sections were stained with hematoxylin and eosin (HE) and examined by Panoramic 250/MIDI (3D HISTECH, Hungary) and CaseViewer 2.0 software for histological observation. The others were subjected to co-staining with DAPI,

rabbit-anti-mouse osteocalcin primary antibody, and goat-anti-rabbit fluorescent secondary antibody (Servicebio, China), and visualized under a fluorescence microscope (Leica, Germany) for immunofluorescence analysis.

Statistical Analysis

All quantitative measurements were calculated from at least three test values and presented as mean ± standard deviation. Statistical analysis was performed using one-way analysis of variance. A p-value of less than 0.05 was considered to indicate a statistically significant difference.

Results

Characterization Of Biomimetic Scaffolds

The scheme used to fabricate the nHA-coated BCP scaffold is illustrated in Figure 1A. Stereo microscopy photos and SEM images (Figure 1B and C) showed a highly interconnected porous structure for both ceramics (Φ10.7×3.0 mm) that replicated the texture of a PU sponge. The images also showed that the BCP and nHA-coated BCP scaffolds exhibited similar trabecular bone-like structures with open macropores (~700 µm); many micropores (<10 µm) were also present on the scaffold structures (~180 µm). However, the surface architecture of the nHA-coated scaffolds was significantly different from that of the BCP substrates, in that the former exhibited a nanoparticle covering and an increased number of nanopores (<100 nm), whereas the latter was composed of micron-scale grains. Cross-section images indicated that an nHA particle surface layer with a thickness of approximate 1 µm was uniformly deposited onto the BCP substrate. The scaffolds were weighed before and after coating to quantify the nHA layer; the nHA coating process increased the total mass by $3.63 \pm 0.6\%$. EDS analysis (Supplementary Figure 1) showed that both the coating layer and substrate were mainly composed of Ca, P, and O, although the ratios of calcium to phosphorus were significantly different in the two cases, with values of about 1.665 and 1.529, respectively. These findings were consistent with the X-ray diffraction results of our previous study,²³ suggesting that the substrate comprised both an HA phase (Ca/P=1.67) and a TCP phase (Ca/P=1.5), whereas the coating layer consisted of pure HA phase.

Representative AFM images revealing the surface micro-nano structure are shown in Figure 2. As can be seen from the topographic 3D view, the BCP specimen had a relatively smooth surface with a flaky texture

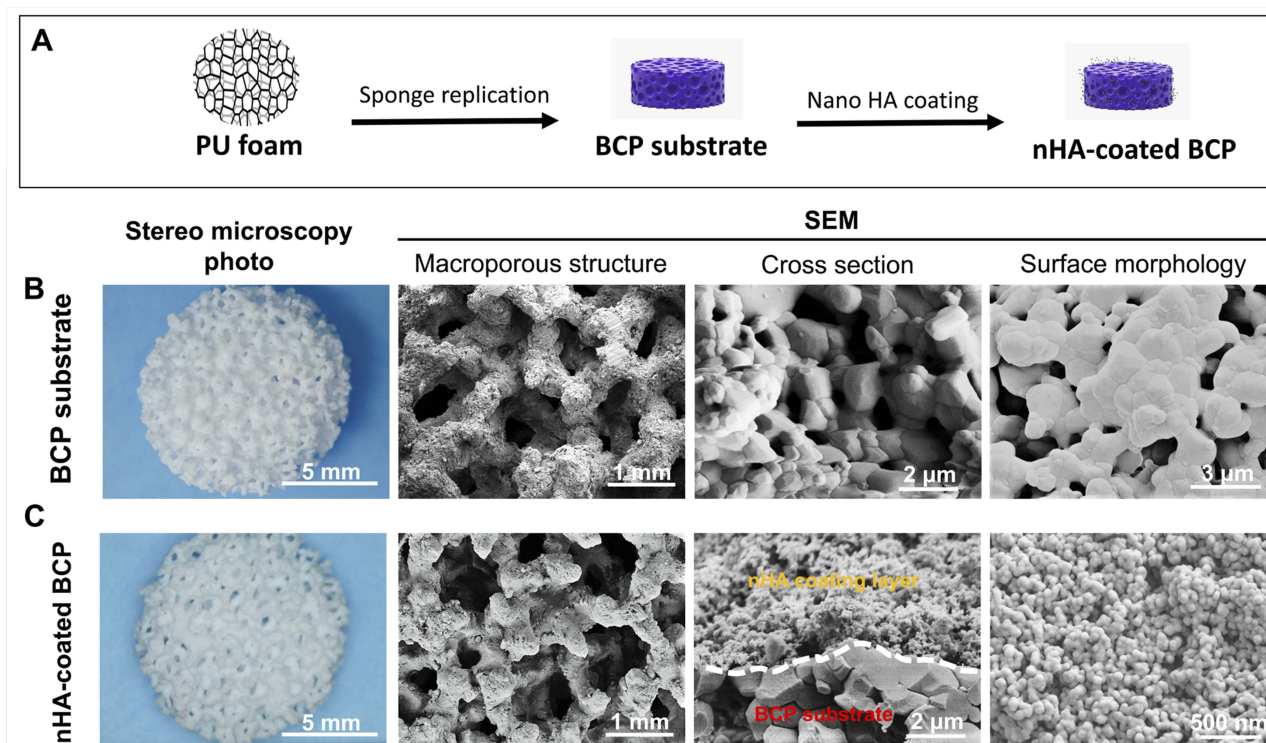


Figure 1 Scheme illustration for the fabrication process of BCP substrate and nHA-coated BCP scaffold (A). Stereo microscopy photos, SEM images of macroporous structure, cross-section, and surface morphology for BCP substrate (B) and nHA-coated BCP (C).

(Figure 2A), whereas the nHA particles resulted in abundant nano-sized peaks and valleys on the nHA-coated specimen (Figure 2C). The topographic top view of the surface given by the AFM images demonstrated that the surface of the nHA-coated specimen (Figure 2D) had a dramatically smaller grain size than that of the BCP scaffold (Figure 2B). The surface grain size and surface roughness of both ceramics are summarized in Table 2. Consistent with the SEM observations (Figure 1), the mean surface grain sizes of the BCP and nHA-coated samples were 961.1 ± 420.1 nm and 55.9 ± 11.7 nm, respectively. AFM analysis also gave a surface roughness of 67 ± 5 nm for the BCP scaffold, and a slightly higher value of 72 ± 4 nm for the nHA-coated sample.

Cell Adhesion And Morphology

As cells can be significantly affected by the surface topography of bioceramics, in particular, nano-scale surface structure, the adhesion and spreading of BMSCs on the two trabecular-like scaffolds (ie, BCP substrates and nHA-coated BCP scaffolds) were observed using SEM after 3 days of culture. As shown in Figure 3, BMSCs presented typical fibroblast-like phenotypes and attached firmly to both scaffolds. High-magnification images (Figure 3C and F) showed

that BMSCs seeded on nHA-coated scaffolds, compared with those on BCP substrates, exhibited more slender extending processes (filopodia), which tightly grasped nano-scale grains.

Osteogenic Differentiation Of BMSCs

Expression Of Osteogenic Genes

qRT-PCR was used to measure the expression levels of six typical osteogenic genes (BMP-2, Runx2, OSX, ALP, BSP, and OPN) in BMSCs co-cultured with BCP and nHA-coated scaffolds for 1, 4, and 7 days (Figure 4). Expression levels of all osteogenic genes in the BCP group dramatically decreased over time, whereas gene expression in the nHA-coated BCP group remained at a relatively high level during the whole culture period. Moreover, the BMP-2, OSX, and OPN genes had the highest expression levels in the nHA-coated BCP group at all time points. Compared with the BCP group and blank controls, the nHA-coated BCP scaffolds showed significantly upregulated expression of the Runx2 gene at day 4, and increased expression of ALP and BSP genes at days 4 and 7. Overall, these results demonstrate that the introduction of an nHA coating onto trabecular

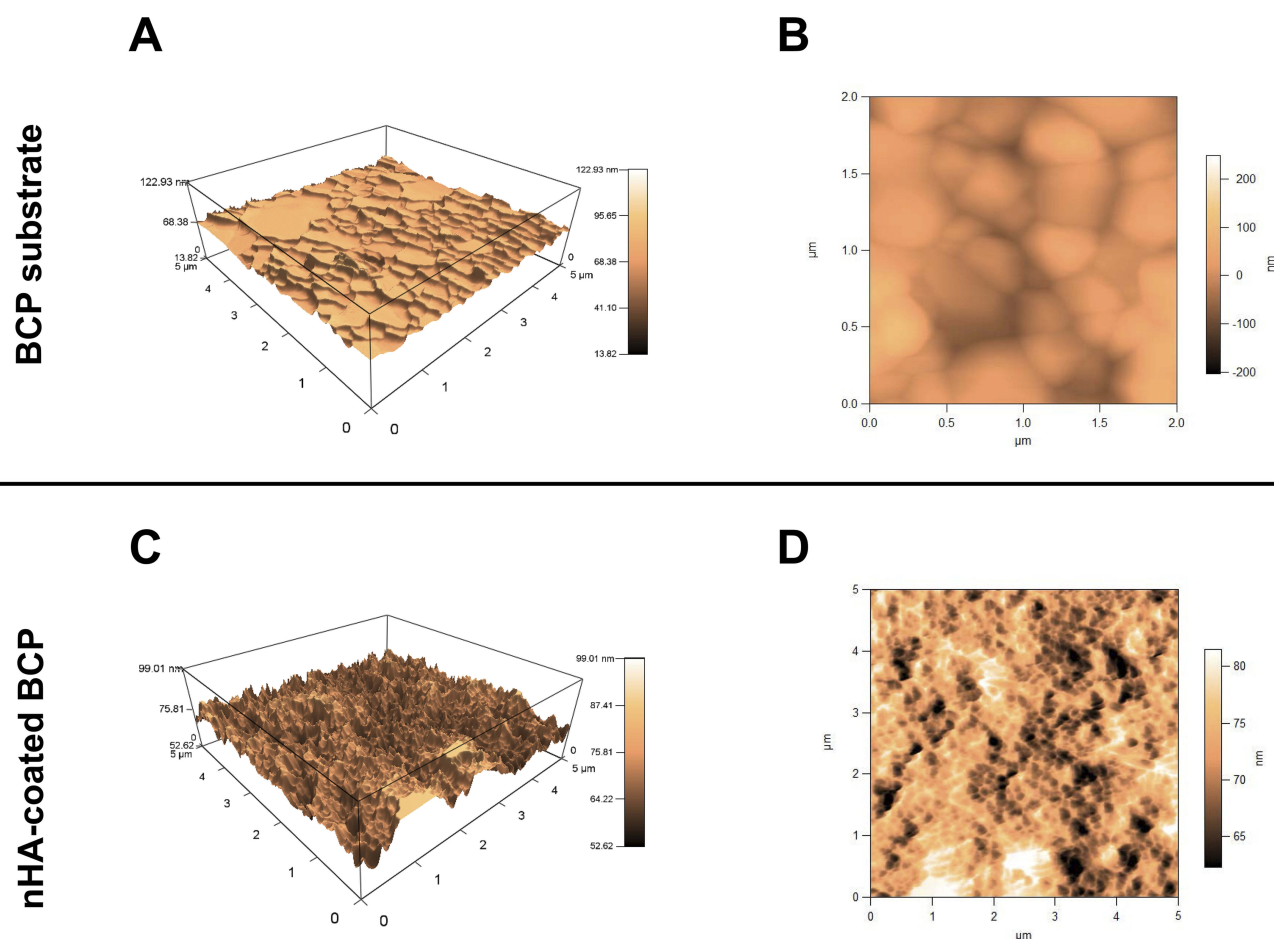


Figure 2 AFM topographic images for 3D view (**A**) and top view (**B**) of BCP substrate; 3D view (**C**) and top view (**D**) of nHA-coated BCP scaffold.

Table 2 Scaffold Characterization Of BCP Substrate And nHA-Coated Scaffold

	BCP Substrate	nHA-Coated Scaffold
Mean grain size (nm)	961.1 ± 420.1	55.9 ± 11.7
Ra (nm)	67 ± 5	72 ± 4

bone-like scaffolds can enhance the expression of several important genes related to osteoblastic differentiation.

ALP Activity And OCN Production

Quantitative analyses were used to determine intracellular ALP activity and OCN production in BMSCs cultured on the two trabecular-like scaffolds. As shown in [Figure 5](#), ALP activity was significantly higher in the nHA-coated BCP group than in the BCP group at day 7 ($p < 0.01$). The control group always had the lowest ALP activity at day 7 and 14 ($p < 0.05$, [Figure 5A](#)). There was

a slight decrease in ALP activity and an increase in OCN production by BMSCs with increasing culture time, showing statistically significant differences among the different groups with ALP and OCN values from high to low: nHA-coated BCP > BCP > Control ($p < 0.05$, [Figure 5B](#)).

Expression Of Genes Related To BMP/Smad Signaling Pathway

Furthermore, to explore the possible mechanism of osteogenesis stimulated by nHA coating layers, mRNA expression levels of several genes related to BMP/Smad signaling (BMPRI, Smad1, Smad4, and Smad5) were investigated using qRT-PCR. As shown in [Figure 6](#), compared with the BCP group, the nHA-coated BCP group showed significantly upregulated expression of the BMPRI gene at day 1 ($p < 0.01$). The Smad genes (Smad1, Smad4, and Smad5) reached their highest expression levels in BMSCs cultured on nHA-coated BCP scaffolds at day 1 and 4 ($p < 0.05$).

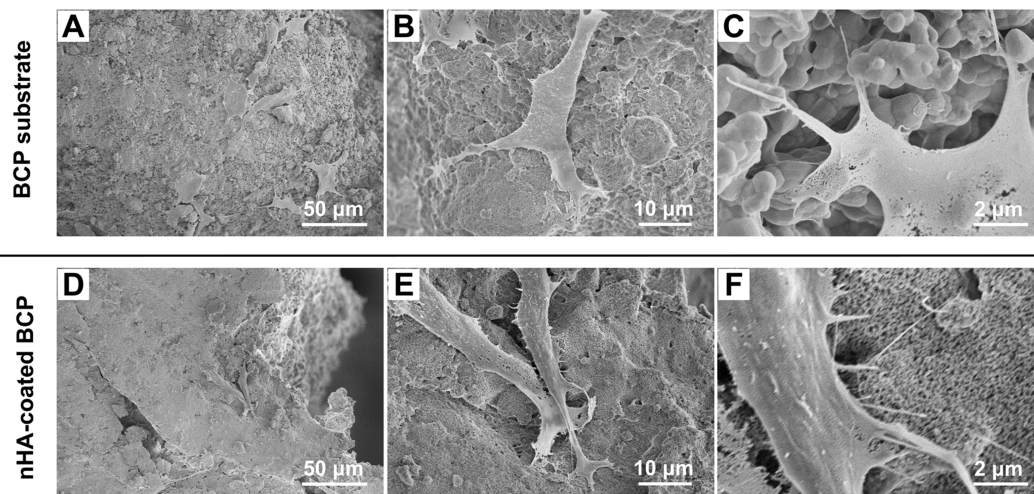


Figure 3 SEM micrographs of BMSC attachment onto the surface of BCP substrates (A–C) and nHA-coated BCP scaffolds (D–F) at day 3.

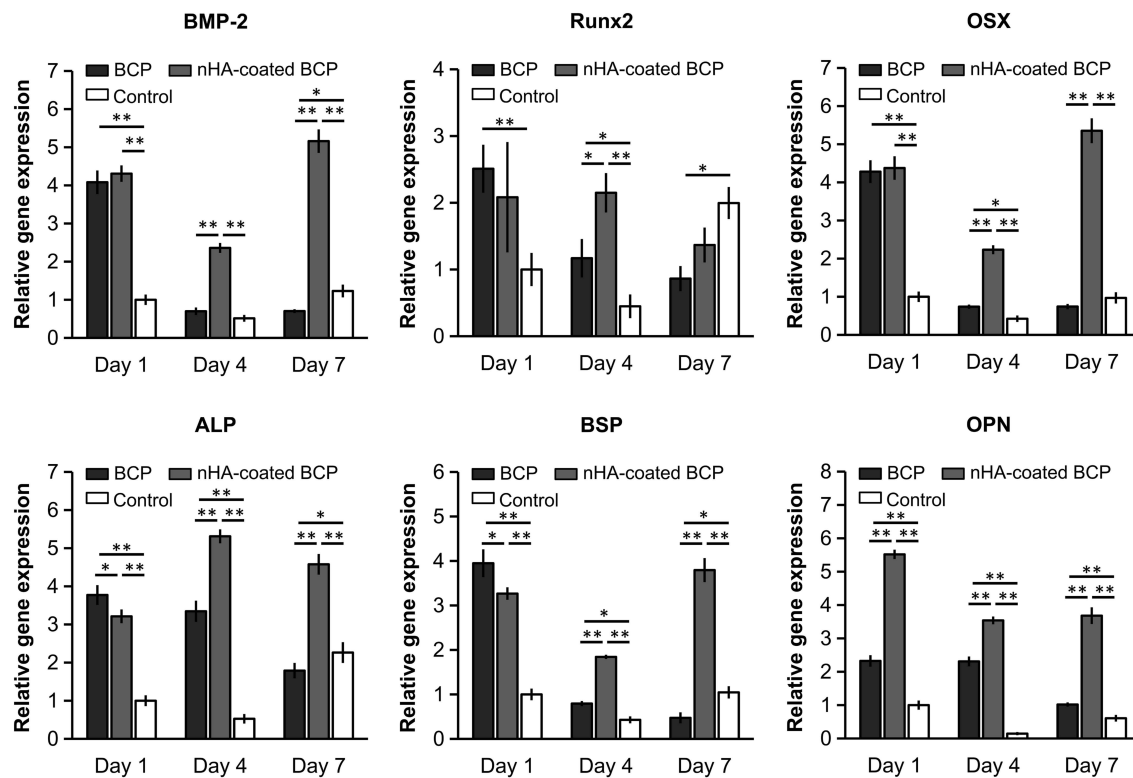


Figure 4 The expression of specific osteogenic genes (BMP-2, Runx2, OSX, ALP, BSP and OPN) in BMSCs cultured in BCP, nHA-coated BCP and control group for 1, 4 and 7 days. * refers to $p < 0.05$, ** refers to $p < 0.01$.

In Vivo Ectopic Bone Formation

To further evaluate the osteoinductive capacity of the two trabecular-like scaffolds, a rabbit intramuscular implantation model was established by implanting two scaffolds (BCP and nHA-coated BCP) into the rabbit's back muscle for 90 days (Figure 7A).

Micro-CT Analysis

Micro-CT (μ -CT) imaging was used to examine structural changes in the BCP and nHA-coated BCP scaffolds before and after implantation (Figure 7B and C). Prior to implantation, both biomimetic scaffolds had architectural structures similar to that of trabecular bone, consistent with the microscopy

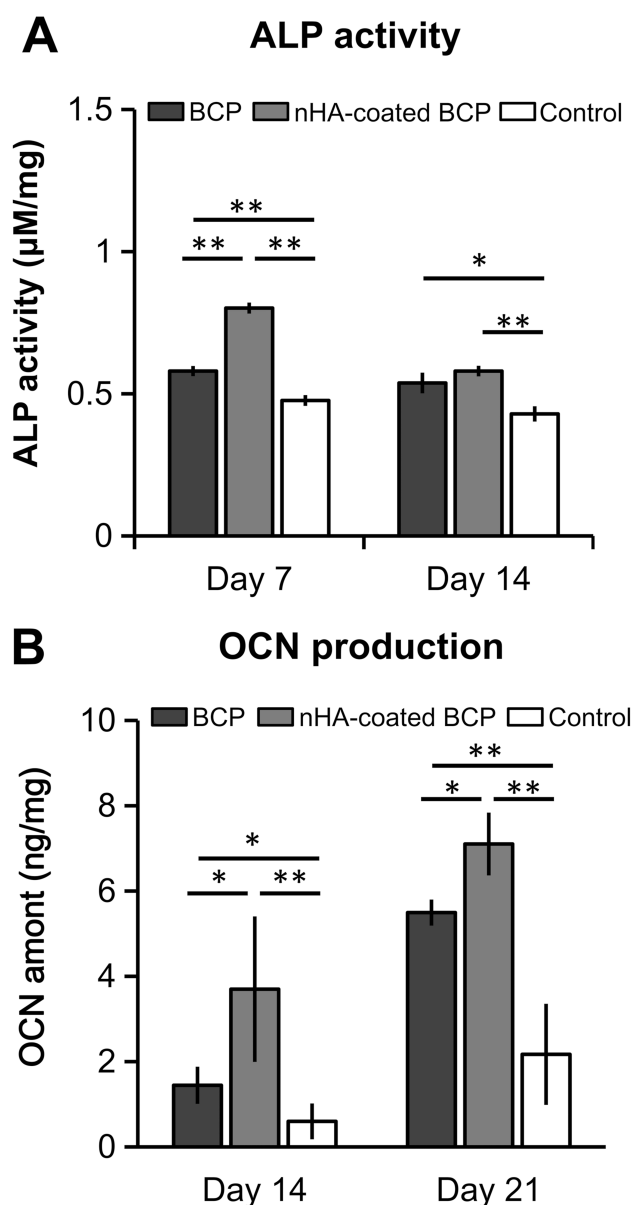


Figure 5 Intracellular ALP activity (A) and OCN production (B) for BMSCs cultured in BCP, nHA-coated BCP and control group. * refers to $p < 0.05$, ** refers to $p < 0.01$.

observations (Figure 1B and C). After intramuscular implantation for 90 days, however, the BCP scaffold had mostly disintegrated and its original macroporous structure was largely destroyed, whereas the nHA-coated BCP scaffold essentially maintained its original geometric shape.

Histological Analysis And Immunofluorescent Staining

The HE staining results also showed that almost all of the BCP scaffolds were degraded after 90-day implantation, with a few sporadic fragments remaining (Figure 8A).

Muscle tissue filled the void resulting from degradation, accompanied by a mass of inflammatory cells in the surrounding area (indicated by a yellow arrow in the figure). However, the nHA-coated BCP samples maintained their original highly porous architecture (Figure 8B), with invasion of blood vessels and the ingrowth of cells, and had formed ectopic bone after 90 days of implantation, as evidenced by the bone lacuna and mature bone cells observed at the periphery of the scaffold struts (indicated by a black arrow in the figure). Immunofluorescence staining images (Figure 8C and D; green fluorescence represents OCN expression and blue indicates cell nuclei) results showed the presence of osteocalcin protein in the nHA-coated BCP group but not in the BCP group.

Discussion

CaP bioceramics have been extensively used as bone substitutes in orthopedic treatment. It is believed that proper material design can endow CaP bioceramics with excellent osteoinductivity that enhances self-renewal and self-repair in patients.^{25,26} However, the bioactivity of conventional CaP ceramics is generally lower than that of human-derived bone grafts (autografts and allografts). This may be because although CaP ceramics resemble the inorganic components of natural bone tissues, it is difficult for them to recapitulate the complex hierarchical structure of bones. Native cancellous bone is composed of collagen fibers and HA nanocrystals, and has a highly interconnected porous structure that contributes to its unique mechanical and biological properties.²⁷ From a biomimetics perspective, it is thought that the biological performance of CaP ceramics could be markedly improved by fully simulating the hierarchical structure of cancellous bone and introducing biomimetic nano-sized HA particles. Therefore, in this study, porous BCP scaffolds were prepared via an improved sponge replication method to mimic the trabecular framework. A surface layer of nHA was uniformly deposited onto this BCP substrate to construct a bionic surface with nanocrystalline structure (ie, nHA-coated BCP). Stereo microscopy and SEM images showed that the BCP substrates and nHA-coated BCP scaffolds had similar trabecular bone-like structures with high porosity and excellent interconnectivity, consistent with previous studies, indicating that both scaffolds had abundant macropores (200–700 μm), minor pores (20–50 μm), micropores (0.1–2 μm), and nanopores (30–100 nm).²³ Micro-to-macropores are thought to benefit nutrient exchange and favor tissue ingrowth.^{3,8,9} However, it has also been

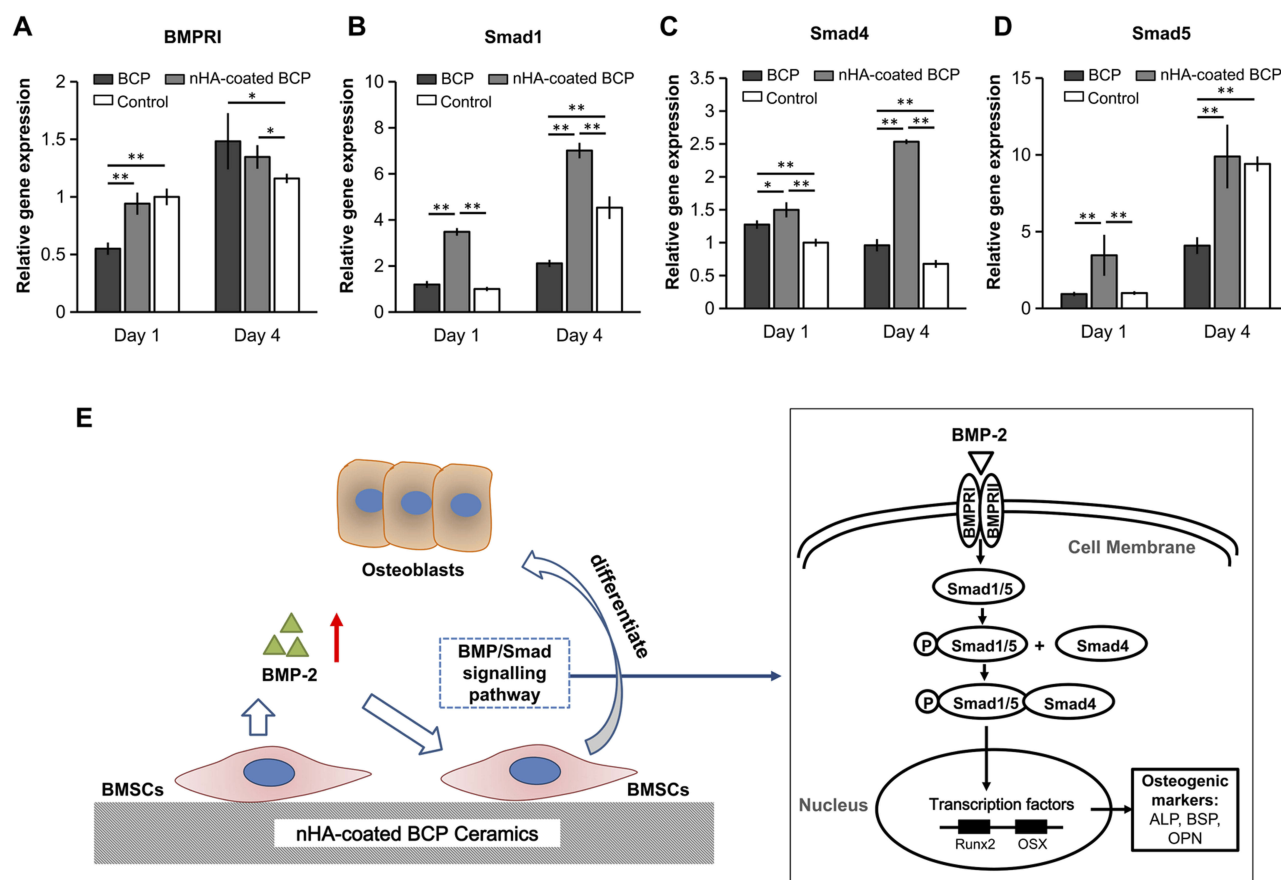


Figure 6 The expression of genes related to BMP/Smad signals – BMPRI (A), Smad1 (B), Smad4 (C), and Smad5 (D). Schematic illustration of the possible involvement of BMP/Smad signaling pathway in pro-osteogenic effects of nanostructured surface (E). * refers to $p<0.05$, ** refers to $p<0.01$.

suggested that the nHA coating process could significantly increase numbers of micropores and nanopores,²³ and indeed the two trabecular bone-like scaffolds had markedly different surface structures: the nHA-coated BCP scaffolds were covered uniformly by a layer of nano-sized HA particles with mean grain sizes of ~55 nm, whereas the BCP substrates had a micron-scale surface architecture with mean grain sizes of ~1 μ m.

In recent years, nanostructured surfaces have received increasing attention. Extensive studies have demonstrated that use of an nHA coating layer confers strong adsorption capacity and provides abundant active sites for cell attachment owing to its large surface area and high surface energy.^{28–30} Thus, the roles of nanostructured surfaces on biomimetic bone grafts in material-induced osteogenesis were evaluated by comparing the osteoinductivity of BCP and nHA-coated BCP in vitro and in vivo. SEM images showed that nHA-coated BCP scaffolds, compared with BCP alone, better enabled BMSCs to stretch out filopodia

to grasp nano-scale grains and to adhere more tightly to the scaffold surface, consistent with previous findings.^{31,32} It has been reported that when the grains of a CaP ceramic were decreased to nano-size, cell morphology was affected more by nanotopography than by chemical composition.^{33,34} Thus, the differential cellular adhesive patterns and morphology of BMSCs could be attributed to the distinct surface structures of the BCP and nHA-coated BCP scaffolds. The nHA-coated group was uniformly covered by nano-sized particles (less than 100 nm in size), resulting in a high specific surface area and abundant active calcium and phosphate sites, which could be beneficial to cell adhesion and spreading. Moreover, nano-scale grains might be more suitable for the anchorage of cellular filopodia, which are also nano-sized.³³

In nature, osteogenesis requires the cascade activation of a series of osteogenic-specific marker genes and proteins, including BMP-2, OSX, Runx2, ALP, and OCN. Runx2, OSX, and ALP are widely considered to be early markers

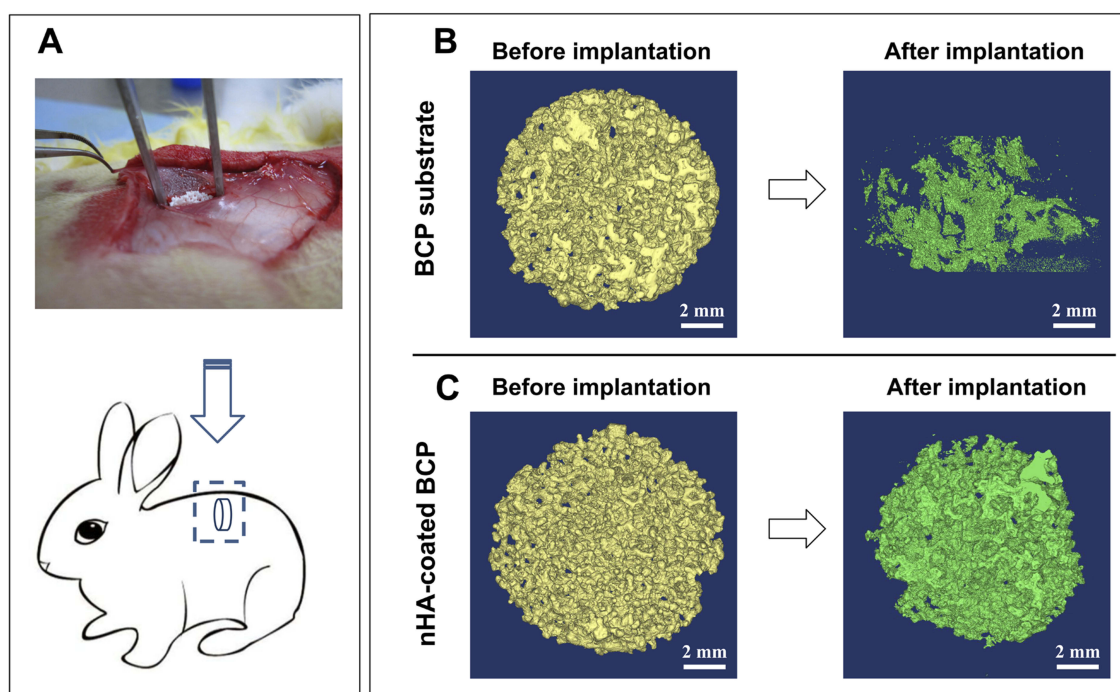


Figure 7 A rabbit intramuscular implantation model for the evaluation of material's osteoinductivity (A); μ -CT images for structural changes of BCP (B) and nHA-coated BCP scaffolds (C) before and after implantation.

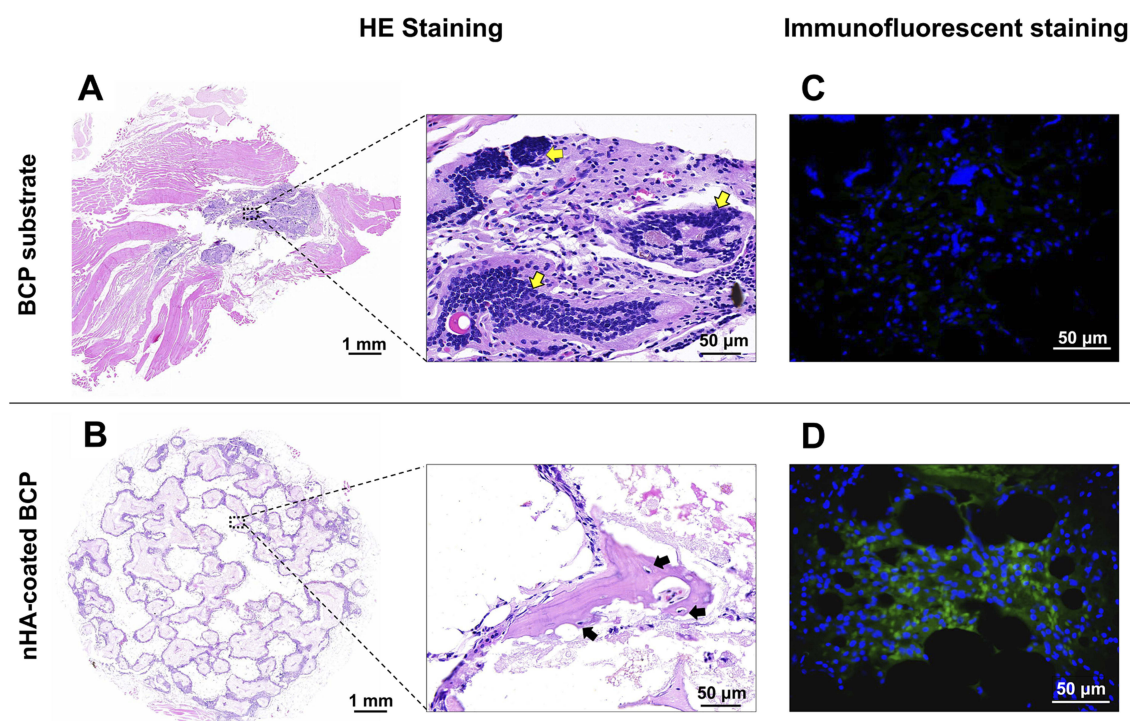


Figure 8 HE histological analysis of in vivo ectopic bone formation ability for BCP (A) and nHA-coated BCP (B) scaffolds after implanted in back muscles of rabbits for 90 days. Yellow arrows: inflammatory cells; black arrows: bone formation as evidenced by osteocytes settled in bone lacuna. Immunofluorescent staining of osteocalcin (OCN) for BCP (C) and nHA-coated BCP (D) groups. Green and blue colors represented osteocalcin and DAPI-stained nuclei, respectively.

for pre-osteoblasts, whereas BSP, OPN, and OCN are late markers for mature osteoblasts.^{35,36} It is well known that

cell differentiation is affected by interactions at the interface between the material surface and cells.³⁷ Here, qRT-PCR

analyses showed that the two trabecular bone-like scaffolds enhanced expression of osteogenic marker genes compared with blank controls. The nHA-coated BCP group with a nano-sized HA coating layer exhibited significantly higher expression of osteogenesis-associated genes than the BCP group with microtopography, in good agreement with previous research.^{29,38–40} Moreover, ALP and OCN are recognized as important specific proteins for osteogenic differentiation of BMSCs at the early and late stage, respectively.^{36,41} In this study, biochemical analyses and ELISA assays showed higher ALP activity in the early stages (day 7), whereas OCN production increased with culture time. Compared with the BCP group, the nHA-coated group dramatically promoted ALP activity and OCN production at each time point. This suggests that the nHA-coated scaffolds with nanostructure surface were more beneficial for direct BMSC differentiation into an osteoblast lineage, compared with BCP substrates with micro-sized grains. These findings are consistent with those of previous studies, which showed that nano-sized ceramics exhibited superior biological ability and induced osteogenic differentiation of stem cells.^{42,43}

The underlying mechanism of the pro-osteogenic effects of the nanostructured surface was further investigated. The qRT-PCR results showed that BMP-2 gene expression in BMSCs cultured on nHA-coated BCP scaffolds, compared with those cultured on BCP alone, increased by 235.5% at day 4 and by 634.4% at day 7. BMP-2 is the best-studied pro-osteogenic growth factor, and the BMP/Smad signaling pathway is known to have a significant role in regulating cellular events such as cell morphogenesis and differentiation.⁴⁴ We also found that the nHA-coated BCP scaffolds could upregulate expression of BMP/Smad signaling-related genes (ie, BMPR1, Smad1, Smad4, and Smad5) in BMSCs. As shown in Figure 6E, BMP-2 promotes the phosphorylation of Smad1/5 proteins, which translocate into the nucleus with the help of Smad4 and trigger further transcription factor cascades to activate the expression of osteogenesis-specific genes (eg, ALP, BSP, OPN, and OCN), finally inducing the osteoblastic differentiation of BMSCs.^{45,46} Therefore, our results indicate that the secretion of BMP-2 proteins stimulated by the nanostructured surface may drive BMSCs into an osteoblastic lineage through the BMP/Smad signaling pathway.

The results of the intramuscular implantation experiment indicated that the nanostructured surface also plays a vital part in maintaining the integration of implants and

inducing ectopic bone formation. The μ -CT scanning and histological examination results showed severe degradation in the BCP group 90 days after implantation, resulting in the loss of primary architecture with only a few sporadic ceramic fragments remaining. By contrast, the nHA-coated BCP scaffolds tended to maintain their original porous structure. This discrepancy may be due to the high porosity of the scaffold, which had numerous interconnected pores and channels within the materials. Blood vessels and cells can completely invade such porous structures as blood flows through penetrating channels, which enables material biodegradation. It has also been shown that sponge replication techniques often cause defects (eg, pores and cracks) on scaffold walls, as well as formation of internal voids in scaffold struts, owing to the removal of the PU foam,⁴⁷ which may lead to fast degradation and even the collapse of the whole structure. For instance, Renghini et al showed that porous scaffolds made by a sponge replication method were already affected by biodegradation after 1 week of immersion in Tris-HCl solution, and that the mean thickness of scaffold walls decreased by one-third after 4 weeks of immersion.⁴⁸ Moreover, in this study, the BCP scaffolds had a high proportion (80%) of β -TCP phase, which might represent another reason for their fast degradation, as previous results have suggested that BCP is a thermodynamically reactive compound, especially when β -TCP accounts for a high proportion.^{49,50} HE staining results also indicated that the fast degradation of BCP scaffolds led to the recruitment of inflammatory cells with no osteogenesis, whereas the nHA-coated BCP scaffolds induced bone formation in non-osseous sites, as evidenced by the presence of mature bone cells settled in bone lacuna. This might be because maintenance of the three-dimensional porous structure provides the necessary environment for osteogenesis. Fast implant degradation can lead to the release of wear debris, which causes chronic inflammation and triggers the secretion of pro-inflammatory factors, chemokines, and pro-osteoclastic factors (eg, RANKL), resulting in the migration, differentiation, and activation of bone-resorbing cells (osteoclasts) that further accelerate implant biodegradation.⁵¹ Interactions between inflammatory cells and implant-derived wear debris are considered to be a key factor in periprosthetic bone resorption (osteolysis) and even implant aseptic loosening.⁵² Moderate inflammation promotes osteogenesis, whereas an excessive inflammatory response is detrimental to osteogenesis. In addition, rapid leaching of the HA layer could lead to weak

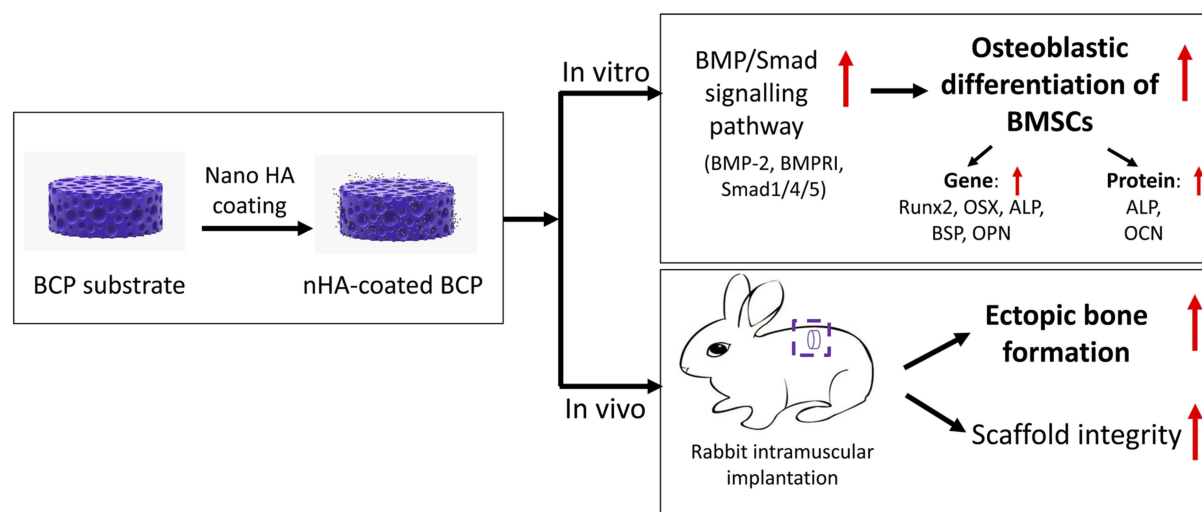


Figure 9 Scheme illustration demonstrated that nHA-coated BCP scaffolds could stimulate osteogenic differentiation via BMP/Smad signaling pathway in vitro, and induce ectopic bone formation in vivo.

bioactivity; previous experiments showed that poor cell adhesion and spreading could be attributed to rapid dissolution and reprecipitation of ceramics,⁵³ whereas osteoblast-like cells were able to adhere and spread on less soluble and more stable crystalline phase surfaces.^{54,55} Thus, compared with BCP substrates, the nHA-coated BCP scaffolds resulted in better bioactivity. However, it was difficult to perform a statistical analysis of new bone production, as previous studies suggest that material-induced osteogenesis is species-dependent and ectopic bone formation in rabbits is relatively poor.^{33,56,57} However, the presence of new bone 90 days post-implantation suggested that the nHA-coated BCP scaffold with both trabecular bone-like architecture and a nanostructured surface might have good osteoinductive capacity. This should be further evaluated in future work using other animal models.

In summary, these findings indicate that the lack of integrity of BCP substrates had an adverse effect on osteogenesis, whereas the nHA-coated BCP scaffolds promoted osteogenic differentiation of BMSCs in vitro and induced ectopic bone formation in vivo (Figure 9), possibly owing to their multiscale porous structure and nanostructured surface. Previous porosimetry analyses showed that this biomimetic bone graft (nHA-coated BCP) has a graded pore gradient, including interconnected macropores (~700 µm) formed by matched replication of PU foam that promote vascularization and bone ingrowth, minor pores (20–50 µm) that facilitate cell migration, micropores (0.8–3 µm) that accelerate nutrient exchange,²³ and abundant

nanopores (<100 nm) resulting from nanoparticle coating that stimulate osteointegration and osteogenesis; these nanopores were reported to have a positive effect on osteoregeneration during early implantation.^{6,58} Moreover, nHA-coated BCP scaffolds were shown to have a biomimetic nanostructured surface with abundant HA nanoparticles, similar to the inorganic component of natural bone, which was believed to promote tissue regeneration.^{42,43} Further research should focus on optimizing scaffold properties to achieve better osteoinductivity, including improving the mechanical behaviors and adjusting the degradation rate of the scaffold to match new bone growth, as well as expanding the use of nanostructured scaffolds to applications such as bone repair, drug delivery, and anti-tumor therapy.

Conclusion

Our current study provides a simple and effective strategy to construct biomimetic bone grafts with both trabecular framework and nano-scale topography by combining modified sponge replication and dip-coating technologies. The trabecular bone-like architecture favored tissue ingrowth and accelerated nutrient exchange, and the introduction of an HA nanoparticle layer endowed implants with nano-scale surface morphology, leading to enhanced osteogenesis via the BMP/Smad signaling pathway. These findings demonstrate that this biomimetic bone graft with a trabecular framework and nanostructured surface have potential clinical applications in bone defect regeneration.

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Disclosure

The authors report no conflicts of interest in this work.

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