REVIEW

Novel nanomaterial-based antibacterial photodynamic therapies to combat oral bacterial biofilms and infectious diseases

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therapy (aPDT) on oral infectious diseases has attracted tremendous interest in the past decade. However, aPDT had a minimal effect on the viability of organized biofilms due to the hydrophobic nature of the majority of the photosensitizers (PSs). Therefore, novel nanotechnologies were rapidly developed to target the delivery of hydrophobic PSs into microorganisms for the antimicrobial performance improvement of aPDT. This review focuses on the state-of-the-art of nanomaterials applications in aPDT against oral infectious diseases. The first part of this article focuses on the cutting-edge research on the synthesis, toxicity, and therapeutic effects of various forms of nanomaterials serving as PS carriers for aPDT applications. The second part discusses nanomaterials applications for aPDT in treatments of oral diseases. These novel bioactive nanomaterials have demonstrated great potential to serve as carriers for PSs to substantially enhance the PDT therapeutic effects. Furthermore, the novel aPDT applications not only have exciting therapeutic potential to inhibit bacterial plaque-initiated oral diseases, but also have a wide applicability to other biomedical and tissue engineering applications.

Keywords: photodynamic therapy, nanomaterials, antibacterial, anti-inflammatory, upconversion nanoparticles, oral diseases

Abstract: Oral diseases such as tooth caries, periodontal diseases, endodontic infections,

etc., are prevalent worldwide. The heavy burden of oral infectious diseases and their

consequences on the patients' quality of life indicates a strong need for developing effective therapies. Advanced understandings of such oral diseases, e.g., inflammatory periodontal

lesions, have raised the demand for antibacterial therapeutic strategies, because these dis-

eases are caused by viruses and bacteria. The application of antimicrobial photodynamic

Introduction

The WHO recently published a global review of oral health which emphasized that, despite great improvements in the oral health of populations in several countries, serious global problems still persisted.¹ The human oral cavity harvests a diverse ecosystem including over 700 species of microorganisms, including bacteria, archaea, fungi, and viruses. Increasing evidence indicated that the microbiota contributes to infectious oral diseases including caries, periodontal diseases, endodontic infections, etc.² Dental caries and periodontal diseases are the most prevalent bacteria-infectious oral diseases in humans worldwide. Severe periodontal diseases (periodontitis) are the leading cause of multiple tooth loss and edentulism in adults. Dental caries is the most common chronic disease of childhood. The data from the

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Global Burden of Disease Study in 2010 showed that the global burden of periodontal diseases, oral cancer, and caries increased markedly, by an average of 45.6% from 1990 to 2010.³ For example, pulpitis was caused by bacteria and their products entering the pulp through a deep caries lesion or a leaked filling.⁴ Moreover, in apical periodontitis, bacteria could invade and colonize the entire root canal system.⁴ In addition, with the infective microorganisms residing in the main canal, the bacteria could penetrate from the main root canal into dentinal tubules, lateral canals, and other canal irregularities, leading to secondary endodontic infections.⁴ Indeed, studies showed that dentin invasion occurred in 50–80% of the teeth with apical periodontitis.^{5,6}

Antibiotics is usually applied in the treatment of such oral infectious diseases. However, it was shown that the minimal concentration of antibiotic for the eradication of bacterial biofilm was difficult to reach in vivo.⁷ Furthermore, there was an increasing concern on the abuse of antibiotics due to bacterial drug-resistance. Recently, antimicrobial photodynamic therapy (aPDT) was investigated as a promising antibacterial therapeutic modality to eliminate the aforementioned shortcomings.

Photodynamic therapy (PDT) was discovered over 100 years ago by observing the killing of microorganisms when harmless dyes and visible light were combined in vitro.⁸ Since then, it was primarily developed as a treatment for cancer. Unlike traditional therapies (surgery, chemotherapy, and radiotherapy), PDT did not have severe side effects and could often be repeated.9 In recent years, the interest in the antimicrobial effects of PDT was revived. PDT was proposed as a therapy for a large variety of localized infections,¹⁰ and it was described as antimicrobial or antibacterial PDT, or photodynamic inactivation (PDI). aPDT employed appropriate excitation light in combination with photosensitizers (PSs) and oxygen, and allowed the non-specific attack against microorganisms by generating cytotoxic ROS, especially singlet oxygen (¹O₂).¹¹ Singlet PS was extremely unstable and could instantaneously release the energy, returning to the triplet state. The released energy was absorbed by the tissue oxygen to form ROS, which had a strong oxidation and high reactivity, thus causing rapid lipid oxidation of the bacteria. This led to the destruction of the vulnerable membrane lipids and eventually the bacterial death.¹¹

Several meritorious reviews on aPDT applications in dentistry had described the antimicrobial effects against oral infectious diseases,^{11–17} which were not repeated in

the present article. The majority disadvantage of PSs was hydrophobic and poorly soluble in water. Therefore, a promising approach to enhancing the performance of PSs was to encapsulate them in nanostructured materials. The application of these nanocarriers would improve the drug availability for parenteral administration, by increasing the nanoparticle (NP) uptake for greater therapeutic efficacy.¹⁸ Therefore, the present article reviewed new developments in nanomaterial-based aPDT and the applications in dentistry, focusing on the bioactive and therapeutic effects against several key oral and dental infectious diseases.

Nanomaterials in aPDT

The aPDT researches attracted tremendous attention recently as an alternative approach to combating the drug-resistant microorganisms. It combined non-toxic PSs and harmless light of suitable wavelengths to produce antimicrobial effects by generating ROS, which eventually caused the cell death. Nevertheless, the drawbacks of PSs, including poor water-solubility, uncontrollable drugrelease profile, poor target selectivity, and low extinction coefficient, had hindered the clinical applications.¹⁹ In addition, photo-bleaching could cause unintended sideeffects and damage the healthy tissues. To overcome these limitations, novel precise designs of nanomaterials were developed to act as vehicles and loaded or embedded with PSs, or to act as PSs themselves. They possessed important properties¹⁹⁻²³: 1) controlled PSs releasing ability, 2) improved water-solubility, 3) preventing aggregation even at high concentrations, 4) good binding, easily uptaken by bacterial cells and generate larger yield of ROS, 5) selective antimicrobial activity at the local-targeted sites, 6) increased bacterial cell wall penetrability, and 7) broadening the photo-therapeutic window to optimize tissue-penetration depth.

The use of nanoparticles to serve as photosensitizers Fullerenes

Fullerenes (C_n, n=60, 70, 72, 76, 84, or 100) were members of the closed-cage carbon molecules family which were entirely composed of sp² hybridized carbon atoms, with pentagons and hexagons arranged in a soccer ball structure.²⁰ Because of their extended π -conjugation, fullerene molecules had a long triplet-excited state lifetime, a high triplet yield and could produce ROS after illumination, indicating they could act as PSs in aPDT.^{24,25} In addition, fullerenes possessed several other advantages over traditional PSs, including:^{20,25} 1) High photostability (resistance to photobleaching), 2) versatile functionalization, 3) ability to carry out both type 1 and type 2 pathways and photochemistry, 4) enhancement of the overall ROS quantum yield, 5) possibility of oxygen-independent photo-killing by electron transfer, 6) potentiated antimicrobial activity by inorganic salts, and 7) formation of selfassembled NPs.

It was reported that fullerene and its derivatives displayed broad-spectrum antimicrobial photodynamic activities against both bacteria and fungi.²⁶ The possible mechanism was the disturbance of the bacterial cell structure and metabolism by generating ${}^{1}O_{2}$, superoxide anions (O_{2} , $\overline{}$), and free radicals (OH). Fullerenes with cationic-functionalized groups can potentiate the aPDT activity against microorganisms.^{27,28} Recently, a novel amine group-modified fullerene derivative C₇₀-(ethylenediamine)₈ showed a strong killing activity against superbacteria, and the ability to mitigate bacterial infection and accelerate wound healing.²⁹ The addition of KI could further potentiate the aPDT effect of the photoactivated fullerenes on Gram-negative bacteria, Gram-positive bacteria, and fungus.^{30,31} In addition, several fullerene composites showed excellent photocatalytic inactivation activity.^{32,33} The irradiated Tetrakis[3-(N-ethylcarbazoyl)]porphyrin-C₆₀ $(TCP-C_{60})$ film showed a 4 log decrease in *Staphylococcus* aureus (S. aureus) and Escherichia coli (E. coli) with 30 and 60 min irradiations. The irradiated C_{60}/C_3N_4 and C_{70}/C_3N_4 hybrid inactivated 86% and 100% of E. coli O157:H7 cells, respectively.33

However, the disadvantages of fullerenes application in aPDT include: 1) highly hydrophobic and easy to accumulate, 2) ultraviolet-visible absorption spectrum, and 3) antioxidants.²⁵ Although these drawbacks could potentially become an obstacle for the application of fullerenes, many approaches could solve these problems. Surface modification (such as anionic or cationic functional groups) and supramolecular approaches (such as PEGylation and encapsulation) could be used to improve the aqueous solubility.³⁴ Light-harvesting antennae³⁵ and dual near-infrared (NIR) two-photon excitation³⁶ can enhance the depth of light penetration. Paradoxically, fullerenes can be strong oxidizing agents or anti-oxidant³⁷ with or without illumination.

Graphene and carbon nanotubes

Graphene and its derivatives were widely investigated due to their excellent conductivity, biocompatibility, non-toxicity, and photothermal activity.³⁸ Recently, much attention was transferred to target microbial pathogens, which was becoming increasingly resistant to conventional therapy.39-42 Graphene oxide (GO) had unique properties such as a large surface area decorated with different functional groups, intrinsic high NIR absorbance and well-dispersed stability in aqueous solution, and was an ideal nanoplatform for PS delivery. Current advancement for PS delivery appeared to concentrate on multifunctional nanostructures for multimodel therapies.⁴³ Tian et al loaded Ce6 onto polyethylene glycol (PEG) functionalized GO via π - π stacking, and the GO-based PDT delivery had significantly increased intracellular Ce6 shuttling, thus enhancing the cancer cell killing efficacy upon light irradiation.⁴⁴ Moreover, when combining photothermal therapy (PTT) with mild local heating using 808 nm laser, the cellular uptake of GO-PEG-Ce6 was increased by 2 folds. Therefore, GO-based nanomaterials showed the potential for targeted PTT and PDT.44 Similar synergistic bacteria-killings through physical action and PDT were also achieved by using GO/Ag/Collagen hybrids.⁴⁰

Carbon nanotubes (CNTs) were another type of carbon-based scaffolds for encapsulating antimicrobial agents. CNTs were allotropes of carbon made by folding a sheet of graphene into a cylindrical nanostructure, with a high aspect ratio (length to diameter) and a large surface area.19,20,45 There were two types of CNTs including single-walled carbon nanotubes (SWNTs), and multi-walled carbon nanotubes (MWNTs). CNTs had excellent capabilities to develop ultrathin films with high surface area for immobilization of PSs, accompanied by their own abilities to yield ROS upon illumination. Therefore, extensive efforts were made to improve the antimicrobial ability of CNTs.⁴⁵⁻⁴⁹ Banerjee et al fabricated MWNT conjugated with protoporphyrin IX, and investigated aPDT efficacy against S. aureus via irradiation with visible light. The colonies of S. aureus was reduced by approximately 15-20%, comparing to the area that was covered by the films (control) after 1 h light exposure.⁴⁶ Since the lifetime and diffusion distance of ¹O₂ was limited, a controllable singlet oxygen generation (SOG) with a high selectivity would be more efficient for aPDT. Zhu et al engineered a molecular complex of Ce6, a-thrombin (Tmb) aptamer, and SWNTs to regulate the SOG.⁵⁰ As shown in Figure 1, no significant difference of fluorescence was shown between buffer solution and aptamer-SWNTs. However, when 2.0 µM Tmb was added, the SOG fluorescence of aptamer-SWNTs exhibited a 13-fold enhancement. Hence, the DNA interaction with SWNTs can be disturbed by the

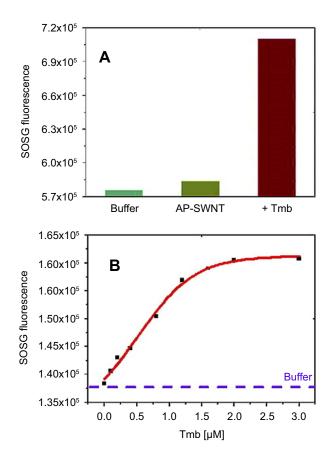


Figure I SOG regulation by aptamer-binding protein. **Notes:** (**A**) SOSG signal readout after 10.5 mins of irradiation with excitation at 404 nm. SWNTs showed great quenching for SOG. After introduction of 2.0 μ M thrombin, SOG was increased. (**B**) SOSG signal plotted as a function of Tmb concentration. The purple line indicates the buffer's SOSG signal. Reprinted with permission from Zhu Z, Tang Z, Phillips JA, Yang R, Wang H, Tan W. Regulation of singlet oxygen generation using single-walled carbon nanotubes. *J Am Chem Soc.* **2008**;130(33):10856–10857.⁵⁰ Copyright (2008) American Chemistry Society. **Abbreviations:** SOG, singlet oxygen generation; SOSG, singlet oxygen sensor green; SWNTs, single-walled carbon nanotubes; Tmb, thrombin.

binding between the aptamer and the target molecule. This can cause the DNA to fall from the SWNT surface, thus promoting a restoration of SOG for PDT applications.⁵⁰

Metal oxides nanoparticles

ZnO and TiO₂ were semi-conducting materials with unique optical, biocompatible, and biodegradable properties.^{51,52} Electrons were excited by ultraviolet (UV) light from a semi-conductor valence band to the conductance band, which then underwent electron transfer to oxygen to form ROS (mainly \cdot OH) to kill the microorganisms. Various surface modifications, such as doping with molecules, could significantly enhance the antibacterial efficiency. For example, Pillai et al studied the photocatalytic antibacterial activity of fluorinated (F-doped) ZnO nanopowders against *E. coli* and *S. aureus*.⁵³ Upon irradiation, F-doped ZnO nanopowders were more effective in inhibiting the growth of microorganisms than the undoped

ZnO nanopowders, showing 99.87% and 99.99% antibacterial activity against E. coli and S. aureus, respectively.⁵³ Recently, Sethi et al fabricated ZnO/TiO₂ nanocomposites to combat E. coli.54 Compared to ZnO or TiO₂ NPs alone, the ZnO/TiO₂ nanocomposite exhibited greater inactivation of E. coli under light irradiation.⁵⁴ Among several different crystalline forms (anatase, rutile, and brookite), the anatase form was a better photocatalyst under UV light.^{55,56} Suketa et al revealed that coating nanostructured anatase TiO2 on the roughened implant surface exerted a bactericidal effect against dental periodontal pathogens Actinobacillus actinomycetemcomitans and Fusobacterium nucleatum.⁵⁷ In addition, as mentioned previously, the addition of KI to TiO₂ NPs that was excited by UVA strongly potentiated the broad-spectrum antimicrobial efficacy by up to 6 logs.58 Therefore, the bactericidal effect of photocatalysis for treatment of localized infections could be enhanced by spraying with a KI solution.⁵⁸

Magnetic NPs (MNPs, Fe₃O₄), possessing a peroxidase-like activity, could be functionalized by conjugatively attaching PSs and other molecules to its surface to enhance the ROS generation at the targeted sites for aPDT applications.^{59,60} Multifunctional MNPs that were conjugated with PSs and vancomycin,⁶¹ and a composite system of reduced GO (rGO)–iron oxide NPs (rGO–IONP)⁶² can synergistically induce damage to methicillin-resistant *Staphylococcus aureus* (MRSA).

Upconversion nanoparticles

Most PSs were optimally excited with UV or short-wavelength blue light, whereas these wavelengths possessed inadequate tissue-penetration ability and may harm the human health. The NIR light ranging from 700 to 1100 nm had advantages of a deeper tissue penetration and lower autofluorescence, and can reduce side effects such as the phototoxicity and photodamage.⁶³ Upconversion was a nonlinear optical process of two or more photons sequential absorption, leading to the emission of light at shorter wavelength than the excitation wavelength (anti-Stokes-type emission).^{20,64} Upconversion NPs (UCNPs) were able to absorb the NIR light and convert it into high energy photons, and could potentially be applied to solve the aforementioned limitations of traditional PDT.

Solid-state materials doped with lanthanide ions were proposed as promising alternatives to yield efficient upconversion energy for optical imaging, organic fluorophosphors, and PDT. Generally, "core-shell" structured composites covered by varieties of PSs were applied for aPDT against microorganisms.^{65–67} Recently, a multifunctionalnanostructured system UCNPs (NaYF₄:Mn/Yb/Er)/ Methylene blue (MB)/CuS-chitosan showed a superior antibacterial activity via synergistic effects of PDT/PTT.⁶⁸ The bacteria-killing efficiency of this nanostructured system was up to 99% with 980 nm at 1 W·cm⁻² for 20 mins. The red emission (650–670 nm) of UCNPs was applied to enhance the effectiveness of efficient energy transfer from UCNPs to MB. Then, the addition of CuS was to obtain a high therapeutic index via the synergistic effects of PDT/PTT. Finally, chitosan was grafted to prevent aggregations among the particles, thus providing an excellent water-solubility and biocompatibility.⁶⁸

Nanoparticles as nanocarriers for photosensitizers

AuNPs-based nanoparticles

Gold NPs (AuNPs) earned significant attention due to its localized surface plasmon resonance (LSPR) upon light exposure. With LSPR having the absorption band of PSs, the absorption rate of the PSs was greatly enhanced by the high instantaneous electric field. This phenomenon could then lead to an increase in the PS excitation rate, which in turn increased the production of ROS.⁶⁹ MB-conjugated AuNPs (MB-AuNPs) displayed excellent photoinactivation to combat 4-day biofilms of MRSA. Upon irradiation, the biofilm was reduced by over 5 log₁₀ using MB-AuNPs, while less than 1 log₁₀ using MB alone.⁷⁰ Therefore, conjugating PSs on the surface of AuNPs was an effective approach to increasing the aPDT efficacy.^{70,71} In another study, core-shell nanostructured gold nanorods/SiO2 with embedded verteporfin (AuNRs@SiO2-VP) were synthesized for aPDT to target E. coli.⁷² Upon repeated irradiation, an infectious dose of 104 CFU/mL E. coli was eradicated by AuNRs@SiO₂-VP at 710 nm (1 W/cm²). In addition, the irradiation time and the laser wavelength affected the efficiency of the nanostructures against pathogens.72

Silica-based nanoparticles

Silica-based NPs were promising nanocarriers for PDT because of their excellent physical and chemical properties, good stability and dispersity, hydrophilicity, as well as bio-compatibility. Functional groups could be attached to the surface of silica NPs (SiNPs), which enhanced them to capture numerous PDT drugs.^{73–75} Recently, Lin et al synthesized Ce6 loaded SiNPs with greater efficacy in inhibiting *S. aureus* and MRSA. The photostability of Ce6 molecules was

significantly improved and maintained the ${}^{1}O_{2}$ generation capacity upon light illumination.⁷⁶

Liposomes

Substantial attention was paid to liposomes as delivery systems for PSs in aPDT, since liposomes could improve the PS loading capacity as well as the PS' safety and selectivity.^{77–80} Liposomes consisted of one or more concentric lipid bilayers with a hydrophilic phase inside and between the bilayers. Therefore, PSs could be loaded into liposomes differently depending on the hydrophilicity or lipophilicity of PSs.^{77,78} Hydrophilic PSs were suspended in the aqueous inner phase of liposomes, whereas lipophilic PSs were dissolved in fatty side-chains in the interior of the lipid bilayer.

Once PSs were loaded into the liposomes, there were three uptake pathways for liposomal PSs into bacteria. The first pathway was the binding of the liposome to the bacterial cell and consequent migration of PSs from the lipid bilayer to the bacterial cell wall. The second pathway was based on the fusion of the liposome with the cell wall, followed by direct release of PSs into the bacteria. A third possibility was the release of PSs from the liposome into the surrounding medium and the diffusion of PSs through the cell wall.^{79,80} However, conventional liposomes had a short half-life circulation, which made it difficult to elevate the antimicrobial efficacy of PSs.⁷⁷

PEG, a water-soluble polymer that exhibits protein resistance, was applied to stabilize and prolong the halflife of liposomes. The relative half-life of PEGylated liposomes was significantly increased (~45 hrs), compared to conventional liposomes (a few hours).⁸¹ Cationic modification of liposomes was another promising way to enhance aPDT, since an overall cationic charge was necessary for PSs to have antimicrobial properties.⁸² Yang et al fabricated cationic liposomes embedded with cetyltrimethyl ammonium bromide (CTAB), in which Ce6 was encapsulated as PS to administer aPDT against clinical isolates of Candida albicans (C. albicans).⁸³ CTAB was a cationic compound with strong antimicrobial activity. However, it was unsuitable for direct usage due to its high affinity to biological membranes. Ce6 loaded CTAB-liposomal compounds showed greater aPDT efficacy against clinical drug-resistant strains of C. albicans in vitro.⁸³ However, a major concern of cationic liposomes serving as antimicrobial PSs could be that the host cells might compete with the microorganisms for binding to the positively charged

liposomes in the infected tissues, causing side effects upon illumination.

Recently, an enzyme-sensitive and PS-quenching lipidbased transfersomal carrier was developed to achieve better target selectivity, greater solubility, and skin penetration efficiency.⁸⁴ In the absence of lipase, pheophorbide A (Pheo A) was self-quenched due to the mechanism of intramolecular interaction. However, when Propionibacterium acnes lipase-mediated disruption of nanocarrier occurred, Pheo A dequenching was selectively induced. Over 99% of Propionibacterium acnes was eliminated by the liposomemediated aPDT.⁸⁴ As shown in Figure 2, free Pheo A was only detected in the stratum corneum because of its low solubility and high molecular weight, whereas Pheo Aloaded liposomes showed greater skin penetration efficiency. Furthermore, in vivo antibacterial effect of the liposomebased transfersomes in nude mice was also confirmed by a significant reduction in the swelling volume and thickness.⁸⁴

Polymer-based nanoparticles

Polymer-based NPs possessed several advantages as drug delivery in aPDT⁸⁵: 1) flexibility in surface modification for better efficiency, 2) ability to prevent degradation of PSs in biological environment, 3) ability to load and deliver large amounts of PS to the target area, 4) excellent biocompatibility and biodegradability, and 5) ability to load multiple components such as targeting agents. Table 1 summarized recent studies using polymer-based NPs to apply in aPDT against different microorganisms.

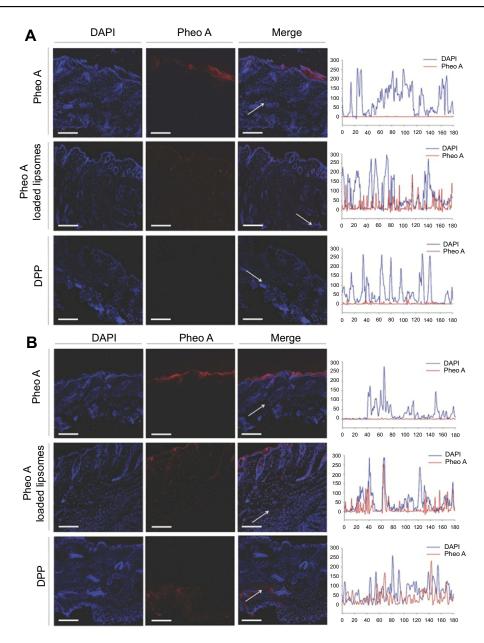
PSs and other therapeutic agents could be co-loaded into the targeted polymer nanocarriers such as nanospheres, nanocapsules, and micelles, which allowed a precise and controlled delivery of the best drug ratio in the same area of the body.⁸⁶ Recently, Xiao et al synthesized ubiquicidin (UBI)-loaded micelle to combat Pseudomonas aeruginosa (P. aeruginosa) in vitro and in vivo.⁸⁷ UBI was a positively charged antimicrobial peptide, which could bind to the negatively charged bacterial cell membrane but not mammalian cells. Therefore, it could selectively adhere to bacteria and depolarize the membrane to combine UBI with poly(butyl methacrylate)-b-poly(2-(dimethylamino) ethyl methacrylate-co-eosin)-b-ubiquicidin [PBMA-b-P(DMAEMA-co-EoS)] (noted as PAM-UBI) to form micelle. Moreover, upon light irradiation, ROS yielded from micelles could kill the bacteria (Figure 3).⁸⁷ Hence, developing such polymeric nanocarriers was a promising alternative to apply in aPDT.

Biosafety of nanomaterial-based aPDT in the oral cavity Non-toxic light source and biocompatible nanomaterial-loaded photosensitizers

Light source and PSs were the two main components of aPDT application. UV light was proven to have excellent antibacterial efficacy on oral microorganisms.¹⁰¹ However, the irradiation of UV light could lead to DNA damage and UV-induced skin damage.¹⁰² In contrast, visible light and NIR light were regarded to be safe. On the other hand, a green and benign method was recommended as the biosynthesis method of PSs,¹⁰³ and PSs would have a good bioactivity and biocompatibility with no dark toxicity or mutagenicity, with a high binding affinity for microorganisms and a low binding affinity and low toxicity for mammalian cells.¹⁰⁴ In addition, the irradiation intensity and dose of PSs should be controlled at a reasonable and effective range, where pathogens could be significantly reduced without damaging the host cells. To obtain a better therapeutic efficacy, a greater irradiation intensity, and a higher concentration of PSs were needed, which could result in thermal damage and tissue toxicity.¹⁰⁵ Therefore, these parameters should be evaluated before biomedical applications in vivo. Commercial PSs such as aminolevulinic acid, Toluidine blue O (TBO), and MB have already been applied in clinical treatments for oral diseases approved by the Food and Drug Administration.

Bioavailability of eco-friendly nanomaterials

The bioavailability and biocatalytic activities of nanomaterials depended on their size and surface area, shape and aspect ratio, surface charge, composition and crystalline structure, aggregation and concentration, surface coating and roughness, and solubility.¹⁰⁶ These physicochemical properties affect the toxicity at the cellular and tissue levels. Similarly, the efficient PS delivery, organ distribution, accumulation, retention, metabolism, degradation, and clearance should be considered at the organ level as well.¹⁰⁷ Although many nanomaterials were developed, and some were used in clinical treatments, the understanding of the potential toxicity and the mechanisms of toxicity were limited. To obtain eco-friendly nanomaterials for a wide range of biomedical applications, all the parameters of nanomaterials should be strictly and repeatedly tested in animals first. Similarly, even for drugs that were effective in the short-term with no toxicity, long-term investigated were needed. Therefore, to address the eco-friendly concerns, good



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Figure 2 Confocal microscopy to compare skin penetration efficiency of free Pheo A, DPPC liposomes, and DPP transfersomes. The line scan data are shown by the white arrows in the images.

Notes: Cross-sections of (**A**) normal skin; and (**B**) *P. acnes*-induced skin were incubated on a Franz diffusion cell (n=3). The blue signal indicates DAPI (nucleus; excitation/ emission: 358/461 nm). The red signal indicates Pheo A (excitation/emission: 410 nm/675 nm). Scale bar = 200 μ m. Reprinted with permission from Park H, Lee J, Jeong S, et al. Lipase-sensitive transfersomes based on photosensitizer/polymerizable lipid conjugate for selective antimicrobial photodynamic therapy of acne. *Adv Healthc Mater.* 2016;5 (24):3139–3147.⁸⁴ Copyright (2016) John Wiley and Sons.

Abbreviations: DAPI, 4',6-diamidino-2-phenylinodle; DPP, DSPE-PEG-Pheo A; DPPC, dipalmitoylphosphatidylcholine; P. acnes, Propionibacterium acnes; Pheo A, pheophorbide A.

biocompatibility, biodegradability, and good solubility were needed for oral medicine applications.

Application of aPDT in oral infectious diseases and treating the associated pathogens at the nanoscale

This section introduced various oral infections and the related pathogens, as shown in Figure 4. Moreover, the

cutting-edge researches on aPDT applications in oral infectious diseases and in treating the associated pathogens at the nanoscale were reviewed.

Dental caries

Dental caries is caused by acid production from biofilms. Among all the studies on aPDT against dental caries, *Streptococcus mutans* (*S. mutans*) is the most widely

Table I Different polymeric nanomaterials-based aPDT	nanomate	rials-based aPDT			
Polymer-based nanomaterials	Ref.	Representative example	Dose of nanomaterials	Light wavelength (i) or light dose (LD) or power density (P)	Type of bacteria and disinfection efficiency
CSNPs	88–92	CSNPs/ICG ⁸⁹	2.0 mg/mL	λ =810 nm	A. Actinomycetemcomitans (91%, 1 min)
CNC	93	CNC-Por (I)	20 µM	LD =118 J/cm ²	P. aeruginosa (99.7%, 30 mins) MRSA (99.999+%, 5 mins)
PLGA NPs	94–96	MB-loaded PLGA NPs ⁹⁴	6.25 µg/mL	λ =665 nm	E. faecalis (96.7%, 10 mins)
Poly HDDA-co-DBPA)-mPEG	26	Ce6 encapsulated Poly (HDDA-co-DBPA)-mPEG	11.93 µg/mL 17.91 µg/mL	λ =664 nm	S. aureus (100%, 30 mins) E. coli (100%, 30 mins)
РММА	98	ТРР/РU/РММА	2 µM	λ =633 nm	S. aureus (99.937%, 30 mins)
CMP NPs	66	Тһ-ВТ-I 00	l mg/mL	P=I.2 W/cm ²	B. subtilis (97%, 120 mins) E. coli (95%, 120 mins)
PAN	00	PAN-Por (*)	34.8 nmol/mg	λ. =400-700 nm	S. aureus (99.9999+%, 30 mins) E. faecium (99.9998+%, 30 mins) E. coli (99.9999+%, 30 mins) A. baumannii (99.9999+%, 30 mins) K. pneumoniae (99.9996+%, 30 mins)
Ahreviations . A actinomycetem comitans Aggregatikarter actinomycetem com	itans Aggreg		haumannii: R subtilis Racillus subtili	is: RT henzorhiadiazola unit	itans: A haumannii Arinetoharter haumannii: R suhtilis Barillus suhtilis BT henzothiadiazola unit: CMP conineated mirronorous polymer: CNC cellulose

Abbreviations: A. actinomycetemcomitans, Aggregatibacter actinomycetemcomitans; A. baumannii, Acinetobacter baumannii, B. subtilis, Bacillus subtilis, BT, benzothiadiazole unit; CMP, conjugated microporous polymer; CNC, cellulose nanocrystals; CSNPs, chitosan nanoparticles; DBPA, 3-(diethylamino)-1-propylamine; E. coli, Escherichia coli; E. faecalis, E. faecilis, E. faecium, Enterococcus faecaim; HDDA, 1,6-hexanediol diacrylate; ICG, indocyanine green; K. pneumoniae, Klebsiella pneumoniae; MRSA, methicillin-resistant Staphylococcus aureus; P. aeruginosa; PAN, polyacrylonitrile nanofibers; PEG, polyethylene glycol; PLGA, poly-lactic-co-glycolic acid; PMMA, poly-methylmethacrylate; Por, porphyrin; PU, polyurethane; S. aureus, Staphylococcus aureus; Th, thiophene; TPR, tetraphenylporphyrin.

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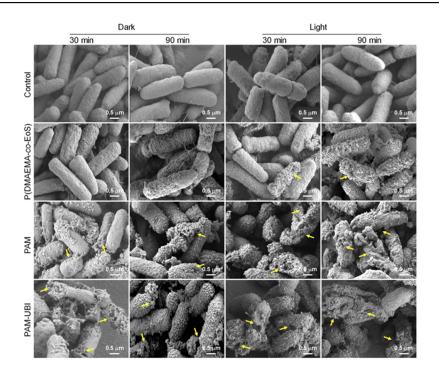


Figure 3 SEM images of *P. aeruginosa* after diverse treatment with different durations with or without light irradiation for 30 mins. Reprinted with permission from Xiao F, Cao B, Wang C, et al. Pathogen-specific polymeric antimicrobials with significant membrane disruption and enhanced photodynamic damage to inhibit highly opportunistic bacteria. The yellow arrows in the images showed the obvious membrane damage of the bacteria. ACS Nano. 2019;13(2):1511–1525.⁸⁷ Copyright (2019) American Chemistry Society.

Abbreviations: P. aeruginosa, Pseudomonas aeruginosa; SEM, scanning electron microscope.

studied pathogens.^{108,109} Recently, conjugate-like nanocarriers were developed for PS loading. TBO-silver NP conjugates (TBO-AgNP) with a 70 s irradiation by 630 nm light showed a reduction of planktonic *S. mutans* by 4 log, compared to 2 log by TBO alone. TBO-AgNP conjugates inhibited biofilm formation, increased the uptake of propidium iodide and increased the leakage of the cellular constituents. In addition, dextran-coated silver NPs (AgNP_{Dex}) were used as an effective broad-spectrum antibacterial with no toxicity (Figure 5A).¹¹⁰

ICG was an effective adjunctive drug applied in wound healing and treatment for chronic infections of mucous membranes and skins with almost no host toxicity.¹¹¹ Gholibegloo et al synthesized various nanocarriers including GO, GOcarnosine (GO-Car), and GO-Car/Hydroxyapatite (HAp) for ICG loading and investigated antibacterial effects in aPDT against *S. mutans* (Figure 5B).¹¹² GO@ICG, GO-Car@ICG, and GO-Car/HAp@ICG mediated aPDT, reduced *S. mutans* counts by 91.2%, 95.5%, and 93.2%, respectively, and suppressed *S. mutans* biofilm by 51.4%, 63.8%, and 56.8%, respectively. GO-Car/HAp@ICG showed the greatest amount of ICG loading as well as the longest time stability, which could promote the efficiency of aPDT for treating local dental infections.¹¹²

Endodontic root canal infections

Endodontic infections include primary or secondary root canal infection, persistent root canal infection, and extraradicular infection.¹¹³ *Enterococcus faecalis* (*E. faecalis*) and *Porphyromonas gingivalis* (*P. gingivalis*) were detected in 49% and 17.6% in primary endodontic infection, respectively.¹¹⁴ *E. faecalis*, the predominant bacteria leading to the failure of endodontic treatments, was also associated with other oral diseases, such as caries, periodontitis, and peri-implantitis.¹¹⁵ Sodium hypochlorite (NaOCI) was an effective irrigating solution to disrupt biofilms in root canals. However, 2.5% NaOCI could not completely eliminate biofilms due to the anatomical complexities. In addition, NaOCI could cause negative effects on dentin hardness,¹¹⁶ bond strength, and marginal adaptation.¹¹⁷

Recently, a novel PS (rose bengal-functionalized chitosan NPs [CSRBnp]) was developed for root canal disinfection to eliminate single bacteria, monospecies, and multispecies bio-film (Figure 5C).^{118,119} *E. faecalis* was reduced by 50–65% with CSRBnp in the dark and was rapidly eradicated with CSRBnp after aPDT. In the presence of inhibitors, dentin, and LPS, the antibacterial efficacy of CSRBnp was reduced in the initial hours, but the bacteria were eradicated after 24 hrs. The pulpal tissues showed the highest inhibitory effect, followed

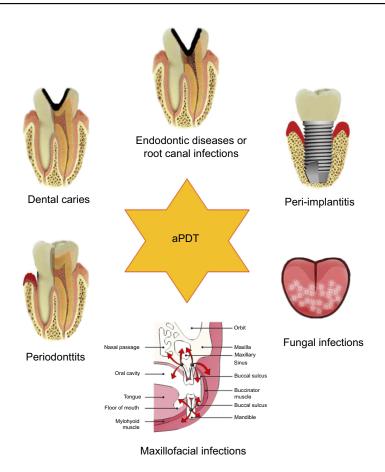


Figure 4 Schematic diagram of the application of aPDT in dentistry. APDT had potential to inhibit many oral infections including caries, endodontic diseases, root canal infections, periodontitis, peri-implantitis, oral fungal infections and maxillofacial infections. Abbreviation: aPDT, antimicrobial photodynamic therapy.

by BSA. CSRBnp had 40–65% bacterial reduction after 24 hrs.¹¹⁸ The CSRBnp-mediated aPDT disturbed the multilayered biofilm architecture of *E. faecalis* and improved the resistance to degradation and mechanical strength of dentinal collagen.¹²⁰ After exposure to aPDT, the bacterial membrane damage was verified by atomic force microscopy.¹¹⁹ Furthermore, multispecies biofilms of *Streptococcus oralis*, *Prevotella intermedia*, and *Actinomyces naeslundii* on the dentin were subjected to 60 J/cm² and were completely disturbed, reducing the biofilm thickness.¹¹⁹ The mechanism of aPDT with CSRBnp was likely that NPs inactivated the endotoxins and lipopolysaccharides, and reduced the expression of inflammatory factors by activating the macrophages, without cytotoxic effect on the cells.¹²¹

In a recent study, ICG was placed into nano-GO (NGO) to produce NGO-ICG, which was more effective for aPDT against *E. faecalis* than using ICG alone (Figure 5D).¹²² NGO-ICG-PDT reduced the viability of *E. faecalis* by 2.8 log, and reduced the biofilm of *E. faecalis* by 99.4% at an energy density of 31.2 J/cm². NGO-ICG-PDT with a lower concentration of ICG (200

ug/mL) showed better antibacterial effect than ICG-PDT with a concentration of ICG (1000 μ g/mL) both in overall antimicrobial and anti-biofilm potential (47% and 21%, respectively). Furthermore, Afkhami et al compared the efficacy of AgNPs/100 ppm (AN), an 810 nm diode laser (DL), ICG-based PDT (ICG/DL), ICG-modified AgNPsbased PDT (AN/ICG/DL), and conventional irrigant NaOCl, against E. faecalis.¹²³ AN/ICG/DL had the greatest reduction of 99% in colony counts, followed by DL group with 97% reduction. AN and NaOCl groups had 94.42% and 94.61% reduction, respectively, whereas ICG/ DL group had only 68.47% reduction.¹²³ In addition, different ICG-loaded metal-organic frameworks (MOF-ICG) including Al-101-ICG, Fe-88-ICG, and Fe-101-ICG were designed to improve the properties of ICG and the antimicrobial activity for aPDT against endodontic infections (Figure 5E).¹²⁴ After 810 nm laser irradiation, the viability of E. faecalis could be suppressed by ICG-free MOFs by 18.1%, 28.8%, and 38.3%, via Al-101, Fe-88, and Fe-101, respectively. MOF-ICG-based aPDT reduced the count of *E. faecalis* by 60.72%, 45.12%, and 62.67%,

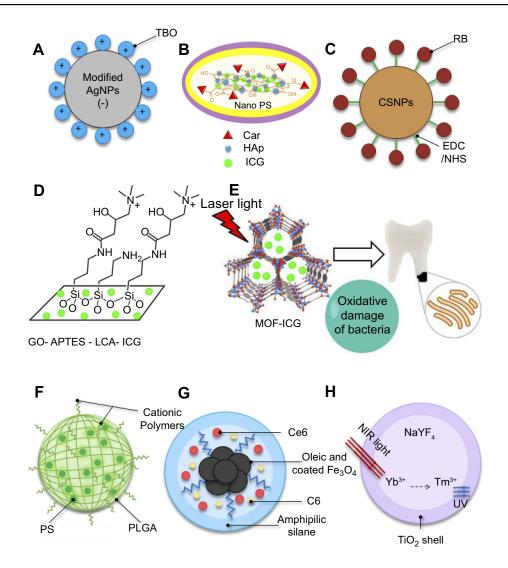


Figure 5 Proposed structures of nanomaterials against dental caries, endodontic infections and periodontal diseases.

Notes: (**A**) TBO-AgNP; (**B**) GO-Car/HAp@ICG; (**C**) CSRBnp with EDC and NHS; (**D**) NGO-ICG; (**E**) MOF-ICG and antibacterial mechanism; (**F**) MB-loaded PLGA cationic NPs; (**G**) antibacterial multifunctional NPs Fe_3O_4 -silane@Ce6/C6; and (**H**) UCNPs@TiO₂ and upconversion processes. Reprinted from Gholibegloo E, Karbasi A, Pourhajibagher M, et al. Carnosine-graphene oxide conjugates decorated with hydroxyapatite as promising nanocarrier for ICG loading with enhanced antibacterial effects in photodynamic therapy against *Streptococcus mutans. J Photoch Photobio B.* 2018;181:14–22,¹¹² Copyright (2018), with permission from Elsevier for Figure 5B. Reprinted from Akbari T, Pourhajibagher M, Hosseini F, et al. The effect of indocyanine green loaded on a novel nano-graphene oxide for high performance of photodynamic therapy against *Enterococcus faecalis. Photodiagn Photodyn.* 2017;20:148–153,¹²² Copyright (2017), with permission from Elsevier for Figure 5D. Reprinted with permission from Golmohamadpour A, Bahramian B, Khoobi M, Pourhajibagher M, Barikani HR, Bahador A. Antimicrobial photodynamic therapy assessment of three indocyanine green-loaded metal-organic frameworks against *Enterococcus faecalis. Photodiagn Photocyn.* 2018;23:31–338.¹²⁴ Copyright (2018), with permission from Elsevier for Figure 5E.

Abbreviations: AgNP, silver nanoparticle; PS, photosensitizer; Car, carnosine; CSRBnp, rose bengal functionalized chitosan nanoparticles; EDC, N-ethyl-N'-(3-dimethyl aminopropyl) carbodiimide; GO, Graphene oxide; HAp, hydroxyapatite; ICG, indocyanine green; MB, Methylene blue; MOF, metal-organic frameworks; NGO, nano-Graphene oxide; NHS, N-Hydroxysuccinimide; NPs, nanoparticles; PLGA, Poly lactic-co-glycolic acid; TBO, Toluidine blue O; UCNPs, upconversion nanoparticles.

respectively. Biofilms were dramatically decreased by 53.68%, 37.54%, and 47.01%, respectively. The expression of biofilm-associated *esp* gene was remarkably reduced by 6.0, 4.4, and 6.2 folds, respectively.¹²⁴

Periodontitis and peri-implantitis

MB-loaded PLGA cationic NPs showed great antimicrobial effects against biofilms of microorganisms from untreated patients with chronic periodontitis (Figure 5F).¹²⁵ After a 5-

min irradiation with red light (100 mW/cm²), these NPs reduced the bacterial activity of microorganisms in planktonic and biofilm phase by approximately 60% and 48%, respectively.¹²⁵ ICG-loaded nanospheres were coated with chitosan (ICG-Nano/c) to eradicate *Porphyromonas gingivalis*.¹²⁶ ICG-Nano/c with positive charge on the surface could adhere to the surface of Gram-negative bacteria, *Porphyromonas gingivalis* and the bacterial viability was reduced by 2 log₁₀ (96.71%) to 4 log₁₀ (99.99%).¹²⁶

Remarkably, the approach of intermittent irradiation with air cooling improved the therapeutic penetration, and the tissue thermal damage was avoided.¹²⁷

Multifunctional NPs Fe₃O₄-silane@Ce6/C6 were developed to trigger the aPDT against periodontitis-related pathogens (Figure 5G).¹²⁸ The NPs were stable and biocompatible without cytotoxicity. The NP-based aPDT could reduce three single-species periodontitis-related biofilm by about 4–5 orders of magnitude after 3 mins irradiation of the 630 nm light (100 mW·cm⁻²). In addition, the addition of C6 and Fe₃O₄ realized the real-time monitoring and magnetic targeting functions.¹²⁸

P. gingivalis was predominant in pockets of deeper than 4 mm.¹²⁹ Therefore, another consideration for aPDT to target periodontitis was for the light source to penetrate deep enough to reach the bottom of the periodontal pocket. In a recent study, we synthesized core-shell-structured NPs of NaYF₄:Yb³⁺, Tm³⁺@TiO₂ (UCNPs@TiO₂) triggered by NIR light in aPDT for treating periodontitis-associated pathogens (Figure 5H). The UCNPs@TiO₂-based aPDT reduced the three single-species biofilm CFU by approximately 4 orders of magnitude. After 5 mins of initial irradiation of 2.5 W·cm⁻², the planktonic bacteria was eliminated via 2 mM at the 4th hour. The UCNPs@TiO₂-mediated aPDT had great potential for effective periodontitis treatment.

Peri-implantitis was defined as an inflammatory process which affected the tissues around an osseointegrated implant and eventually contributed to the loss of supporting bone. Although multiple factors could result in implant failure, a large number of evidences showed that anaerobic plaque bacteria had the detrimental effect on peri-implant tissue health.¹³⁰ Like periodontitis, plaque was the initiator of peri-implantitis.

Although there were few reports about nanomaterialsbased aPDT for treating peri-implantitis, nanosurface layer showed promising in infection-prevention. Surface nanocoatings on implants with silver, titanium dioxide, and hydroxyapatite (HA) were created by silver plating, anodization, and sintering techniques. The dual-layered silvernHA nanocoating completely inhibited the bacterial growth in the surrounding media and reduced the biofilm on the surface of implants by 97.5%. This novel nanocoating to titanium alloy implants offered a lower infection risk, osseointegration, and accelerated bone healing.¹³¹

Oral fungal infections

Oral candidiasis was an oral mucosal disease caused by fungal candidiasis. *Candida* species were the most

common fungi isolated from the oral cavity of both healthy and sick people. Under certain conditions, the

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healthy and sick people. Under certain conditions, the Candida species in the symbiotic relationship could turn into a pathogenic state, which could contribute to its growth. The pathogenesis of candidiasis involved three factors: host factors, fungus, and oral microenvironment changes.¹³² Oral microenvironment changes included an improper denture, overuse of antibiotics, poor oral hygiene, smoking, and alcohol abuse.¹³² Oral infections caused by C. albicans were usually superficial and had different clinical manifestations. However, if not treated, it could spread to the throat, digestive tract and respiratory tract, and could be complicated by fungal sepsis, endocarditis, meningitis, and other serious complications.¹³³ Currently, broad-spectrum antibiotics were used to treat oral candidosis. However, Candida biofilms showed resistance to both antimicrobial mouthwashes and traditional antifungal agents.134

Various conjugates of PSs with AuNPs were developed to combat *C. albicans*.¹³⁵ The conjugates of aluminum phthalocyanine (complex 1) with gold nanorods (complex 1-AuNRs) and bipyramids (complex 1-AuBPs) showed a strong antifungal and bactericidal activity.¹³⁵ Under a power density of 92 mW·cm⁻² at 691 nm, complex 1 alone had a 1.78 CFU log reduction, whereas complex 1-AuBPs and complex 1-AuNRs had 2.08 and 2.53 log reductions, respectively.¹³⁵ AuNPs conjugated to MB or TBO were found to effectively combat both *C. albicans* planktonic cells and biofilms. In vivo, the mixture of AuNPs conjugated to two different PSs depleted the hyphal *C. albicans* burden against superficial skin and oral *C. albicans* infection in mice with a total light dose of 21.6 J/cm² at 662 nm as well as 635 nm, respectively.¹³⁶

Erythrosine (Ery) was used to kill oral microorganisms such as *S. aureus, S. mutans*, and *C. albicans*.¹³⁷ TiO₂ NPs, as a catalyst, enhanced the activity of PSs. Ery gel, when combined with TiO₂ stimulated by blue light (BL) with 15 J/cm², had greater fungicidal activity than 0.12% chlorhexidine. The treatments of 220 μ M Ery gel+1% TiO₂+BL and of 440 μ M Ery gel+1% TiO₂+BL reduced the *Candida* CFU by 1.27 and 1.85 logarithmic orders, respectively.¹³⁷

Maxillofacial infections

Traditionally, maxillofacial infections were treated with antibiotic therapy. However, the overuse of antibiotics could cause bacterial resistance and even multidrug-resistant (MDR) species. *S. aureus* and *P. aeruginosa* were the

two most common organisms in both acute and chronic wounds, and MRSA were the majority of all superficial skin infections.¹³⁸ Recently, a novel organic-inorganic hybrid coating (Tannic Acid/Fe³⁺/AgNPs) was developed mitigate implant-related bacterial infection to (Figure 6).¹³⁹ Under 660 nm visible light for 20 mins, the antibacterial ratio of TA/Fe3+/AgNPs-5 was 92% and 87% against E. coli and S. aureus, respectively. The corresponding values of TA/Fe3+/AgNPs-10 were nearly 100% and 94%, respectively.¹³⁹ Upconversion nanoconstructs, consisting of the cationic N-octyl chitosan (OC), NaYF₄:Yb,Er@NaYF₄ (UCNPs), and zinc phthalocyanine (ZnPc) (OC-UCNP-ZnPc), showed excellent antibacterial efficacy with NIR irradiation for the MDR bacteria including MRSA and β-lactamase-producing E. coli in vitro (Figure 7).¹⁴⁰ OC-UCNP-ZnPc-based aPDT had better antibacterial activity than the well-known positive drug vancomycin. OC-UCNP-ZnPc-based PDT against MRSA showed 0% viability at 250 μ g mL⁻¹ and had similar antibacterial activity against β -lactamase-producing *E. coli*. Significant reduction in the abscess volume of the subjected mice using 980 nm irradiation was observed in vivo.¹⁴⁰

Novel multifunctional NIR-sensitive NO delivery nanoplatform Roussin's black salt@NaGdF₄:Yb/ Tm@mSiO₂@quaternized ammonium chitosan (RBS @UCNP@mSiO₂@qC) under NIR irradiation was developed for eradication of MRSA biofilms (Figure 8).¹⁴¹ The eradication of drug-resistant bacterial biofilms was up to 78% by treating with RBS@UCNP@mSiO₂@qC via NIR irradiation at 200 μ g·mL⁻¹. In vivo, 13 medical catheters pre-colonized with the biofilms of *S. aureus* were implanted subcutaneously in rats to simulate implant-related peri-

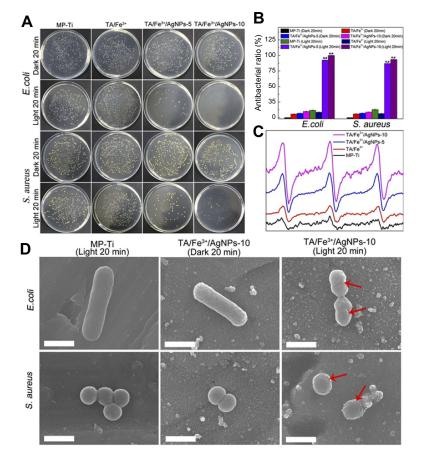


Figure 6 (**A**) Plate samples showing colonies of *E. coli* and *S. aureus* under 660 nm visible light for 20 mins or in the dark. (**B**) Antibacterial ratio determined according to the number of colonies in the plate samples **P < 0.01 vs Ti group as control. (**C**) ESR signals of ${}^{1}O_{2}$ obtained upon irradiation of different samples for 20 mins in the presence of TEMP. (**D**) Morphologies of *E. coli* and *S. aureus* seeded on MP-Ti and TA/Fe³⁺/AgNPs-10 under the illumination of 660 nm visible light for 20 min (scale bar = 1 µm). The red arrows in (**D**) indicated the portions of the membrane were shrunk, and several parts were completely broken. Reprinted with permission from Xu Z, Wang X, Liu X, et al. Tannic acid/Fe³⁺/Ag nanofilm exhibiting superior photodynamic and physical antibacterial activity. *ACS Appl Mater Inter.* 2017;9(45):39657–39671.¹³⁹ Copyright (2017) American Chemistry Society.

Abbreviations: E. coli, Escherichia coli; ESR, electron spin resonance; MP-Ti, mechanically polished Ti plates; S. aureus, Staphylococcus aureus; TEMP, 2,2,6,6-tetramethylpiperidine.

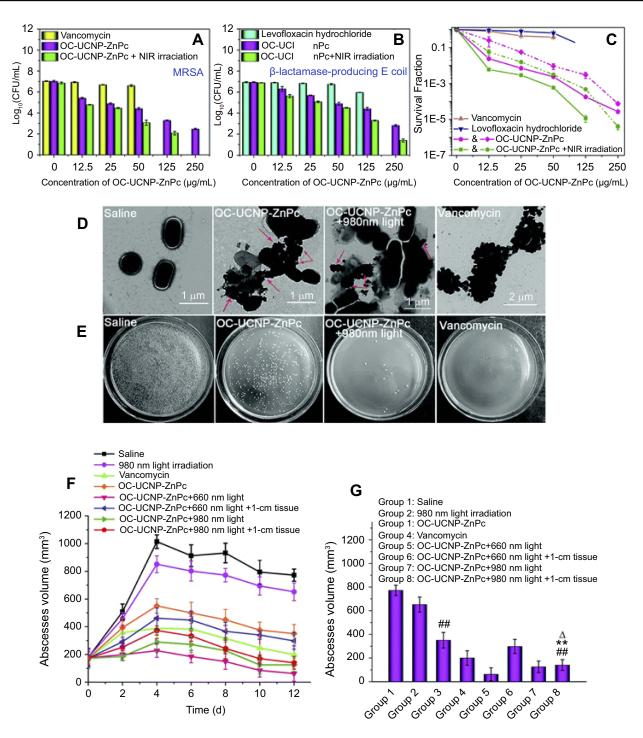


Figure 7 (**A**–**E**) Antibacterial effects of OC-UCNP-ZnPc on MDR bacteria in vitro. Viability of (**A**) MRSA and (**B**) β -lactamase-producing *E. coli* after incubation with OC-UCNP-ZnPc at different concentrations with/without NIR irradiation; (**C**) antibacterial effects induced by the nanoconstructs on MRSA (solid line) and β -lactamase-producing *E. coli* (dotted line); (**D**) TEM images, and (**E**) photographs of plate samples of MRSA in control, OC-UCNP-ZnPc (125 µg mL⁻¹) group without NIR irradiation, 980 nm light-triggered PDT group, and vancomycin (125 µg mL⁻¹) positive control. OC-UCNP-ZnPc nanoconstructs could be observed in TEM of OC-UCNP-ZnPc groups (the red arrows). (**F**–**G**) Anti-MRSA efficacy of OC-UCNP-ZnP cnanoconstructs in vivo. (**F**) Changes in the volume of MRSA-infected skin abscesses; (**G**) in vivo anti-MRSA efficacy of OC-UCNP-ZnPc nanoconstructs in vivo. (**F**) Changes in the volume of MRSA-infected skin abscesses; (**G**) in vivo anti-MRSA efficacy of different groups at day 12. Statistical significance was determined by a Student's t test (##P< 0.01 for the OC-UCNP-ZnPc treatment group and the 980 nm light triggered deep-tissue PDT group without \$80 nm light irradiation; $\Delta P < 0.05$ for the 980 nm light triggered deep-tissue PDT group compared with the control group; **P< 0.01 for the 980 nm light triggered with the 660 nm light induced deep-tissue PDT group without \$80 nm light irradiation; $\Delta P < 0.05$ for the 980 nm light triggered deep-tissue PDT group compared with the conversion photodynamic therapy system against drug-resistant bacteria in deep tissue. *Nanoscale*. 2017;9(11):3912–3924.¹⁴⁰ Copyright (2017) Royal Society of Chemistry. Permission conveyed through Copyright Clearance Center, Inc.

Abbreviations: E. coli, Escherichia coli; MDR, multi-drug resistant; MRSA, methicillin-resistant Staphylococcus aureus; NIR, near-infrared; OC, N-octyl chitosan; TEM, transmission electron microscope; UCNP, upconversion nanoparticle; ZnPc, zinc phthalocyanine.

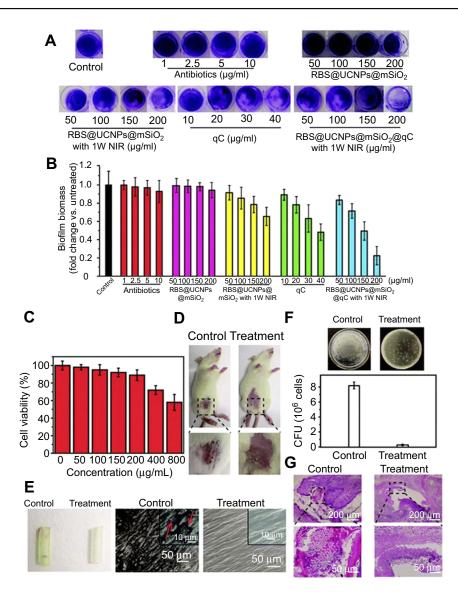


Figure 8 (**A**) Photographs of biofilms. (**B**) Quantifying the eradication of antibiotic-resistant biofilms by crystal violet staining treatment with various agents (n = 5). (**C**) Viabilities of HeLa cells vs increasing concentrations of RBS@UCNP@mSiO₂@qC NPs. (**D**) Synergistic dispersal of biofilms in vivo. (**E**) Implanted catheters and the surface of embedded catheters observed by SEM. The inset of (**E**) shows the magnified images. The red arrows indicate the biofilms on the embedded catheters. (**F**) CFU in vivo in the control and treatment group. (**G**) Histological analysis of tissues around implants. Each group had 5 rats (n = 5). Republished with permission of Royal Society of Chemistry, from Dong K, Ju E, Gao N, Wang Z, Ren J, Qu X. Synergistic eradication of antibiotic-resistant bacteria based biofilms in vivo using a NIR-sensitive nanoplatform. *Chem Commun.* 2016;52(30):5312–5315.¹⁴¹ Permission conveyed through Copyright Clearance Center, Inc.

Abbreviations: CFU, colony-forming units; qC, quaternized ammonium chitosan; RBS, Roussin's black salt; SEM, scanning electron microscope; UCNP, upconversion nanoparticle.

prosthetic infections. The wounds of rats in the nanocompounds-based aPDT group formed scabs and gradually healed after 7 days. In addition, there were few bacteria and no biofilms on the embedded catheters in the nanocompounds-based aPDT group, suggesting that the biofilm was indeed eradicated. Neither detectable signs of infection nor bacteria were observed in the implanted site in the histological slices of hematoxylin and eosin staining.¹⁴¹ Therefore, the UCNP-based NIR light system offered significantly improved penetration depth and was effective for potential therapeutic applications to prevent maxillofacial infections.

Conclusion

This article reviewed recent cutting-edge research on antibacterial PDT based on nanostructured materials in the field of dentistry. This included the nanostructured PS modification for aPDT enhancement and the therapeutic effects on oral infectious diseases, including caries, periodontitis, peri-implantitis, endodontic infections, and oral were located in deep sites, the challenge was for the drugs to reach the deep pockets and be maintained at an effective concentration for the infection. Different types of nanostructured materials were designed to be triggered by UV light, visible light, and NIR light for effective ROS generation. Based on these technologies, the antibacterial PDT exerted superior effects on microorganism-based oral diseases such as caries, periodontitis, peri-implantitis, oral fungal infections, and endodontic infections. Therefore, this novel therapeutic approach demonstrated the exciting potential for applications in dentistry, especially for infections located in deep pockets.

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Author contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors declare no conflicts of interest in this work.

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