

Exercise restores impaired endothelium-derived hyperpolarizing factor–mediated vasodilation in aged rat aortic arteries via the TRPV4-K_{Ca}2.3 signaling complex

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Background: Aging leads to structural and functional changes in the vasculature characterized by arterial endothelial dysfunction and stiffening of large elastic arteries and is a predominant risk factor for cardiovascular disease, the leading cause of morbidity and mortality in modern societies. Although exercise reduces the risk of many age-related diseases, including cardiovascular disease, the mechanisms underlying the beneficial effects of exercise on age-related endothelial function fully elucidated.

Purpose: The present study explored the effects of exercise on the impaired endothelium-derived hyperpolarizing factor (EDHF)–mediated vasodilation in aged arteries and on the involvement of the transient receptor potential vanilloid 4 (TRPV4) channel and the small-conductance calcium-activated potassium (K_{Ca}2.3) channel signaling in this process.

Methods: Male Sprague-Dawley rats aged 19–21 months were randomly assigned to a sedentary group or to an exercise group. Two-month-old rats were used as young controls.

Results: We found that TRPV4 and K_{Ca}2.3 isolated from primary cultured rat aortic endothelial cells pulled each other down in co-immunoprecipitation assays, indicating that the two channels could physically interact. Using ex vivo functional arterial tension assays, we found that EDHF-mediated relaxation induced by acetylcholine or by the TRPV4 activator GSK1016790A was markedly decreased in aged rats compared with that in young rats and was significantly inhibited by TRPV4 or K_{Ca}2.3 blockers in both young and aged rats. However, exercise restored both the age-related and the TRPV4-mediated and K_{Ca}2.3-mediated EDHF responses.

Conclusion: These results suggest an important role for the TRPV4-K_{Ca}2.3 signaling undergirding the beneficial effect of exercise to ameliorate age-related arterial dysfunction.

Keywords: endothelium, EDHF, exercise, TRPV4, K_{Ca}2.3, aging

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Introduction

Cardiovascular disease (CVD) is the leading cause of morbidity and mortality in modern societies, and aging is the dominant risk factor for the development of CVD.^{1,2} Currently, aging is an irreversible biological process and leads to structural and functional changes in the vasculature characterized by stiffening of large elastic arteries and arterial endothelial dysfunction.³ In contrast to the effects of aging, regular exercise has been shown to greatly improve vascular structure and function.^{4–6} Although it is well recognized that exercise reduces the risk of many age-related diseases, including CVD, the mechanisms underlying the beneficial

effects of exercise on age-related endothelial dysfunction are not completely understood.

Normal endothelial function is essential to maintain a healthy homeostasis of the vascular wall, and endothelial dysfunction is associated with a number of pathological conditions, including aging, diabetes, and coronary heart disease.^{7–9} Vasoactive substances, such as acetylcholine (ACh) and bradykinin, induce vasodilation by activating endothelium to release nitric oxide (NO), prostacyclin, and endothelium-derived hyperpolarizing factor (EDHF). EDHF plays an important role in the regulation of vascular tone, and impaired EDHF has been associated with endothelial dysfunction.¹⁰ Increasing evidence has shown that EDHF-mediated vasodilation decreases with aging, hypertension, and diabetes.^{9,11,12}

EDHF-mediated relaxation and the associated hyperpolarization involve intermediate-conductance and small-conductance calcium-activated potassium channels ($K_{Ca2.3}$) in the endothelium.¹⁰ Recently, an important role for the transient receptor potential vanilloid 4 (TRPV4) channel in EDHF-associated functions has been shown. TRPV4 physically and functionally interacts with the $K_{Ca2.3}$ in endothelial cells.⁹ This channel coupling enables Ca^{2+} influx via TRPV4 to activate $K_{Ca2.3}$, stimulating EDHF-mediated signaling and subsequent vasodilation. However, in pathological conditions, including diabetes and hypertension, the functional role of the endothelial TRPV4- $K_{Ca2.3}$ signaling is impaired, which may be a mechanism underlying decreased EDHF-mediated vasodilation.^{9,11} But whether the function of TRPV4- $K_{Ca2.3}$ coupled channels is altered with aging has not been addressed.

Therefore, the present study was designed to explore the functional role of TRPV4- $K_{Ca2.3}$ signaling in the regulation of EDHF-mediated relaxation and to examine the effects of exercise on the response of this channel coupling in aged arteries.

Materials and methods

Materials

Phenylephrine,⁹ ACh,⁹ RN1734,⁷ GSK1016790A,¹³ NG-nitro-L-arginine (L-NNA),⁸ indomethacin,⁸ and apamin⁸ were purchased from Sigma-Aldrich (St. Louis, MO, USA). Anti- $K_{Ca2.3}$ (catalog No. APC-025) and anti-TRPV4 (catalog No. ACC-034) antibodies were purchased from Alomone Labs.

Animals and the exercise training protocol

Male Sprague-Dawley rats were purchased from the Guangdong Medical Laboratory Animal Center (Guangzhou, Guangdong, China). The rats (aged 19–21 months) were randomly assigned to a sedentary group (Old) and an exercise-trained group (Old-ET). Two-month-old rats were used as young controls. The Old-ET rats were subjected to a motor-driven treadmill training protocol (15 m/min without inclination, 60 min/day, 5 days/week for 12 weeks) as previously described.¹⁴ All animal experiments were conducted in accordance with the guidelines of the US National Institutes of Health (NIH publication No. 8523) and were approved by the Animal Experimentation Ethics Committee of Guangzhou Sport University (Guangzhou, Guangdong, China).

Vessel tension measurement

Vessel tension measurement was performed as previously reported.^{15–18} Briefly, we dissected vessel segments of the rat thoracic aorta in a petri dish, which was filled with ice-cold Krebs solution (118 mmol/L NaCl, 4.7 mmol/L KCl, 2.5 mmol/L $CaCl_2$, 1.2 mmol/L KH_2PO_4 , 1.2 mmol/L $MgSO_4 \cdot 7 H_2O$, 25.2 mmol/L $NaHCO_3$, and 11.1 mmol/L glucose, pH 7.4) that was continuously bubbled with 95% oxygen (O_2) and 5% carbon dioxide (CO_2) mixed gas. The vessel segments were cut into approximately 2 mm rings and were then transferred to an organ bath. One end of the vessel ring was fastened to a hook on the bottom of the organ bath. The other end was connected to an isometric force transducer that was connected to an amplifier and recorder. The isometric tension was recorded and analyzed by a data acquisition and analysis system (BL-420S, Chengdu Taimeng Technology). In each experiment, the vessel rings were given a preload tension of 1 g and allowed to equilibrate for 1 h. The Krebs solution was changed every 15 min, and the tension was adjusted as needed to maintain the same preload tension. After equilibration, the vessel rings were first precontracted with phenylephrine (1 μ mol/L) and then relaxed in response to the addition of ACh (10 μ mol/L). The EDHF-mediated relaxation was studied in the presence of the cyclooxygenase inhibitor indomethacin (7 μ mol/L) and the NO synthase inhibitor L-NNA (300 μ mol/L).

Cell preparation and culture

The isolation and primary culture of rat aortic endothelial cells has been published elsewhere.^{9,11} Briefly, we dissected

the rat thoracic aorta and then digested the endothelial cells with 0.02% collagenase type I (Sigma-Aldrich) in phosphate-buffered saline (PBS) at 37 °C for 45 min. The digested endothelial cells were collected and centrifuged at 250×g for 5 min. Cell pellets were then resuspended in Dulbecco's Modified Eagle's Medium containing 10% fetal bovine serum, 100 U/mL penicillin, and 100 µg/mL streptomycin. After incubation at 37 °C for 1 h, the culture medium was replaced to remove nonadherent cells. The remaining adherent endothelial cells were continuously cultured at 37 °C with 5% CO₂ for 3–5 days before experiments.

Immunoprecipitation and immunoblots

Immunoprecipitation and immunoblotting were conducted as previously described.^{9,15,16} Briefly, TRPV4 or K_{Ca}2.3 proteins were immunoprecipitated by incubating 800 µg of the extracted proteins with 5 µg of anti-TRPV4 or anti-K_{Ca}2.3 antibody at 4 °C overnight. Protein A magnetic beads were then added, followed by an additional incubation at 4 °C overnight. The immunoprecipitates were washed with saline three times and then resolved via sodium dodecyl sulfate polyacrylamide gel electrophoresis using an 8% gel. The resulting proteins were transferred to a polyvinylidene difluoride membrane using a wet transfer system (Bio-Rad Laboratories). The membrane containing the transferred proteins was incubated at 4 °C overnight with the primary antibody (at a dilution of 1:200) in PBS containing 0.1% Tween 20 and 5% nonfat dry milk. The next day, immunodetection was performed using horseradish peroxidase-conjugated secondary antibodies. Protein binding was detected using an enhanced chemiluminescence system and recorded with imaging capture equipment.

Statistical analysis

Data are presented as the mean ± SEM. Statistical analyses were conducted using one-way analysis of variance followed by the Tukey post hoc test or by Student's unpaired *t* tests when appropriate. A two-sided value of *P* < 0.05 was considered statistically significant.

Results

TRPV4 physically associates with K_{Ca}2.3 in endothelial cells of rat aortic arteries

In a co-immunoprecipitation experiment using lysates freshly prepared from rat aortic endothelial cells, an anti-K_{Ca}2.3 antibody pulled down TRPV4 proteins (Figure 1A) and an anti-TRPV4 antibody pulled down K_{Ca}2.3 (Figure 1B). In the control pull-down experiments conducted using preimmune IgG, no bands were detected (Figure 1). Taken together, these results indicate that TRPV4 can physically associate with K_{Ca}2.3 and suggest that they may form a signaling complex in endothelial cells of rat aortic arteries.

Role of TRPV4-K_{Ca}2.3 signaling in EDHF-mediated vasodilation in rat aortic arteries

In an ex vivo arterial tension study, ACh induced significant relaxation in rat aortic arteries (Figure 2A and B). In the presence of the cyclooxygenase inhibitor indomethacin and the NO synthase inhibitor L-NNA, ACh-induced EDHF-mediated relaxation was significantly decreased by approximately 20% of the control value (Figure 2A and B). However, the EDHF-mediated response was markedly inhibited by approximately 70% of the control response

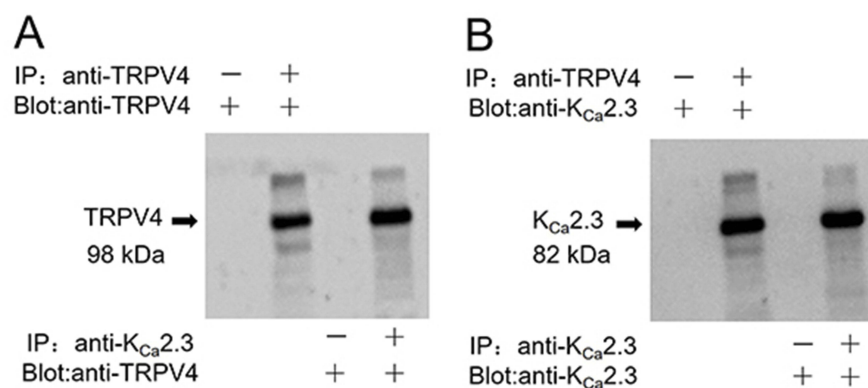


Figure 1 Physical interaction of TRPV4 with K_{Ca}2.3 in primary cultured rat aortic endothelial cells. Representative images showing co-immunoprecipitation (IP) and immunoblotting (blot) results: (A) immunoblot with anti-TRPV4 antibody; (B) immunoblot with anti-K_{Ca}2.3 antibody. Proteins from rat aortic endothelial cells were immunoprecipitated with indicated antibody (+) or preimmune IgG (-); n=3 experiments.

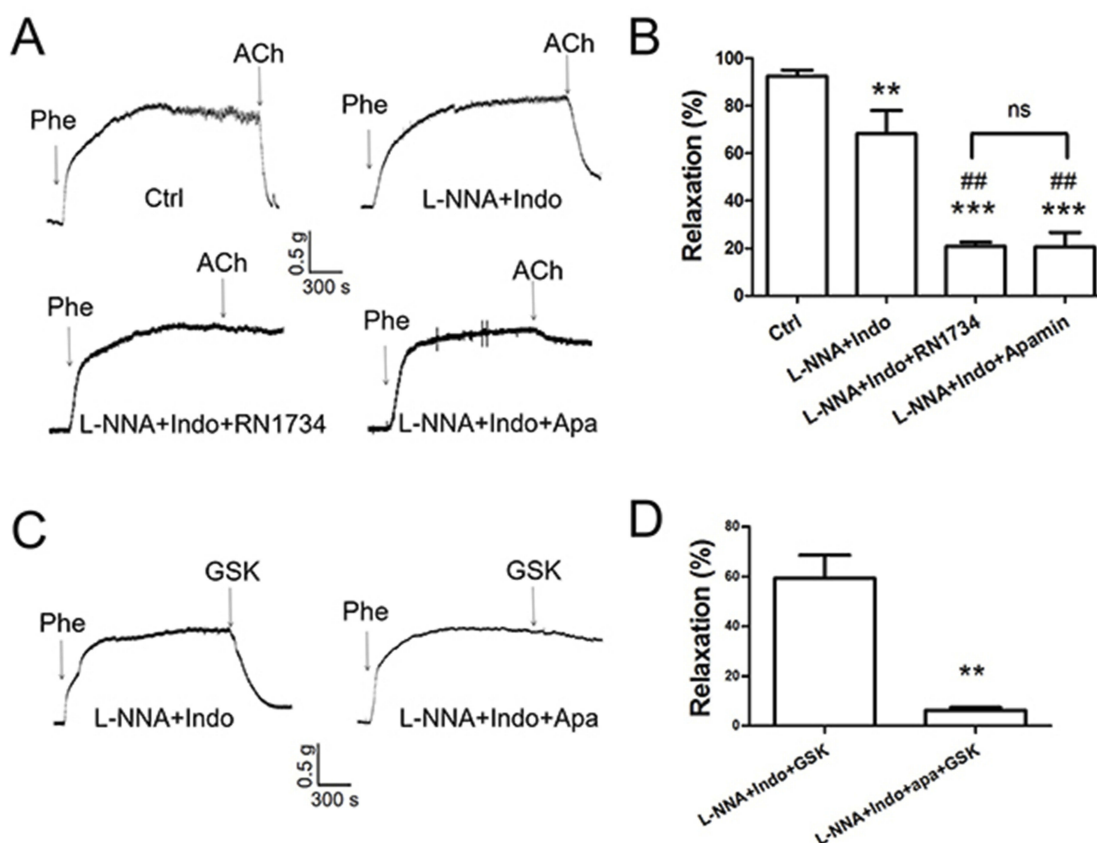


Figure 2 Role of TRPV4-K_{Ca}2.3 signaling in acetylcholine (ACh)-induced, endothelium-derived hyperpolarizing factor (EDHF)-mediated relaxation in rat aortic arteries. Representative traces (**A**, **C**) and summary data (**B**, **D**) for ACh-induced, EDHF-mediated relaxation and the effect of transient receptor potential vanilloid 4 (TRPV4) or small-conductance calcium-activated potassium channel (K_{Ca}2.3) inhibition. The EDHF response was induced by 10 μmol/L of ACh (n=10) or 300 nmol/L of the TRPV4 activator GSK1016790A (n=3) in the presence of 7 μmol/L of indomethacin and 300 μmol/L of NG-nitro-L-arginine (n=5). Preincubation with 20 μmol/L of the TRPV4 inhibitor RN1734 (n=4) or with 200 nmol/L of the K_{Ca}2.3 inhibitor apamin (n=4) decreased the ACh-induced, EDHF-mediated relaxation in the presence of indomethacin and L-NNA. Values are the mean ± SEM. **P<0.01, ***P<0.001 vs. Ctrl; ###P<0.01 vs. L-NNA + Indo. Phe, indicates phenylephrine; Ctrl, control; Indo, indomethacin; L-NNA, NG-nitro-L-arginine; Apa, apamin; and GSK, GSK1016790A.

by L-NNA plus indomethacin combined with RN-1734 (a selective blocker of TRPV4; 20 μmol/L) or apamin (a K_{Ca}2.3 selective inhibitor; 200 nmol/L). Moreover, no significant difference was found in the degree of inhibition by RN-1734 and apamin (Figure 2A and B). To further explore the role of TRPV4 in EDHF-mediated relaxation, GSK1016790A, a selective activator of TRPV4, was used. The results showed that GSK1016790A (300 nmol/L) induced relaxation even when indomethacin and L-NNA were present. However, the GSK1016790A-induced EDHF-mediated relaxation was significantly inhibited by apamin in the presence of indomethacin and L-NNA compared with that in the control, as expected (Figure 2C and D). Taken together, these results indicate that TRPV4 functionally interacts with K_{Ca}2.3 and suggest

that this interaction plays a critical role in regulating EDHF-mediated vasodilation.

Role of TRPV4-KCa2.3 signaling on the age-related decrease in EDHF-mediated relaxation in rat aortic arteries and the effect of exercise

Aging reduces normal endothelial function. As shown in Figure 3A and B, ACh-induced EDHF-mediated relaxation was markedly attenuated in older rats compared with that in young rats; however, this attenuation was less apparent in older rats after exercise training. In the presence of the TRPV4 blocker RN1734 (Figure 3A and B), EDHF-mediated relaxation was markedly and significantly

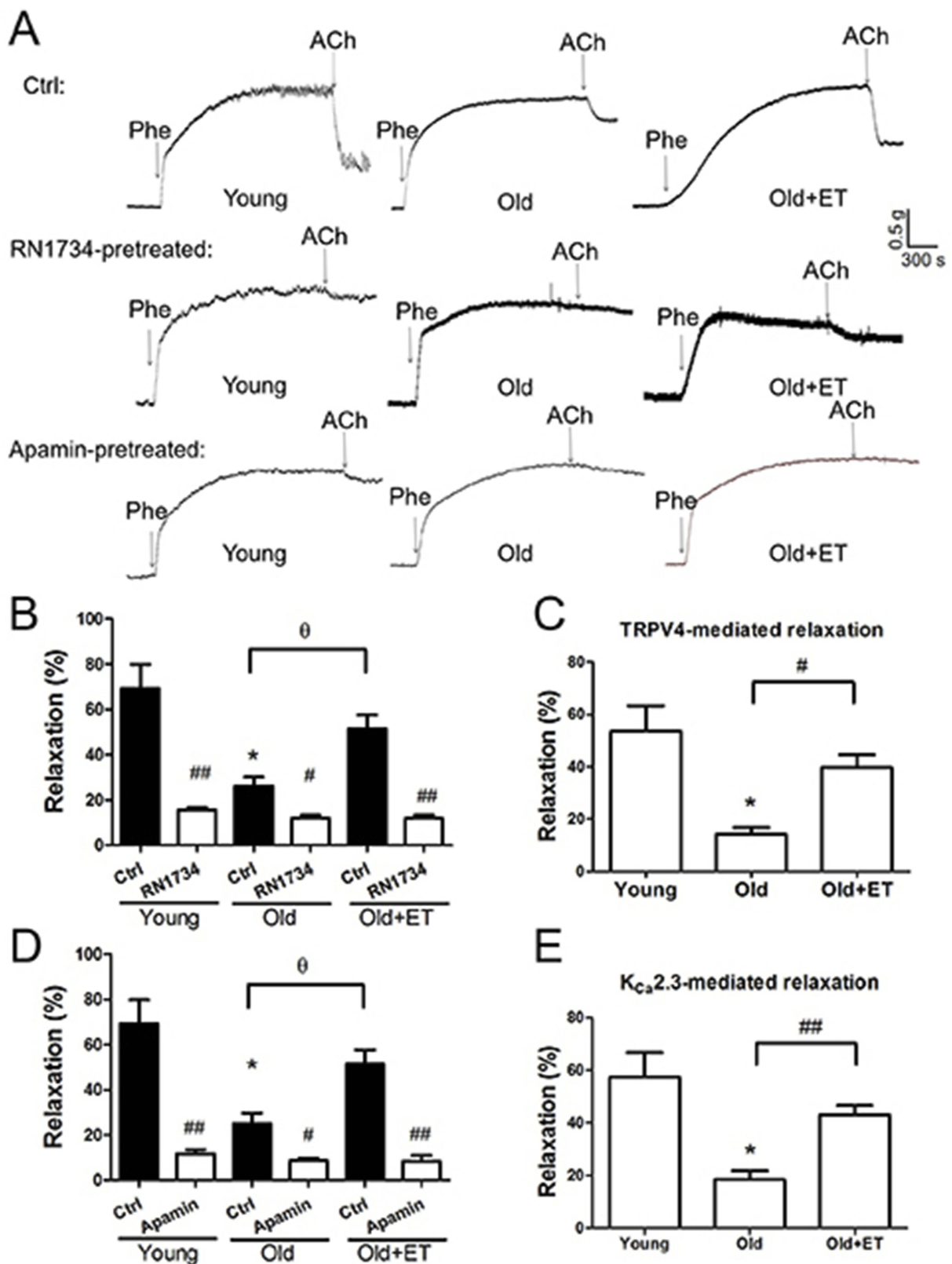


Figure 3 Role of TRPV4-KCa2.3 signaling in age-related decrease in endothelium-derived hyperpolarizing factor (EDHF)-mediated relaxation in rat aortic arteries and the effect of exercise training (ET). Representative traces (**A**) and summary data (**B**, **D**) for acetylcholine (ACh)-induced EDHF-mediated relaxation in the presence or absence of the transient receptor potential vanilloid 4 channel (TRPV4) selective inhibitor RN1734 (20 μ mol/L) or the small-conductance calcium-activated potassium channel (KCa2.3) selective inhibitor apamin (200 nmol/L) in aortic arteries derived from rats in the Young, Old, and Old + ET groups. (**C**, **E**) Summary data for TRPV4-mediated (**C**) or KCa2.3-mediated (**E**) EDHF response in aortic arteries derived from rats in the Young, Old, and Old + ET groups. Values are the mean \pm SEM (n=3 experiments). * P <0.05 vs. Young; # P <0.05, ### P <0.01 vs. Ctrl; θ P <0.05 vs. Old. Phe indicates phenylephrine; Ctrl, control.

inhibited in all groups (Figure 3A and B). Subtraction of the relaxation observed in the presence of RN1734 (Figure 3B) from that in the absence of RN1734 revealed the TRPV4-mediated EDHF response (Figure 3C). Our results showed that the TRPV4-mediated EDHF response was significantly decreased with aging compared with that in young rats; however, this response was restored to levels nearer those observed in young rats by exercise training (Figure 3C). Similarly, in the presence of the $K_{Ca2.3}$ blocker apamin, EDHF-mediated relaxation was significantly decreased in all three groups (Figure 3D). Compared with that in the young group, the $K_{Ca2.3}$ -mediated EDHF response was significantly attenuated in the aged group, whereas this attenuation was almost completely reversed after exercise training (Figure 3E). Additionally, the GSK1016790A-induced EDHF-mediated relaxation was markedly and significantly decreased with aging compared with that in young rats (Figure 4A–D), indicating that the functional role of TRPV4 in regulating EDHF-mediated vasodilation

was decreased in aged rats. However, exercise restored the TRPV4-mediated EDHF response.

Discussion

The main findings of the present study were as follows. (1) Co-immunoprecipitation data demonstrated a physical association between TRPV4 and $K_{Ca2.3}$ in primary cultured rat aortic endothelial cells; (2) ACh and GSK1016790A mainly acted through endothelial TRPV4- $K_{Ca2.3}$ signaling to induce EDHF-mediated vasodilation in rat aortic artery segments; (3) In isolated aged rat arteries, the EDHF response to either ACh or GSK1016790A was impaired; (4) Exercise training in aged rats restored both the age-related TRPV4-mediated and the $K_{Ca2.3}$ -mediated EDHF responses. Taken together, our findings suggested that TRPV4- $K_{Ca2.3}$ signaling might play an important role in the beneficial effect of exercise to improve age-related impaired arterial function.

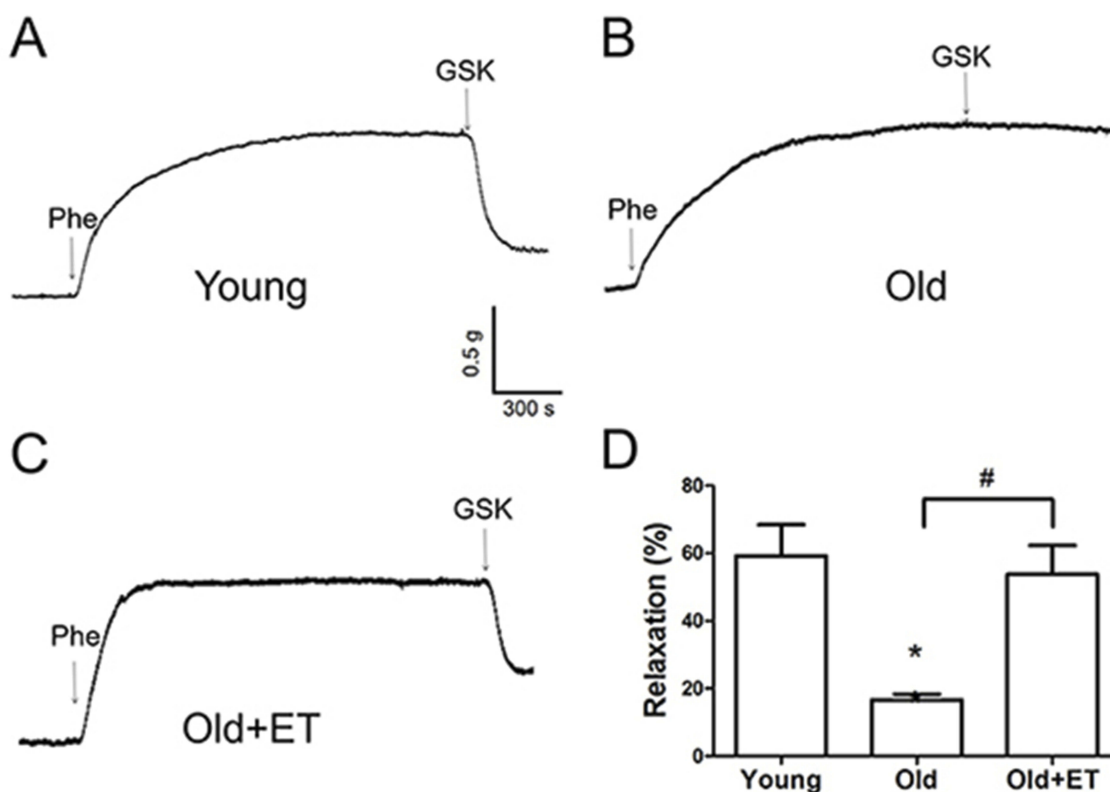


Figure 4 Exercise training (ET) restores the age-related decrease in the GSK1016790A-induced endothelium-derived hyperpolarizing factor (EDHF) response in rat aortic arteries. Representative traces (A–C) and summary data (D) for 300 nmol/L GSK1016790A-induced EDHF-mediated relaxation in aortic arteries derived from rats in the Young, Old, and Old + ET groups. The EDHF response was induced by GSK1016790A (300 nmol/L) in the presence of indomethacin (7 μ mol/L) and NG-nitro-L-arginine (300 μ mol/L). Values are the mean \pm SEM (n=3 experiments). * P <0.05 vs. Young; # P <0.05 vs. Old. Phe indicates phenylephrine; GSK, GSK1016790A.

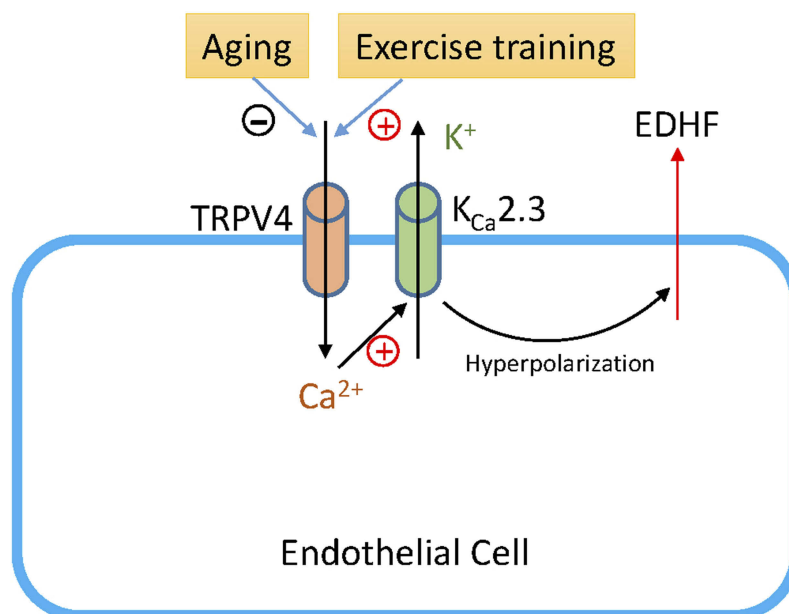
Accumulating evidence suggests that Ca^{2+} -activated potassium channels may interact with Ca^{2+} -permeable transient receptor potential channels to generate signal transduction between these channels in the vasculature.¹⁹ For example, TRPC1 is physically associated with BK_{Ca} in vascular smooth muscle cells, and Ca^{2+} entry via TRPC1 activates BK_{Ca} to cause membrane hyperpolarization.²⁰ In addition, endothelial TRPA1 forms a signaling complex with K_{Ca} channels and K_{IR} channels to mediate endothelium-dependent cerebral artery dilation.²¹ Recent studies have also shown both physical and functional interactions of TRPV4 with $\text{K}_{\text{Ca}2.3}$ in rat mesenteric endothelial cells.⁹ Consistent with the results of previous studies, the present study identified a physical association between TRPV4 and $\text{K}_{\text{Ca}2.3}$ in rat aortic endothelial cells. A pivotal role for TRPV4- $\text{K}_{\text{Ca}2.3}$ signaling was observed in regulating the ACh-induced and the GSK1016790A-induced EDHF-mediated vasodilation in rat aortic arteries. Taken together, the results from the present study and previous findings suggest that TRPV4 physically and functionally associates with $\text{K}_{\text{Ca}2.3}$ to form a signaling complex in vascular endothelial cells that enables Ca^{2+} influx via TRPV4 to activate $\text{K}_{\text{Ca}2.3}$, inducing the EDHF-mediated response of vasodilation.

CVD remains the number one cause of death in the world, and age-related vascular endothelial dysfunction is a key risk factor for the development of CVD.³ EDHF plays a central role in the regulation of endothelial function for vasodilation and coordination of blood flow in the vasculature.¹⁰ The EDHF-mediated response is altered with aging, hypertension, diabetes, and hypoxia-reoxygenation injury.^{9,11,12,22} These alterations may either contribute to endothelial dysfunction or compensate for the loss of NO bioavailability, depending on the vascular bed.¹⁰ In the present study, we identified an important role of TRPV4- $\text{K}_{\text{Ca}2.3}$ signaling in regulating EDHF-mediated vasodilation. Decreased expression of TRPV4 and $\text{K}_{\text{Ca}2.3}$ has been reported in endothelial cells with aging, diabetes, or hypertension.^{7,9} Therefore, the present results indicate that impaired TRPV4- $\text{K}_{\text{Ca}2.3}$ signaling might be the mechanism underlying the decreased EDHF responses in aged arteries.

It has been shown that regular exercise has a prominent effect on vascular endothelial function by improving factors such as the vasodilator-to-vasoconstrictor balance, inflammation, and oxidative stress.²³ Exercise has a comprehensive effect on the cardiovascular

system, including resistance arteries. Thus, the effect of exercise on elastic arteries, including the aorta, may be also important for some diseases, for example, aortic dissection.²⁴ Although exercise can reduce or even reverse the detrimental effects of aging on vascular function, the mechanisms underlying the favorable effects of exercise on age-related endothelial dysfunction have not been fully evaluated. A previous study indicated that exercise restores EDHF responses in aged arteries;²⁵ however, whether TRPV4- $\text{K}_{\text{Ca}2.3}$ signaling is involved in this effect is not known. To our knowledge, the present study showed for the first time that exercise training restored the function of TRPV4- $\text{K}_{\text{Ca}2.3}$ signaling in regulating EDHF-mediated relaxation. TRPV4, the mechanosensitive Ca^{2+} -permeable cation channel, is essential for the shear stress-induced intracellular Ca^{2+} concentration increase.^{26,27} Our previous study found that an impairment of TRPV4-mediated Ca^{2+} signaling in endothelial cells contributes to the decreased flow-induced vasodilation in aged mesenteric arteries.⁷ Interestingly, irisin, an exercise-induced myokine, induces an increase in endothelial intracellular Ca^{2+} concentration via TRPV4 channels in rat mesenteric arteries, suggesting a critical role of TRPV4 in the vasodilation effect of irisin.²⁸ However, whether exercise exerts a direct effect on TRPV4 channels in endothelial cells needs to be explored in further studies. Calcium-activated potassium channels play a critical role in regulating vascular tone. It has been reported that aerobic exercise restores the age-related reduction in BK_{Ca} channel expression on mesenteric arteries.¹⁴ Here, we demonstrated that $\text{K}_{\text{Ca}2.3}$ was involved in the EDHF-mediated aortic artery relaxation and that exercise restored the impaired function of $\text{K}_{\text{Ca}2.3}$ in aged rat aortic endothelial cells. Taken together, our results suggest that TRPV4- $\text{K}_{\text{Ca}2.3}$ coupling might play an important role in the beneficial effect of exercise on improving the age-related impairment in arterial function (Schematic 1).

In conclusion, the results of the present study indicated that exercise training significantly ameliorated the impaired TRPV4- $\text{K}_{\text{Ca}2.3}$ signaling in aged arteries, and this amelioration might underlie the favorable effect of exercise to improve age-related inhibition of EDHF-mediated vasodilation. These findings extended our knowledge of the molecular mechanisms undergirding



Schematic I Schematic diagram showing how TRPV4-K_{Ca}2.3 signaling may regulate the release of endothelium-derived hyperpolarizing factor (EDHF) in the vascular endothelium. Aging decreases TRPV4-K_{Ca}2.3 signaling, whereas exercise training recovers the signaling reversely.

the beneficial effects of exercise on the vascular function that is impaired with aging.

Abbreviations

Ach, acetylcholine; EDHF, endothelium-derived hyperpolarizing factor; TRPV4, transient receptor potential vanilloid 4; K_{Ca}2.3, small-conductance calcium-activated potassium channel; CVD, cardiovascular disease; NO, nitric oxide; L-NNA, NG-nitro-L-arginine.

Acknowledgments

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Disclosure

The authors report no conflicts of interest in this work.

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