Increasing resistance to quinolones: A four-year prospective study of urinary tract infection pathogens

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Abstract: A four-year prospective study was carried out to determine the incidence and rate of development of resistance by common urinary tract infection (UTI) pathogens to quinolone antimicrobial agents. Results show that there is high intrinsic resistance to the quinolones among strains of *Pseudomonas aeruginosa* (43.4%), *Escherichia coli* (26.3%), and *Proteus* spp. (17.1%). Over four years, rising rates of resistance were observed in *P. aeruginosa* (14.6% increase), *Staphylococcus aureus* (9.8%), and *E. coli* (9.7%). The highest potency was exhibited by ciprofloxacin (91.2%), levofloxacin (89.2%), and moxifloxacin (85.1%), while there were high rates of resistance to nalidixic acid (51.7%) and pefloxacin (29.0%). Coliforms, particularly *E. coli* (>45%), remain the most prevalent causative agents of UTI while females within the age range of 20–50 years were most vulnerable to UTI.

Keywords: UTI, microorganisms, antibiotics, resistance

Introduction

Urinary tract infections (UTI) could be defined as the persistent presence within the urinary tract of actively multiplying microorganisms. UTI implies both microbial colonization of the urine and invasion of the lower or upper urinary tract by microorganisms.¹ It is an infection with more than 100,000 organisms per milliliters in the mid-stream urine samples.²

Urinary tract infection is the most common disease of the urinary tract and it is a major cause of morbidity in both the hospital and the community.³ The most common causes of UTI are bacteria, and less often viruses, yeasts, or other intracellular microorganisms.⁴ According to Bohnson,⁵ bacteriuria may be completely asymptomatic or remain localized in the bladder without the development of renal infection. Urine secreted by normal kidneys is sterile and remains so while it travels to the bladder, however, normal urine is known to have microbial flora and any voided urine in normal patients may therefore contain thousands of bacteria per milliliter derived from this normal flora. In order to differentiate this smaller number of microorganisms from the larger number of microorganisms commonly found in UTIs, it is essential to count the number of bacteria present in freshly collected specimens by appropriate methods.⁶

Therapy of infectious diseases with antimicrobial agents is very dynamic as a result of increase in bacterial resistance to most available antimicrobial agents. Hence most bacterial infections, especially hospital-acquired infections, are becoming more and more difficult to treat. Bacterial resistance has been reported to almost every antimicrobial drug available; hence this study evaluated the susceptibility pattern of
the newer and presumed more efficient antibiotics which have been reported to be effective in the treatment of lots of ailments including UTI.6 A report on the susceptibility pattern of bacteria isolated from UTI cases to quinolones will further enhance the treatment of UTI, which has become difficult to treat due to multidrug resistance. Such multidrug resistance has become increasingly common throughout the world and has become expensive to treat.7 It therefore becomes imperative to check the potency of quinolones against some principal pathogens isolated in UTI cases. In vitro tests of the potency of antimicrobial agents provide a means of obtaining rapid and inexpensive information on the treatment of UTI.8,9

In Nigeria, UTI is prevalent among men and women, but is more common among women, especially during pregnancy.9 Unrecognized UTI in infancy and childhood may have serious long-term effects and chronic pyelonephritis may occur in adults. However, the infection occurs in all persons regardless of sex or age with particular impact on the young and the very elderly.10 Sexually active females are also more predisposed to UTI than their male counterparts.9,11 In later life, UTI is more common among men until the age of prostatic hypertrophy (above 40 years of age).6

For many years, pathogens associated with uncomplicated UTI have remained constant, with Escherichia coli identified as the etiological agent in about 75%–90% of infections.12 Five to 10% of uncomplicated cases are caused by Staphylococcus saprophyticus13 with Klebsiella, Proteus, Enterococcus, and Pseudomonas species seen in much smaller percentages.14-16

Etiological agents of infectious disease and their sensitivity to antimicrobial drugs used for their control have always been of interest in clinical medicine. Various studies have reported the causative bacterial agents of many cases of UTIs seen in Nigerian hospitals and their in vitro sensitivity to antimicrobial agents.17,18 One common concern amongst these reports has been the increasing resistance of pathogenic bacteria isolated from clinical specimens to antibiotics commonly used within the hospital communities.19

The World Health Organization (WHO) has called antibiotic resistance an emerging disease.20 Bacteria may be innately resistant or may acquire resistance to antibiotics. The rapid spread of bacterial resistance to antimicrobial agents has led to the search for newer and more potent drugs. However, as soon as new drugs appear, irrational use and abuse of them causes bacterial resistance. An extensive body of clinical research confirms that the fluoroquinolones are extremely effective for the treatment of UTIs ranging in severity from uncomplicated cystitis to urosepsis.21 The superiority of ciprofloxacin compared to trimethoprim/sulfamethoxazole has also been confirmed in patients with acute pyelonephritis.22

Of the newer fluoroquinolones, only levofloxacin is approved for both upper and lower UTIs. It should be emphasized that some other quinolones may demonstrate high activity against some Gram-negative organisms, and hence may be indicated for the treatment of these organisms implicated in UTI. Based on the emerging abuse of fluoroquinolone drugs and bacterial resistance in recent times, it becomes increasingly imperative to monitor drug resistance over a period of time.

**Materials and methods**

**Sampling and ethical consideration**

Mid-stream urine samples were collected from patients attending clinics for treatment of UTI. Clinical information on the patients included pain during urination, penile and vaginal discharge, urethral and vaginal itching, foul vaginal smell, and urethral irritation. Informed consent of all the patients were obtained by the heads of the various units in all the clinics sampled. All the patients during the four years of the study agreed to participate. Other necessary data regarding the patients were obtained from the clinical register. Samples were collected randomly from different hospitals and clinics in and around the Sapele Delta State of Nigeria during the years 2001, 2002, 2003, and 2004. The sample size was determined by dividing the average number of patients visiting each clinic for UTI per year by the total number of the clinics where samples were collected.

**Procedure**

The urine samples were immediately cultured onto nutrient agar, McConkey agar, blood agar, chocolate agar, Mannitol salt agar, and cystine lactose electrolyte-deficient (CLED) agar plates (Oxoid Ltd., Basingstoke, UK). All the samples were cultured in duplicate and incubated both aerobically and anaerobically at 37 °C (Weiss Gallenkamp, Loughborough, UK) for 24 hours. Bacterial count and identification were done according to the standard methods of Bergey’s manual,23 Harrigan and McCane,24 and Cowan and colleagues.25

Antibiotic sensitivity tests were performed using the filter paper disc diffusion method.26 The quinolone drugs used were moxifloxacin, levofloxacin, ciprofloxacin, ofloxacin (Sigma-Aldrich, St. Louis, MO, USA), pefloxacin, and nalidixic acid (Fidson Healthcare Plc, Lagos, Nigeria). They were all commercially obtained (Fidson Healthcare; Sigma-Aldrich).
The bacterial isolates were subcultured into peptone water before being used for antibiotic sensitivity tests as described by Cheesbrough. However, the plates with more than three different types of culture were regarded as contaminants and hence eliminated from the study. Only 15 of the samples had mixed growth of *E. coli* and Klebsiella spp., and 10 *S. faecalis* and *E. coli*. Zones of inhibition were read according to the method of Stroke and Ridway.

An Oxford strain of *S. aureus*, NCTC 6571, was used as control for Gram-positive bacteria while an *E. coli* strain, NCTC 10418, was used as control for Gram-negative bacteria. The results were statistically analyzed using regression analysis using SPSS software (version 11; SPSS Inc., Chicago, IL, USA). The total number of bacterial isolates in each year was used as the dependent variable while the number of isolates per year was used as independent variables.

## Results

A total of 1,100 patients within the age ranges of <20, 20–50 and >50 years comprising of 520 (32.5%) males and 739 (67.2%) females with clinical cases of UTI were sampled (Table 1). Age distribution shows that persons aged 20–50 years frequented general practice clinics for UTI treatment more often (822; 74.7%), which was followed by persons aged <20 (165; 15%). From the samples, a total of 1,906 pure bacterial cultures were obtained with Gram-negative bacteria being predominant (Table 2). *E. coli* was the highest isolate (871; 45.7%). Other isolates were Proteus spp., Klebsiella spp., and *P. aeruginosa*. The Gram-positive bacteria isolated were relatively fewer and were mainly *S. aureus* and *Streptococcus* spp. However, there were 87 samples with mixed culture. Only 36 of these gave the same result after a repeated sample collection and were regarded as positive cultures. Of the total bacterial isolates, 80% grew aerobically while only 20% grew anaerobically.

The mean percentage of the isolates in all years sampled are shown in Table 3. The mean percentage of *E. coli* isolates resistant to all the quinolones used increased from 21.6% in 2001 to 24.0% in 2002. In 2003, the mean resistance was 28.1%, which rose to 68.7% in 2004, showing a 9.7% rise in resistance of *E. coli* to quinolones from 2001 to 2004. Also, for *S. aureus* there was a steady rise in resistance from 2001 to 2004. All other organisms showed increased resistance except for Klebsiella spp., which showed a minimal decrease in resistance (0.9%) between 2001 and 2004. For *P. aeruginosa*, there was a marked increase in resistance to quinolones from 35.6% to 49.8%, showing a 14.2% rise in resistance. The summary of susceptibility and resistance profiles for all identified isolates (Table 4) shows that *P. aeruginosa* exhibited the highest resistance to all the quinolones with a mean of 43.4% resistant strains while *Streptococcus* spp. was most susceptible to the quinolones. Among the quinolone drugs, nalidixic acid was least efficient in preventing the growth of UTI pathogens with 51.7% of all isolates resistant, while ciprofloxacin was most potent, being able to prevent the growth of 91.2% of all the UTI pathogens tested. Statistical analysis showed that there was a relationship between isolates per year (*R* = 1.00; 95% confidence interval). There was also a significant difference in all the bacteria isolates per year (F = 369.404, 95% CI: *p* = 0.39).

## Discussion

In spite of the reported efficacy of the quinolones tested in this study, some isolates were found to be resistant to some of these quinolones. However, some of the quinolones, particularly ciprofloxacin and levofloxacin proved to be of high therapeutic value. The inclusion of nutrient agar and McConkey agar in this study was because these agars are routinely used for isolation in the average Nigerian hospital laboratory, and we tried to mimic as much as possible the routine Nigerian laboratory processes. This study was also able to show that there is increase in resistance to the quinolone drugs in Nigeria and this will continue if not urgently arrested. The results obtained from the four years sampled showed increase in the number of resistant isolates over the years sampled (Table 3). This represents a major public health threat.
health problem that requires drastic action for it to be properly controlled as earlier reported by Ozerek and Roa.\textsuperscript{30} This study also reveals that \textit{P. aeruginosa}, though isolated in fewer numbers, is gradually taking over as a major public health problem since this bacteria presents the highest resistance of all the bacteria isolated in this part of Nigeria. There have also been other studies which show the high drug resistance of \textit{P. aeruginosa} in Nigeria\textsuperscript{19} (Table 4) as well as showing the most rapid rise in resistance (Table 3). This does not undermine \textit{E. coli} as the most prevalent bacteria in UTI as earlier reported by Orenstein and Wong\textsuperscript{28} and Lutters and colleagues,\textsuperscript{29} who reported that the UTI may be endogenous or exogenous. \textit{S. aureus} was the most prevalent Gram-positive bacteria isolated with a mean percentage of 10.1% (Table 2). The advantage of this study is that it looked at positive bacteria isolated with a mean percentage of 10.1% or exogenous.

Our study shows that resistance to quinolones continues to increase. If urgent measures are not taken to arrest the situation, we may see the return of the era of the search for new drugs to fight bacterial infections.

Age group distribution showed that UTI was high among the group aged <20, which continued into the group aged 20–50 (Table 1), probably because of the high sexual activity within this age group. More females were affected with UTI than their male counterparts (Table 1). This has been attributed to the shortness of the female urethra and the closeness of the vagina to the perianal region of the urethral meatus in females, and also due to the antibacterial activity of the prostatic fluid in males and the relative longer length of the male urethra.\textsuperscript{9,11,30}

In summary, the susceptibility and resistance profiles of all isolates in this study have shown that ciprofloxacin and levofloxacin possess the highest efficacy among the quinolones while nalidixic acid is least therapeutic. Despite this efficacy, there was a general increase in the resistance pattern of isolates to all the quinolones used in this study. This was

### Table 3 Percentage (%) mean of quinolone resistant bacteria isolates obtained from the studied population from 2001 to 2004

<table>
<thead>
<tr>
<th></th>
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<th></th>
<th></th>
<th></th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>Escherichia coli SPF</td>
<td>21.6</td>
<td>24.0</td>
<td>28.1</td>
<td>31.3</td>
<td>9.7% Increase</td>
</tr>
<tr>
<td>2</td>
<td>Proteus spp SPF</td>
<td>12.4</td>
<td>16.7</td>
<td>18.5</td>
<td>21.2</td>
<td>8.8% Increase</td>
</tr>
<tr>
<td>3</td>
<td>Klebsiella spp SPF</td>
<td>11.3</td>
<td>12.0</td>
<td>11.2</td>
<td>10.4</td>
<td>0.9% Decrease</td>
</tr>
<tr>
<td>4</td>
<td>Pseudomonas aeruginosa</td>
<td>35.6</td>
<td>40.1</td>
<td>47.8</td>
<td>49.8</td>
<td>14.2% Increase</td>
</tr>
<tr>
<td>5</td>
<td>Staphylococcus aureus</td>
<td>5.5</td>
<td>8.2</td>
<td>12.7</td>
<td>15.3</td>
<td>9.8% Increase</td>
</tr>
<tr>
<td>6</td>
<td>Streptococcus faecalis</td>
<td>4.5</td>
<td>4.9</td>
<td>5.2</td>
<td>5.1</td>
<td>0.8% Increase</td>
</tr>
</tbody>
</table>

| Abbreviations: SPF, sensitive; R, resistant; S, sensitive. |

### Table 4 Summary of results of sensitivity/resistance profiles of all identified bacterial isolates to the quinolones

<table>
<thead>
<tr>
<th>Bacterial isolates</th>
<th>Sensitivity profiles</th>
<th>Moxi</th>
<th>Levo</th>
<th>Cipro</th>
<th>Oflo</th>
<th>Peflo</th>
<th>Oflo</th>
<th>Mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>Escherichia coli (871)</td>
<td>R</td>
<td>125 (14.4)</td>
<td>62 (7.1)</td>
<td>122 (14.0)</td>
<td>249 (28.6)</td>
<td>311 (35.7)</td>
<td>503 (57.7)</td>
<td>229 (26.3)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>746 (85.6)</td>
<td>809 (92.9)</td>
<td>749 (86.0)</td>
<td>622 (71.4)</td>
<td>560 (64.3)</td>
<td>368 (42.3)</td>
<td>642 (73.7)</td>
</tr>
<tr>
<td>Proteus spp (286)</td>
<td>R</td>
<td>12 (4.3)</td>
<td>11 (3.7)</td>
<td>37 (12.9)</td>
<td>36 (12.6)</td>
<td>71 (24.8)</td>
<td>126 (44.2)</td>
<td>49 (17.1)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>274 (95.7)</td>
<td>275 (96.3)</td>
<td>249 (87.1)</td>
<td>250 (87.4)</td>
<td>215 (75.2)</td>
<td>160 (55.8)</td>
<td>237 (82.9)</td>
</tr>
<tr>
<td>Klebsiella spp (235)</td>
<td>R</td>
<td>18 (7.6)</td>
<td>12 (4.9)</td>
<td>4 (1.6)</td>
<td>22 (9.2)</td>
<td>20 (8.5)</td>
<td>78 (33.3)</td>
<td>26 (11.1)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>217 (92.4)</td>
<td>223 (95.1)</td>
<td>231 (98.4)</td>
<td>213 (90.8)</td>
<td>215 (91.3)</td>
<td>157 (66.7)</td>
<td>209 (88.9)</td>
</tr>
<tr>
<td>Pseudomonas aeruginosa (219)</td>
<td>R</td>
<td>127 (58.2)</td>
<td>119 (54.9)</td>
<td>2 (0.9)</td>
<td>23 (10.5)</td>
<td>126 (57.5)</td>
<td>173 (79.1)</td>
<td>95 (43.4)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>92 (41.8)</td>
<td>100 (45.6)</td>
<td>217 (99.1)</td>
<td>196 (89.5)</td>
<td>93 (42.5)</td>
<td>46 (20.9)</td>
<td>124 (56.6)</td>
</tr>
<tr>
<td>Staphylococcus aureus (193)</td>
<td>R</td>
<td>1 (0.5)</td>
<td>&gt;1 (0.7)</td>
<td>&lt;1 (0.3)</td>
<td>11 (5.5)</td>
<td>17 (9)</td>
<td>88 (45.5)</td>
<td>20 (10.4)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>192 (99.5)</td>
<td>192 (99.3)</td>
<td>192 (99.7)</td>
<td>182 (94.5)</td>
<td>176 (91)</td>
<td>173 (89.6)</td>
<td></td>
</tr>
<tr>
<td>Streptococcus spp (102)</td>
<td>R</td>
<td>1 (1.2)</td>
<td>1 (1.0)</td>
<td>1 (1.1)</td>
<td>4 (3.6)</td>
<td>7 (7.1)</td>
<td>17 (16.4)</td>
<td>5 (4.9)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>101 (98.8)</td>
<td>101 (99.0)</td>
<td>101 (99.9)</td>
<td>98 (96.4)</td>
<td>95 (92.9)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total (1906)</td>
<td>R</td>
<td>284 (14.9)</td>
<td>206 (10.8)</td>
<td>167 (88)</td>
<td>345 (18.1)</td>
<td>552 (29.0)</td>
<td>985 (51.7)</td>
<td>423 (22.2)</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>1622 (85.1)</td>
<td>1700 (89.2)</td>
<td>1739 (91.2)</td>
<td>1561 (81.9)</td>
<td>1354 (71.0)</td>
<td>921 (48.3)</td>
<td>1483 (77.8)</td>
</tr>
</tbody>
</table>

Abbreviations: Cipro, ciprofloxacin; Levo, levofloxacin; Moxi, moxifloxacin; Oflo, ofloxacin; Peflo, pefloxacin; R, resistant; S, sensitive.
further confirmed by SPSS statistical analysis. However, in the course of treatment, cost, and side effects must also be considered especially concerning the low-income earners as high costs might discourage them from visiting clinics for proper examination and treatment. Drug abuse, which is a major source of multidrug resistance, must be discouraged and there should be continuous education of the general public on the need for rational use of drugs, particularly antimicrobial drugs. However, this is the first study of its type in Nigeria. The advantage of this study is that clinicians now know that some bacteria are becoming resistant to quinolones, which used to be the most valued drugs in this part of the world. This study also states the importance of carrying out sensitivity tests prior to treatment against the use of empirical treatment currently in practice in this locality.

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References